

## **Appendix**

### **Upcycling waste protein and heavy metal into single-atom catalytic gas diffusion electrode for CO<sub>2</sub> reduction**

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## Experimental procedures

### Synthesis of electrode for oxygen evolution

In the MEA, the counter electrode was a 4 cm<sup>2</sup> titanium mesh (100 mesh) with IrO<sub>2</sub> coating. Briefly, a titanium mesh was heated in 6 M HCl at 85°C to sweep surface impurity. Then the mesh was drop-cast by H<sub>2</sub>IrCl<sub>6</sub>·xH<sub>2</sub>O solution (1 mL solution consisted 10mg H<sub>2</sub>IrCl<sub>6</sub>·xH<sub>2</sub>O, 900 μL isopropanol and 100 μL 6 M HCl), and followed with heating at 100°C for 10 min. The dried mesh was further calcinated at 500°C for 10 min to form IrO<sub>2</sub> coating. This process was triplicated to achieve a suitable 2 mg/cm<sup>2</sup> H<sub>2</sub>IrCl<sub>6</sub>·xH<sub>2</sub>O loading.

### Evaluation of turnover frequency (TOF)

TOFs/h for ECO<sub>2</sub>RR products were evaluated based on 2 calculation methods. The first is the electrochemical active surface area (ECSA) normalization. Proposed by Jiang *et al.* [1] and Chen *et al.* [2], the ECSA of SP<sub>GDE</sub> and NP<sub>GDE</sub> were calculated to 404.6 cm<sup>2</sup> and 357.90 cm<sup>2</sup> according to 21 μF/cm<sup>2</sup> of reported electrochemical double layer capacity (EDLC) of graphene (Fig. S17, Ref. 1 and 2). Then the moles of C atoms were estimated by the formula: ECSA/(specific surface area)×12 (11.97 m<sup>2</sup>/g for SP<sub>GDE</sub> and 16.39 m<sup>2</sup>/g for NP<sub>GDE</sub>). And considering the Ni atomic content in the membrane electrode (XPS result, Fig. S7 and S8), accordingly number of Ni sites in SP<sub>GDE</sub> and NP<sub>GDE</sub> were 1.06×10<sup>-8</sup> and 1.02×10<sup>-8</sup>, respectively. Then TOFs based on ESCA normalization was calculated by  $(j_{\text{total}} \times t \times FE_{\text{CO}}) / (2F \times \text{the number of Ni sites})$ .

Mass loading normalization TOFs were calculated by mass loading of Ni in the catalyst. The number of Ni sites were calculated by (catalyst loading mass × Ni mass content)/58, Ni mass contents were 3.75 wt% and 2.41 wt% for SP<sub>GDE</sub> and NP<sub>GDE</sub>, respectively. Accordingly, TOFs were calculated as the formula aforementioned.

### Calculation of potential extraction amounts of protein from agriculture & food waste

According to the Food and Agriculture Organization of the United Nations (FAO) statics, 7 kinds of agriculture & food waste amount were reported in the latest *Global Food Losses and Wastes* [3].

The calculation in our study referred statics provided by FAO. But due to the inexact data provided, we provided some approximate data according to the statics charts in FAO's report [3]. We also gave the protein content and annual yield of several agriculture & food representative in each category; the comprehensive protein content of each category was calculated from weighted average of main subdivision of each category (Table S2 to S3) since no exact wasted agriculture & food amount was provided by FAO [3]. Total wasted agriculture & food amounts were calculated via comprehensive lost & wasted rate of 7 macro regions multiply approximate data of 7 kinds of agriculture & food production. All data were summarized in Table S2 to S3.

Lost & wasted amount was calculated as bellow:

Lost & wasted amount =  $\sum_{i=1}^7$  Each category production of each macro region  $\times$  comprehensive lost & wasted rate

Where,  $i$  represents 7 macro regions mentioned in Table S3.

Potential extracted protein amount was calculated based on the following:

Potential protein extraction amount = production  $\times$  comprehensive protein content

Comprehensive protein content =  $\sum_{i=1}^n x_i \times$  average protein content

Where,  $x_i$  represents the main subdivision production in each category.

### **Calculation of potential CO<sub>2</sub> capture when using SP<sub>GDE</sub> in practice under renewable electricity driving**

Considering the durability of each SP<sub>GDE</sub> was 8 h, and average 90% FE<sub>CO</sub> (Fig. 5f) during operation under 50 mA/cm<sup>2</sup>, the annual CO<sub>2</sub> capture amount per ton of extracted protein was as follows:

CO<sub>2</sub> capture amount =  $x_{CO} \times v \times 60 \times 24 \times 365 /$  [extracted protein amounts in per cm<sup>2</sup>  $\times$  (365  $\times$  24 / 8)]

Where,  $x_{CO}$  represents the volume fraction of CO;  $v$  is the flow rate; extracted protein amount in per cm<sup>2</sup> was calculated as below:

Extracted protein amount in per cm<sup>2</sup> = protein addition during electrospinning  $\times$  pristine SP<sub>GDE</sub> area  $\times$  contraction factor

Where, the pristine  $SP_{GDE}$  was around  $450\text{ cm}^2$  with 10 g extracted protein as the feedstock. After preoxidation and carbonization, the final  $SP_{GDE}$  shrank to around 0.7 area of the pristine one, thus the contraction factor was 0.7.

### **Energy input to synthesize $SP_{GDE}$**

In this study, we only consider the extracted protein listed in Table S3 and S4. According to the energy limitation to produce common graphene ( $6440\text{ kW}\cdot\text{h/t}$ ), converting all extracted protein into  $SP_{GDE}$  needs  $125.5\text{ million tons} \times 6440\text{ kW}\cdot\text{h/t} = 808.22\text{ billion kW}\cdot\text{h}$ , which equals to 99.33 million tons standard coal combustion (1  $\text{kW}\cdot\text{h}$  electricity generation needs 0.1229 kg standard coal combustion). The total C combustion reaches to 66.18 million tons and correspond 123.51 billion  $\text{m}^3$   $\text{CO}_2$  emission (1 ton standard coal contains 0.67 tons C, this data was suggested by Nation Development and Reform Commission of China. And 1 ton C combustion equals 3.67 tons or 1866.33  $\text{m}^3$   $\text{CO}_2$  emission).

### **Implications toward carbon reduction and heavy metal recovery**

The annual generation of vast wastes worldwide has underscored the need for mitigating both environmental pollutants and  $\text{CO}_2$  emissions effectively. In this context, we introduce  $\text{ECO}_2\text{RR}$  by  $SP_{GDE}$  as a promising approach to reuse organic wastes for carbon mitigation. Due to the incredible magnitude of production annually, we investigate the potential of agriculture & food wastes as the protein-enriched resources to synthesize  $SP_{GDE}$ . A 2011 report by the Food and Agriculture Organization of the United Nations (FAO) notes that around 1.3 billion tons global agriculture & food wastes are produced annually, including 7 categories — cereals, roots and tuber, oilcrops and pulses, fruits and vegetables, meat, fish and diary (Gustavsson et al., 2011). Based on the protein content of these 7 categories (summarized in Table S2 and S3), available waste protein amounts are expected to be ~251 million tons, corresponding to ~125 million tons C (considering 50 wt% C content in protein) that can be sequestered by carbonization of waste protein to produce the electrode material. The corresponding energy required to produce  $SP_{GDE}$  is estimated to 808 billion  $\text{kW}\cdot\text{h}$  annually, which equals to 99 million tons standard coal combustion. Totally, if the strategy of this study is used, more than 353 million tons C can be converted into several

chemicals and more than 125 million tons C are fixed into  $SP_{GDE}$  itself. Moreover, 5 million tons heavy metals can be recovered for SACs synthesis.

