

$\gamma\delta$ T cells in liver diseases

Xuefu Wang (✉)¹, Zhigang Tian^{2,3}

¹School of Pharmacy, Anhui Medical University, Hefei 230032, China; ²Institute of Immunology and CAS Key Laboratory of Innate Immunity and Chronic Disease, School of Life Sciences and Medical Center, University of Science and Technology of China, Hefei 230027, China; ³Collaborative Innovation Center for Diagnosis and Treatment of Infectious Diseases, State Key Laboratory for Diagnosis and Treatment of Infectious Diseases, First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou 310003, China

© Higher Education Press and Springer-Verlag GmbH Germany, part of Springer Nature 2018

Abstract $\gamma\delta$ T cells display unique developmental, distributional, and functional patterns and can rapidly respond to various insults and contribute to diverse diseases. Different subtypes of $\gamma\delta$ T cells are produced in the thymus prior to their migration to peripheral tissues. $\gamma\delta$ T cells are enriched in the liver and exhibit liver-specific features. Accumulating evidence reveals that $\gamma\delta$ T cells play important roles in liver infection, non-alcoholic fatty liver disease, autoimmune hepatitis, liver fibrosis and cirrhosis, and liver cancer and regeneration. In this study, we review the properties of hepatic $\gamma\delta$ T cells and summarize the roles of $\gamma\delta$ T cells in liver diseases. We believe that determining the properties and functions of $\gamma\delta$ T cells in liver diseases enhances our understanding of the pathogenesis of liver diseases and is useful for the design of novel $\gamma\delta$ T cell-based therapeutic regimens for liver diseases.

Keywords $\gamma\delta$ T cells; liver infection; non-alcoholic fatty liver disease; autoimmune hepatitis; liver fibrosis and cirrhosis; liver cancer; liver regeneration

Introduction

$\gamma\delta$ T cells belong to a unique innate lymphocyte subset. Research on the differentiation, distribution, function, and application of $\gamma\delta$ T cells has achieved considerable progress since the discovery of the $\gamma\delta$ T-cell lineage approximately 30 years ago. T cells develop from multipotent progenitor CD4⁺CD8⁻double-negative (DN) thymocytes in the thymus and undergo four stages (DN1, CD44⁺CD25⁻; DN2, CD44⁺CD25⁺; DN3, CD44⁻CD25⁺; and DN4, CD44⁻CD25⁻) [1]. T-cell receptor (TCR) rearrangement begins at the DN2–DN3 stages, and $\gamma\delta$ TCRs or preTCRs are expressed at the DN3–DN4 stages [2]. Cell differentiation into $\alpha\beta$ or $\gamma\delta$ T-cell lineages is determined at the DN3 stage [3]. $\gamma\delta$ T cells can produce at least two distinct subsets, namely, interferon (IFN)- γ - and interleukin (IL)-17-producing $\gamma\delta$ T cells. The segregation of $\gamma\delta$ T-cell functional subsets can be distinguished by cell surface markers. CD27 segregates $\gamma\delta$ T cells into IL-17-producing CD27⁻ $\gamma\delta$ T cells, and IFN- γ -producing CD27⁺ $\gamma\delta$ T cells [4]. CCR6⁺ $\gamma\delta$ T cells exclusively produce IL-

17A, whereas NK1.1⁺ $\gamma\delta$ T cells readily produce IFN- γ [5]. Increasing evidence suggests that the functional polarization of $\gamma\delta$ T cells is developmentally programmed in the thymus rather than in localized peripheral tissues [6]. Therefore, TCR signals play an essential role, beyond lineage commitment, in the functional polarization of $\gamma\delta$ T cells. In contrast to conventional adaptive T cells, which recognize peptide antigens presented by antigen-presenting cells in a MHC-dependent manner, $\gamma\delta$ T cells can recognize nonpeptide antigens and stress-induced ligands [7]. Moreover, $\gamma\delta$ T cells are preferentially located in peripheral mucosal tissues [8] and play a protective role in pathogen clearance, tumor surveillance, and tissue repair and a deleterious role in autoimmunity, allergy, and carcinogenesis through cytokine secretion and/or cytotoxicity [9]. Thus, $\gamma\delta$ T cells contribute not only to immune balance and tissue homeostasis but also to immune disorders and tissue pathogenesis. In fact, the effector functions of $\gamma\delta$ T cells are determined by developmental polarization, tissue localization, and environment cues. Growing evidence reveals that $\gamma\delta$ T cells in the liver respond to liver-targeted insults and modulate the development of liver diseases [10,11]. As such, this review focuses on the roles of $\gamma\delta$ T cells in liver diseases (Fig. 1).

