

## Supporting Information

### Text S1 Biochar preparation and characterization

After pyrolysis of carp residues in muffle furnace, the biochars were ground and filtered by a 0.17 mm sieve, then stored for characterization analysis and adsorption experiments. To prepare the demineralized biochars (DRCBs), the pristine biochars were washed in constant temperature oscillator (150 r/min, 25 °C) with 0.1 mol/L HCl. Subsequently, the DRCBs were washed with pure water when the pH of the leaching solution kept unchanging. Studies from Cui et al. (2016) have showed that the acid leaching process did not change the oxygen-containing functional groups (OFGs) on biochar. Then, the DRCBs were dried at 75 °C for 12 h, then ground and filtered by a 0.17 mm sieve. Finally, these DCRBs were labeled as DCRB450, DCRB550, and DCRB650, respectively.

The zeta potentials of CRBs were determined by a Zetasizer (SZ-100, Japan). The pH of all CRBs were obtained by adding CRBs (1 mg) to deionized water (pH 7.0, 10 mL) and shaking for 1 h at 150 r/min (Zhao et al., 2022). The specific surface area and porosity of CRBs were detected using micromeritics (ASAP2020C, USA). Total of the CRBs were determined by an elemental analyzer (Micro Cube, Elementar, Germany). Thermal analysis of CRBs was measured by STA 449 (NETZSCH, Germany). The morphology and elemental composition (C, H, N, and O) of biochars were analyzed by using JSM-7800F scanning electron microscopy (SEM) coupled with electron dispersive X-ray analysis (EDX) and thermogravimetric analyzer (STA449F5, Netzsch, Germany). The surface functional groups were detected by X-ray diffraction and X-ray photoelectron spectroscopy (XPS, ESCALAB 250Xi, USA) and Fourier transform infrared spectroscopy (FTIR) (Bruker, TENSOR 27, Germany).

### Text S2 The adsorption kinetics and isotherms of Cu on CRBs

The adsorption kinetics process of Cu<sup>2+</sup> adsorption on biochars were fitted by using the pseudo-first-order equation (Eq. (S1)), the pseudo-second-order equation (Eq. (S2)), the Elovich equation (Eq. (S3)) and the intra-particle diffusion equation (Eq. (S4)), and the equations were as follows (Tran et al., 2017; Deng et al., 2019):

$$dQ_t/dt = k_1 (Q_e - Q_t) \quad , \quad (S1)$$

$$dQ_t/dt = k_2 (Q_e - Q_t)^2 \quad , \quad (S2)$$

$$Q_t = (\ln(ab) + \ln t)/b \quad , \quad (S3)$$

$$Q_t = k_d t^{1/2} + a_i \quad , \quad (S4)$$

where  $Q_t$  (mg/g) and  $Q_e$  (mg/g) are the corresponding adsorptive quantities at time  $t$  and adsorption equilibrium, respectively.  $K_1$  (min) and  $k_2$  (g/mg/min) are the rate constant of the pseudo-first-order equation and the pseudo-second-order equation, respectively.  $K_d$  (mg/g/min<sup>1/2</sup>) is the apparent diffusion rate constants.  $a$  (mg/g/min) and  $b$  (g/mg) are the initial adsorption rate constant and a parameter of the Elovich equation, respectively.  $B$  is related to the extent of surface coverage and activation energy for chemisorption.  $a_i$  (mg/g) is a constant representing the thickness of the liquid film.

The isothermal adsorption process of Cu<sup>2+</sup> adsorption on CRBs were fitted to the Langmuir model (Eq. (S5)), the Freundlich model (Eq. (S6)), the Sips model (Eq. (S7)) and the Temkin model (Eq. (S8)), and the equations were as follows (Tran et al., 2017; Deng et al., 2019):

$$Q_e = Q_m K_L C_e / (1 + K_L C_e) \quad , \quad (S5)$$

$$Q_e = K_F C_e^{1/n} \quad , \quad (S6)$$

$$Q_e = Q_m K_S C_e^{1/n} / (1 + K_S C_e^{1/n}) \quad , \quad (S7)$$

$$Q_e = A \ln K_T C_e \quad , \quad (S8)$$

where  $C_e$  (mg/L) is the Cu<sup>2+</sup> concentration.  $Q_e$  (mg/g) is the sorption capacity of Cu<sup>2+</sup> at equilibrium.  $Q_m$  (mg/g) is the saturated sorption capacity for monolayer adsorption.  $K_L$  (L/mg),  $K_F$  (mg/g),  $K_S$  (L<sup>1/n</sup>/mg<sup>1/n</sup>) and  $K_T$  (L/mg) are the adsorption coefficient of the Langmuir, the Freundlich, Sips, and Temkin model. The  $n$  represents the dimensionless constant, which indicates the adsorption intensity.  $A$  (kJ/mol) is the coefficient associated to the heat of adsorption.