**Table S1** XPS result of Ni content (atom ratio) in GDEs

NP <sub>GDE</sub>	SP <sub>GDE</sub>
0.26%	0.39%



**Table S3** Wasted agriculture & food amount and rate in 7 macro regions, and potential protein extraction amount

Lost & wasted rate						
Europe	North America, Oceania	Industrialize Asia	Sub-Sahara Africa	North Africa, West and Central Asia	South and Southeast Asia	Latin America
~52%	~60%	~48%	~45%	~33%	~41%	~40%
Lost & wasted amount						
Cereals	Roots and tubers	Oilcrops and pulses	Fruit and vegetables	Meat	Fish	Diary
~1114 (6.99%)	~372 (1.85%)	~255 (38.83%)	~749 (1%)	~120 (19.54%)	~76 (18%)	~350 (3.3%)
Potential protein extraction amount						
Cereals	Roots and tubers	Oilcrops and pulses	Fruit and vegetables	Meat	Fish	Diary
~78	~7	~99	~7	~23	~14	~12

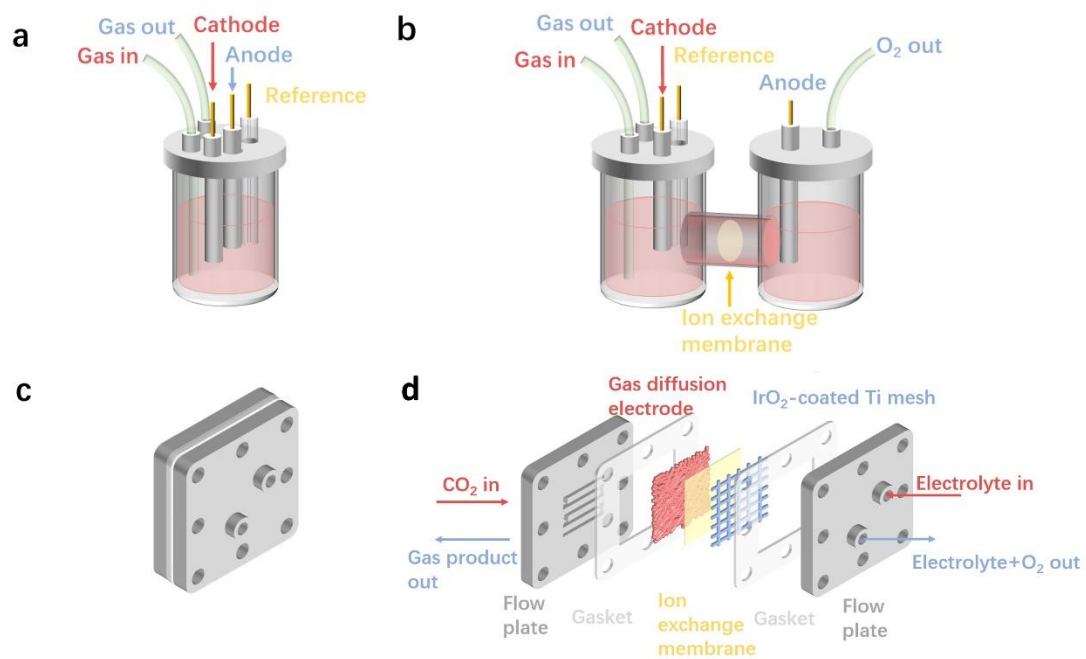
Note: 1. Unit: million tons; 2. Statistics in brackets are comprehensive protein content.

**Table S4** Compositions of SP processing wastewater (mg/L)

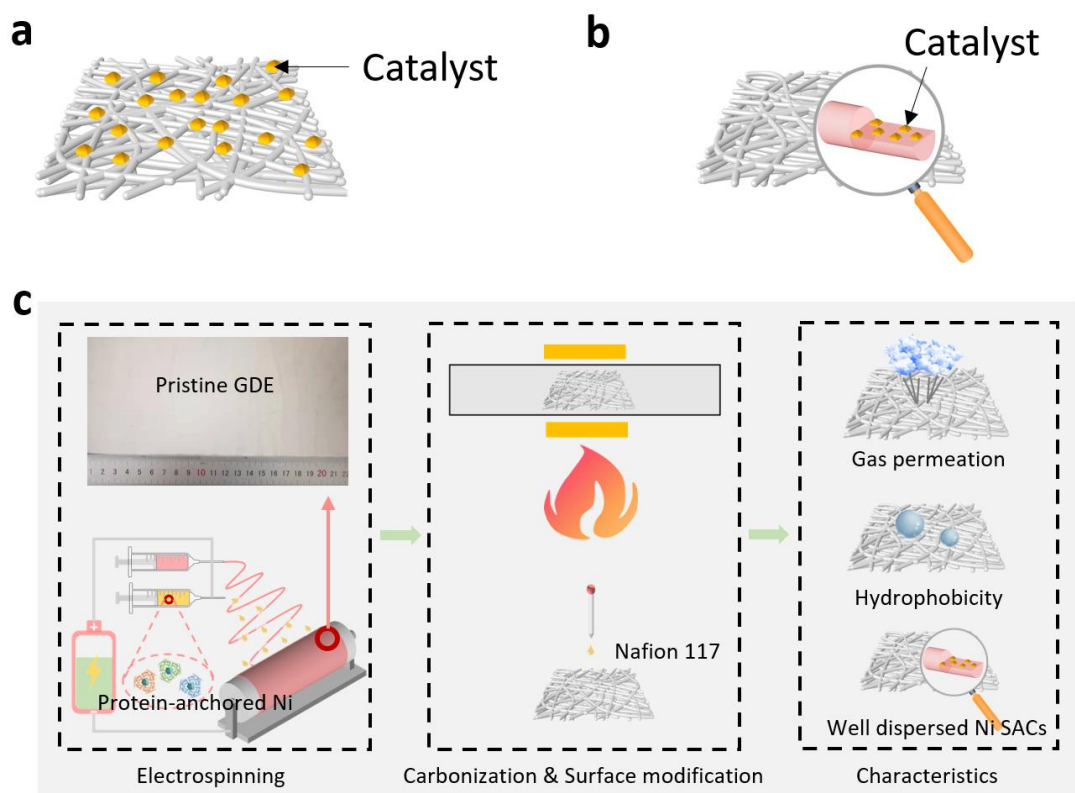
COD <sub>Cr</sub>	BOD <sub>5</sub>	pH	Total soluble protein
15000 ± 125	8500 ± 80	5.0 ± 0.1	10000 ± 100
Na	K	Ca	Mg
271 ± 22	455 ± 161	74 ± 19	81 ± 33
Fe	S		
17 ± 5	368 ± 82		

**Table S5** Compositions of electroplating wastewater (mg/L)

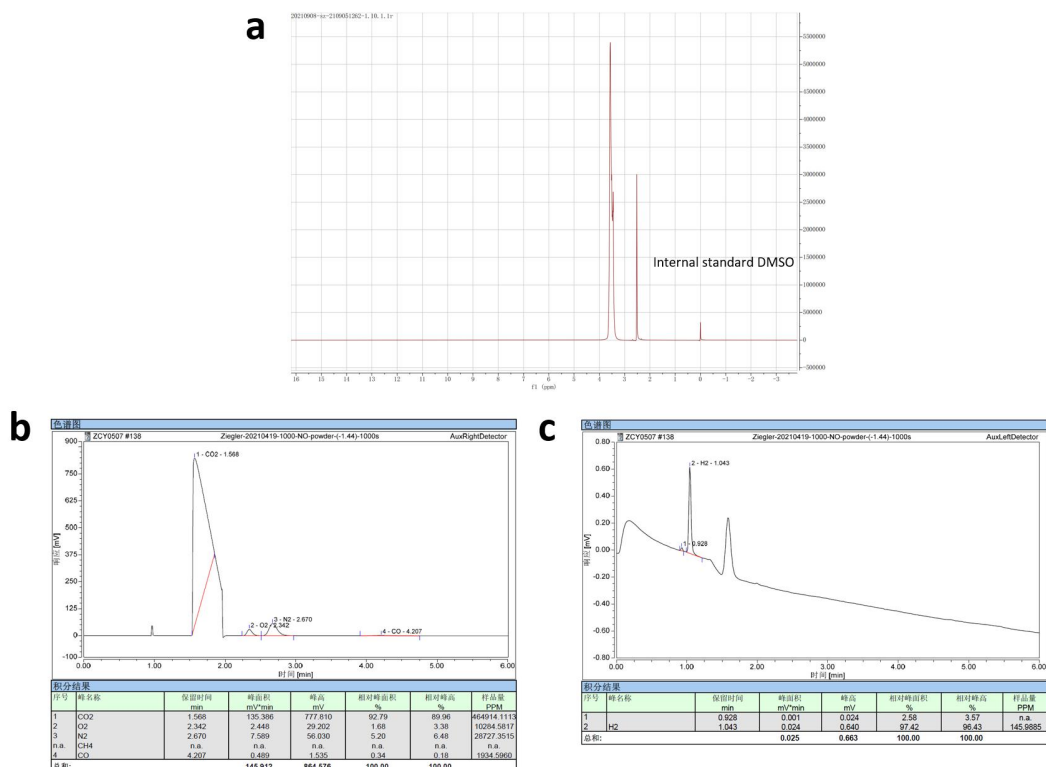
Ni	Al	Cd	Co	Cu	Fe	Mn	Pd	Zn
182.94	9.40	1.58	0.20	1.26	12.59	0.12	0.29	4.48



