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Figure S1

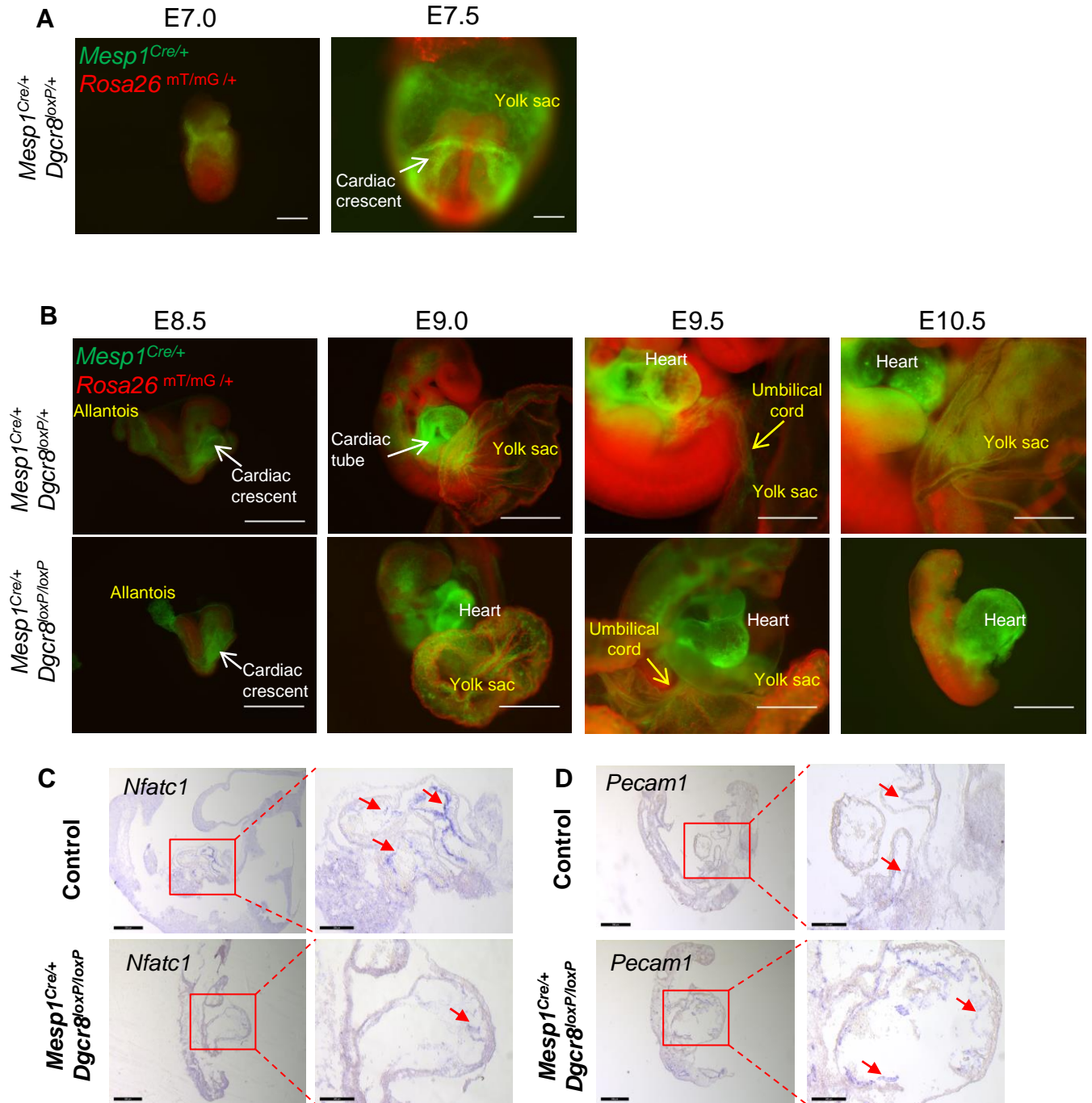


Figure S1. Phenotype of *Mesp1^{Cre/+}*; *Dgcr8^{loxP/loxP}*; *Rosa26^{mT/mG/+}* embryos at different developmental stages, related to Figure 1. A, Fluorescence images of *Mesp1^{Cre/+}*; *Rosa26^{mT/mG/+}* embryos at E7.0 and E7.5. *Mesp1* lineage cells in green and other cells in red. B, Fluorescence images of *Mesp1^{Cre/+}*; *Dgcr8^{loxP/+}*; *Rosa26^{mT/mG/+}* and *Mesp1^{Cre/+}*; *Dgcr8^{loxP/loxP}*; *Rosa26^{mT/mG/+}* embryos at E8.5, E9.0, E9.5, and E10.5. *Mesp1* lineage cells in green and other cells in red. Note that by E10.5, the *Dgcr8* cKO embryo already degenerated but the heart was still severely enlarged. Scale bar: 1 mm. C-D, *Nfatc1* (C) and *Pecam1* (D) expression in Control and cKO heart. Arrow highlights gene expression. Scale bar: 500 μ m. Magnified view scale bar: 200 μ m.

