

Emerging immunological strategies: recent advances and future directions

Hongyun Zhao², Fan Luo³, Jinhui Xue², Su Li², Rui-Hua Xu (✉)¹

¹Department of Medical Oncology, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou 510060, China; ²Department of Clinical Research, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou 510060, China; ³Department of Experimental Research, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou 510060, China

© The Author(s) 2021. This article is published with open access at link.springer.com and journal.hep.com.cn

Abstract Immunotherapy plays a compelling role in cancer treatment and has already made remarkable progress. However, many patients receiving immune checkpoint inhibitors fail to achieve clinical benefits, and the response rates vary among tumor types. New approaches that promote anti-tumor immunity have recently been developed, such as small molecules, bispecific antibodies, chimeric antigen receptor T cell products, and cancer vaccines. Small molecule drugs include agonists and inhibitors that can reach the intracellular or extracellular targets of immune cells participating in innate or adaptive immune pathways. Bispecific antibodies, which bind two different antigens or one antigen with two different epitopes, are of great interest. Chimeric antigen receptor T cell products and cancer vaccines have also been investigated. This review explores the recent progress and challenges of different forms of immunotherapy agents and provides an insight into future immunotherapeutic strategies.

Keywords cancer immunotherapy; bispecific antibodies; small molecules; chimeric antigen receptor T therapy; cancer vaccines

Introduction

Immunotherapy has brought tumor therapy into a new era. From surgery, to radiotherapy, chemotherapy, and targeted therapy, immuno-oncology therapy is almost within reach. However, many obstacles remain for this treatment. Although immune checkpoint inhibitors (ICIs) are now widely studied and have shown promising clinical data, many patients receiving ICIs fail to achieve clinical benefits, show varying response rates among different tumor types [1,2], and suffer from risk of immune-related adverse events (irAEs) [3].

New approaches that promote anti-tumor immunity have recently been developed, such as small molecules, bispecific antibodies (bsAbs), chimeric antigen receptor (CAR) T cell products, and even cancer vaccines. These new drugs can be used alone or in conjunction with

existing biological antibodies and traditional therapies (radiotherapy or chemotherapy) to affect various members of the immune system and microenvironment, promote antitumor effectiveness, and benefit many patients.

This review explores the mechanisms and recent advances of small molecule drugs, bsAbs, cancer vaccines, and CAR T cell therapy. Challenges and future directions of these novel immunotherapy strategies are also discussed.

Small molecules in immunotherapy

An overview

With deepened understanding of innate immunity and tumor microenvironment (TME), many small molecules and their importance in cancer immunity have been discovered. Small molecule drugs include agonists and inhibitors that can reach the intracellular or extracellular targets of immune cells participating in specific immune pathways, enhancing anti-tumor immunity, or reducing

immune suppression. These substances also have potential complementary or synergistic effects with existing immunotherapy. Compared with therapeutic antibodies, small molecule drugs are more permeable to tissues and the TME, and can cross the blood–brain barrier and other physiologic barriers, thus providing new options for the treatment of brain tumors and brain metastases. By adjusting the pharmacokinetic and pharmacodynamic parameters, small molecule drugs may provide the best bioavailability and avoid some of the irAEs associated with long-lasting antibody therapies. These medications also have relatively low production costs and are usually taken orally which enables easy administration.

Although a growing number of small molecules have entered early phase clinical trials, many challenges remain to be solved. Specific issues relate to understanding their

mechanisms of action in the immune system and the theoretical basis for further clinical applications, as well as, the need for more safety and efficacy evaluations.

Mechanisms of small molecule drugs

Over the past decade, more than 50 small molecule drugs have been produced as single agents or in combination with monoclonal antibodies for tumor immunotherapy [4], and over 100 clinical trials are currently underway (Table 1 and Fig. S1). Small molecule agonists and inhibitors target specific pathways participating in innate or adaptive immunity through different mechanisms (Fig. 1). Understanding the mechanism of small molecule drugs and their current clinical research progress will aid in exploring their role in immunotherapy.

Table 1 Small molecules under global clinical development as of August 2020

Small molecule	Target	Clinical studies	Phase	Cancer type
CA-170	PD-L1/VISTA	NCT02812875	Phase 1	Advanced solid tumors or lymphomas
Imiquimod	TLR7		Approved	
Motolimod	TLR7/8	NCT02431559	Phase 1/2	Ovarian cancer
		NCT03906526	Phase 1	Head and neck cancer
		NCT04272333	Early phase 1	Head and neck squamous cell carcinoma
		NCT02650635	Phase 1	Metastatic, persistent, recurrent, or progressive solid tumors
		NCT02124850	Phase 1	Head and neck squamous cell carcinoma
Resiquimod	TLR7/8	NCT00821652	Phase 1	Tumors
		NCT00948961	Phase 1/2	Advanced malignancies
		NCT00960752	Phase 2	Melanoma
		NCT01204684	Phase 2	Brain tumors
		NCT01808950	Phase 1/2	Nodular basal cell carcinoma
		NCT00470379	Phase 1	Melanoma (skin)
		NCT01748747	Phase 1	Melanoma
		NCT02126579	Phase 1/2	Melanoma
		NCT01676831	Phase 1/2	Cutaneous T cell lymphoma
VTX-2337	TLR8	NCT01666444	Phase 1/2	Epithelial ovarian cancer, fallopian tube cancer, primary peritoneal cancer
		NCT01334177	Phase 1	Locally advanced, recurrent, or metastatic squamous cell cancer of head and neck
		NCT03906526	Phase 1	Head and neck cancer
		NCT01836029	Phase 2	Head and neck squamous cell carcinoma
		NCT02124850	Phase 1	Head and neck squamous cell carcinoma
Epacadostat	IDO1	NCT03322540	Phase 2	Metastatic non-small cell lung cancer
		NCT03348904	Phase 3	Non-small cell lung cancer
		NCT03322566	Phase 2	Metastatic non-small cell lung cancer
		NCT02959437	Phase 1/2	Advanced solid tumors
		NCT03085914	Phase 1/2	Advanced or metastatic solid tumors
		NCT02318277	Phase 1/2	Advanced solid tumors
		NCT03347123	Phase 1/2	Advanced or metastatic malignancies
		NCT03006302	Phase 2	Metastatic pancreas cancer
		NCT03361865	Phase 3	Urothelial carcinoma
		NCT02327078	Phase 1/2	B cell malignancies, colorectal cancer, head and neck cancer, lung cancer, lymphoma, melanoma, ovarian cancer, glioblastoma

