

S1 Ordinary differential equations (ODEs) of in AA metabolic pathway

Based on the rules in Materials and Methods, we have constructed ODEs for all species in the AA

metabolic pathway, as follows:

$$\begin{aligned}
 1) \frac{d[AA]}{dt} &= \frac{K_{cat,PLA2} \times [PLA_2][PL]}{k_{m,PLA2} \times \left(1 + \frac{[AA]}{ki}\right) + [PL]} - \frac{K_{cat,COX2} \times [COX2][AA]}{k_{m,COX2} \times \left(1 + \frac{[PGH_2]}{ki}\right) + [AA]} - \frac{K_{cat,12-LO} \times [12-LO][AA]}{k_{m,12-LO} \times \left(1 + \frac{[12-HpETE]}{ki}\right) + [AA]} \\
 &\quad - \frac{K_{cat,5-LO} \times [5-LO][AA]}{k_{m,5-LO} \times \left(1 + \frac{[AA]}{k_{AA \rightarrow 5-LO}} + \frac{[15-HETE]}{k_{15-HETE \rightarrow 5-LO}} + \frac{[5-HETE]}{k_{5-HETE \rightarrow 5-LO}} + \frac{[LTA_4]}{k_{LTA_4 \rightarrow 5-LO}} + \frac{[5-HpETE]}{ki} + \frac{[JH-2]}{k_{inh15LO}}\right) + [AA]} \\
 &\quad - \frac{K_{cat,12-LO} \times [12-LO][AA]}{k_{m,12-LO} \times \left(1 + \frac{[12-HpETE]}{ki}\right) + [AA]} - \frac{K_{cat,15-LO} \times [15-LO][AA]}{k_{m,15-LO} \times \left(1 + \frac{[15-HpETE]}{ki} + \frac{[JH-2]}{k_{inh15LO}}\right) + [AA]} \\
 &\quad - kd_{AA} \times [AA] \\
 2) \frac{d[15-HpETE]}{dt} &= \frac{K_{cat,15-LO} \times [15-LO][AA]}{k_{m,15-LO} \times \left(1 + \frac{[15-HpETE]}{ki} + \frac{[JH-2]}{k_{inh15LO}}\right) + [AA]} - \frac{K_{cat,PHGPx} \times [PHGPx][15-HpETE]}{k_{m,PHGPx} \times \left(1 + \frac{[15-HETE]}{ki}\right) + [15-HpETE]} \\
 3) \frac{d[15-HETE]}{dt} &= \frac{K_{cat,PHGPx} \times [PHGPx][15-HpETE]}{k_{m,PHGPx} \times \left(1 + \frac{[15-HETE]}{ki}\right) + [15-HpETE]} - kd_{15-HETE} \times [15-HETE] \\
 4) \frac{d[12-HpETE]}{dt} &= \frac{K_{cat,12-LO} \times [12-LO][AA]}{k_{m,12-LO} \times \left(1 + \frac{[12-HpETE]}{ki}\right) + [AA]} - \frac{K_{cat,PHGPx} \times [PHGPx][12-HpETE]}{k_{m,PHGPx} \times \left(1 + \frac{[12-HETE]}{ki}\right) + [12-HpETE]} \\
 5) \frac{d[12-HETE]}{dt} &= \frac{K_{cat,PHGPx} \times [PHGPx][12-HpETE]}{k_{m,PHGPx} \times \left(1 + \frac{[12-HETE]}{ki}\right) + [12-HpETE]} - kd_{12-HETE} \times [12-HETE] \\
 6) \frac{d[5-HpETE]}{dt} &=
 \end{aligned}$$