The adsorption efficiency of adsorbents can be analyzed by the separation factor ( $R_L$ ). The calculation equation is as follows (Zhao et al., 2010):

$$R_L = 1 / (1 + K_L C_0) \quad , \quad (S9)$$

where  $K_L$  is the langmuir constant.  $C_0$  is the initial Cu<sup>2+</sup> concentration.  $0 < R_L < 1$  indicates that the adsorption process is favorable,  $1 < R_L$  indicates that the adsorption process is unfavorable,  $R_L = 1$  indicates that the adsorption process is linear, and  $R_L = 0$  indicates that the adsorption process is irreversible.

**Table S1** Pyrolytic temperature selected to prepare different biowaste-derived biochars in literature.

Adsorbents	Temperature (°C)	References
silvergrass, <i>Enteromorpha prolifera</i> , and seaweed residue-derived biochar	300, 500, 700	Zhao et al., 2022
Pig carcass-derived biochar	650	Yang et al., 2022
Wheat straw biochar	300, 450, 600	Yin et al., 2017
Fish bone-based biochar	550, 700, 900	Módenes et al., 2020
Fish-scale biochar	450, 600, 750	Xia et al., 2022
Carp residues-derived biochar	450, 550, 650	This study

**Table S2** Intra-particle diffusion model parameters for Cu<sup>2+</sup> sorption on CRBs.

Sample <sup>α)</sup>	$a_1^{\beta)}$ (mg/g)	$k_{d1}$ (mg/g/min <sup>1/2</sup> )	$R^2$	$a_2$ (mg/g)	$k_{d2}$ (mg/g/min <sup>1/2</sup> )	$R^2$
CRB450	1.308	3.939	0.9375	17.37	0.639	0.9924
CRB550	4.599	7.345	0.9931	32.72	1.639	0.9876
CRB650	7.648	6.843	0.9955	29.86	2.541	0.9896

Notes: α) CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450°C, 550°C and 650°C, respectively. β)  $a_i$ : the thickness of the liquid film, mg/g.  $k_{di}$ : the rate constant of apparent diffusion rate constants, in the intra-particle diffusion equation, mg/g/min<sup>1/2</sup>.