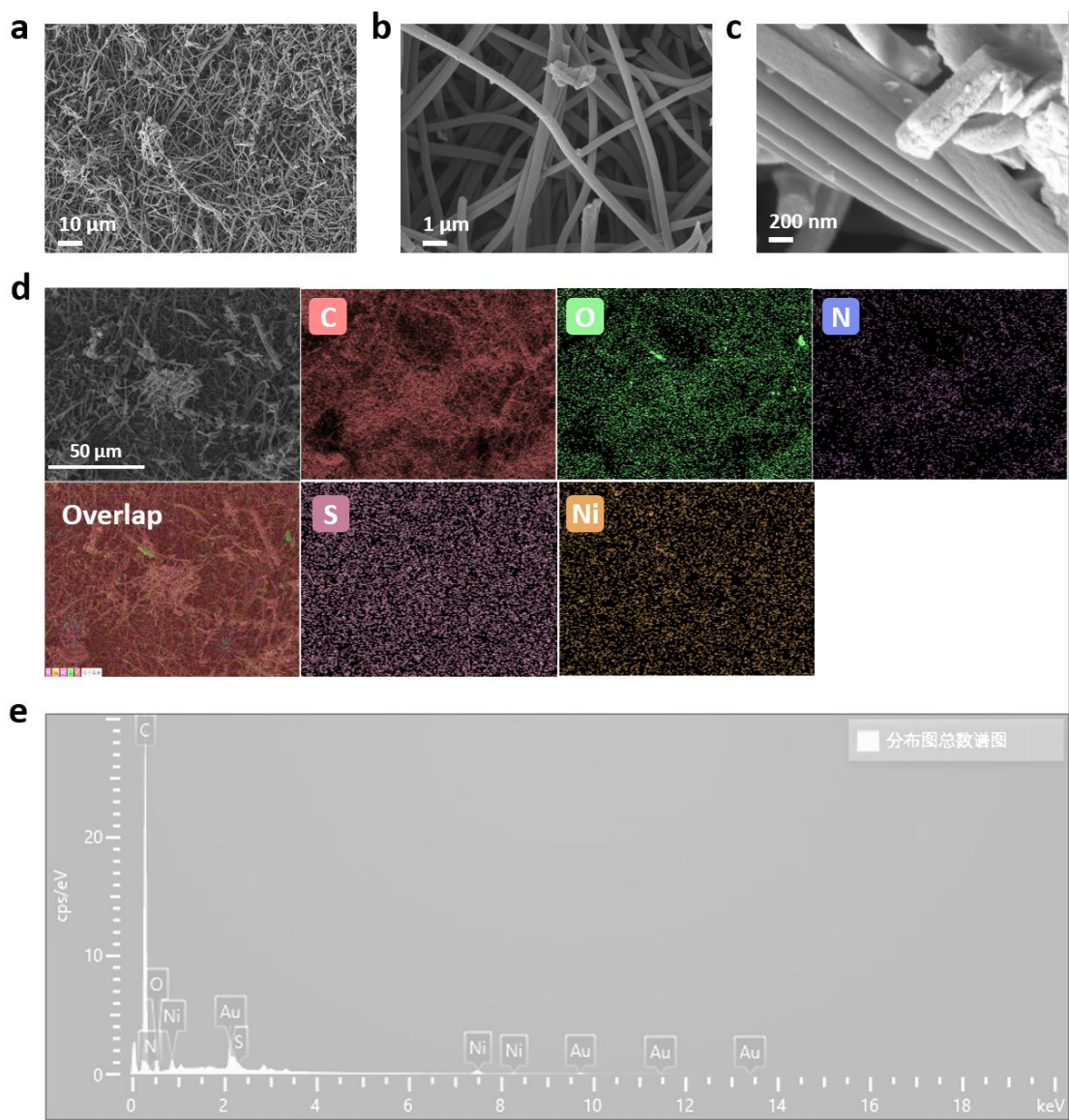
**Fig. S1** Electrochemical CO<sub>2</sub>RR electrolysis cell. **(a)** Traditional single chamber cell. **(b)** Traditional H-type cell. **(c)** and **(d)** Membrane electrode assembly.



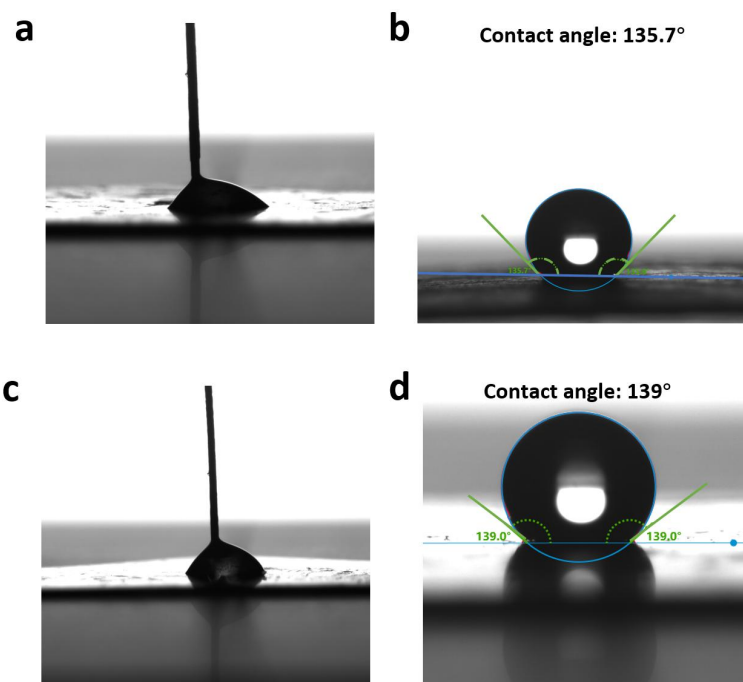
**Fig. S2** Gas diffusion electrode (GDE). **(a)** Traditional GDE with catalysts slathering onto a gas diffusion layer (GDL). **(b)** An integrated gas diffusion electrode with Ni SACs inserting into the nanofibers of GDL, proposed by Yang *et al.* [10] **(c)** An overview of protein as the carbon matrix for GDE synthesis and its characteristics.