(Continued)

Small molecule	Target	Clinical studies	Phase	Cancer type
		NCT03374488	Phase 3	Recurrent or progressive metastatic urothelial carcinoma
		NCT03196232	Phase 2	Metastatic or unresectable gastresophageal junction or gastric cancer
		NCT03358472	Phase 3	Recurrent or metastatic head and neck squamous cell carcinoma
		NCT02364076	Phase 2	Thymic carcinoma
		NCT03493945	Phase 1/2	Metastatic prostate cancer, prostate cancer, prostate neoplasm, advanced solid tumors, solid tumor
		NCT03823131	Phase 2	Unresectable head and neck cancer
		NCT03414229	Phase 2	Advanced sarcoma
		NCT03260894	Phase 3	Renal cell carcinoma
		NCT02752074	Phase 3	Melanoma
		NCT03532295	Phase 2	Recurrent gliomas
Navoximod (GDC-0919)	IDO1	NCT02048709	Phase 1	Advanced solid tumors
		NCT02471846	Phase 1	Locally advanced or metastatic solid tumors
BMS-986205	IDO1	NCT03519256	Phase 2	Bladder cancer
		NCT03792750	Phase 1/2	Advanced malignant solid tumors
		NCT03192943	Phase 1	Advanced cancer
		NCT03661320	Phase 3	Muscle-invasive bladder cancer
		NCT02658890	Phase 1/2	Advanced cancer
		NCT04106414	Phase 2	Endometrial cancer or endometrial carcinosarcoma
		NCT03329846	Phase 3	Advanced melanoma
		NCT03854032	Phase 2	Head and neck squamous cell carcinoma
		NCT03695250	Phase 1/2	Liver cancer
		NCT04047706	Phase 1	Glioblastoma
PF-06840003	IDO1	NCT02764151	Phase 1	Malignant gliomas
CB-1158 (INCB001158)	ARG	NCT03910530	Phase 1	Advanced solid tumors
		NCT02903914	Phase 1/2	Advanced/metastatic solid tumors
		NCT03837509	Phase 1/2	Multiple myeloma
AT-38	ARG	NCT01109004	Phase 3	Multiple myeloma
CB-839	Glutaminase 1	NCT03263429	Phase 1/2	Ras wildtype colorectal cancer
		NCT02771626	Phase 1/2	Clear cell renal cell carcinoma, melanoma, non-small cell lung cancer
CPI-444 (V81444; ciforadenant)	A2A receptor	NCT02655822	Phase 1	Renal cell cancer, metastatic castration resistant prostate cancer
		NCT04280328	Phase 1	Multiple myeloma
		NCT03454451	Phase 1	Advanced cancers
Preladenant	A2A receptor	NCT03099161	Phase 1	Advanced solid tumors
PBF 509	A2A receptor	NCT02403193	Phase 1/2	Non-small cell lung cancer
AZD4635	A2A receptor	NCT04089553	Phase 2	Prostate cancer
		NCT04495179	Phase 2	Prostate cancer
		NCT03980821	Phase 1	Advanced solid malignancies
		NCT02740985	Phase 1	Advanced solid malignancies
		NCT03381274	Phase 1/2	Non-small cell lung cancer
ADU-S100	STING	NCT03937141	Phase 2	Head and neck cancer
		NCT02675439	Phase 1	Advanced/metastatic solid tumors or lymphomas
		NCT03172936	Phase 1	Solid tumors and lymphomas
MK1454	STING	NCT03010176	Phase 1	Solid tumors, lymphoma
		NCT04220866	Phase 2	Head and neck squamous cell carcinoma
Turalio (pexidartinib) (PLX3397)	CSF1R	NCT02777710	Phase 1	Metastatic/advanced pancreatic or colorectal cancers
		NCT02734433	Phase 1	Advanced solid tumors
		NCT01525602	Phase 1	Advanced solid tumors
		NCT02452424	Phase 1/2	Advanced melanoma and other solid tumors
		NCT01349036	Phase 2	Recurrent glioblastoma
		NCT02975700	Not applicable	Melanoma

(Continued)

Small molecule	Target	Clinical studies	Phase	Cancer type
		NCT01790503	Phase 1/2	Glioblastoma
LYC-55716	ROR γ t	NCT02929862	Phase 1/2	Advanced or metastatic cancer
		NCT03396497	Phase 1	Non-small cell lung cancer
TNO155	SHP2	NCT04000529	Phase 1	Non-small cell lung carcinoma, head and neck squamous cell carcinoma, esophageal SCC, gastrointestinal stromal tumors, colorectal cancer
		NCT03114319	Phase 1	Advanced solid tumors
RMC-4630 (SAR442720)	SHP2	NCT03989115	Phase 1/2	Solid tumor
		NCT03634982	Phase 1	Relapsed/refractory solid tumors
		NCT04418661	Phase 1	Metastatic neoplasm
JAB-3068	SHP2	NCT03518554	Phase 1	Advanced solid tumors
		NCT03565003	Phase 1/2a	Advanced solid tumors
JAB-3312	SHP2	NCT04121286	Phase 1	Advanced solid tumors
		NCT04045496	Phase 1	Advanced solid tumors
Idelalisib	PI3K- δ		Approved	
IPI-549	PI3K- γ	NCT03961698	Phase 2	Breast cancer, renal cell carcinoma
		NCT03719326	Phase 1	Triple-negative breast cancer, ovarian cancer
		NCT02637531	Phase 1	Advanced solid tumors
		NCT03980041	Phase 2	Advanced urothelial carcinoma
		NCT03795610	Phase 2	Head and neck squamous cell carcinoma
Ibrutinib	BTK		Approved	
Plerixafor (AMD3100)	CXCR4		Approved	
SX-682	Dual CXCR1/2	NCT04599140	Phase 1/2	Metastatic colorectal cancer
		NCT04574583	Phase 1/2	Advanced solid tumors
		NCT04477343	Phase 1	Metastatic pancreatic ductal adenocarcinoma
		NCT03161431	Phase 1	Melanoma
		NCT04245397	Phase 1	Myelodysplastic syndromes
AZD5069	CXCR2	NCT03177187	Phase 1/2	Metastatic castration resistant prostate cancer
		NCT02499328	Phase 2	Advanced solid tumors, metastatic head and neck squamous cell carcinoma
		NCT02583477	Phase 1/2	Metastatic pancreatic ductal adenocarcinoma
X4P-001	CXCR4	NCT02823405	Phase 1	Melanoma
		NCT02923531	Phase 1/2	Clear cell renal cell carcinoma
		NCT02667886	Phase 1/2	Clear cell renal cell carcinoma
Maraviroc	CCR5	NCT01785810	Phase 2	Metastatic colorectal cancer
		NCT03274804	Phase 1	Colorectal cancer
BMS-813160	Dual CCR2/5	NCT03184870	Phase 1/2	Colorectal cancer, pancreatic cancer
		NCT04123379	Phase 2	Non-small cell lung cancer, hepatocellular carcinoma
		NCT02996110	Phase 2	Advanced renal cell carcinoma
		NCT03767582	Phase 1/2	Locally advanced pancreatic ductal adenocarcinomas
		NCT03496662	Phase 1/2	Pancreatic ductal adenocarcinoma
FLX-475	CCR4	NCT03674567	Phase 1/2	Advanced cancer

