

Effect of down-regulating VEGF on proliferation of colon carcinoma cell HT-29

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Abstract We designed specific small interfering RNA (siRNA) targeting vascular endothelial growth factor (VEGF) mRNA and synthesized oligo fragments, then siRNA was obtained by in vitro transcription and transfected into cultured human colon carcinoma cell line HT-29 with lipofectamine. We also analyzed the effect of the siRNA on proliferation of HT-29 cells by methyl thiazolyl tetrazolium (MTT) assay and expression level of VEGF mRNA of transfected cells by RT-PCR as well as amounts of secreted VEGF protein in the supernatant by enzyme linked immunosorbent assay (ELISA). Two groups of siRNA targeting human VEGF effectively inhibited proliferation of HT-29 cells after transfection. The secretion of VEGF protein also notably decreased, but the control scramble siRNA showed no effect.

Keywords siRNA, HT-29, VEGF, gene therapy

1 Introduction

Colon cancer is one of the most frequent neoplasms in the world. Its incidence is the second highest in Western countries (Kerr, 2003) and the fourth highest in China (Zhang and Zhang, 2001). Therefore, an efficient and well-tolerated therapy for colon carcinoma is urgently needed.

Compelling evidences indicate that uncontrolled angiogenesis is a major contributing factor in both tumor growth and metastasis (Folkman, 1995; Zetter, 1998). A number of growth factors have been identified as potential positive regulators of angiogenesis. Among them, vascular endothelial growth factor (VEGF) is the only growth factor most consistently found under a wide variety of conditions

associated with angiogenesis (Leung et al., 1989). VEGF plays a critical role in normal embryonic angiogenesis and also in the pathological angiogenesis that occurs in a number of diseases, including cancer. Despite its limited expression in normal tissues, VEGF is over-expressed in various human tumors, such as colon cancer, breast cancer, prostate carcinoma, and pancreatic cancer. VEGF165 is the main protein synthesized and secreted by colon carcinoma cells (Lin et al., 1999; Ma and Deng, 2004; Yang and Zhang, 2004).

RNAi is one of the most exciting discoveries in the past decade in functional genomics. RNAi is the sequence-specific gene silencing induced by double-stranded RNA (dsRNA) that is conserved in a variety of organisms: *Caenorhabditis elegans*, *Drosophila*, plants, and mammals (Williams, 1997; Fire et al., 1998; Elbashir et al., 2001). In this study, we designed three pairs of siRNAs according to the sequence of VEGF165, synthesized by T7 RNA polymerase transcription in vitro. We transfected siRNA with lipofectamine 2000TM into human colon carcinoma cells HT-29, which over-expressed VEGF165, to study the inhibitory effect of RNAi on VEGF165 gene expression and cell proliferation and to explore the applying prospect of RNAi in tumor gene therapy.

2 Materials and methods

2.1 Materials

Human colon carcinoma HT-29 cells were obtained from China Peking Union Cell Culture Center. LipofectamineTM 2000, TRIzol Reagent and RPMI Medium 1640 were obtained from Invitrogen Corporation. T7RiboMAXTM Express RNAi System kit and Access RT-PCR Introductory System were obtained from Promega Corporation. Human VEGF ELISA kit was obtained from Boster Corporation.

2.2 siRNAs synthesized by *in vitro* transcription

With Promega software, we selected the target VEGF165 (GenBank: ACCESSION: AB021221) and designed three pairs of DNA oligonucleotides for the *in vitro* transcription. These DNA oligonucleotides were ordered from Sangon Bio Corp, China. The DNA template for *in vitro* transcription of siRNAs was a short duplex oligonucleotide that was consisted of the target sequence plus the T7 RNA polymerase promoter sequence and six extra nucleotides upstream of the minimal promoter sequence to allow more efficient T7 RNA polymerase binding. Additional adenine-adenine nucleotides were added to the 5' end of oligo to allow addition of the two uridine 3' overhangs in the siRNA strands. Two oligonucleotides were annealed to generate separate templates for synthesis of each strand of the siRNA. The separate short RNA strands were synthesized and then annealed to form siRNA. We used T7RiboMAX™ Express RNAi System kit to synthesize two siRNAs and one scrambled siRNA (used for a negative control) (Takei et al., 2004; Yang et al., 2004) with the following sense and antisense sequences.