Figure S2

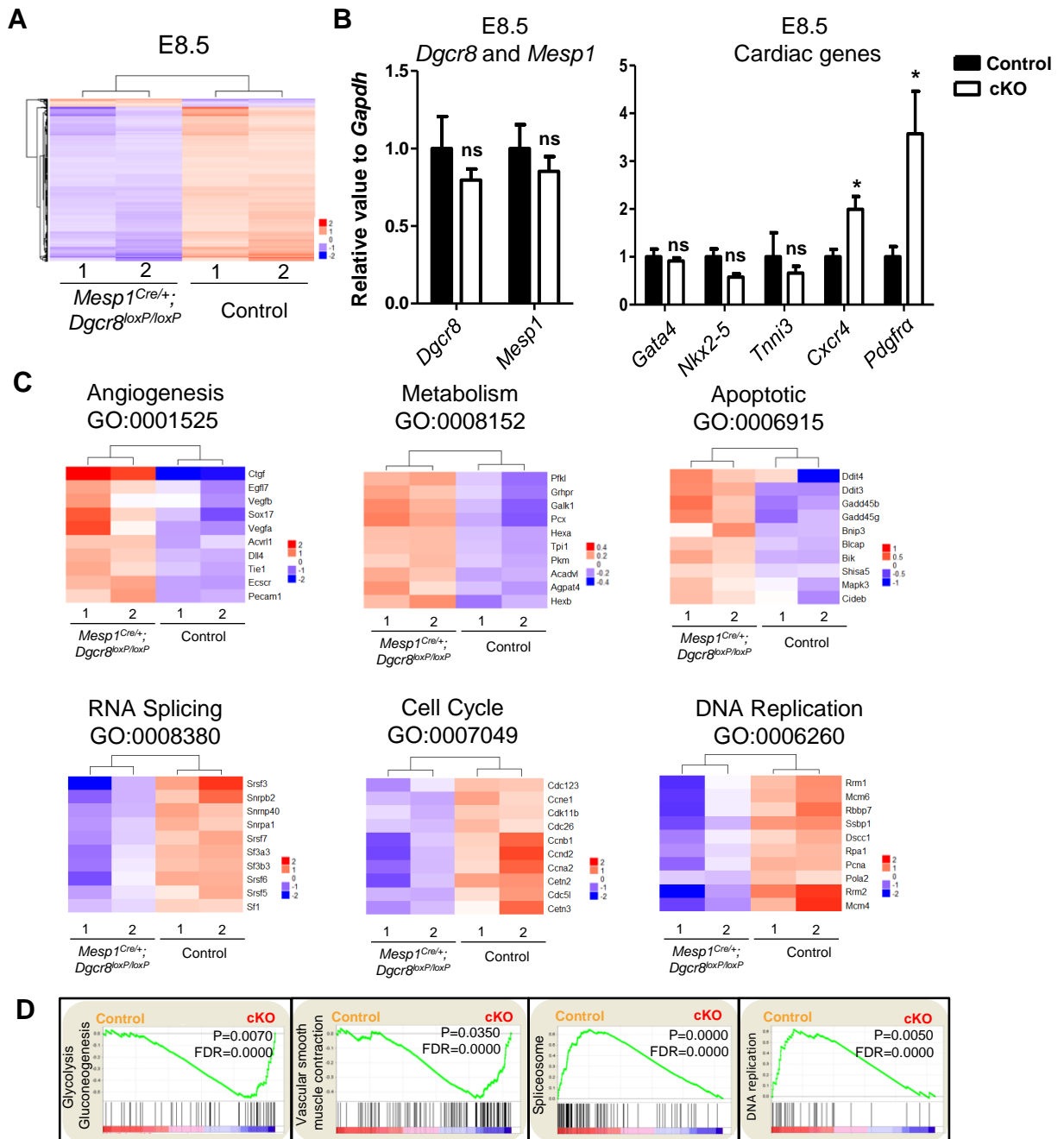
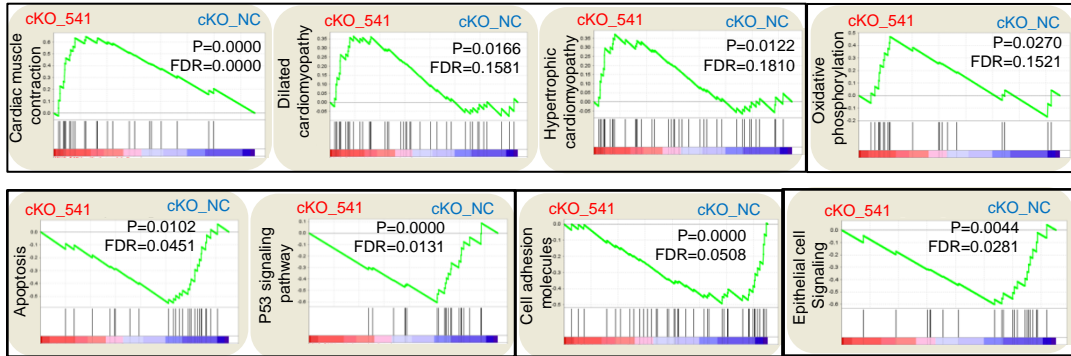