Data were collected from ClinicalTrials.gov. Abbreviations: PD-L1, programmed death protein-ligand 1; VISTA, V-domain Ig suppressor of T cell activation; NCT, clinicaltrials.gov identification number; TLR, toll-like receptor; IDO1, indoleamine-2,3-dioxygenase-1; ARG, arginase; A2A, Adora2a; STING, stimulator of interferon genes; CSF1R, colony stimulating factor 1 receptor; ROR γ t, receptor-related orphan receptor gamma t; SHP2, Src homology-2-containing protein tyrosine phosphatase 2; PI3K, phosphoinositide-3 kinase; BTK, Brutons tyrosine kinase; CXCR, C-X-C chemokine receptor; CCR, C-C chemokine receptor.

Targeting immune checkpoints

Programmed death protein 1 (PD-1) or programmed death

protein-ligand 1 (PD-L1) antibodies have a long-half life and only act on extracellular PD-1/PD-L1, that is, they cannot penetrate the tissue barrier. Therefore, the

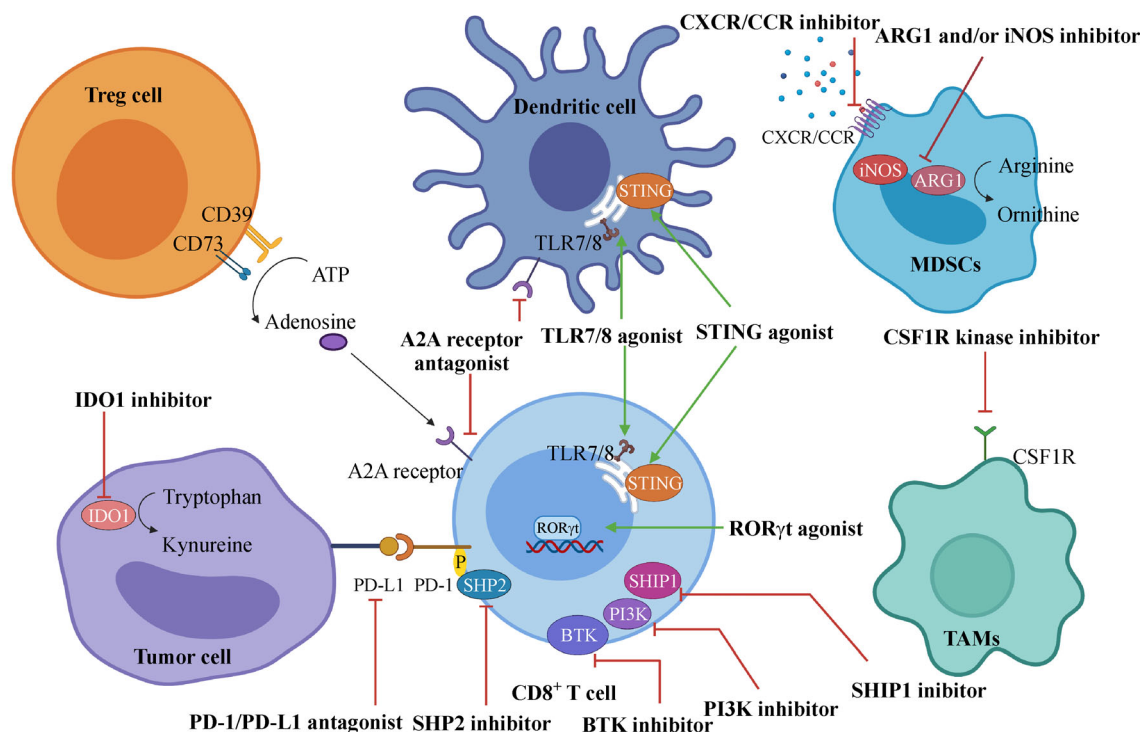


Fig. 1 Small molecule drugs and their targets in immunotherapy. This figure was created with BioRender.com. ATP, adenosine triphosphate; STING, stimulator of interferon genes; TLR, toll-like receptor; A2A, Adora2a; MDSCs, myeloid-derived suppressor cells; CXCR, C-X-C chemokine receptor; ARG1, arginase 1; IDO1, indoleamine-2,3-dioxygenase-1; PD-1, programmed death protein 1; PD-L1, programmed death protein-ligand 1; SHP2, Src homology-2-containing protein tyrosine phosphatase 2; SHP1, SH2 domain-containing inositol-5'-phosphatase 1; PI3K, phosphoinositide-3 kinase; BTK, Brutons tyrosine kinase; ROR γ t, receptor-related orphan receptor gamma t; TAMs, tumor-associated macrophages; CSF1R, colony stimulating factor 1 receptor.

occurrence of irAEs must be anticipated and monitored. The advantages of small molecules are permeabilization, oral delivery, and dose modulation, which promote the development of small molecule inhibitors acting on the PD-1/PD-L1 pathway [5–7].