$$\begin{aligned}
& \frac{K_{cat,5-LO} \times [5-LO][AA]}{k_{m,5-LO} \times \left(1 + \frac{[AA]}{k_{AA \rightarrow 5-LO}} + \frac{[15-HETE]}{k_{15-HETE \rightarrow 5-LO}} + \frac{[5-HETE]}{k_{5-HETE \rightarrow 5-LO}} + \frac{[LTA_4]}{k_{LTA_4 \rightarrow 5-LO}} + \frac{[5-HpETE]}{ki} + \frac{[JH-2]}{k_{inh5LO}} \right) + [AA]} \\
& \frac{K_{cat,5-LO} \times [5-LO][5-HpETE]}{k_{m,5-LO} \times \left(1 + \frac{[AA]}{k_{AA \rightarrow 5-LO}} + \frac{[15-HETE]}{k_{15-HETE \rightarrow 5-LO}} + \frac{[5-HETE]}{k_{5-HETE \rightarrow 5-LO}} + \frac{[LTA_4]}{k_{LTA_4 \rightarrow 5-LO}} + \frac{[5-HpETE]}{ki} + \frac{[JH-2]}{k_{inh5LO}} \right) + [5-HpETE]} \\
& \frac{K_{cat,PHGPx} \times [PHGPx][5-HpETE]}{k_{m,PHGPx} \times \left(1 + \frac{[5-HETE]}{ki} \right) + [5-HpETE]} \\
7) \quad \frac{d[5-HETE]}{dt} &= \frac{K_{cat,PHGPx} \times [PHGPx][5-HpETE]}{k_{m,PHGPx} \times \left(1 + \frac{[5-HETE]}{ki} \right) + [5-HpETE]} - kd_{5-HETE} \times [5-HETE] \\
8) \quad \frac{d[LTA_4]}{dt} &= \frac{K_{cat,5-LO} \times [5-LO][5-HpETE]}{k_{m,5-LO} \times \left(1 + \frac{[AA]}{k_{AA \rightarrow 5-LO}} + \frac{[15-HETE]}{k_{15-HETE \rightarrow 5-LO}} + \frac{[5-HETE]}{k_{5-HETE \rightarrow 5-LO}} + \frac{[LTA_4]}{k_{LTA_4 \rightarrow 5-LO}} + \frac{[5-HpETE]}{ki} + \frac{[JH-2]}{k_{inh5LO}} \right) + [5-HpETE]} \\
& \frac{K_{cat,LTA4H} \times [LTA4H][LTA_4]}{k_{m,LTA4H} \times \left(1 + \frac{[LTA_4]}{k_{LTA4 \rightarrow LTA4H}} + \frac{[LTB_4]}{ki} \right) + [LTA_4]} - \frac{K_{cat,LTC4S} \times [LTC4S][LTA_4]}{k_{m,LTC4S} \times \left(1 + \frac{[LTA_4]}{k_{LTA4 \rightarrow LTC4S}} + \frac{[LTC_4]}{ki} \right) + [LTA_4]} \\
9) \quad \frac{d[LTB_4]}{dt} &= - \frac{K_{cat,LTA4H} \times [LTA4H][LTA_4]}{k_{m,LTA4H} \times \left(1 + \frac{[LTA_4]}{k_{LTA4 \rightarrow LTA4H}} + \frac{[LTB_4]}{ki} \right) + [LTA_4]} - kd_{LTB_4} \times LTB_4 \\
10) \quad \frac{d[LTC_4]}{dt} &= - \frac{K_{cat,LTC4S} \times [LTC4S][LTA_4]}{k_{m,LTC4S} \times \left(1 + \frac{[LTA_4]}{k_{LTA4 \rightarrow LTC4S}} + \frac{[LTC_4]}{ki} \right) + [LTA_4]} - kd_{LTC_4} \times LTC_4 \\
11) \quad \frac{d[PGH_2]}{dt} &= \frac{K_{cat,COX2} \times [COX2][AA]}{k_{m,COX2} \times \left(1 + \frac{[PGH_2]}{ki} \right) + [AA]} - \frac{K_{cat,PGDS} \times [PGDS][PGH_2]}{k_{m,PGDS} \times \left(1 + \frac{[PGD_2]}{ki} \right) + [PGH_2]} \\
& \frac{K_{cat,PGFS} \times [PGFS][PGH_2]}{k_{m,PGFS} \times \left(1 + \frac{[PGF_{2\alpha}]}{ki} \right) + [PGH_2]} - \frac{K_{cat,TXAS} \times [TXAS][PGH_2]}{k_{m,TXAS} \times \left(1 + \frac{[PGH_2]}{k_{PGH_2 \rightarrow TXAS}} + \frac{[TXA_2]}{ki} \right) + [PGH_2]} \\
& \frac{K_{cat,PGES} \times [PGES][PGH_2]}{k_{m,PGES} \times \left(1 + \frac{[AA]}{k_{AA \rightarrow PGES}} + \frac{[LTC_4]}{k_{LTC_4 \rightarrow PGES}} + \frac{[PGE_2]}{ki} \right) + [PGH_2]} - kd_{PGH_2} \times PGH_2
\end{aligned}$$

$$12) \frac{d[PGE_2]}{dt} = \frac{K_{cat,PGES} \times [PGES][PGH_2]}{k_{m,PGES} \times \left(1 + \frac{[AA]}{k_{AA \rightarrow PGES}} + \frac{[LTC_4]}{k_{LTC_4 \rightarrow PGES}} + \frac{[PGE_2]}{ki}\right) + [PGH_2]} - kd_{PGE_2} \times PGE_2$$