Figure S2. Gene expression difference between Control and *Dgcr8* cKO embryos, related to Figure 2.

A, Heatmap showing difference of gene expression in E8.5 Control and *Dgcr8* cKO hearts. Values represent normalized mean centered log₂ of FPKM for each sample. Significantly up and down-regulated transcripts are shown in red and blue colors respectively (n = 2). **B**, Q-PCR analysis of expression levels of *Dgcr8* and *Mesp1*, cardiac marker and progenitor cell marker genes in E8.5 Control and *Dgcr8* cKO heart tubes. Mean \pm s.e.m. are shown from three biological repeats. Student's t test was used: * P<0.05. **C**, Heatmaps of E9.5 cKO hearts enriched genes belonging to the following GO classes: angiogenesis, metabolism, apoptotic; and Control hearts enriched genes of the GO classes: RNA splicing, cell-cycle, DNA-replication. Values represent normalized mean centered log₂ of FPKM for each cKO sample. **D**, GSEA graphs of selective pathway genes enriched in E9.5 Control and *Mesp1^{Cre/+} Dgcr8^{loxP/loxP}* embryonic hearts. P < 0.05, FDR < 0.25.

Figure S4

A



B

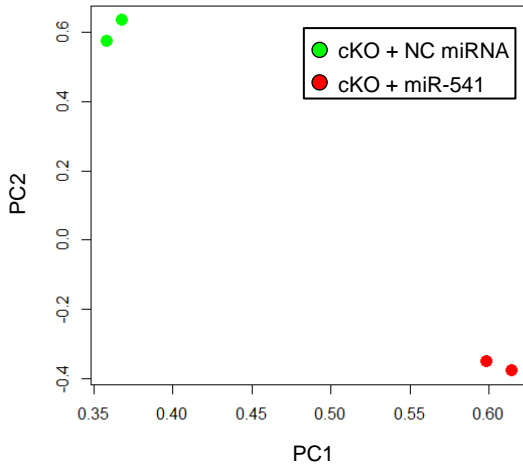


Figure S4. Introducing MiR-541 back into *in vitro* cultured cKO CMs lead to change in global gene expression, related to Figure 6. A, GSEA graph of selective pathway genes highly represented in cKO CMs transfected with miR-541(cKO_541) or NC-miRNA (cKO_NC). $P < 0.05$, $FDR < 0.25$. B, PCA of global gene expression in cKO cardiomyocytes transfected with miR-541 and NC-miRNA.