Companies Bristol-Myers Squibb (BMS) and Aurigene are leading the development of small molecule PD-L1 inhibitors, with molecules such as BMS-103, BMS-142, BMS-1166, CA-327, and CA170. Small PD-L1 inhibitors developed by BMS can induce the PD-L1 dimer by filling a deep hydrophobic channel-like pocket between two PD-L1 molecules and then blocking PD-1 binding [8,9]. Oral molecule, CA-327, shows anti-tumor activity in preclinical cancer models by inhibiting the PD-L1 and T cell immunoglobulin domain and mucin domain-3 (TIM-3) [10].

Developed by Aurigene, CA170 is an oral inhibitor that targets PD-L1 and the V-domain Ig suppressor of T cell activation (VISTA), and was reported as the pioneer of oral immunotherapy drugs among small molecule checkpoint inhibitors [11]. A CA-170 phase 2 clinical study is currently ongoing with data obtained from 15 non-small cell lung cancer cases and notable tumor reductions noted

in six patients [12]. However, the affinity of small molecules to the target is worse than that of antibodies. Hence, off-targeting may occur and result in reduced efficacy and toxicity. Further mechanism explorations and clinical efficacy evaluations are needed. Although small molecule immune checkpoint inhibitors are mostly in preclinical and early clinical stages, these drugs will open a new avenue for tumor immunotherapy because of their pharmacokinetics and druggability advantages.

Targeting innate immunity

Pattern recognition receptors are key members in innate immunity that can distinguish pathogen-associated molecular patterns and promote T cell effector function [13]. Toll-like receptor (TLR) 7/8 is located in the endosome of cells. By improving the identification of foreign organisms, small molecule TLR agonists activate immune response. Imiquimod, a TLR7 agonist developed as topical cream by the Minnesota Mining & Manufacturing Company (the United States) has been used for superficial basal cell carcinoma [14]. This drug has also shown anti-tumor activity in a phase 2 clinical trial for patients with bladder

cancer [15]. Motolimod (VTX-2337), an agonist of TLR8, can mediate the release of IL-18 and activate natural killer (NK) cells [16]. Resiquimod (R848), a TLR7/8 agonist, helps macrophages acquire an anti-tumorigenic phenotype [17]. These TLR7/8 agonists are mostly in phase 1/2 clinical trials (Table 1).

Stimulator of interferon genes (STING) participate in the innate immune recognition of immunogenic tumors [18]. The activation of the STING pathway contributes to tumor regression in mouse models [19]. STING agonists might also improve the activation of dendritic cells (DCs) and T cells [20]. In June of 2019, Aduro announced the results of a phase 1b clinical trial for a small molecule STING antagonist (ADU-S100) combined with spartalizumab. However, only 6 out of the 83 patients with lymphoma or advanced solid tumors exhibited remarkable responses [21]. In hope of achieving relatively improved results, Aduro is currently preparing to combine ADU-S100 and Keytruda for head and neck cancers in a phase 2 clinical trial. The STING small molecule antagonist MK-1454 is also in a phase 2 clinical trial (NCT04220866).

In addition to antibodies for checkpoint modulation and cell therapy, pattern recognition receptor agonists and STING agonists provide a new approach for small molecules to prompt innate immune members to contribute to anti-tumor immune strategies. Although TLR agonists are promising targets that may exhibit synergistic effects with existing immunotherapy strategies, future research must consider that the TLR pathway is associated with gastric and pancreatic tumorigenesis [22,23]. Additional studies are required to further assess the safety of these small molecule agonists.

Targeting amino acid metabolism

The TME contains diverse immunocytes. Tumor-associated macrophages (TAMs) support tumor invasion and metastasis. Treg cells and myeloid-derived suppressor cells (MDSCs) are linked to immunosuppression in the TME. Small molecule drugs navigating metabolic pathways might strengthen the anti-tumor immunity by metabolic reprogramming of tumor and immune cells in the TME [24].

Indoleamine-2,3-dioxygenase-1 (IDO1) participates in the degradation of tryptophan to kynurenine, and selective inhibition of IDO1 enhances NK cell proliferation and reduces conversion to Treg cells [25]. BMS-986205 is one highly-efficient oral IDO1 inhibitor that can shrink bladder tumors when combined with ICIs from a phase 1/2a study [26]. IDO1 inhibitor navoximod has also shown acceptable safety and tolerance in a phase 1 clinical trial of advanced solid tumors, but its combination with atezolizumab was not beneficial [27]. A recent phase 3 trial, ECHO301,

tested the efficacy of IDO1 inhibitor epacadostat combined with pembrolizumab in melanoma; however, the reaction was not better than that for pembrolizumab alone [28].

Small molecule arginase 1 (ARG1) or inducible nitric oxide synthase (iNOS) inhibitors targeting MDSCs or TAMs might overcome immunosuppression and aid the restoration of immune function [29]. ARG1 inhibitor CB-1158 promotes the production of inflammatory cytokines and increases CD8⁺ T cell tumor infiltration [30]. CB-1158 is now under phase 1/2 clinical trials and is also being combined with a small molecule PD-1 blockade (Table 1). Transient treatment with CB-839, an inhibitor of glutaminase 1, also enhances cytotoxic lymphocyte-mediated anti-tumor responses [31].

Treatments targeting the amino acid metabolism of tumor and/or immune cells in the TME can produce a synergistic effect with existing immunotherapy approaches. However, the unexpected efficacy of IDO1 inhibitor epacadostat combined with pembrolizumab in clinical trial suggests that much efforts are need to further understand the metabolic mechanisms of immune cells to improve the effectiveness of combination therapies.

Targeting adenosine signaling

Ectonucleotidases CD73 and CD39 participate in the dephosphorylation of adenosine triphosphate to produce adenosine, which binds to the Adora2a (A2A) receptor, activates adenosine signaling, and amplifies the immunosuppressive effects of Treg cell [32]. In preclinical studies, the efficacy of ICIs have been enhanced using a combination of A2A receptor antagonists [33]. Preliminary evidence from a phase 1b clinical trial showed that A2A receptor inhibitor CPI-444 combined with atuzumab exhibits disease control in refractory renal cell carcinoma [34]. Other phase 1/2 studies have also assessed the safety of A2A receptor antagonists used alone or combined with ICIs in advanced tumors (Table 1). Given the immunosuppressive role of adenosine signaling in the TME, small molecule antagonists targeting A2A receptor show potential as therapeutics.