Although the liver is recognized as an important organ in

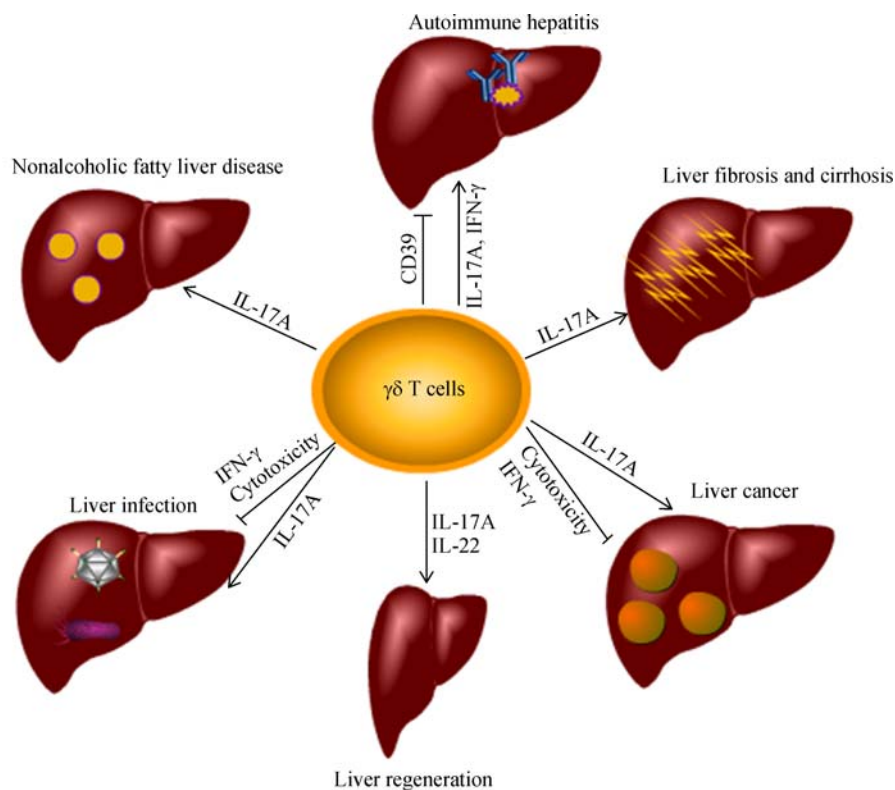


Fig. 1 $\gamma\delta$ T cells in liver diseases. The effector functions of hepatic $\gamma\delta$ T cells are context-dependent, although IL-17A-producing $\gamma\delta$ T cells predominantly localize in the liver. On the one hand, hepatic IL-17A-producing $\gamma\delta$ T cells not only play pathogen-promoting roles in infection-induced liver injury, non-alcoholic fatty liver disease, autoimmune hepatitis, liver fibrosis/cirrhosis, and liver cancer but also promote liver regeneration (shown as \rightarrow). On the other hand, hepatic IFN- γ -producing $\gamma\delta$ T cells play protective roles in liver infection and liver cancer (shown as \leftarrow) and also mediate autoimmune hepatitis (shown as \rightarrow). In addition, hepatic $\gamma\delta$ T cells suppress the inflammatory response by expressing inhibitory molecules (such as CD39, shown as \leftarrow).

the defense against blood-borne infections, liver injury can also be triggered by drugs, toxins, pathogen infections, over-eating, and genetic disorders [12]. Moreover, persistent liver damage is likely to induce the progression of mild chronic liver disease to liver fibrosis, liver cirrhosis, liver cancer, and liver failure [13]. The liver displays tissue-specific features that need to be emphasized in this study because prior knowledge of these features enhances the understanding of the induction, development, and end stage of liver disease. First, the liver is a barrier organ that segregates the digestive tract from the rest of the body. It possesses a special blood circulation system, wherein 80% of blood supply from the portal vein is rich in bacterial products, environment toxins, and food-derived antigens that are purified by the liver from the intestines; meanwhile, the remaining blood from the hepatic artery provides nutrients and oxygen [14,15]. Second, the liver is an immune-tolerant organ. Liver tolerance is manifested by immune hyporesponsiveness in the context of allogeneic liver transplants and liver infections and is maintained and modified by diverse intrahepatic cells, including immune cells and non-immune cells [16,17]. Third, the liver is an organ with predominant innate immunity. It is enriched with Kupffer cells, natural killer

cells, natural killer T (NKT) cells, and $\gamma\delta$ T cells [18–22]. These liver features reflect the close relationship among its anatomical location, immune status, immune components, functions, and liver disease. In this study, we highlight advances in the understanding of the properties and roles of hepatic $\gamma\delta$ T cells in liver diseases and discuss the potential of $\gamma\delta$ T cells as therapeutic targets in treating liver diseases.

Hepatic $\gamma\delta$ T cells

$\gamma\delta$ T cells constitute approximately 2%–10% of the total T cells in the peripheral blood, whereas hepatic $\gamma\delta$ T cells account for 3%–5% of the total liver lymphocytes and 15%–25% of the total number of liver T cells [18]. In terms of phenotype, hepatic $\gamma\delta$ T cells exhibit mixed V γ chains (including V γ 1, V γ 4, and V γ 6 in mice and V δ 1 and V δ 3 in humans) [23]. Hepatic $\gamma\delta$ T cells in the murine liver contain a high fraction of V γ 4⁺ cells with enhanced functional activation; meanwhile, hepatic $\gamma\delta$ T cells in the human liver are more mature than their counterparts in peripheral blood [24]. Recently, our group carefully analyzed hepatic $\gamma\delta$ T cells [25]. Hepatic $\gamma\delta$ T cells are highly localized to the

liver and rarely transported. Hepatic $\gamma\delta$ T cells also display active and mature phenotype characterized by high frequency of CD44^{high}CD62L^{low} $\gamma\delta$ T cells in the liver. In terms of function, IL-17A-producing $\gamma\delta$ T cells increase in number and become predominant in the liver with age, accompanied by a decrease in IFN- γ -producing $\gamma\delta$ T cells. Therefore, hepatic $\gamma\delta$ T cells exhibit a unique phenotype and composition. Commensal microbes in the gut maintain the homeostasis of hepatic IL-17A-producing $\gamma\delta$ T cells in a lipid antigen/CD1d-dependent manner; this phenomenon may explain the steady increase in hepatic IL-17A-producing $\gamma\delta$ T cells with age [25]. Accumulating evidence suggests that hepatic $\gamma\delta$ T cells play important roles in maintaining hepatic physiological homeostasis and modulating hepatic pathological progression. Thus, we review the roles of hepatic $\gamma\delta$ T cells in diverse liver diseases.