Figure S5

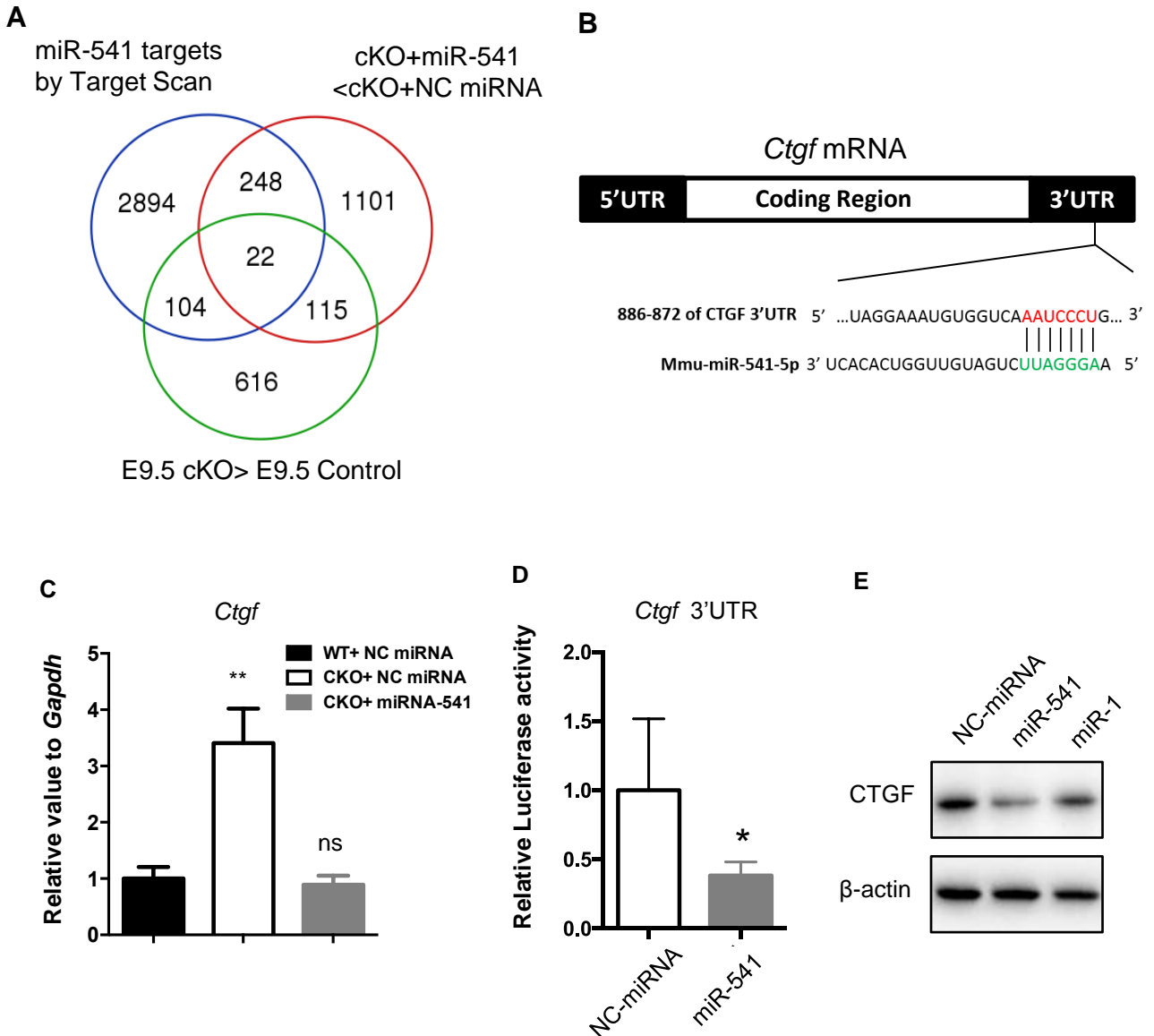


Figure S5. MiRNA-541 target gene analysis , related to Figure 6. A, Venn diagram depicting number of putative target genes of miR-541 predicted by TargetScan (blue) that overlapped with genes upregulated in E9.5 *Dgcr8* cKO heart (green), and downregulated in miR-541 transfected cKO CMs (red). **B,** A putative mmu-miR-541-5p targeting site on the 3' UTR of *Ctgf* predicted by TargetScan. **C,** Q-PCR analysis of *Ctgf* expression in Control and *Dgcr8* cKO CMs 48 h after NC-miRNA or miR-541 transfection. Mean \pm s.e.m. from three biological repeats are shown. ** $P < 0.01$. **D,** Bar graph showing luciferase activity of a reporter construct with the *Ctgf* 3' UTR containing the miR-541 putative target site. E9.5 *Dgcr8* cKO CMs were transfected with reporter plasmids and either miR-541 mimics or control mimics. Values were normalized with blank psiCheck2 vector for each mimic group, * $P < 0.05$. **E,** Western blot for CTGF expression in SVEC4-10 cells 48h after NC-miRNA, miR-541 and miR-1 transfection.