Targeting cytokine signaling

Small molecules can regulate the tumor immune response by influencing specific cytokine-mediated pathways. Retinoic acid receptor-related orphan receptor gamma t (ROR γ t) is a member of the nuclear receptor superfamily of transcription factors and plays an important role in the differentiation of cytokine interleukin-17 expressing immune cells [35]. ROR γ t agonists enhance anti-tumor immunity by activating Th17 cells and reducing Treg proliferation [36]. ROR γ t agonist, LYC-55716 in

combination with an ICI, is currently undergoing a phase 1 clinical trial (Table 1).

Galunisertib, a transforming growth factor-beta (TGF- β) receptor I inhibitor, suppresses Smad family member 2 phosphorylation and was granted orphan drug designation for the treatment of liver cancer by the European Medical Agency and the FDA in the United States in 2013 [37]. Galunisertib combined with a PD-L1 blockade can enhance the expression of immune-related genes and modulate T cell immunity in colorectal and breast cancer mouse models [38].

Although the relationship between these cytokines and immune regulation has been established, only a few of these drugs are currently undergoing clinical trials, possibly because they mediate complex signaling pathways. Their effects on tumor cells and immune cells in the TME and the risks of combination drugs must be paid attention.

Targeting oncogenic phosphatases and kinases

Phosphatases and kinases that regulate signal transduction are potential targets for small molecule drugs. Src homology-2-containing protein tyrosine phosphatase 2 (SHP2) is involved in the downstream signaling of PD-1, which suppresses T cell function [39]. Owing to its crucial role in T cell activation, SHP2 has emerged as a treatment strategy. In colon cancer xenograft models, SHP2 inhibitor SHP099 combined with an anti-PD-1 antibody showed better reducing ability for tumor load than monotherapy [40]. The SHP2 inhibitor RMC-4630s is currently under phase 1/2 clinical trials, and its pharmacokinetic profile and safety are also being evaluated (Table 1).

Colony stimulating factor 1 receptor (CSF1R) is activated by phosphorylation; pexidartinib, an oral CSF1R inhibitor, decreases TAMs and increases CD8⁺ T cells when used in combination with a dendritic cell cancer vaccine in mesothelioma mouse models [41]. Two clinical trials of pexidartinib monotherapy and two clinical trials of pexidartinib combined with monoclonal antibodies in advanced tumors are currently ongoing.

3- α -Aminocholestane, a small molecule inhibitor of lipid phosphatases SH2 domain-containing inositol-5'-phosphatase 1 (SHIP1), can strengthen the antitumor response of NK and T cells in mouse models [42]. IPI-549, a phosphoinositide-3 kinase (PI3K)- γ inhibitor, can inhibit neutrophil migration and increase the antitumor efficacy of CD8⁺ T cells [43,44]. IPI-549 used alone or in combination with ICIs is currently under investigation (Table 1). Ibrutinib, an inhibitor of Brutons tyrosine kinase (BTK), can also enhance T cell function in leukemia [45].

These small molecule drugs targeting phosphorylases and kinases usually affect tumor cell signal transduction. Additional research is needed to clarify their overall

influence on tumor and immune cells prior to clinical trials.

Targeting chemokine receptors

The chemokine superfamily consists of a large number of ligands and receptors that participate in the homing, retention, circulation, and activation of immune cells [46]. C-C chemokine receptor (CCR) 2 inhibitor PF-04136309 depletes macrophages and inflammatory monocytes from the primary lesion and premetastatic liver, thereby enhancing antitumor immunity, depressing tumor growth, and reducing metastasis [47]. Inhibiting C-X-C chemokine receptor (CXCR) 4 may also reduce the accumulation of macrophages in the TME [48]. Plerixafor, a CXCR4 antagonist, has achieved good results as a chemosensitizer in phase 1/2 leukemia clinical trials [49]. Other ongoing clinical trials of small molecule drugs targeting chemokine receptors have focused on CCR2/5 antagonist BMS-813160, CCR4 inhibitor FLX475, and CXCR2 antagonist AZD5069 (Table 1).

The small molecule targeting of chemokine receptors is often used in combination with ICIs and chemotherapeutics in clinical trials. Given the important role of chemokines in the TME, the combinational strategies may provide meaningful clinical benefits. At present, numerous small molecule drugs have been developed to target the extracellular or intracellular pathways in adaptive or innate immunity; however, most of them are in the early stage of clinical trials. Additional basic experiments and clinical trials are urgently required to clarify their mechanism, clinical efficacy, and pharmacokinetics.

Bispecific antibodies (bsAbs)

An overview

First described in the 1960s [50], bsAbs are special molecules that can bind two antigens or one antigen with different epitopes. The technological innovation of bsAbs subsequently developed in antibody engineering and biology (Fig. 2) [51]. At present, only three bsAbs are approved for global marketing: catumaxomab (CD3/EpCAM) [52], blinatumomab (CD3/CD19) [53], and emicizumab (FIXa/FX or Hemlibra) [54].