$\gamma\delta$ T cells in liver infection

Viral hepatitis is an inflammatory liver disease induced mainly by hepatitis B virus (HBV) and hepatitis C virus (HCV) infections. In HBV infection, approximately 5% of infected individuals develop chronic hepatitis B (CHB), whereas the remaining 95% of adult patients achieve resolution of hepatitis B infection [26,27]. Clinical studies show that the frequencies of peripheral and hepatic V δ 2 T cells decrease with disease progression, from tolerance to activation [28]. In addition, the ability of V δ 2 T cells with an activated memory phenotype to produce IFN- γ and induce cytotoxicity is impaired in CHB patients, although it can be enhanced by IFN- α treatment [29]. Therefore, $\gamma\delta$ T cells in CHB patients protect the host against HBV infection. Nonetheless, $\gamma\delta$ T cells mediate liver injury and contribute to HBV-associated acute-on-chronic liver failure [30]. Hepatic IL-17A-producing $\gamma\delta$ T cells also induce CD8⁺ T cell exhaustion by recruiting myeloid-derived suppressor cells (MDSC) in the HBV-induced immunotolerance mouse model [31]. Similar conclusions concerning patients with chronic HBV infection still require further clinical studies. As suggested by the aforementioned observations, distinct $\gamma\delta$ T subsets produce unique effects as disease progress after HBV infection.

In contrast to HBV infection, HCV infection induces chronic hepatitis at a high rate [32]. V δ 2 T cells exhibit an activated phenotype and enhanced cytotoxicity in HCV-infected patients. Moreover, hepatic V δ 2 T cells display more cytotoxic functions than peripheral V δ 2 T cells [33]. However, the elevated hepatic $\gamma\delta$ T cells exhibit strong cytotoxicity against hepatocytes and mediate liver injury in HCV-infected patients [34]. V δ 2 T cells also exhibit a markedly impaired capacity to produce IFN- γ , the key molecule in combating HCV infection, although the activation of V δ 2 T cells by nonpeptide antigens can inhibit HCV replication through IFN- γ release [35]. In

HBV- and HCV-infected individuals, $\gamma\delta$ T cells display different functions. This condition may result in different outcomes.

In addition to viral infection, $\gamma\delta$ T cells also participate in other hepatotropic pathogen infections. For instance, hepatic $\gamma\delta$ T cells remarkably expand and protect the liver against *Trypanosoma cruzi* infection by producing IFN- γ [36]. Hepatic $\gamma\delta$ T cells limit inflammation induced by *Listeria monocytogenes* infection and necrotic liver lesions by killing chemokine-producing macrophages in a Fas ligand (FasL)-dependent manner [37]. Hepatic $\gamma\delta$ T cells contribute to granulomatous inflammatory and hepatic lesions by producing IL-17A in mice infected with *Schistosoma japonicum* [38]. Thus, improved therapeutic strategies based on $\gamma\delta$ T cells for liver infections should be explored to enhance the antipathogenic potential while attenuating the risk of tissue damage.

$\gamma\delta$ T cells in non-alcoholic fatty liver disease

Non-alcoholic fatty liver disease (NAFLD) is a spectrum of chronic liver disorders ranging from simple steatosis to non-alcoholic steatohepatitis and cirrhosis [39]. IL-17 accelerates NAFLD progression by recruiting neutrophils and inducing reactive oxygen species [40]. Thus, IL-17 neutralization efficiently attenuates high-fat diet-induced NAFLD [41]. Hepatic $\gamma\delta$ T cells are major providers of IL-17A in HFD/HFHCD-induced NAFLD [25]. IL-17A-producing $\gamma\delta$ T cells are elevated in mouse livers with NAFLD. $\gamma\delta$ T cell deficiency protected mice from NAFLD, characterized by reduced steatohepatitis and liver damage. The adoptive transfer of hepatic $\gamma\delta$ T cells into HFHCD-fed mice accelerates NAFLD. Similar to the findings in mice, IL-17A-producing CD161⁺ $\gamma\delta$ T cells are enriched in the livers of patients with NAFLD [10]. Therefore, IL-17A-producing $\gamma\delta$ T cells are a major regulating factor in NAFLD progression. However, the mechanism of $\gamma\delta$ T cell subset activation requires further exploration. Therefore, blocking or deleting IL-17A-producing $\gamma\delta$ T cells may be a feasible therapeutic regimen for NAFLD treatment.