Figure S6

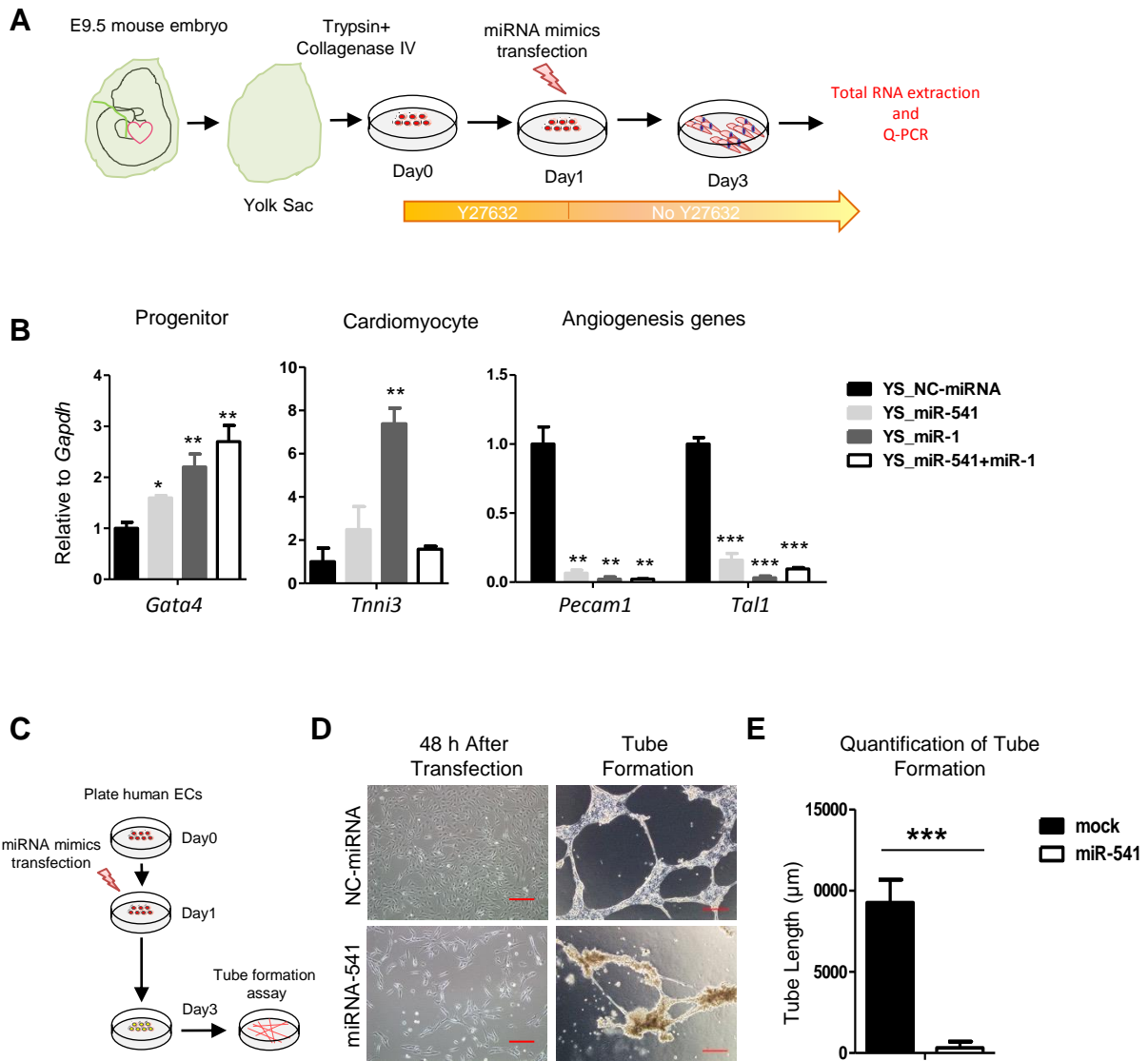


Figure S6. MiR-541 perturbed endothelial gene expression and function, related to Figure 7. **A**, Schematic view of miR-1 and miR-541 transfection into *in vitro* cultured Control yolk sac cells. **B**, Q-PCR analysis of gene expression in Control yolk sac cells after transfection with NC-miRNA, miR-1, or miR-541 and cultured for 48 h. Data represent mean \pm s.e.m. from three biological repeats. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.0001$ when compared to YS_NC-miRNA using Student's unpaired t-test. **C**, Schematic view of miR-541 transfection and endothelial function analysis in human iPSC derived ECs. **D**, Images of human EC tube formation assay. ECs were differentiated from human iPSCs. Note that miR-541 also strongly inhibited human EC tube formation. **E**, Quantification of tube length in (D). Mean \pm s.e.m. from three biological repeats are shown. *** $P < 0.0001$.