**Table S3** Summary of the literature data on adsorption of Cu<sup>2+</sup> on different adsorbents.

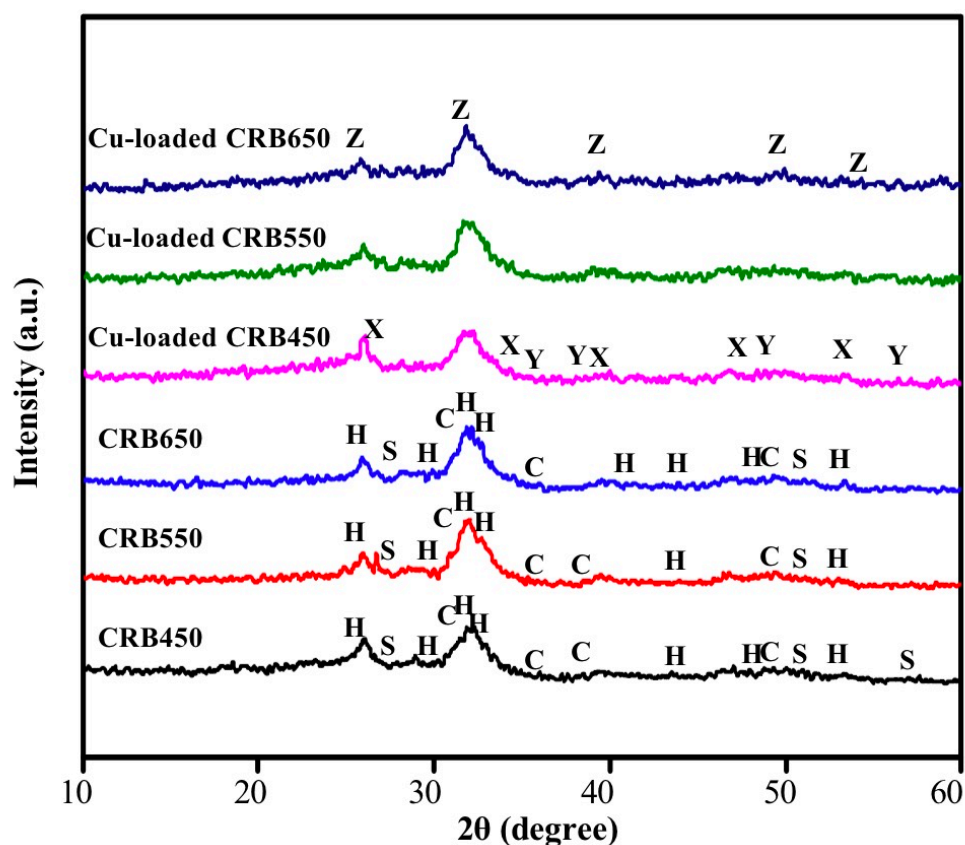
Adsorbents	$T_p^{(a)}$ (°C)	$t_p^{(b)}$ (h)	Conditions of adsorption experiments					$Q_m^{(d)}$ (mg/g)	References
			adsorbent dose (g/L)	$C_0^{(\gamma)}$ (mg/L)	pH	$T$ (°C)	$t$ (h)		
Chitosan saturated montmorillonite	NA <sup>(e)</sup>	NA <sup>(e)</sup>	0.4	2–96	5.0	25	24	34.91	Hu et al., 2017
Cellulose nanofibrils	NA <sup>(e)</sup>	NA <sup>(e)</sup>	1	1–300	NA <sup>(e)</sup>	30	24	52.32	Zhang et al., 2016
Acai								5	
Brazil nut	700	1	0.2	0–224	4.5	25	24	6	Dias et al., 2019
Palm kernel cake								6.5	
Modified cellulose	NA <sup>(e)</sup>	NA <sup>(e)</sup>	0.5	0–50	4.0	25	48	6.86	Huang et al., 2018
Date-palm waste biomass biochar	800	NA <sup>(e)</sup>	1	50–250	5.5	30	0.5	52.08	Amin et al., 2019
	300							28.60	
Wheat straw biochar	450	1	5	20–500	6.0	25	72	38.70	Yin et al., 2017
	600							44.30	
Date seed biochar	550	3	10	19–254	6.0	25	24	26.75	Mahdi et al., 2018
Cattle biochar								66.72	
Swine biochar								74.35	
Cattle manure biochar	500	4	5	0–635.5	NA <sup>(e)</sup>	25	24	49.57	Lei et al., 2019
Swine manure biochar								44.48	
Poultry manure biochar								43.21	
	450							26.95	
Carp residues-derived biochar	550	6	1	20–150	5.5	25	5	58.82	This study
	650							62.50	

Notes:  $\alpha$ )  $T_p$ : the pyrolytic temperature in the preparation of biochar, °C.  $\beta$ )  $t_p$ : the pyrolytic time, h.  $\gamma$ )  $C_0$ : the initial concentration of Cu<sup>2+</sup> in the adsorption process, mg/L.  $\delta$ )  $Q_m$ : the maximum adsorption capacity of Cu<sup>2+</sup> on biochars obtained by Langmuir model, mg/g.  $\epsilon$ ) NA: indicates that the data is not obtained.