$$13) \frac{d[PGD_2]}{dt} = \frac{K_{cat,PGDS} \times [PGDS][PGH_2]}{k_{m,PGDS} \times \left(1 + \frac{[PGD_2]}{ki}\right) + [PGH_2]} - kd_{PGD_2} \times PGD_2$$

$$14) \frac{d[PGF_{2\alpha}]}{dt} = \frac{K_{cat,PGFS} \times [PGFS][PGH_2]}{k_{m,PGFS} \times \left(1 + \frac{[PGF_{2\alpha}]}{ki}\right) + [PGH_2]} - kd_{PGF_{2\alpha}} \times PGF_{2\alpha}$$

$$15) \frac{d[TXA_2]}{dt} = \frac{K_{cat,TXAS} \times [TXAS][PGH_2]}{k_{m,TXAS} \times \left(1 + \frac{[PGH_2]}{k_{PGH_2 \rightarrow TXAS}} + \frac{[TXA_2]}{ki}\right) + [PGH_2]} - kd_{TXA_2} \times TXA_2$$

$$16) \frac{d[TXB_2]}{dt} = kd_{TXA_2} \times TXA_2 - kd_{TXB_2} \times TXB_2$$

$$17) \frac{d[PLA_2]}{dt} = 0$$

$$18) \frac{d[15-LO]}{dt} = -k_{inh15LO} \times [JH - 2] \times [15-LO]$$

$$19) \frac{d[PHGPx]}{dt} = 0$$

$$20) \frac{d[12-LO]}{dt} = 0$$

$$21) \frac{d[5-LO]}{dt} = -k_{inh5LO} \times [JH - 2] \times [5-LO]$$

$$22) \frac{d[LTA4H]}{dt} = 0$$

$$23) \frac{d[LTC4S]}{dt} = 0$$

$$24) \frac{d[COX2]}{dt} = 0$$

$$25) \frac{d[PGES]}{dt} = 0$$

$$26) \frac{d[PGDS]}{dt} = 0$$

$$27) \frac{d[PGFS]}{dt} = 0$$

$$28) \frac{d[TXAS]}{dt} = 0$$

$$29) \frac{d[JH-2]}{dt} = -k_{on5LO} \times [JH-2] \times [5-LO] + k_{on5LO} \times k_{inht5LO} \times [comp5LO] - k_{on15LO} \times [JH-2] \times [15-LO] + k_{on15LO} \times k_{inht15LO} \times ([itotal] - [comp5LO] - [JH-2])$$

$$30) \frac{d[comp5LO]}{dt} = k_{on5LO} \times [JH-2] \times [5-LO] - k_{on5LO} \times k_{inht5LO} \times [comp5LO]$$

*JH-2 is the code name of HOEC, comp5LO is the complex of HOEC and 5-LO.

S2.1 Initial values of parameters

The initial values of the metabolites which have been measured were taken from the experimental values, and the others were given a default value directly. The initial value of the kinetic parameters of the enzyme was queried from the BRENDA [1] database, as shown in Table S1. Since no query results were available of 5-LOX and PHGPx, the initial values of them were based on the fitting values of the AA metabolic pathway model of human neutrophils established by Lai Luhua et al. In addition, we have also collected and collated the feedback pathways included in the AA pathway, as shown in Table S2.

Table S1 Enzyme kinetic parameters involved in AA metabolic pathway (experimental data)

Enzymes	K_{cat} (min^{-1})	K_m (μM)	K_{cat}/K_m	Data sources
PLA ₂ [2]	35	8.24	4.24	Rattus norvegicus
COX-2 [3]	1620	5.14	315.18	Mus musculus, (N580A COX2)
PGDS [4]	124.3	0.8	155.38	Mus musculus, (L-PGDS)
PGES [5]	3000	160	18.75	Homo sapiens
PGFS [6]	14.8	6.9	2.14	Mus musculus
TXAS [7]	14.79	10	1.48	Homo sapiens
5-LOX	/	/	/	/

LTA4H [8]	51	6	8.50	Homo sapiens
LTC4S [9]	4860	36	135.00	Mus musculus
12-LOX [10]	693.72	8.2	84.60	Oryctolagus cuniculus
15-LOX [11]	595.8	3.7	161.03	Homo sapiens
PHGPx	/	/	/	/

Table S2 Feedback parameters involved in AA metabolic pathway (experimental data)