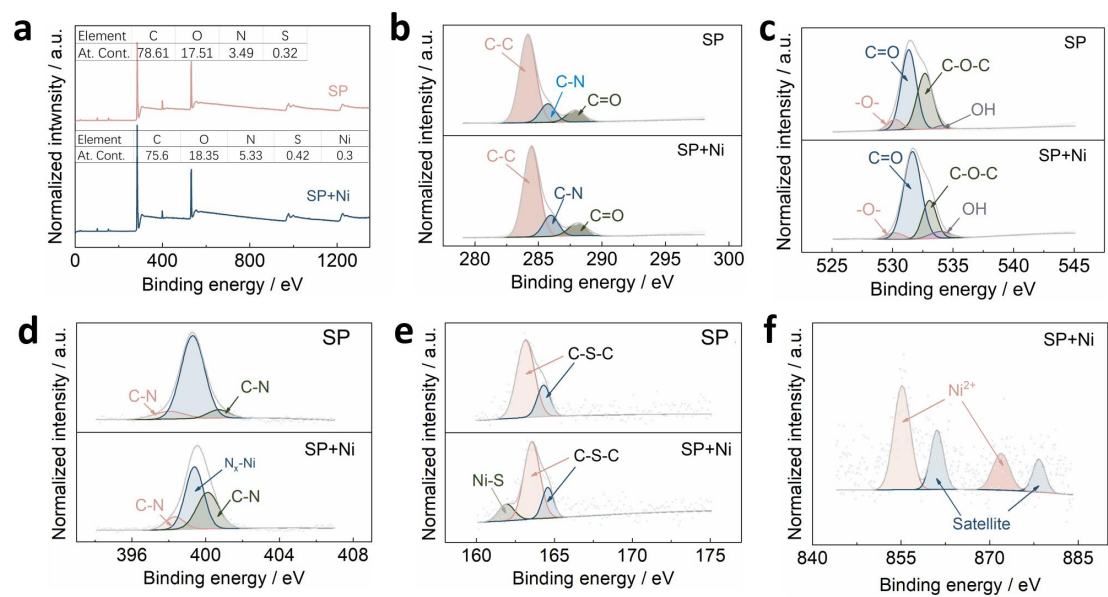
**Fig. S3** ECO<sub>2</sub>RR products analysis. **(a)** <sup>1</sup>H NMR spectra of 0.5M KHCO<sub>3</sub> undergoing 10 h ECO<sub>2</sub>RR with powdered SP<sub>GDE</sub> as the working electrode under -0.8 V vs RHE. **(b)** and **(c)** Gas chromatography analysis of the main gas products under -0.8 V vs RHE.



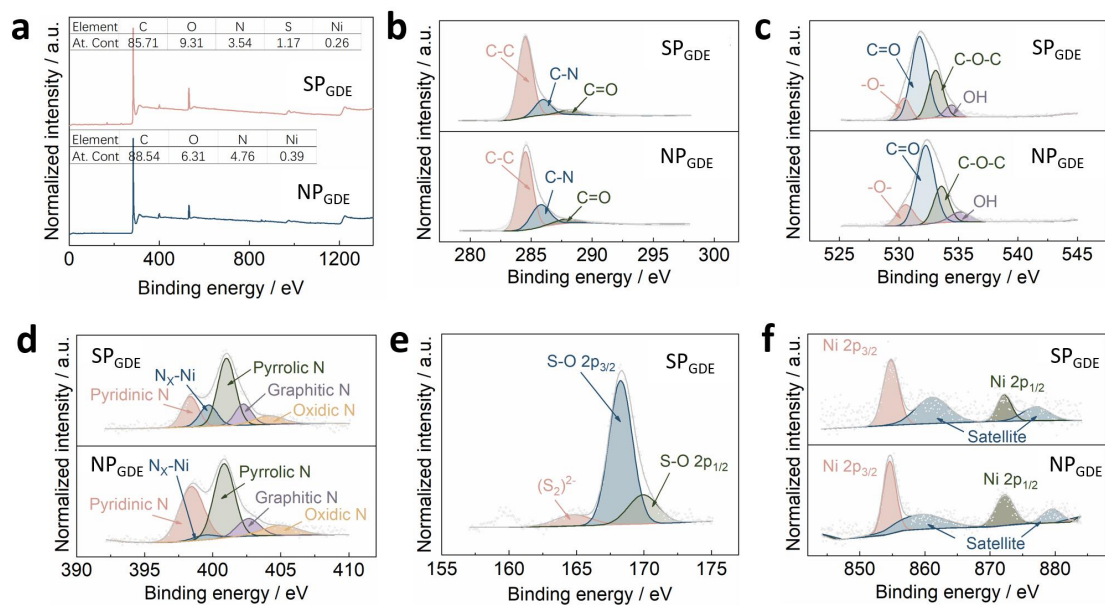
**Fig. S4** Surface texture of SP<sub>GDE</sub> and elemental distribution. **(a)** to **(c)** SEM image. **(d)** Elemental mapping. **(e)** EDS spectra.



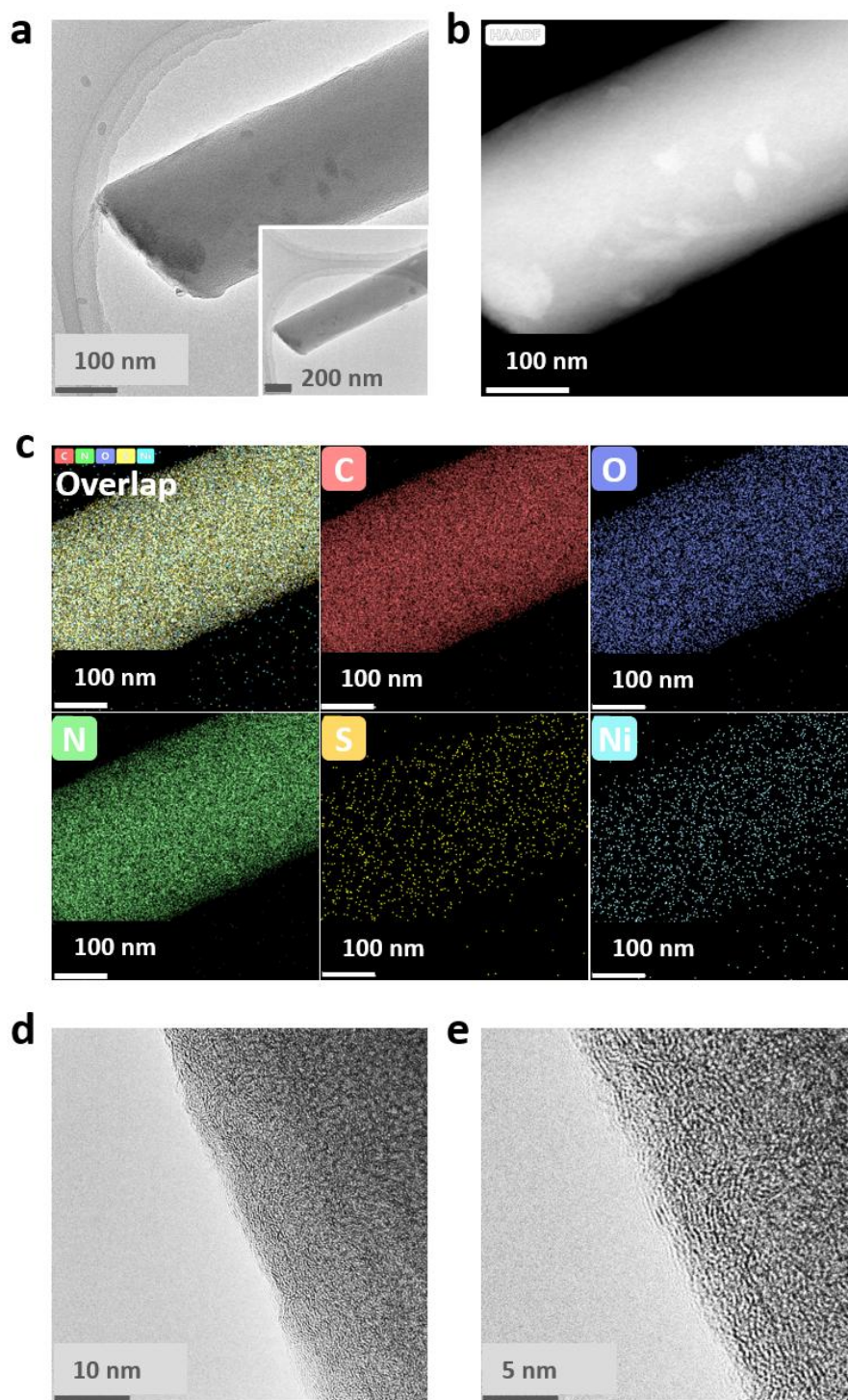
**Fig. S5** Contact angle measurement of SP<sub>GDE</sub> surface and NP<sub>GDE</sub> surface. **(a)** Hydrophilic nature of SP<sub>GDE</sub> surface. **(b)** Hydrophobic nature of SP<sub>GDE</sub> surface with 5 wt% Nafion 117 solution modification, the contact angle is 135.7°. **(c)** Hydrophilic nature of NP<sub>GDE</sub> surface. **(d)** Hydrophobic nature of NP<sub>GDE</sub> surface with 5 wt% Nafion 117 solution modification, the contact angle is 139°.