BsAbs are utilized in various ways, including receptor-activation, receptor-blocking, receptor-internalization, receptor-clustering, or retargeting of cytotoxic effector cells [55]. Cancer is a complicated and polyfactorial disease. Compared with monospecific monoclonal antibodies, bsAbs can synchronously bind two individual epitopes or antigens for greater impact and better treatment effects. Multi-combined regions in one antibody could help regulate diverse functional pathways in cancer, thus

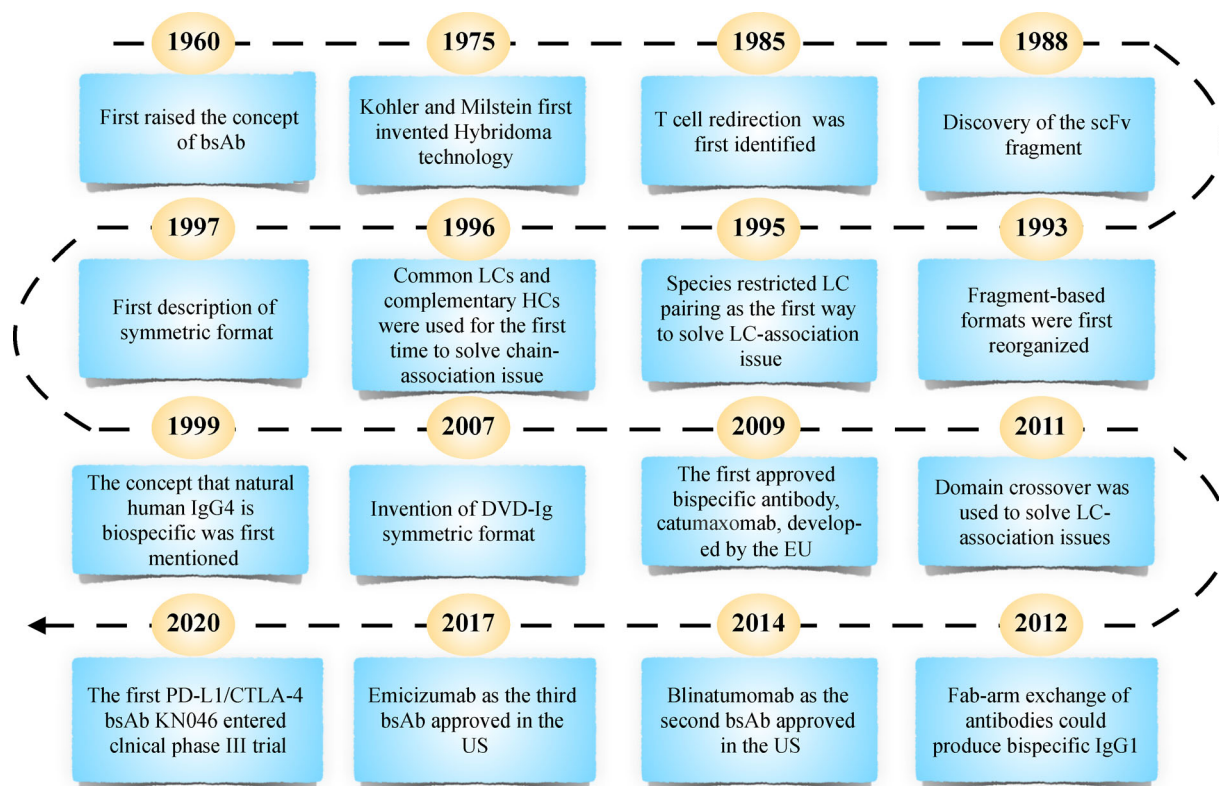


Fig. 2 Timeline of the conceptual and technical innovations contributing to the therapeutic bsAb landscape. bsAb, bispecific antibodies; DVD, dual variable domain; EU, European Union; Fab, antigen binding fragment; HC, heavy chain; Ig, immunoglobulin; LC, light chain; scFv, single-chain variable fragment.

avoiding drug resistance and decreasing the side effects on intravital tissues [56–59].

With the rapid development of gene engineering antibodies and immunology, the construction, technology platform, product research, and development of bsAbs are continuously being innovated at high speed. BsAbs are expected to be the next generation of biological therapeutics for tumors, autoimmune illness, contagious diseases, diabetes, Alzheimer’s disease, and osteoporosis [51,60]. However, several challenges have been encountered during their development, namely, how to prevent poisoning and immunogenicity due to neo-antigenic determinants, how to meet the threshold for sensitizing diverse molecular mechanisms, and how to ensure the manufacturing quality [61].

Preparation method of bsAbs

BsAbs contain two different antigen binding domains that cannot be found in nature and can only be prepared artificially. Chemical coupling [62], two-hybrid method [63], and genetic engineering [64] are the most common preparation techniques for bsAbs. The most attractive application is the realization of new biological functions

and therapeutic mechanism of action (MOA). However, new MOAs pose undiscovered risks that cannot be estimated in preclinical research. The indeterminacy over their safety is the major hurdle in the exploration of bsAbs. Molecular imaging studies could be used to create predictive models for the pharmacokinetic parts of bispecific constructs and to develop optimal dosing strategies [65].

Structure types

The basic structure of bsAbs consists of two pairs of heavy-light polypeptide chains connected by interchain disulfide and noncovalent bonds resembling a “Y” shape compound, including antigen binding fragments (Fabs) and a fragment crystallizable region (Fc). BsAbs could help immune cells target tumor cells by binding to one surface antigen expressed on cancer cells and to a second antigen expressed on immune cells, such as NK cells or effector T cells. The fusing of the antitumor binding domain with the Fc receptor (FcR) or the anti-CD3 binding domain may help produce bsAbs that can recruit immune cells. FcR is the terminal area of the antibody that interplays with the neonatal receptor, which results in

lethal immune-mediated effects [66,67].

BsAbs can be divided into two categories according to their structure: one contains the Fc region, and the other lacks the Fc region. These types can be further classified into asymmetric IgG-like bsAbs, symmetric IgG-like bsAbs, and non-IgG-like bsAbs [68]. IgG-like bsAbs can achieve effector functions, and non-IgG-like binding antibody (bAbs) are diminutive, which can improve penetration. IgG-like bAbs contain three arms/binding sites: two Fab arms and an Fc arm. The IgG-like bsAb structure promotes Fc domain-mediated effects and defends the physical properties endowed by the FcR [69,70]. A unique kind is asymmetric IgG-like bsAbs that possess an integrated Fc and a couple of distinguishing arms combining different antigens; some examples include M802, M701 [51], KN026 [71], MBS301 [72], IBI318 [73], IBI315 [74], and KN046 [75].

Symmetric IgG-like bsAbs are composed of an IgG-like Fc and a pair of symmetric arms formed by the association between different Fabs, single-chain variable fragment (scFV), and variable domain of heavy chain (VHH); these include EMB-01 [76], ES101 [77], K193 [78], AK104 [79], SI-B001 [80], and MGD013 [81]. Non-IgG-like bsAbs lack the Fc domain and exert the corresponding effect mechanism mainly through the characteristics of antigen binding; these include SHR-1701 [82], IMM0306 [83], and HX009 [84].