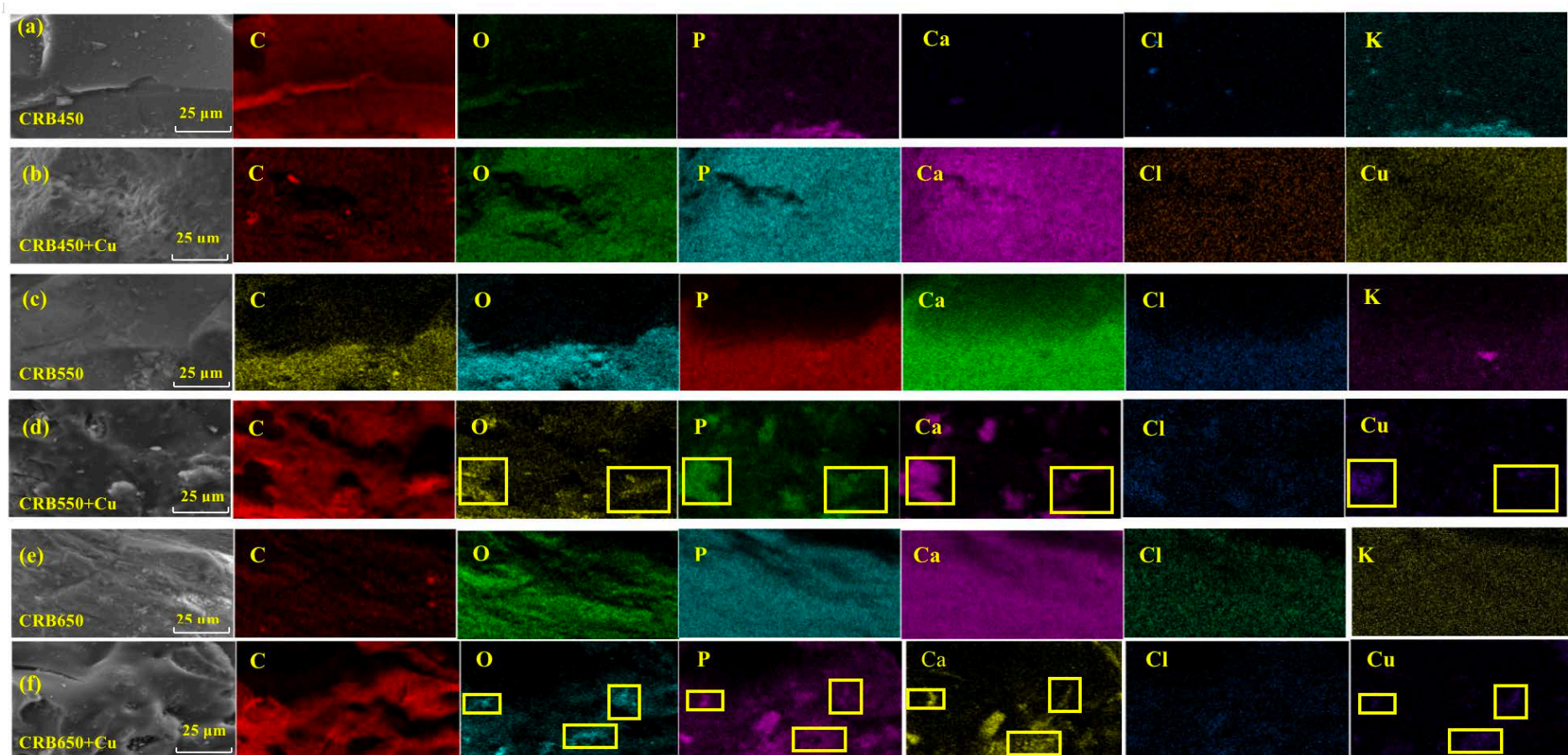
**Table S4** The atomic ratio (%) of the main elements in SEM-EDS elemental dot maps.

Element	Atomic (%)					
	CRB450 <sup>(a)</sup>	Cu-loaded CRB450	CRB550 <sup>(a)</sup>	Cu-loaded CRB550	CRB650 <sup>(a)</sup>	Cu-loaded CRB650
C	79.60	37.47	35.07	75.66	28.19	62.92
O	8.43	45.93	42.78	10.72	43.69	26.71
P	0.68	6.09	6.54	1.49	9.36	3.83
Ca	0.14	7.39	13.32	1.09	16.34	4.59
Cu	0	2.77	0	0.31	0	0.74