VEGF siRNA No.1 (siRNA₁):

5'-GAUCAAACCU CACCAAGGCUU-3' (sense)

5'-GCCUUGGUGAGGUUUGAUCUU-3' (antisense)

VEGF siRNA No.2 (siRNA₂):

5'-GGAGUACCCUGAUGAGAUCUU-3' (sense)

5'-GAUCUCAUCAGGGUACUC CUU-3' (antisense)

VEGF siRNA No.2-_{SCR} (siRNA_{SCR}):

5'-GCGUAAACGCGGGAAUUUACUU-3' (sense)

5'-GUAAAUUCCGCGGUACGCUU-3' (antisense)

Each freeze-dried siRNA was reconstituted with RNase-free water to prepare a 200 μM stock solution. Finally, we determined by absorption at 260 nm, the concentration of siRNA separated on 20% polyacrylamide gel electrophoresis.

2.3 Cell culture and transfection

Cells were cultured in RPMI Medium 1640 supplemented with 100 U/mL penicillin and 100 mg/mL streptomycin and 10% heat-inactivated new bovine serum (NBS) at 37°C in a humidified atmosphere with 5% CO₂. At 24 h before transfection at 50%–80% confluency, HT-29 cells were trypsinized and diluted to appropriate concentration (8×10^7 cells/L) with fresh medium without antibiotics. Transfections of VEGF-siRNA or siRNA_{SCR} were performed with Lipofectamine 2000™ according to the manufacturer's instructions. Cells were then treated with various concentrations of siRNA (100, 300, 600, and 900 nmol/L) and 300 nmol/L siRNA_{SCR} (Liu et al., 2004). Cells were treated with OPTI-MEM medium alone for normal control group and with Lipofectamine 2000™- OPTI-MEM for lipofectamine for control group.

2.4 MTT assay

A total of 4×10^4 cells were seeded in each well of 24-well plate and allowed to attach for 24 h. At 24 h after transfection, 100 μL 10 g/L MTT was added to each well, incubated for 4 h, and 1000 μL DMSO was added and lysed for 15 min. The optical density was determined with a microculture plate reader at 492 nm. Absorbance values were normalized to the values obtained for the lipofectamine-treated cells.

2.5 RT-PCR analysis mRNA levels of VEGF

Total RNA was prepared from culture cells (2×10^6) transfected with 300 nmol/L VEGF-siRNA for 24 h and was extracted from HT-29 cells with Trizol reagent. The RNA precipitate was centrifuged and dissolved with 20 μL of RNase-free water. UV spectrophotometer analysis at 260 nm and electrophoresis detection showed the good quality of purified RNA. A measure of 1 μg of total RNA was reversely transcribed to synthesize cDNA at 45°C for 45 min, then the cDNA was subjected to PCR amplification with specific primers in 50 μL mixtures.

The primers for VEGF were

5'-AATCGAGACCCTGGTGGACA-3' (forward)

5'-TTAACTCAAGCTGCCTCGCC-3' (reverse)

and its amplification product was 378 bp. GAPDH was analyzed as internal control.

The primers for GAPDH were

5'-CGTGAAGGACTCATGACCA-3' (forward)

5'-TCCAGGGGTCTTACTCCTTG-3' (reverse)

and its amplification product was 512 bp.

VEGF was amplified through 40 cycles of denaturing at 94°C for 30 s, annealing at 52.5°C for 1 min and extending at 68°C for 2 min with a 10 min extension. The PCR products were electrophoresed on 2% agarose gel, stained with ethidium bromide, and revealed by UV irradiation. The digital images were analyzed with Tanon image analysis software.