$\gamma\delta$ T cells in autoimmune liver diseases

Autoimmune hepatitis (AIH), primary biliary cirrhosis (PBC), and primary sclerosing cholangitis (PSC) are the three major autoimmune liver diseases (ALDs). AIH is an inflammation of the liver and characterized by periportal hepatitis, hypergammaglobulinemia, and the presence of serum autoantibodies [42]. PBC is a chronic cholestatic liver disease characterized by the destruction of small intrahepatic bile ducts and the presence of antimitochondrial antibodies [43]. PSC is the hepatobiliary

manifestation of inflammatory bowel disease, characterized by chronic inflammation and bile duct fibrosis [44]. The percentages and absolute numbers of $\gamma\delta$ T cells are elevated in the peripheral blood and in the portal areas of patients with AIH, PSC, or PBC [45,46]. Meanwhile, $\gamma\delta$ T cells in the peripheral blood of patients with ALD are activated [47]. $\gamma\delta$ T cells in patients with AIH produce more IFN- γ and granzyme B, contributing to liver damage [48]. In addition, $\gamma\delta$ T cell-derived IL-17A mediates hepatocyte damage in *Ja18* knockout (KO) mice with AIH [49]. The roles of IL-17A in autoimmune liver diseases have been reviewed [50], suggesting therapeutic potential for ALD treatment by targeting the IL-17 signaling pathway. Nonetheless, $\gamma\delta$ T cells can produce suppressor cytokines to prevent hepatitis induced by autoreactive T cells [51]. Moreover, in ConA-induced NKT cell-mediated ALD, IL-17A-producing $V\gamma4^+$ $\gamma\delta$ T cells protect mice from liver damage by inhibiting the pathogenic effect of NKT cells [52]. Therefore, distinct subsets of $\gamma\delta$ T cells may exist simultaneously during ALD progression. Furthermore, the roles of specific $\gamma\delta$ T cell subsets in ALD are context dependent. Thus, the modulation of $\gamma\delta$ T cells may be a useful strategy in treating ALD.

$\gamma\delta$ T cells in liver fibrosis and cirrhosis

Chronic inflammation promotes liver fibrosis and subsequently leads to cirrhosis, which is characterized by the excessive accumulation of extracellular matrix production by hepatic stellate cells (HSCs) [13]. CCL20 is strongly upregulated upon chronic liver injury in mice and in human patients with cirrhosis, which results in the recruitment of IL-17-producing $\gamma\delta$ T cells to the liver in a CCR6-dependent manner to restore hepatic fibrosis by promoting HSC apoptosis in a FasL-dependent manner [53]. However, HSC-derived CCL20 induced by exosomes recruiting IL-17A-producing CCR6⁺ $\gamma\delta$ T cells to exacerbate liver fibrosis was reported [54]. IL-17A stimulates α -smooth muscle actin, tumor growth factor (TGF)- β , IL-6, and collagen expression in HSCs in liver fibrosis [55,56]. In fact, the contradictory roles of $\gamma\delta$ T cells in liver fibrosis are a result of different mechanisms. The cytotoxicity against HSCs induces $\gamma\delta$ T cells to play a protective role, whereas IL-17A confers $\gamma\delta$ T cells to exhibit a deleterious role.

$\gamma\delta$ T cells in liver cancer

Hepatocellular carcinoma (HCC) is the third leading cause of cancer-related deaths and shows a poor five-year survival rate [57]. $\gamma\delta$ T cells can efficiently kill liver tumor cell lines *in vitro* [4]. Nonetheless, infiltration by $\gamma\delta$ T cells and associated IFN- γ secretion and cytotoxicity in

HCC tumor tissues are significantly compromised compared with paired peritumoral tissues, which is partially mediated by Treg cells in a TGF- β - and IL-10-dependent manner [58]. However, the levels of peritumoral $\gamma\delta$ T cells are negatively correlated with tumor size and incidence of recurrence in HCC [59]. This phenomenon indicates that peritumoral $\gamma\delta$ T cells can still exert an antitumor effect in patients with HCC. Therefore, enhancing the infiltration and function of $\gamma\delta$ T cells in HCC tumor tissues is a promising immunotherapy for HCC. However, IL-17A-producing $V\gamma4$ $\gamma\delta$ T cells in HCC recruit MDSCs, resulting in the suppression of the CD8⁺T-cell response and the promotion of tumor growth [60]. Moreover, HCC formation following the introduction of Hepa1-6 cells is suppressed in $\gamma\delta$ TCR KO mice [61]. The antitumor ability of $\gamma\delta$ T cells cannot be surpassed. Human $V\gamma9V\delta2$ T cells display a strong cytotoxic activity against HCC [62,63], and CD226 promotes $V\gamma9V\delta2$ T-cell-mediated death of human HCC cells [64]. Zoledronate increases HCC cell sensitivity to $V\gamma9V\delta2$ T lymphocyte-mediated killing [65]. Although the roles of different subsets of $\gamma\delta$ T cells are extensively explored in HCC, the potential immunotherapies based on $\gamma\delta$ T cells require further investigation.