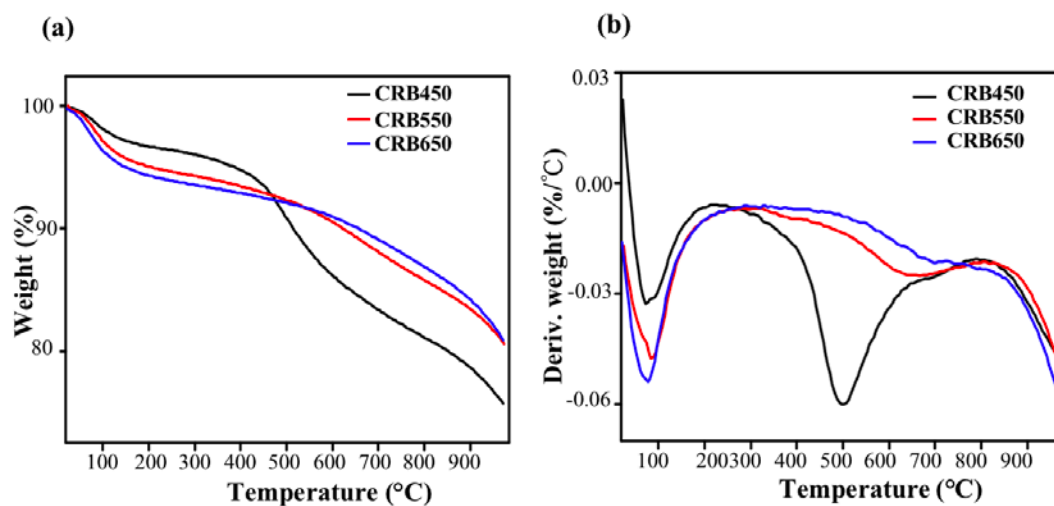
Notes: <sup>a)</sup> CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450°C, 550°C and 650°C, respectively.



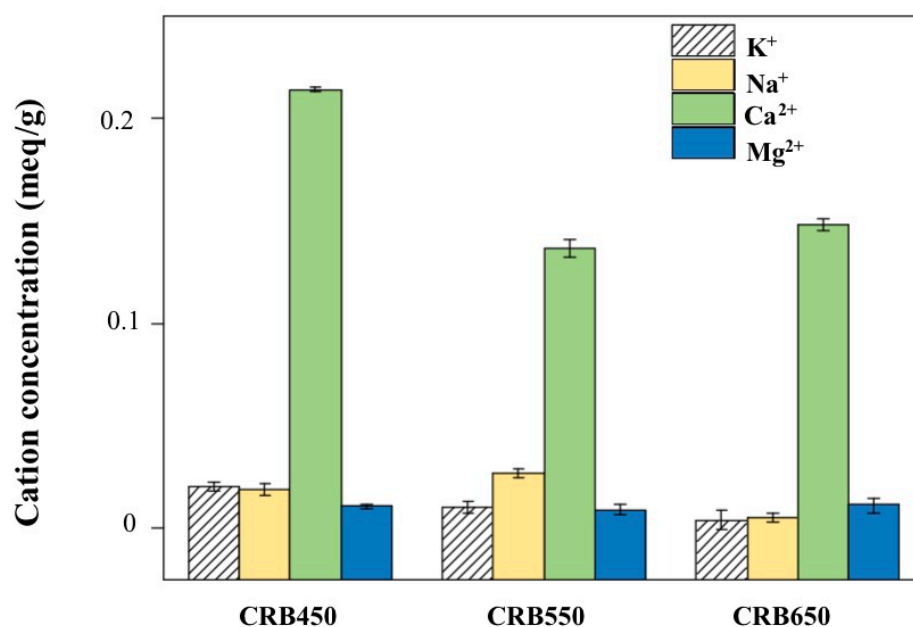
**Fig. S1** XRD patterns of biochars prepared at different temperatures before and after adsorption of Cu. CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450°C, 550°C and 650°C, respectively. The letters in the figure represent characteristic peaks corresponding to different substances, C represents the CaCO<sub>3</sub>, S represents the KCl, X corresponds to the cupric hydroxide, Y corresponds to the cupric oxide, and Z represents the Cu-phosphate precipitation. H contains the following substances: hydroxyapatite (HAP), Ca<sub>5</sub>(PO<sub>4</sub>)<sub>3</sub>X, Ca<sub>10</sub>(PO<sub>4</sub>)<sub>6</sub>X<sub>2</sub>, X=Cl<sup>-</sup>, or OH<sup>-</sup>.