2.6 VEGF ELISA analysis

Cells were seeded in 24-well plates treated with 300 nmol/L siRNA for 24 h. Secretion of VEGF into the cell culture supernatant of the HT-29 cells was determined using a human VEGF ELISA kit according to the manufacturer's instructions. All of the analyses and calibrations were carried out in duplicate. The calibration on each microplate used recombinant human VEGF standards (7.8, 15.6, 31.2, 62.5, 125, 250, 500, and 1000 pg/mL). Optical densities were determined using a microplate reader (Rayto2100C) at 450 nm. The blank was subtracted from the readings for each standard and sample. A standard curve was created by plotting the logarithm of the mean absorbance of each standard *versus* the logarithm of the VEGF concentrations. VEGF concentrations were reported as pg/mL.

2.7 Data analysis

Data were described as means $\pm$ SD of the indicated number of separate experiments. A one-way analysis of variance

was performed for multiple comparisons. Data were analyzed using Statistical Program for Social Sciences (SPSS) software.

3 Results

3.1 Proliferation of HT-29 cells s after siRNA transfection

The absorption values in siRNA₁ and siRNA₂ treated HT-29 cells were significantly lower than those in the lipofectamine control group ($P < 0.01$, Table 1, Fig. 1). To determine the effect of different siRNA concentrations on the inhibition of cancer cells growth, we evaluated the effect of different siRNA concentrations against VEGF mRNA molecule. The results showed that transfected VEGF siRNA markedly inhibited the growth of MCF-7 cells, while siRNA_{SCR} and lipofectamine group had no effect. Meanwhile, 900 nmol/L siRNA had lower effect than 300 nmol/L siRNA to HT-29 cells did (Fig. 2). Thus, to suppress the proliferation of MCF-7 cells effectively and decrease the toxicity of siRNAs, we used 300 nmol/L VEGF-siRNA for the following experiments.

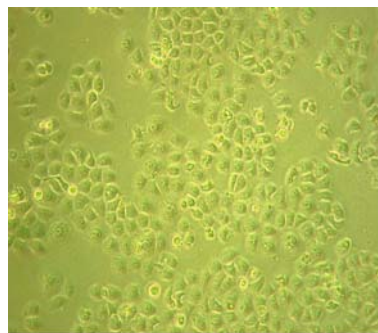
Table 1 MTT metabolic ratio of each group siRNA transfected HT-29 cell ($\bar{x} \pm s$, $n = 6$)

Groups	A ₄₉₂ of siRNA ₁	A ₄₉₂ of siRNA ₂
100 nmol/L group	0.331 ± 0.052**	0.334 ± 0.048**
300 nmol/L group	0.305 ± 0.073**	0.303 ± 0.071**
600 nmol/L group	0.326 ± 0.055**	0.323 ± 0.052**
900 nmol/L group	0.486 ± 0.059*	0.480 ± 0.060*
Scramble group	/	0.538 ± 0.034
Lipofectamine group	/	0.555 ± 0.023
Normal control group	/	0.570 ± 0.048

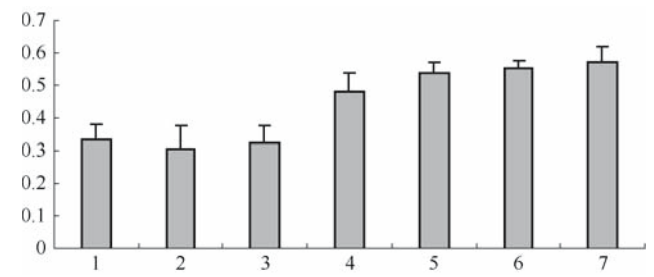
** $: P < 0.01$; * $: P < 0.05$, compared with lipofectamine group

3.2 The changes of VEGF mRNA levels after siRNA transfection

Compared with the control, the mRNA expression of VEGF in HT-29 cells was reduced clearly after transfection with



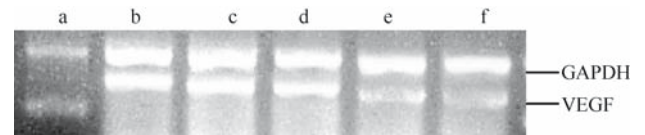
(a)



1: 100 nmol/L group; 2: 300 nmol/L group; 3: 600 nmol/L group; 4: 900 nmol/L group; 5: scramble group; 6: lipofectamine group; 7: normal control group (compared with lipofectamine control)