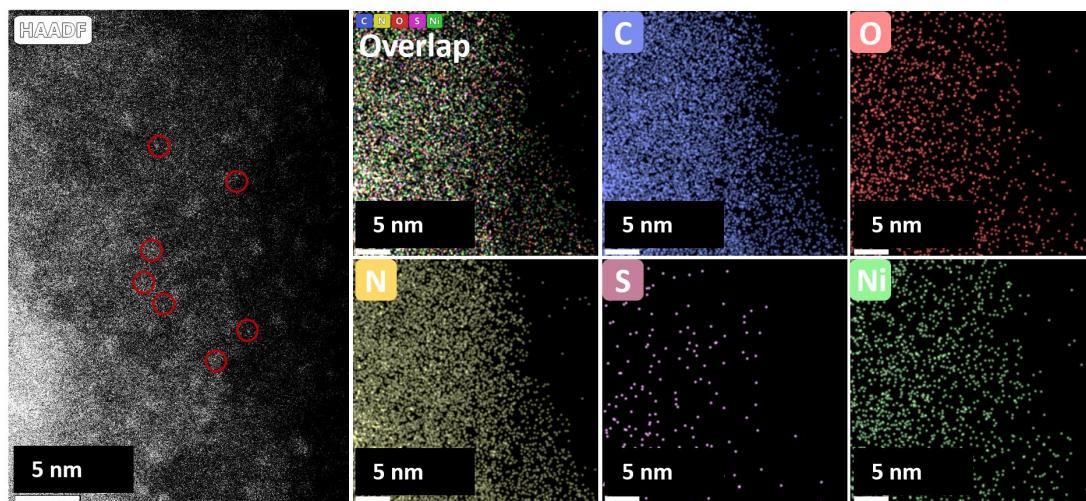
**Fig. S6** XPS spectra of reclaimed SP and SP anchoring Ni (SP+Ni). **(a)** Full spectrum. **(b)** C 1s. **(c)** O 1s. **(d)** N 1s. **(e)** S 2p. **(f)** Ni 2p.



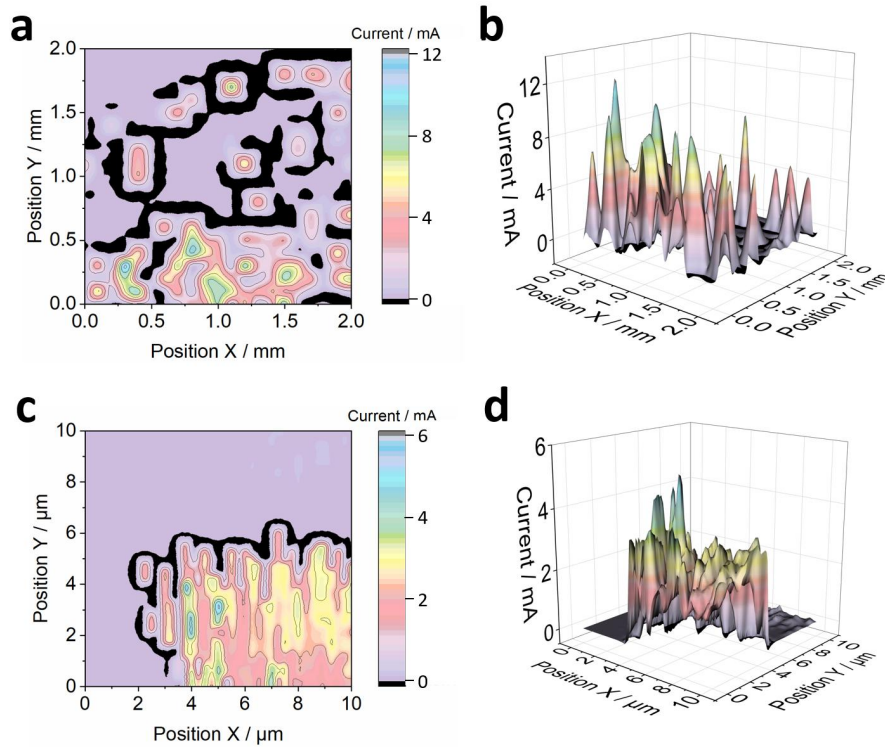
**Fig. S7** XPS spectra of SP<sub>GDE</sub> and NP<sub>GDE</sub>. **(a)** Full spectrum. **(b)** C 1s. **(c)** O 1s. **(d)** N 1s. **(e)** S 2p. **(f)** Ni 2p.



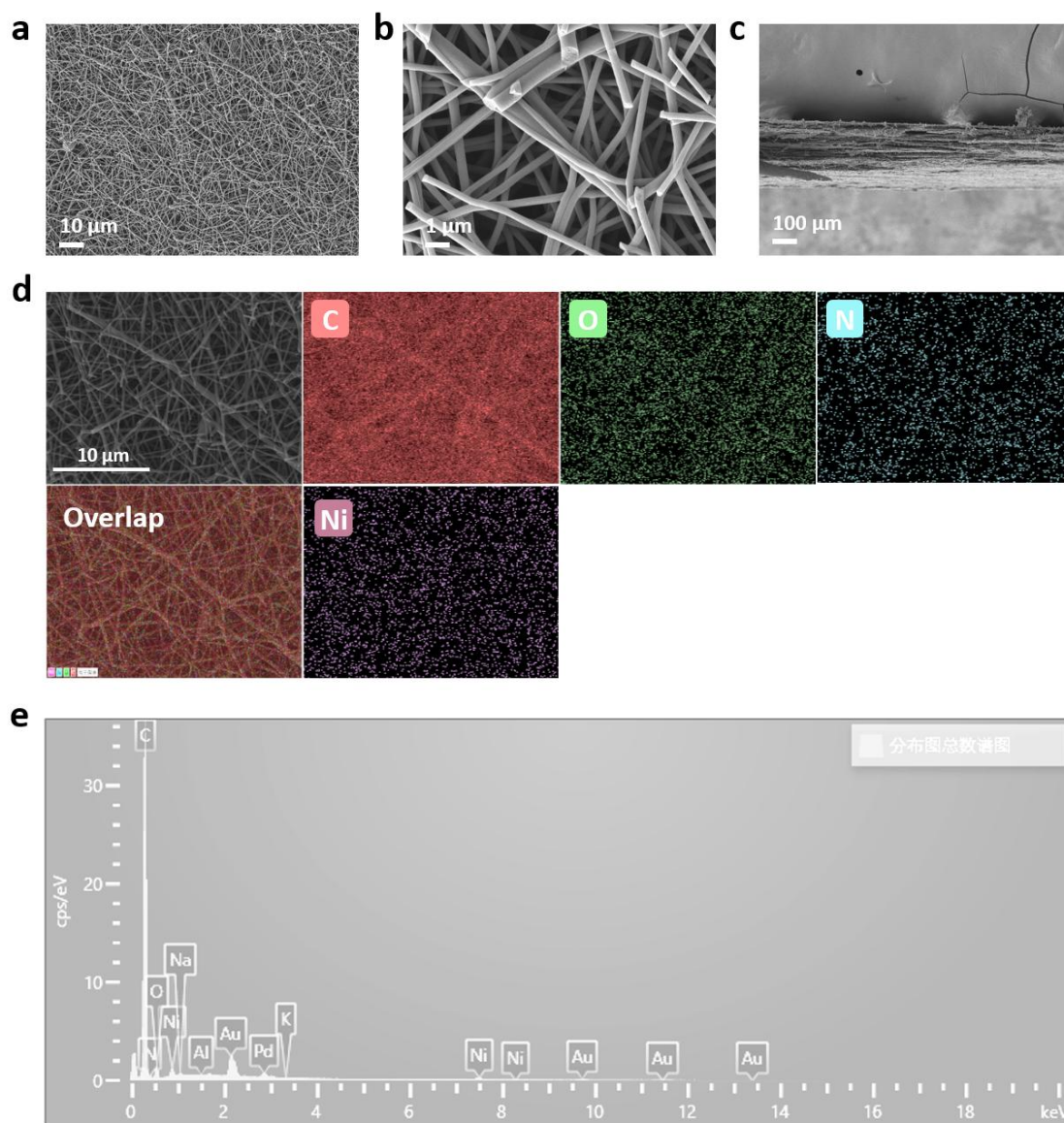
**Fig. S8** TEM images of SP<sub>GDE</sub>. (a) to (c) TEM, HAADF-TEM and corresponding elemental mapping of a SP<sub>GDE</sub> nanofiber. (d) and (e) TEM images of nanofiber's carbon matrix.