$\gamma\delta$ T cells in liver regeneration

Regeneration is a unique feature of the liver compared with other organs and is important for rescuing liver function after hepatic resection or in acute and chronic liver disease. Multiple cytokines, including IL-6, IL-22, and TNF- α , act to promote liver regeneration [66]. Recently, liver regeneration is significantly impaired in IL-17A KO mice compared with the wild-type mice following partial hepatectomy [24,67]. Hepatic $\gamma\delta$ T cells increase within the first three hours but return to baseline six hours after partial hepatectomy. Hepatocyte proliferation is markedly impaired in TCR $\delta^{-/-}$ mice and is accompanied by low levels of IL-17A and IL-22. Mechanistically, hepatic $\gamma\delta$ T cells induce a proregenerative phenotype in inflammatory hepatic cells through IL-17A and accelerate liver regeneration through IL-22. Moreover, dectin-1 is upregulated in hepatic $\gamma\delta$ T cells in a regenerating liver. Dectin-1 ligation induces IL-22 and IL-17A production in hepatic $\gamma\delta$ T cells. Therefore, the dectin-1- $\gamma\delta$ T cells-IL-17A/IL-22 axis promotes liver regeneration. These findings may be helpful in understanding the roles of hepatic $\gamma\delta$ T cell in the maintenance of physiological homeostasis in the liver. Interestingly, evidence suggests that the microbiota not only enhance hepatic IL-17A-producing $\gamma\delta$ T cell homeostasis but also promote liver regeneration [25,68]. Therefore, the role of the microbiota in liver regeneration may depend on hepatic $\gamma\delta$ T cells.

Conclusions and perspectives

$\gamma\delta$ T cells exhibit strong immune defenses against tumors and viral infections and have been employed in treating cancers and infectious diseases [69,70]. In the liver, $\gamma\delta$ T cells not only protect the liver against infections and carcinogenesis but also contribute to immune-mediated liver damage. Although these findings have significantly enhanced our understanding of the roles of $\gamma\delta$ T cells in liver diseases, further studies are still required to elucidate the following: (1) subsets of hepatic $\gamma\delta$ T cells other than IFN- γ - and IL-17A-producing $\gamma\delta$ T cells, (2) the interaction of hepatic $\gamma\delta$ T cells with other intrahepatic cells during the response, (3) the contribution of hepatic $\gamma\delta$ T cells to hepatic immune-tolerant status in physiology, (4) the presence and the specific marker(s) of liver-resident $\gamma\delta$ T cells, (5) the origin and development of liver-resident $\gamma\delta$ T cells, and (6) the tissue-specific regulators of functional polarization of liver-resident $\gamma\delta$ T cells. Clarification on the basic biology and functions of hepatic $\gamma\delta$ T cells is essential for the development of improved $\gamma\delta$ T cell-based therapeutic strategies for liver diseases.

Acknowledgements

This work was supported by the Anhui Natural Science Foundation (No. 1708085QH183), Natural Science Foundation of China (Nos. 81302863, 31390433, and 91542000), and the Ministry of Science and Technology of China (973 Program, No. 2013CB944902).

Compliance with ethics guidelines

Xuefu Wang and Zhigang Tian declare no conflicts of interests. This manuscript is a review article and does not involve a research protocol requiring approval by the relevant institutional review board or ethics committee.

References

- Godfrey DI, Kennedy J, Suda T, Zlotnik A. A developmental pathway involving four phenotypically and functionally distinct subsets of CD3⁺ CD4⁺ CD8⁻ triple-negative adult mouse thymocytes defined by CD44 and CD25 expression. *J Immunol* 1993; 150(10): 4244–4252
- Germain RN. T-cell development and the CD4-CD8 lineage decision. *Nat Rev Immunol* 2002; 2(5): 309–322
- Ciofani M, Knowles GC, Wiest DL, von Boehmer H, Zúñiga-Pflücker JC. Stage-specific and differential notch dependency at the alphabeta and $\gamma\delta$ T lineage bifurcation. *Immunity* 2006; 25(1): 105–116
- Hoh A, Dewerth A, Vogt F, Wenz J, Baeuerle PA, Warmann SW, Fuchs J, Armeanu-Ebinger S. The activity of $\gamma\delta$ T cells against paediatric liver tumour cells and spheroids in cell culture. *Liver Int* 2013; 33(1):127–136
- Haas JD, González FH, Schmitz S, Chennupati V, Föhse L, Kremmer E, Förster R, Prinz I. CCR6 and NK1.1 distinguish between IL-17A and IFN- γ -producing $\gamma\delta$ effector T cells. *Eur J Immunol* 2009; 39(12): 3488–3497
- Muñoz-Ruiz M, Sumaria N, Pennington DJ, Silva-Santos B. Thymic determinants of $\gamma\delta$ T cell differentiation. *Trends Immunol* 2017; 38(5): 336–344
- Groh V, Steinle A, Bauer S, Spies T. Recognition of stress-induced MHC molecules by intestinal epithelial $\gamma\delta$ T cells. *Science* 1998; 279(5357): 1737–1740
- Fay NS, Larson EC, Jameson JM. Chronic inflammation and $\gamma\delta$ T cells. *Front Immunol* 2016; 7: 210
- Vantourout P, Hayday A. Six-of-the-best: unique contributions of $\gamma\delta$ T cells to immunology. *Nat Rev Immunol* 2013; 13(2): 88–100
- Rajoriya N, Fergusson JR, Leithead JA, Klenerman P. $\gamma\delta$ T-lymphocytes in hepatitis C and chronic liver disease. *Front Immunol* 2014; 5: 400
- Wang X, Sun R, Wei H, Tian Z. High-mobility group box 1 (HMGB1)-Toll-like receptor (TLR)4-interleukin (IL)-23-IL-17A axis in drug-induced damage-associated lethal hepatitis: interaction of $\gamma\delta$ T cells with macrophages. *Hepatology* 2013; 57(1): 373–384
- Protzer U, Maini MK, Knolle PA. Living in the liver: hepatic infections. *Nat Rev Immunol* 2012; 12(3): 201–213
- Pellicoro A, Ramachandran P, Iredale JP, Fallowfield JA. Liver fibrosis and repair: immune regulation of wound healing in a solid organ. *Nat Rev Immunol* 2014; 14(3): 181–194
- Robinson MW, Harmon C, O'Farrelly C. Liver immunology and its role in inflammation and homeostasis. *Cell Mol Immunol* 2016; 13(3): 267–276
- Shuai Z, Leung MW, He X, Zhang W, Yang G, Leung PS, Eric Gershwin M. Adaptive immunity in the liver. *Cell Mol Immunol* 2016; 13(3): 354–368
- Horst AK, Neumann K, Diehl L, Tiegs G. Modulation of liver tolerance by conventional and nonconventional antigen-presenting cells and regulatory immune cells. *Cell Mol Immunol* 2016; 13(3): 277–292
- Crispe IN. Immune tolerance in liver disease. *Hepatology* 2014; 60(6): 2109–2117
- Gao B, Jeong WI, Tian Z. Liver: an organ with predominant innate immunity. *Hepatology* 2008; 47(2): 729–736
- Bandyopadhyay K, Marrero I, Kumar V. NKT cell subsets as key participants in liver physiology and pathology. *Cell Mol Immunol* 2016; 13(3): 337–346
- Peng H, Wisse E, Tian Z. Liver natural killer cells: subsets and roles in liver immunity. *Cell Mol Immunol* 2016; 13(3): 328–336
- Ju C, Tacke F. Hepatic macrophages in homeostasis and liver diseases: from pathogenesis to novel therapeutic strategies. *Cell Mol Immunol* 2016; 13(3): 316–327
- Zhou Z, Xu MJ, Gao B. Hepatocytes: a key cell type for innate immunity. *Cell Mol Immunol* 2016; 13(3): 301–315
- Bonneville M, O'Brien RL, Born WK. $\gamma\delta$ T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol* 2010; 10(7): 467–478
- Rao R, Graffeo CS, Gulati R, Jamal M, Narayan S, Zambirinis CP, Barilla R, Deutsch M, Greco SH, Ochi A, Tomkötter L, Blobstein R, Avanzi A, Tippens DM, Gelbstein Y, Van Heerden E, Miller G. Interleukin 17-producing $\gamma\delta$ T cells promote hepatic regeneration in