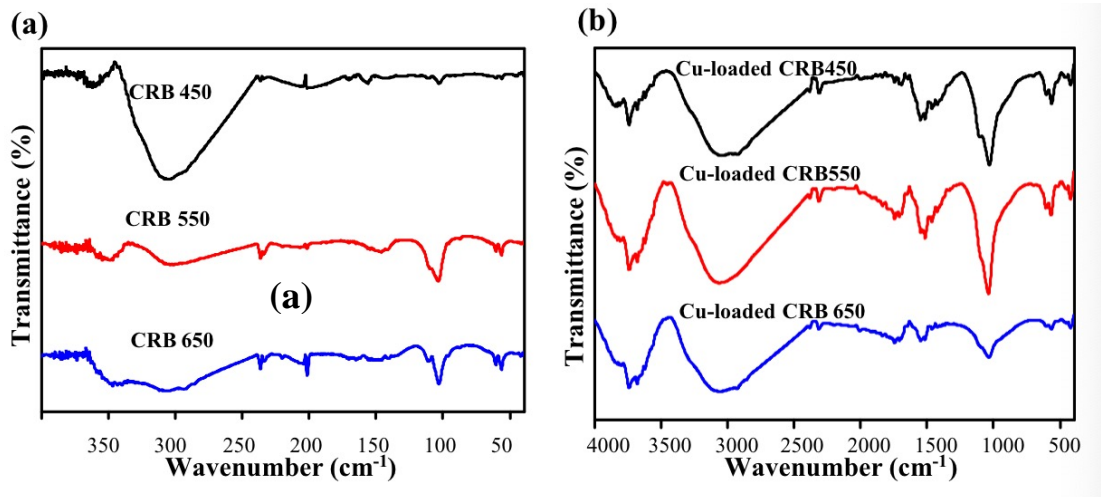
**Fig. S2** SEM images and elemental dot maps of CRB450 (a, b), CRB550 (c, d), and CRB650 (e, f) before and after  $\text{Cu}^{2+}$  adsorption. The elemental mapping of CRBs in the panel: C, O, P, Ca, Cl, and K (a, c, e), while the elemental mapping of CRBs after  $\text{Cu}^{2+}$  adsorption in the panel: C, O, P, Ca, Cl, and Cu (b, d, f). The yellow rectangle indicated that the Cu adsorption was related to the relevant functional groups or the release of cations and anions, such as the exchange of  $\text{Cu}^{2+}$  and  $\text{Ca}^{2+}$  on CRBs. CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at  $450^\circ\text{C}$ ,  $550^\circ\text{C}$  and  $650^\circ\text{C}$ , respectively.



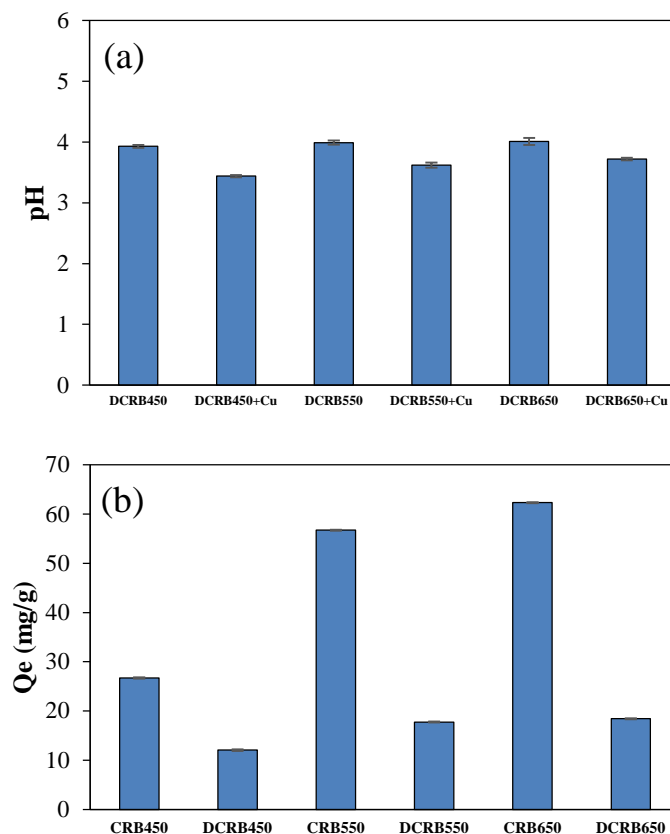
**Fig. S3** Thermogravimetric analysis (TGA) (a) and differential thermal gravimetric curves (b) of CRBs derived at 450°C–650°C. CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450°C, 550°C and 650°C, respectively.