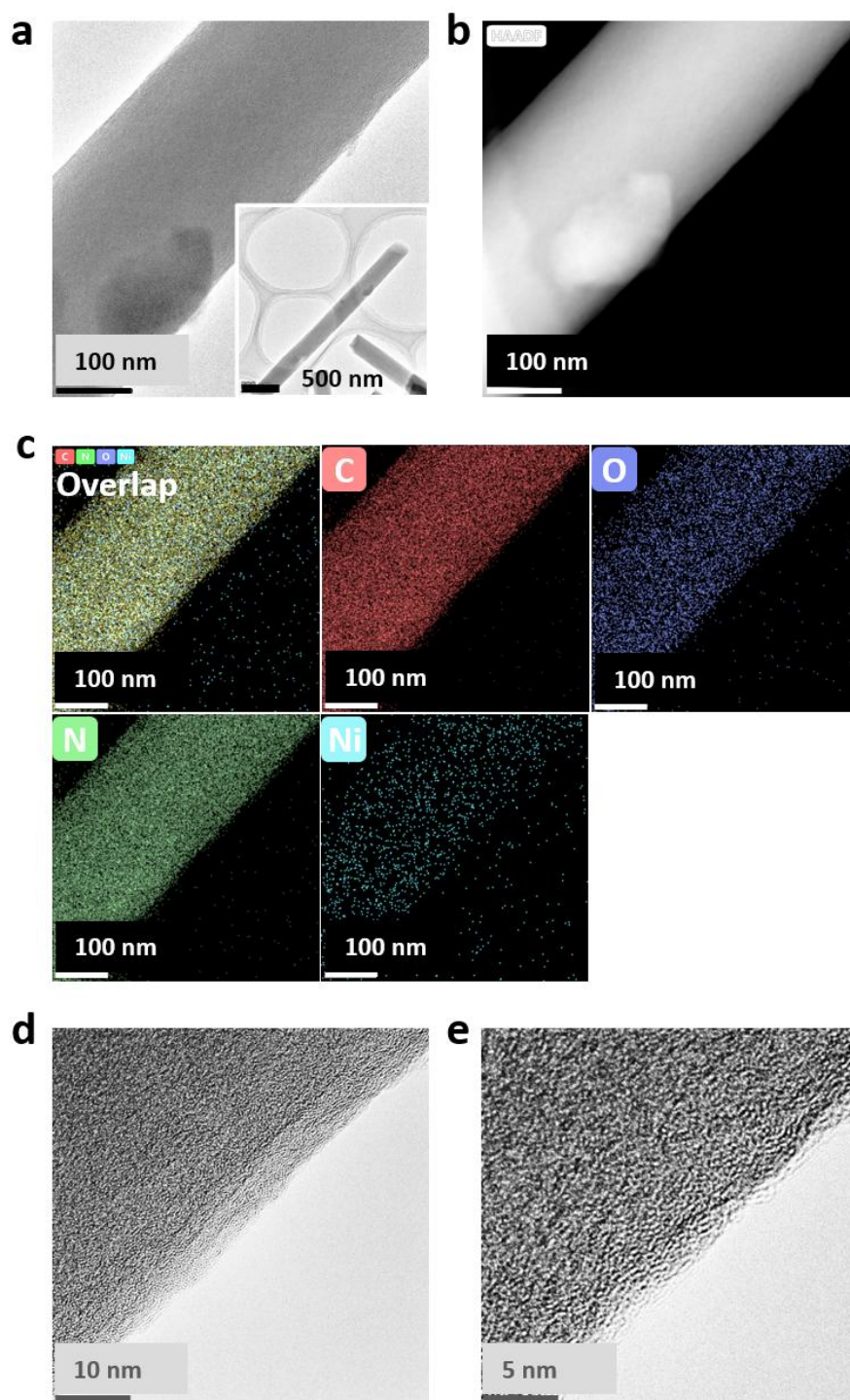
**Fig. S9** AC-HAADF-STEM image and corresponding elemental mapping of SP<sub>GDE</sub>.



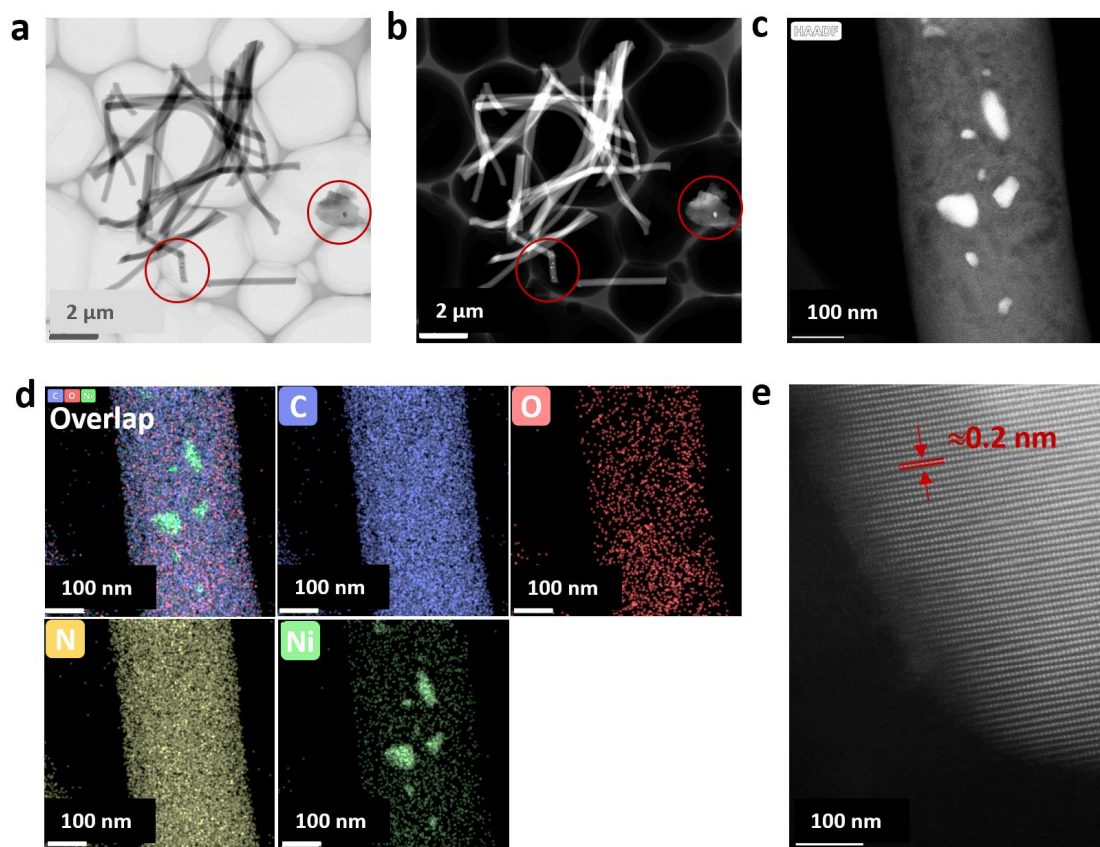
**Fig. S10** Scanning Electrochemical Microscopy images of boarder between SP<sub>GDE</sub> and epoxy resin. Size: 2 mm×2 mm (a and b), 10 μm×10 μm (c and d). Step length: 100 μm (a and b), 250 nm (c and d).



**Fig. S11** Surface texture of NP<sub>GDE</sub> and element distribution. **(a)** and **(b)** Surface SEM image. **(c)** Sectional SEM image. **(d)** Elemental mapping. **(e)** EDS spectra.



**Fig. S12** TEM images of  $\text{NP}_{\text{GDE}}$ . **(a)** to **(c)** TEM, HAADF-TEM and corresponding elemental mapping of  $\text{NP}_{\text{GDE}}$  nanofiber. **(d)** and **(e)** TEM images of nanofiber's carbon matrix.