Fig. 2 Contrast figure of MTT metabolic ratio of each group siRNA₂ transfected HT-29 cell

VEGF-siRNA, but without altering with siRNA_{SCR}. The GAPDH has no changes (Fig. 3, Table 2).



a: DNA Marker; b: Normal control group; c: Lipofectamine group; d: Scramble group; e: siRNA₁ group; f: siRNA₂ group

Fig. 3 Expression of VEGF mRNA analyzed by RT-PCR, GAPDH used as a control

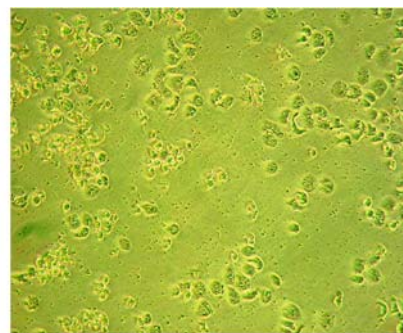
Table 2 RT-PCR OD contrasts of each group HT-29 cell ($\bar{x} \pm s$, $n = 4$)

Group	OD _{VEGF} /OD _{GAPDH}
Normal control group	0.894 ± 0.099
Lipofectamine group	0.872 ± 0.101
siRNA _{SCR} group	0.853 ± 0.145
siRNA ₁ group	0.543 ± 0.106*
siRNA ₂ group	0.532 ± 0.122*

* $: P < 0.01$, compared with lipofectamine group

3.3 ELISA analysis protein level of VEGF after siRNA transfection

VEGF siRNA₁ and siRNA₂ significantly suppressed secretion of VEGF in human colon carcinoma HT-29 cells after



(b)

(a): Lipofectin control HT-29 cell; (b): 300 nmol/L siRNA₂ transfected HT-29 cell

Fig. 1 HT-29 cell photos 24 hours after siRNA transfected ($\times 100$)

transfection with siRNA compared with lipofectamine group. There was no difference between the VEGF siRNA_{SCR} group and lipofectamine control group (Table 3).

Table 3 VEGF level in the medium of each HT-29 cell group ($\bar{x} \pm s$, $n = 3$)

Group	VEGF/(pg·mL ⁻¹)	Inhibition efficiency/%
siRNA ₁ group	240.00 ± 43.1*	40.3
siRNA ₂ group	206.00 ± 55.2*	48.8
siRNA _{SCR} group	382.00 ± 76.2	5.0
Lipofectamine group	402.00 ± 58.6	/
Normal control group	414.00 ± 61.5	/

*: $P < 0.05$, compared with lipofectamine control;

Inhibition efficiency = (lipofectamine group — siRNA group)/lipofectamine group

4 Discussion

Overexpression of VEGF has been observed in some solid human tumours, and correlated with tumour angiogenesis, progression, metastasis, and poor outcome (Tokunaga et al., 1998). In animal experiments it was reported that elevated VEGF can accelerate tumour growth (Ferrara and Davis-Smyth, 1997). Therefore, specific downregulation of VEGF might be a potential therapeutic strategy against human cancers, including colon cancer. Some studies found that RNAi seemed to be quantitatively more efficient and durable in cell culture and in nude mice than antisense RNA (Sohail et al., 2003; Harborth et al., 2001). Specific downregulation of genes that promote proliferation, prevent apoptosis or chemotherapy resistance by RNAi is a promising treatment approach. In this study, we have shown that two VEGF siRNA molecules can effectively downregulate VEGF overexpression with great specificity and suppressed cell proliferation of colon carcinoma *in vitro*. The negative control siRNAs-siRNA_{SCR} did not induce similar effects in HT-29 cells.

We have shown that siRNA synthesized by *in vitro* transcription with T7 RNA polymerase, provides an economical alternative to chemical synthesis of siRNA. The synthesis of siRNAs by T7 RNA polymerase significantly reduces the cost of RNAi in mammalian cells and paves the way for application of RNA interference on a whole genome scale. Lipofectamine is an effective transfected agent (Donze and Picard, 2002; Martinez et al., 2002). We successfully found effective siRNA sequences to suppress VEGF expressing in HT-29 cells and thus shed light on the treatment of neovascularization.

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