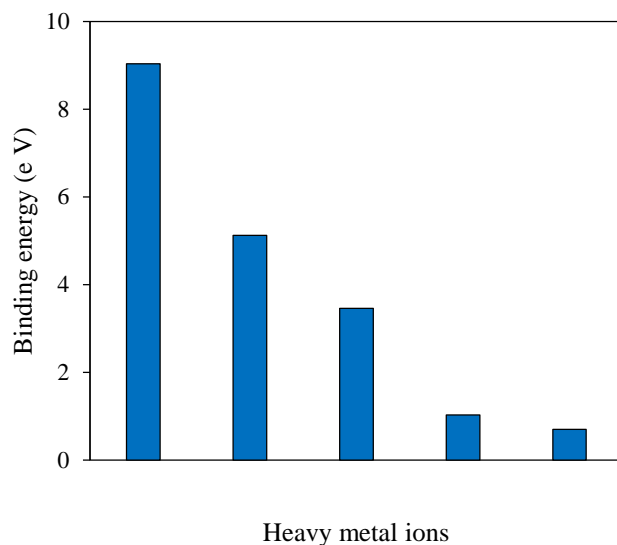
**Fig. S4** The content of released cations ( $K^+$ ,  $Ca^{2+}$ ,  $Na^+$ , and  $Mg^{2+}$ ) from CRBs. CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450°C, 550°C and 650°C, respectively. The error bars represent standard deviation ( $n=3$ ).



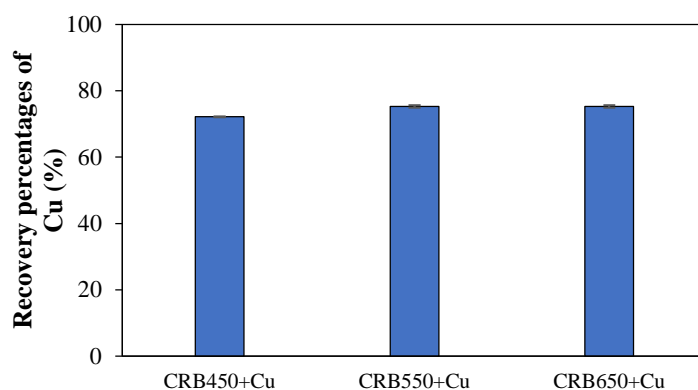
**Fig. S5** FTIR spectra of CRBs (a) and Cu-loaded CRBs (b). The bands at 2133, 1029, 563 and 468  $\text{cm}^{-1}$  were attributed to the  $\nu_1$ ,  $\nu_3$ ,  $\nu_4$  and  $\nu_2$  modes of  $\text{PO}_4^{3-}$  (Markovic et al., 2004; Lei et al., 2019), respectively. The bands at 1458 and 875  $\text{cm}^{-1}$  were attributed to the  $\nu_3$  and  $\nu_2$  modes of  $\text{CO}_3^{2-}$  (Lei et al., 2019), respectively. In addition, biochars also contained more organic functional groups, including phenolic-OH (3498  $\text{cm}^{-1}$ ), CO-OH (3034  $\text{cm}^{-1}$ ), C-H bonds in aromatic group (2916  $\text{cm}^{-1}$ ),  $\text{C}\equiv\text{C}$  (2200  $\text{cm}^{-1}$ ), N-H (2362  $\text{cm}^{-1}$ ), C=C (1649  $\text{cm}^{-1}$ ), C-O-C (1103  $\text{cm}^{-1}$ ), and C=O (1560  $\text{cm}^{-1}$ ) (Lei et al., 2019).



**Fig. S6** The pH change of the mixed solution of demineralized biochars with and without  $\text{Cu}^{2+}$  after equilibration with the initial pH of all solution was 5.0 (a). The adsorption capacity of  $\text{Cu}^{2+}$  on the pristine biochars (CRBs) and demineralized biochars (DCRBs) (b). CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450  $^{\circ}\text{C}$ , 550  $^{\circ}\text{C}$  and 650  $^{\circ}\text{C}$  respectively. The error bars represent standard deviation ( $n = 3$ ).



**Fig. S7** The binding energy between the heavy metal ions ( $K^+$ ,  $Na^+$ ,  $Ca^{2+}$ ,  $Mg^{2+}$  and  $Cu^{2+}$ ) and the benzene ring (electron-rich  $\pi$ -systems) through the G09 package.



**Fig. S8** The sequential extraction recovery percentages of  $Cu^{2+}$  from Cu-loaded biochars. CRB450, CRB550 and CRB650 were the carp residue-derived biochars produced at 450°C, 550°C and 650°C, respectively. The error bars represent standard deviation ( $n=3$ ).

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