Feedbacks	Value (μM)	Type
AA \rightarrow PGES [12]	0.3	IC ₅₀
15-HETE \rightarrow 5-LOX [13]	4	IC ₅₀
5-HETE \rightarrow 5-LOX [14]	6.3	K_i
5-HpETE \rightarrow 5-LOX [14]	0.5	K_i
PGH ₂ \rightarrow TXAS [15]	18	K_i
LTA ₄ \rightarrow LTA4H [16]	3.7 (2°C)	K_i (irreversible)
LTC ₄ \rightarrow PGES [17]	1.2	IC ₅₀
LTA ₄ \rightarrow 5-LOX [18]	2	K_i (irreversible)
LTA ₄ \rightarrow LTC4S [19]	2.3	K_i
AA \rightarrow 5-LOX [20]	13.7	K_i

S2.2 Fitted value of parameter

We used the parameter fitting tool of MATLAB and manual debug to obtain a set of parameters to reproduce the changes of metabolites with time in AA pathway after stimulated. The fitted enzyme kinetic parameters, non-enzymatic catalysis and the initial concentrations of enzymes and small molecules in the model are shown in the Table S3-S5

Table S3 Enzyme kinetic parameters involved in AA metabolic network (fitting data)

Enzymes	K_{cat} (1/min)	K_m (μM)
PLA ₂	35	8.24
COX-2	5620	5.14
PGDS	224.3	0.8

PGES	5760	122.252
PGFS	14.8	6.9
TXAS	27.79	9.991
5-LOX	2436.16	12.8
LTA4H	343	0.6
LTC4S	4860	3.6
12-LOX	693.7	8.2
15-LOX	595.8	4.7
PHGPx	500	70

Table S4 Non-enzyme kinetic parameters involved in AA metabolic network (fitting data)

Parameter	Reaction	Rate of decay (1/min)
kd1	AA→	0.74
kd2	LTB ₄ →	0.104
kd3	LTC ₄ →	0.258
kd4	PGE ₂ →	0.094
kd5	PGD ₂ →	0.08
kd6	PGF _{2α} →	0.11
kd7	TXA ₂ →	5.38
kd8	TXB ₂ →	0.049
kd9	PGH ₂ →	0.019
kd10	5-HETE→	0.0099
kd11	15-HETE→	0.00045
kd12	12-HETE→	0.0048

Table S5 Initial concentrations of enzymes and molecules involved in AA metabolic network

(fitting data)

Parameter	Initial concentration (nM)	Parameter	Initial concentration (nM)
PGH2 (0)	0.00075	TXB₂ (0)	0.857
AA (0)	334.102	PLA₂ (0)	350.3
15-HETE (0)	3.772	15-LOX (0)	1053.23
5-HETE (0)	0.3	5-LOX (0)	560.93

12-HETE (0)	0.13	12-LOX (0)	65.64
15-HpETE (0)	0.000128	LTA4H (0)	1230.8
5-HpETE (0)	0.00078	LTC4S (0)	58.52
12-HpETE (0)	0.00019	PHGPx (0)	539.78
LTA₄ (0)	0.000176	COX (0)	2700.63
LTB₄ (0)	0.1273	PGDS (0)	6708.96
LTC₄ (0)	0.336	PGES (0)	203.97
PGD₂ (0)	113.316	PGFS (0)	378.23
PGE₂ (0)	13.134	TXAS (0)	36.72
PGF_{2α} (0)	3.56	HOEC (0)	0
TXA₂ (0)	0.000241	COMP5LO (0)	0