- mice. *Gastroenterology* 2014; 147(2):473–84.e2
25. Li F, Hao X, Chen Y, Bai L, Gao X, Lian Z, Wei H, Sun R, Tian Z. The microbiota maintain homeostasis of liver-resident $\gamma\delta$ T-17 cells in a lipid antigen/CD1d-dependent manner. *Nat Commun* 2017; 7: 13839
 26. Liaw YF, Chu CM. Hepatitis B virus infection. *Lancet* 2009; 373 (9663): 582–592
 27. Chyuan IT, Tsai HF, Tzeng HT, Sung CC, Wu CS, Chen PJ, Hsu PN. Tumor necrosis factor- α blockage therapy impairs hepatitis B viral clearance and enhances T-cell exhaustion in a mouse model. *Cell Mol Immunol* 2015; 12(3): 317–325
 28. Chen M, Zhang D, Zhen W, Shi Q, Liu Y, Ling N, Peng M, Tang K, Hu P, Hu H, Ren H. Characteristics of circulating T cell receptor $\gamma\delta$ T cells from individuals chronically infected with hepatitis B virus (HBV): an association between V δ 2 subtype and chronic HBV infection. *J Infect Dis* 2008; 198(11): 1643–1650
 29. Chen M, Hu P, Ling N, Peng H, Lei Y, Hu H, Zhang D, Ren H. Enhanced functions of peripheral $\gamma\delta$ T cells in chronic hepatitis B infection during interferon α treatment *in vivo* and *in vitro*. *PLoS One* 2015; 10(3): e0120086
 30. Chen M, Hu P, Peng H, Zeng W, Shi X, Lei Y, Hu H, Zhang D, Ren H. Enhanced peripheral $\gamma\delta$ T cells cytotoxicity potential in patients with HBV-associated acute-on-chronic liver failure might contribute to the disease progression. *J Clin Immunol* 2012; 32(4): 877–885
 31. Kong X, Sun R, Chen Y, Wei H, Tian Z. $\gamma\delta$ T cells drive myeloid-derived suppressor cell-mediated CD8⁺ T cell exhaustion in hepatitis B virus-induced immunotolerance. *J Immunol* 2014; 193 (4): 1645–1653
 32. Rehermann B, Nascimbeni M. Immunology of hepatitis B virus and hepatitis C virus infection. *Nat Rev Immunol* 2005; 5(3): 215–229
 33. Yin W, Tong S, Zhang Q, Shao J, Liu Q, Peng H, Hu H, Peng M, Hu P, Ren H, Tian Z, Zhang D. Functional dichotomy of V δ 2 $\gamma\delta$ T cells in chronic hepatitis C virus infections: role in cytotoxicity but not for IFN- γ production. *Sci Rep* 2016; 6(1): 26296
 34. Tseng CT, Miskovsky E, Houghton M, Klimpel GR. Characterization of liver T-cell receptor $\gamma\delta$ T cells obtained from individuals chronically infected with hepatitis C virus (HCV): evidence for these T cells playing a role in the liver pathology associated with HCV infections. *Hepatology* 2001; 33(5): 1312–1320
 35. Agrati C, Alonzi T, De Santis R, Castilletti C, Abbate I, Capobianchi MR, D’Offizi G, Siepi F, Fimia GM, Tripodi M, Poccia F. Activation of V γ 9V δ 2 T cells by non-peptidic antigens induces the inhibition of subgenomic HCV replication. *Int Immunol* 2006; 18(1): 11–18
 36. Sardinha LR, Elias RM, Mosca T, Bastos KR, Marinho CR, D’Império Lima MR, Alvarez JM. Contribution of NK, NK T, $\gamma\delta$ T, and $\alpha\beta$ T cells to the γ interferon response required for liver protection against *Trypanosoma cruzi*. *Infect Immun* 2006; 74(4): 2031–2042
 37. Tramonti D, Rhodes K, Martin N, Dalton JE, Andrew E, Carding SR. $\gamma\delta$ T cell-mediated regulation of chemokine producing macrophages during *Listeria monocytogenes* infection-induced inflammation. *J Pathol* 2008; 216(2): 262–270
 38. Chen D, Luo X, Xie H, Gao Z, Fang H, Huang J. Characteristics of IL-17 induction by *Schistosoma japonicum* infection in C57BL/6 mouse liver. *Immunology* 2013; 139(4): 523–532