Mechanisms of bsAbs

BsAbs have manifold targets and special MOA.

T cell redirection

BsAbs characteristically target the antigen connected to T cells. By bonding to T cells and cancer cells, they can redirect the toxicity of effector T cells and obliterate cancer cells [85,86].

Double checkpoint inhibition

BsAbs can block PD-1 or lymphocyte-activation gene 3 (LAG3), PD-L1, TIM-3, cytotoxic T lymphocyte-associated antigen-4 (CTLA-4), and T cell immunoreceptor with immunoglobulin and ITIM domain (TIGIT) interaction, thereby activating tumor immune response [87–90]. A number of current clinical trials are targeting the above two immune checkpoints [91–94].

Co-localized blockade

SHR-1701 can simultaneously block the PD-L1 immune checkpoint and TGF- β on cancer cells. The aforementioned combination therapy could also increase the

antineoplastic effect compared with mono-treatment in cancer cell pathways [95,96].

Dual signaling inhibitions

EMB-01 (EGFR/MET) has shown promising effectiveness in numerous preclinical tests. EGFR and MET signaling paths are partly complementary and mediate the restriction of signal pathways [97–99]. SI-B001 (EGFR/HER3) activates the downstream pathways and inhibits tumorigenesis [100,101].

Tumor targeted immune-modulators

Tumor-targeted immune-modulators are intended to be combined with tumor-associated antigen (TAA) and immune-regulating receptors (PD-1/CD47) to improve immune-treatment by orientating cancer cells. Such modulators include IBI315 (HER2/PD-1) [74] and IMM0306 (CD47/CD20) [102].

Biparatopic bsAbs (bpAbs)

BpAbs combine two non-overlapping sites of identical antigen to cement Ab-Ag reciprocity and enhance the cancer cellular targeting of monoclonal antibodies [103]; these include KN026 (HER2/HER2) [104] and MBS301 (HER2/HER2) [105].

Research status

Many multinational pharmaceutical companies and biotechnology companies have committed to developing bsAb-related drugs. Many Chinese companies are also involved in the research and development of bsAbs, some of which have entered the clinical or clinical application stage.

More than 100 bsAb constructions and 200 clinical trials and over 30 technology research platforms, including CrossMab (Roche), CRIBTM (China), ItabTMv (China), and FIT-IgTM (China), have been conducted over the past decade [61]. Despite starting later than other countries, Chinese bsAb development has rapidly progressed. By using the aforementioned bsAb technology research platforms, China has created 18 bsAb structures and initiated 25 clinical trials (Table 2). As of August 2020, PD-L1 and CTLA-4 are the most commonly studied targets in China [106]. In particular, 10 bsAb and 41 clinical trials were noted for both China and other countries (Table 3) [106]. 90 bsAb structures and 149 clinical trials are currently being studied outside of China (Table 4).

AK104 (PD-1/CTLA-4) is under a recent phase 2 multicenter study on advanced gastric adenocarcinoma. The common targeted cancer bsAb simultaneously blocks the

Table 2 BsAbs under clinical development in China as of August 2020

Antibody name	Targets	Clinical studies	Phase	Cancer type
MBS-301	HER2 × HER2	NCT03842085	Phase 1	Her2 positive recurrent or metastatic malignant solid tumor
IBI-318	PD-1 × PD-L1	NCT03875157	Phase 1	Advanced malignancy
IBI-322	PD-L1 × CD47	NCT04338659	Phase 1	Advanced malignancies
IBI-315	PD-1 × HER2	NCT04162327	Phase 1	Advanced solid tumor
A-319	CD3 × CD19	NCT04056975	Phase 1	Relapsed or refractory B cell lymphoma
		CTR20190205	Phase 1	Relapsed or refractory B cell lymphoma
M701	CD3 × EpCAM	NCT04501744	Phase 1	Malignant ascites
M802	HER2 × CD3	NCT04501770	Phase 1	Her2 positive advanced solid tumor
IMM0306	CD47 × CD20	CTR20192612	Phase 1	Refractory or recurrent CD20 positive B cell non-Hodgkin's lymphoma
KN-026	HER2 × HER2	CTR20190853	Phase 2	Her2 positive advanced solid tumor
EMB-01	EGFR × c-MET	CTR20190241	Phase 2	Advanced or metastatic solid tumors
KN-046	PD-L1 × CTLA-4	NCT04469725	Phase 2	Thymic carcinoma
		NCT04474119	Phase 3	Non-small cell lung cancer
		NCT04521179	Phase 2	Her2 positive solid tumors
AK-104	PD-1 × CTLA-4	CTR20182027	Phase 1/2	Advanced solid tumor and advanced or metastatic gastric adenocarcinoma or gastroesophageal junction adenocarcinoma
		CTR20200779	Phase 2	Hepatocellular carcinoma
		CTR20202184	Phase 2	Locally advanced unresectable or metastatic highly unstable satellite or mismatch repair defective solid tumor, gastric carcinoma and colorectal cancer
MGD-013	PD-1 × LAG-3	NCT04009460	Phase 1	Solid tumors
		CTR20200549	Phase 2	Advanced hepatocellular carcinoma
HX-009	PD-1 × CD47	CTR20192299	Phase 1	Advanced solid tumor
M7824	PD-L1 × TGF-β	NCT04396886	Phase 2	Recurrent or metastatic carcinoma
SHR-1701	PD-L1 × TGF-β	CTR20182404	Phase 1	Advanced solid tumor
		CTR20181823	Phase 1	Advanced solid tumor
SI-B001	HER3 × EGFR	CTR20200502	Phase 1	Locally advanced or metastatic epithelial tumors
K193	CD3 × CD19	CTR20191955	Phase 1	Refractory or recurrent B cell non-Hodgkin's lymphoma

Abbreviations: HER, human epidermal growth factor receptor; NCT, clinicaltrials.gov identification number; PD-1, programmed death protein 1; PD-L1, programmed death protein-ligand 1; CTR, Clinical Trial Registry; EpCAM, epithelial cell adhesion molecule; c-MET, cellular-mesenchymal epithelial transition; CTLA-4, cytotoxic T lymphocyte-associated antigen-4; LAG-3, lymphocyte-activation gene 3; TGF, transforming growth factor; EGFR, epidermal growth factor receptor.