**Fig. S13** AC-HAADF-STEM images of NP<sub>GDE</sub> nanofibers. **(a)** STEM of nanofibers of NP<sub>GDE</sub>. **(b)** to **(d)** HAADF-STEM of nanofibers of and corresponding elemental mapping. **(e)** Fringe pattern of Ni (111) originated from Ni nanoparticles onto the surface of NP<sub>GDE</sub> surface.

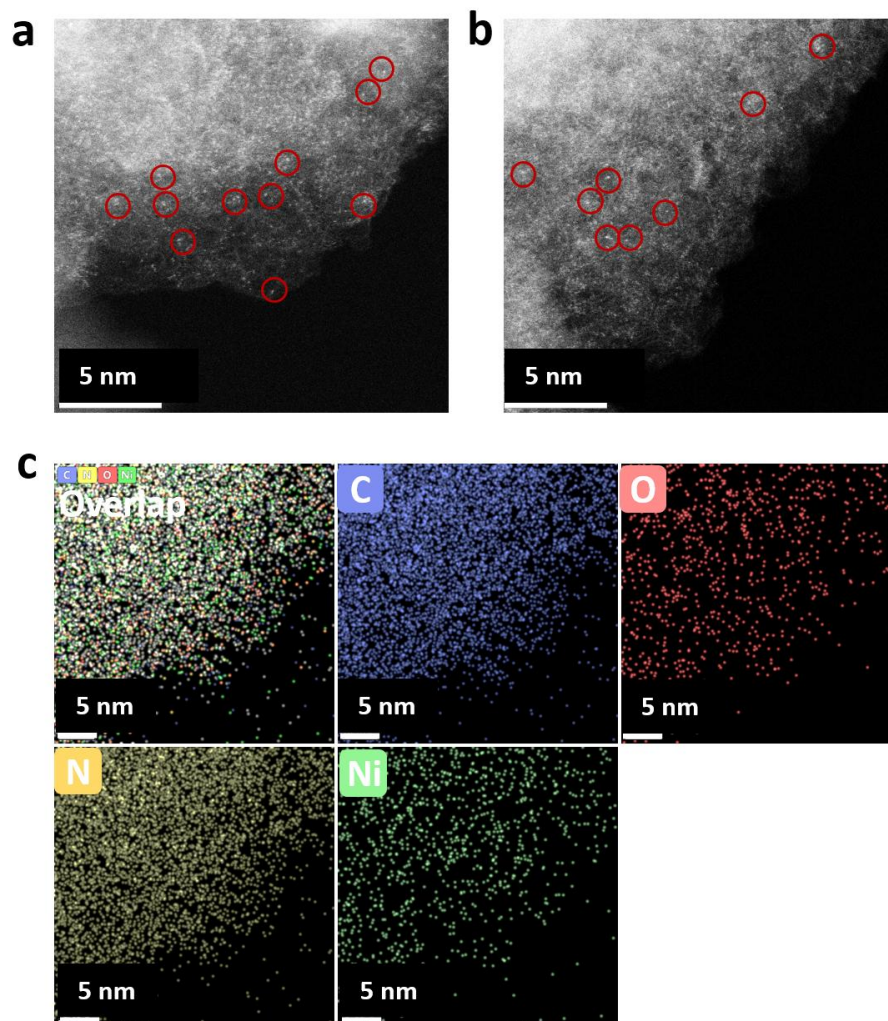
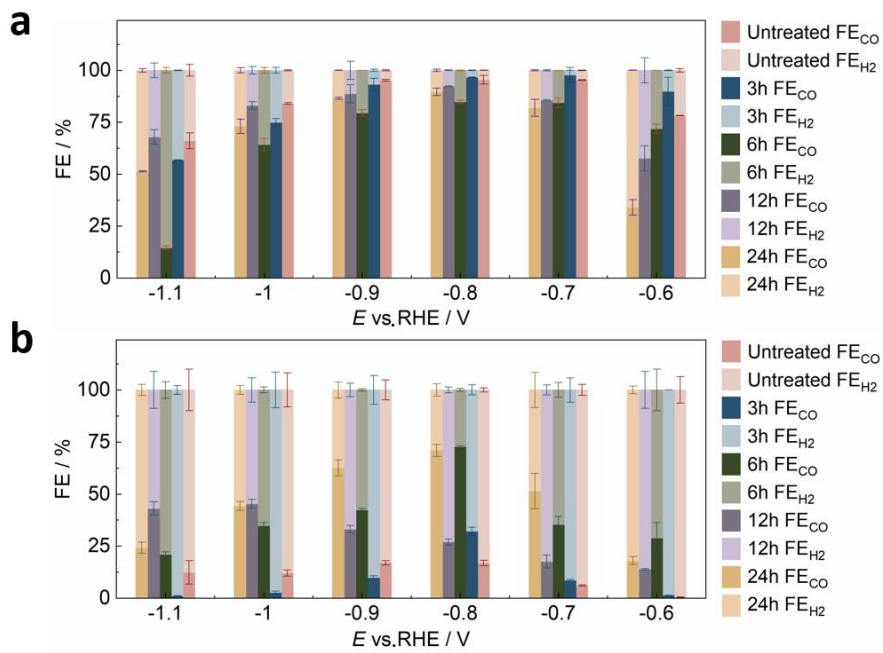
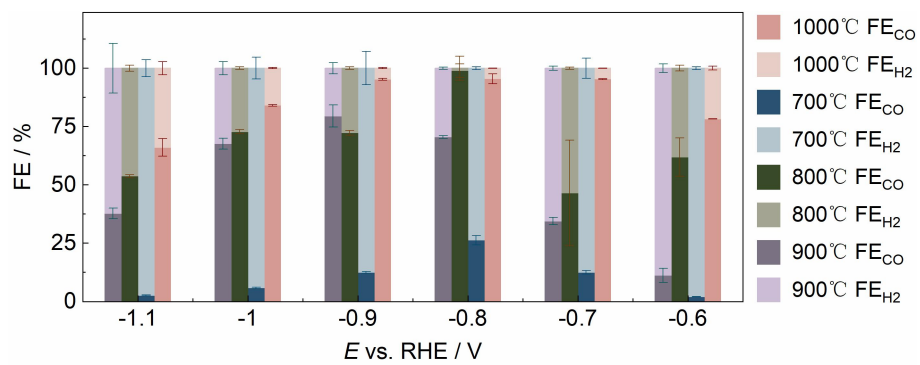


Fig. S14 AC-HAADF-STEM image on different sites and corresponding elemental mapping of NP<sub>GDE</sub>.

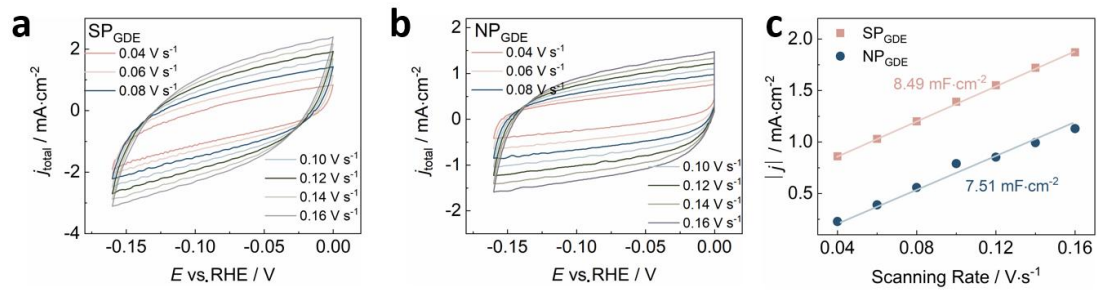


**Fig. S15** ECO<sub>2</sub>RR performance of SP<sub>GDE</sub> and NP<sub>GDE</sub> when treated with different acid (2 M H<sub>2</sub>SO<sub>4</sub>) soaking time.