 39. Rinella ME. Nonalcoholic fatty liver disease: a systematic review. *JAMA* 2015; 313(22): 2263–2273
 40. Harley IT, Stankiewicz TE, Giles DA, Softic S, Flick LM, Cappelletti M, Sheridan R, Xanthakos SA, Steinbrecher KA, Sartor RB, Kohli R, Karp CL, Divanovic S. IL-17 signaling accelerates the progression of nonalcoholic fatty liver disease in mice. *Hepatology* 2014; 59(5): 1830–1839
 41. Xu R, Tao A, Zhang S, Zhang M. Neutralization of interleukin-17 attenuates high fat diet-induced non-alcoholic fatty liver disease in mice. *Acta Biochim Biophys Sin (Shanghai)* 2013; 45(9): 726–733
 42. Aizawa Y, Hokari A. Autoimmune hepatitis: current challenges and future prospects. *Clin Exp Gastroenterol* 2017; 10: 9–18
 43. Carey EJ, Ali AH, Lindor KD. Primary biliary cirrhosis. *Lancet* 2015; 386(10003): 1565–1575
 44. Singh S, Talwalkar JA. Primary sclerosing cholangitis: diagnosis, prognosis, and management. *Clin Gastroenterol Hepatol* 2013;11 (8):898–907
 45. Martins EB, Graham AK, Chapman RW, Fleming KA. Elevation of $\gamma\delta$ T lymphocytes in peripheral blood and livers of patients with primary sclerosing cholangitis and other autoimmune liver diseases. *Hepatology* 1996; 23(5): 988–993
 46. Hua F, Wang L, Rong X, Hu Y, Zhang JM, He W, Zhang FC. Elevation of V δ 1 T cells in peripheral blood and livers of patients with primary biliary cholangitis. *Clin Exp Immunol* 2016; 186(3): 347–355
 47. Wen L, Peakman M, Mieli-Vergani G, Vergani D. Elevation of activated $\gamma\delta$ T cell receptor bearing T lymphocytes in patients with autoimmune chronic liver disease. *Clin Exp Immunol* 1992; 89(1): 78–82
 48. Ferri S, Longhi MS, De Molo C, Lalanne C, Muratori P, Granito A, Hussain MJ, Ma Y, Lenzi M, Mieli-Vergani G, Bianchi FB, Vergani D, Muratori L. A multifaceted imbalance of T cells with regulatory function characterizes type 1 autoimmune hepatitis. *Hepatology* 2010; 52(3): 999–1007
 49. Nishio K, Miyagi T, Tatsumi T, Mukai K, Yokoyama Y, Yoshioka T, Sakamori R, Hikita H, Kodama T, Shimizu S, Shigekawa M, Nawa T, Yoshihara H, Hiramatsu N, Yamanaka H, Seino K, Takehara T. Invariant natural killer T cell deficiency leads to the development of spontaneous liver inflammation dependent on $\gamma\delta$ T cells in mice. *J Gastroenterol* 2015; 50(11): 1124–1133
 50. Zhang H, Bernuzzi F, Lleo A, Ma X, Invernizzi P. Therapeutic potential of IL-17-mediated signaling pathway in autoimmune liver diseases. *Mediators Inflamm* 2015; 2015: 436450
 51. Ujiiie H, Shevach EM. $\gamma\delta$ T cells protect the liver and lungs of mice from autoimmunity induced by scurfy lymphocytes. *J Immunol* 2016; 196(4): 1517–1528
 52. Zhao N, Hao J, Ni Y, Luo W, Liang R, Cao G, Zhao Y, Wang P, Zhao L, Tian Z, Flavell R, Hong Z, Han J, Yao Z, Wu Z, Yin Z. V γ 4 $\gamma\delta$ T cell-derived IL-17A negatively regulates NKT cell function in Con A-induced fulminant hepatitis. *J Immunol* 2011; 187(10): 5007–5014
 53. Hammerich L, Bangen JM, Govaere O, Zimmermann HW, Gassler N, Huss S, Liedtke C, Prinz I, Lira SA, Luedde T, Roskams T, Trautwein C, Heymann F, Tacke F. Chemokine receptor CCR6-dependent accumulation of $\gamma\delta$ T cells in injured liver restricts hepatic inflammation and fibrosis. *Hepatology* 2014; 59(2): 630–642
 54. Seo W, Eun HS, Kim SY, Yi HS, Lee YS, Park SH, Jang MJ, Jo E,