Reference

1. Schomburg, I., Chang, A., Ebeling, C., Gremse, M., Heldt, C., Huhn, G., Schomburg, D. (2004) BRENDA, the enzyme database: updates and major new developments. *Nucleic acids research*, 32, D431-D433
2. Duncan, R. E., Sarkadinagy, E., Jaworski, K., Ahmadian, M., Sul, H. S. (2008) Identification and Functional Characterization of Adipose-specific Phospholipase A2 (AdPLA). *Journal of Biological Chemistry*, 283, 25428
3. Vecchio, A. J., Simmons, D. M., Malkowski, M. G. (2010) Structural Basis of Fatty Acid Substrate Binding to Cyclooxygenase-2. *Journal of Biological Chemistry*, 285, 22152-22163
4. Kumasaka, T., Aritake, K., Ago, H., Irikura, D., Tsurumura, T., Yamamoto, M., Miyano, M., Urade, Y., Hayaishi, O. (2009) Structural Basis of the Catalytic Mechanism Operating in Open-Closed Conformers of Lipocalin Type Prostaglandin D Synthase. *Journal of Biological Chemistry*, 284, 22344
5. Pettersson, P. L., Thorén, S., Jakobsson, P. J. (2005) Human microsomal prostaglandin E synthase 1: a member of the MAPEG protein superfamily. *Methods in Enzymology*, 401, 147
6. Moriuchi, H., Koda, N., Okudaashitaka, E., Daiyasu, H., Ogasawara, K., Toh, H., Ito, S., Woodward, D. F., Watanabe, K. (2008) Molecular characterization of a novel type of prostamide/prostaglandin F synthase, belonging to the thioredoxin-like superfamily. *Journal of Biological Chemistry*, 283, 792-801
7. Nüsing, R., Schneider-Voss, S., Ullrich, V. (1990) Immunoaffinity purification of human thromboxane synthase. *Archives of Biochemistry & Biophysics*, 280, 325
8. Mueller, M. J., Blomster, M., Jörnvall, H., Samuelsson, B., Haeggström, J. Z. (1996) Leukotriene A4 Hydrolase: Protection from Mechanism-Based Inactivation by Mutation of Tyrosine-378. *Proceedings of the National Academy of Sciences of the United States of America*, 93, 5931
9. Niegowski, D., Kleinschmidt, T., Ahmad, S., Qureshi, A. A., Mårback, M., Rinaldomatthis, A., Haeggström, J. Z. (2014) Structure and Inhibition of Mouse Leukotriene C4 Synthase. *Plos One*, 9, e96763
10. Almerinda, D. V., Thomas, H., Sabine, S., Giampiero, M., Laura, M., Angels, G. L., Hartmut, K., Igor, I. (2013) Role of Arg403 for thermostability and catalytic activity of rabbit 12/15-lipoxygenase. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, 1831, 1079-1088
11. Jacquot, C., Weckler, A. T., McGinley, C. M., Segreaves, E. N., Holman, T. R., Donk, W. A. V. D. (2008) Isotope sensitive branching and kinetic isotope effects in the reaction of deuterated arachidonic acids with human 12- and 15-lipoxygenases. *Biochemistry*, 47, 7295
12. Quraishi, O., Mancini, J. A., Riendeau, D. (2002) Inhibition of inducible prostaglandin E(2) synthase by 15-deoxy-Delta(12,14)-prostaglandin J(2) and polyunsaturated fatty acids. *Biochemical Pharmacology*, 63, 1183-1189
13. Petrich, K., Ludwig, P., Kühn, H., Schewe, T. (1996) The suppression of 5-lipoxygenation of arachidonic acid in human polymorphonuclear leucocytes by the 15-lipoxygenase product (15S)-hydroxy-(5Z,8Z,11Z,13E)-eicosatetraenoic acid: structure-activity relationship and mechanism of action. *Biochemical Journal*, 314 (Pt 3), 911-916
14. Aharony, D., Redkar-Brown, D. G., Hubbs, S. J., Stein, R. L. (1987) Kinetic studies on the inactivation of 5-lipoxygenase by 5(S)-hydroperoxyeicosatetraenoic acid. *Prostaglandins*, 33, 85
15. Jones, D. A., Fitzpatrick, F. A. (1990) "Suicide" inactivation of thromboxane A2 synthase. Characteristics of mechanism-based inactivation with isolated enzyme and intact platelets. *Journal of*

Biological Chemistry, 265, 20166-20171

16. Orning, L., Gierse, J., Duffin, K., Bild, G., Krivi, G., Fitzpatrick, F. A. (1992) Mechanism-based inactivation of leukotriene A4 hydrolase/aminopeptidase by leukotriene A4. Mass spectrometric and kinetic characterization. *Journal of Biological Chemistry*, 267, 22733
17. Friesen, R. W., Mancini, J. A. (2008) Microsomal prostaglandin E2 synthase-1 (mPGES-1): a novel anti-inflammatory therapeutic target. *Journal of Medicinal Chemistry*, 51, 4059-4067
18. Lepley, R. A., Fitzpatrick, F. A. (1994) Irreversible inactivation of 5-lipoxygenase by leukotriene A4. Characterization of product inactivation with purified enzyme and intact leukocytes. *Journal of Biological Chemistry*, 269, 2627-2631
19. Gupta, N., Gresser, M. J., Ford-Hutchinson, A. W. (1998) Kinetic mechanism of glutathione conjugation to leukotriene A4 by leukotriene C4 synthase. *Biochimica Et Biophysica Acta*, 1391, 157
20. Mcmillan, R. M., Masters, D. J., Vickers, V. C., Michael, D. P., Jacobs, V. N. (1989) Metabolism of unsaturated fatty acids by RBL-1 5-lipoxygenase: influence of substrate solubility and product inactivation. *Biochimica Et Biophysica Acta*, 1005, 170-176