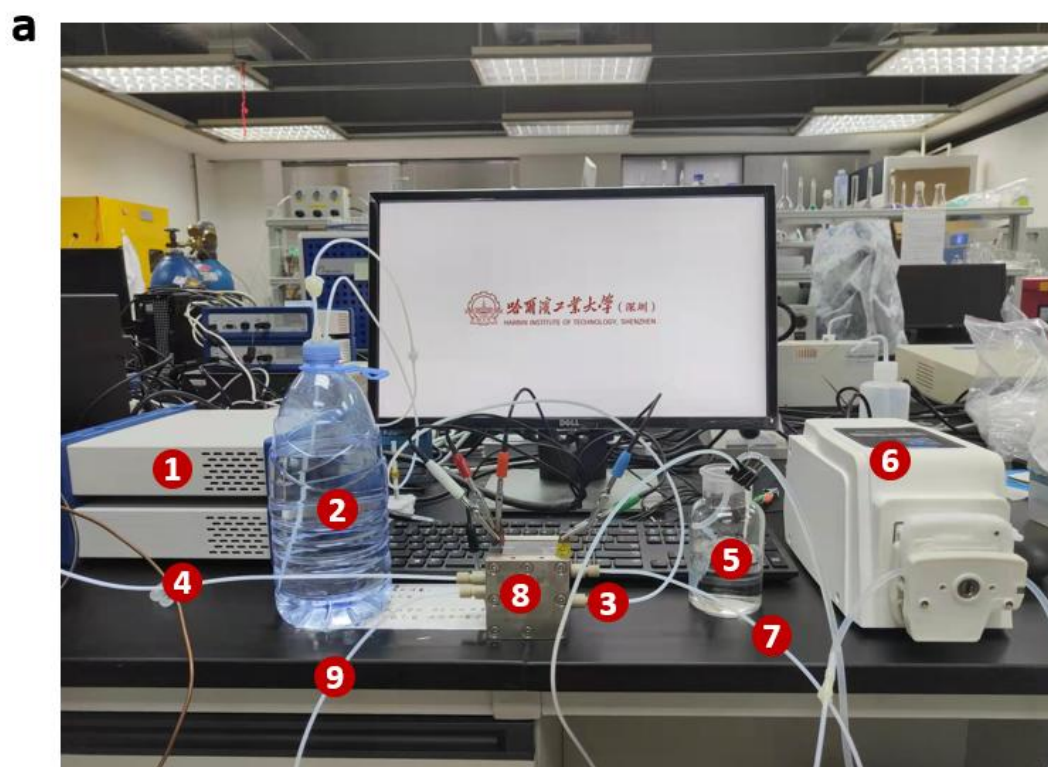
**(a)** SP<sub>GDE</sub>. **(b)** NP<sub>GDE</sub>.



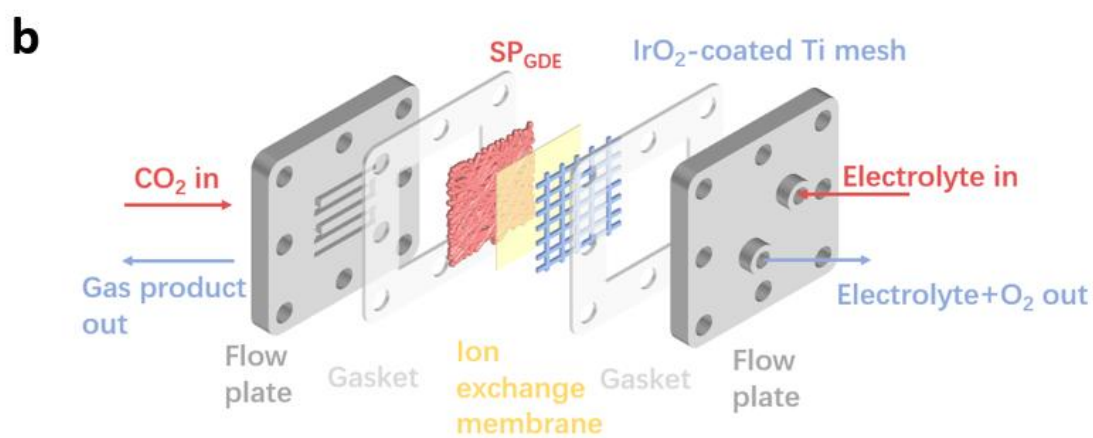
**Fig. S16** ECO<sub>2</sub>RR performance of SP<sub>GDE</sub> carbonized at 700°C, 800°C, 900°C and 1000°C.



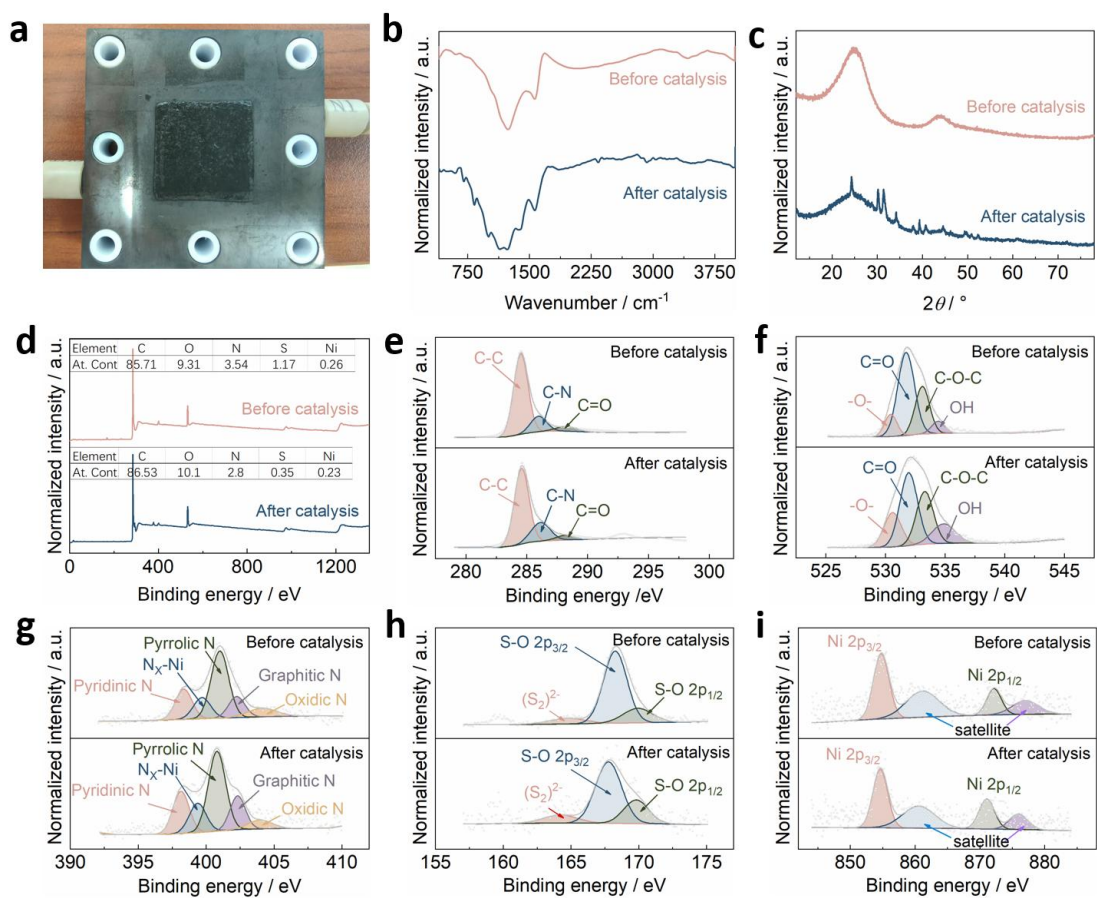
**Fig. S17** Electrochemical double layer capacitance (EDLC) measurements.



- |                           |                                     |
|---------------------------|-------------------------------------|
| 1. Potentiostat           | 6. Peristaltic pump                 |
| 2. Gas humidifier         | 7. Electrolyte in                   |
| 3. CO <sub>2</sub> gas in | 8. MEA                              |
| 4. Gas products out to GC | 9. Electrolyte + O <sub>2</sub> out |
| 5. Electrolyte tank       |                                     |



**Fig. S18** A set of facility for conducting ECO<sub>2</sub>RR in MEA. **(a)** The entire equipment, **(b)** Composition details of MEA.



**Fig. S19** SP<sub>GDE</sub> properties comparison before and after electrolysis. **(a)** Photograph of SP<sub>GDE</sub> after electrolysis. **(b)** FTIR spectra. **(c)** XRD spectra. **(d)** to **(i)** XPS spectra of overview, C 1s, O 1s, N 1s, S 2p and Ni 2p.

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