- Kim SC, Han YM, Park KG, Jeong WI. Exosome-mediated activation of toll-like receptor 3 in stellate cells stimulates interleukin-17 production by $\gamma\delta$ T cells in liver fibrosis. *Hepatology* 2016; 64(2): 616–631
55. Meng F, Wang K, Aoyama T, Grivennikov SI, Paik Y, Scholten D, Cong M, Iwaisako K, Liu X, Zhang M, Österreicher CH, Stickel F, Ley K, Brenner DA, Kisseleva T. Interleukin-17 signaling in inflammatory, Kupffer cells, and hepatic stellate cells exacerbates liver fibrosis in mice. *Gastroenterology* 2012; 143(3):765–776.e3
56. Tan Z, Qian X, Jiang R, Liu Q, Wang Y, Chen C, Wang X, Ryffel B, Sun B. IL-17A plays a critical role in the pathogenesis of liver fibrosis through hepatic stellate cell activation. *J Immunol* 2013; 191(4): 1835–1844
57. Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ, He J. Cancer statistics in China, 2015. *CA Cancer J Clin* 2016; 66(2): 115–132
58. Yi Y, He HW, Wang JX, Cai XY, Li YW, Zhou J, Cheng YF, Jin JJ, Fan J, Qiu SJ. The functional impairment of HCC-infiltrating $\gamma\delta$ T cells, partially mediated by regulatory T cells in a TGF β - and IL-10-dependent manner. *J Hepatol* 2013; 58(5): 977–983
59. Cai XY, Wang JX, Yi Y, He HW, Ni XC, Zhou J, Cheng YF, Jin JJ, Fan J, Qiu SJ. Low counts of $\gamma\delta$ T cells in peritumoral liver tissue are related to more frequent recurrence in patients with hepatocellular carcinoma after curative resection. *Asian Pac J Cancer Prev* 2014; 15(2): 775–780
60. Ma S, Cheng Q, Cai Y, Gong H, Wu Y, Yu X, Shi L, Wu D, Dong C, Liu H. IL-17A produced by $\gamma\delta$ T cells promotes tumor growth in hepatocellular carcinoma. *Cancer Res* 2014; 74(7): 1969–1982
61. Zhang BN, Watanabe S, Kohyama M, Saijo K, Kusakabe M, Ohno T. Tumor formation suppressed in $\gamma\delta$ T knock-out mice. *Cancer Lett* 2000; 153(1-2): 63–66
62. Silva-Santos B, Serre K, Norell H. $\gamma\delta$ T cells in cancer. *Nat Rev Immunol* 2015; 15(11): 683–691
63. Wu D, Wu P, Qiu F, Wei Q, Huang J. Human $\gamma\delta$ T-cell subsets and their involvement in tumor immunity. *Cell Mol Immunol* 2017; 14(3): 245–253
64. Toutirais O, Cabillic F, Le Fricc G, Salot S, Loyer P, Le Gallo M, Desille M, de La Pintièrre CT, Daniel P, Bouet F, Catros V. DNAX accessory molecule-1 (CD226) promotes human hepatocellular carcinoma cell lysis by V γ 9V δ 2 T cells. *Eur J Immunol* 2009; 39(5): 1361–1368
65. Sugai S, Yoshikawa T, Iwama T, Tsuchiya N, Ueda N, Fujinami N, Shimomura M, Zhang R, Kaneko S, Uemura Y, Nakatsura T. Hepatocellular carcinoma cell sensitivity to V γ 9V δ 2 T lymphocyte-mediated killing is increased by zoledronate. *Int J Oncol* 2016; 48(5): 1794–1804
66. Forbes SJ, Newsome PN. Liver regeneration — mechanisms and models to clinical application. *Nat Rev Gastroenterol Hepatol* 2016; 13(8): 473–485
67. Furuya S, Kono H, Hara M, Hirayama K, Tsuchiya M, Fujii H. Interleukin-17A plays a pivotal role after partial hepatectomy in mice. *J Surg Res* 2013; 184(2): 838–846
68. Wu X, Sun R, Chen Y, Zheng X, Bai L, Lian Z, Wei H, Tian Z. Oral ampicillin inhibits liver regeneration by breaking hepatic innate immune tolerance normally maintained by gut commensal bacteria. *Hepatology* 2015; 62(1): 253–264
69. Wu YL, Ding YP, Tanaka Y, Shen LW, Wei CH, Minato N, Zhang W. $\gamma\delta$ T cells and their potential for immunotherapy. *Int J Biol Sci* 2014; 10(2): 119–135
70. Legut M, Cole DK, Sewell AK. The promise of $\gamma\delta$ T cells and the $\gamma\delta$ T cell receptor for cancer immunotherapy. *Cell Mol Immunol* 2015; 12(6): 656–668