PD-1 and CTLA-4 immune regulatory checkpoints, resulting in the potential suppression of double checkpoints and antineoplastic activity [107]. Developed by Alphamab, KN046 (PD-L1/CTLA-4) is currently in a phase 3 trial. Some studies have recently reported treatment-related toxic side effects of anti-CTLA-4 antibody [108,109]. Compared with each parental mAbs, KN046 can improve the safety and efficacy [110].

Developed by Biokin, SI-B001 is an anti-HER3 × anti-EGFR bsAb that is currently in a phase 1 trial and could firsthand activate the downstream paths and inhibit tumorigenesis [100,101].

BsAbs have great clinical potential because of their unique characteristics that cannot be found in monoclonal antibodies. Most bsAbs are in clinical or preclinical research. Adverse reactions, such as cytokine storms, neurotoxicity, and production processing, are the main problems for this therapy. Designing a reasonable antibody

structure according to different effect mechanisms is the focus of bsAb research and development. The continued development of clinical studies and advances in upstream and downstream technology will hopefully help to solve these bsAb-related problems.

Chimeric antigen receptor (CAR) T cell therapy

CAR T cells are T cells designed to express an artificial receptor that redirects the T cell toward tumor cell antigen. CAR T cell therapy is one of the most encouraging therapeutic strategies and has remarkable clinical potential. CARs are composed of four domains including the extracellular domain, the transmembrane (TM) domain, the intracellular domain, and an activation domain. The first-generation of CARs comprise an extracellular domain

Table 3 BsAbs under clinical development in both China and other countries as of August 2020

Antibody name	Targets	Clinical studies	Phase	Cancer type
KN-026	HER2 × HER2	NCT04165993	Phase 2	Metastatic breast cancer
		NCT03847168	Phase 1	Breast cancer
		NCT04040699	Phase 1	Her2 positive solid tumors
		NCT03619681	Phase 1	Breast cancer, gastric cancer
		NCT03925974	Phase 2	Gastric, gastroesophageal junction cancer
EMB-01	EGFR × c-MET	NCT03797391	Phase 1/2	Neoplasm metastasis, non-small cell lung cancer
JNJ-61186372, JNJ-6372	EGFR × c-MET	NCT02609776	Phase 1	Non-small cell lung cancer
		NCT04077463	Phase 1	Carcinoma, non-small-cell lung
KN-046	PD-L1 × CTLA-4	NCT04040699	Phase 1	Her2 positive solid tumors
		NCT03838848	Phase 2	Advanced non-small cell lung cancer
		NCT03927495	Phase 2	Esophageal squamous cell carcinoma
		NCT03925870	Phase 2	Esophageal squamous cell carcinoma
		NCT03733951	Phase 1	Advanced solid tumors
		NCT04054531	Phase 2	Non-small cell lung cancer
		NCT03872791	Phase 1/2	Triple-negative breast cancer
AK-104	PD-1 × CTLA-4	NCT03529526	Phase 1	Advanced solid tumors
		NCT04380805	Phase 2	Recurrent or metastatic cervical cancer
		NCT04172454	Phase 1/2	Advanced solid tumors
		NCT04220307	Phase 2	Nasopharyngeal carcinoma
		NCT03261011	Phase 1	Advanced cancer
MGD-013	PD-1 × LAG-3	NCT03852251	Phase 1/2	Advanced solid tumors
		NCT04212221	Phase 1/2	Advanced hepatocellular carcinoma
		NCT03219268	Phase 1	Advanced solid tumors
		NCT04178460	Phase 1	Gastric cancer
INBRX-105-1, INBRX-105, ES-101	PD-L1 × 4-1BB	NCT04082364	Phase 2/3	Her2 positive gastric cancer, breast cancer
		NCT03809624	Phase 1	Metastatic solid tumors
		NCT04009460	Phase 1	Solid tumors
HX-009	PD-1 × CD47	NCT04097769	Phase 1	Advanced solid tumors
M7824	PD-L1 × TGF-β	NCT04246489	Phase 2	Uterine cervical neoplasms
		NCT04066491	Phase 2/3	Biliary tract cancer
		NCT04396535	Phase 2	Advanced lung non-small cell carcinoma
		NCT04220775	Phase 1/2	Recurrent head and neck squamous cell carcinoma
		NCT03631706	Phase 3	Non-small cell lung cancer
		NCT02517398	Phase 1	Solid tumors
		NCT03840915	Phase 1/2	Non-small cell lung cancer
		NCT03840902	Phase 2	Non-small cell lung cancer
		NCT03833661	Phase 2	Biliary tract cancer
		SHR-1701	PD-L1 × TGF-β	NCT03710265
NCT03774979	Phase 1			Solid tumors
NCT04282070	Phase 1			Nasopharyngeal carcinoma
NCT04324814	Phase 1			Advanced solid tumors

Abbreviations: HER, human epidermal growth factor receptor; NCT, clinicaltrials.gov identification number; EGFR, epidermal growth factor receptor; c-MET, cellular-mesenchymal epithelial transition; PD-L1, programmed death protein-ligand 1; CTLA-4, cytotoxic T lymphocyte-associated antigen-4; PD-1, programmed death protein 1; LAG-3, lymphocyte-activation gene 3; TGF, transforming growth factor.

linked to an intracellular domain without any co-stimulatory domain. However, no promising antitumor response was observed largely due to the lack of adequate activation [111]. As a solution, second- and third-generation CARs are being developed by adding one or

two co-stimulatory domains, respectively, to enhance their activity [112,113].

Second-generation autologous (patient-derived) CAR T cell therapy has changed the treatment of hematologic malignancies; four CD19-targeting CARs have achieved

Table 4 BsAbs under clinical development excluding China as of August 2020

Antibody name	Targets	Clinical studies	Phases	Cancer type
Dilpacimab, ABT-165	VEGF × DLL4	NCT01946074	Phase 1	Advanced solid tumors
		NCT03368859	Phase 2	Neoplasms
MP0250	VEGF × HGF	NCT03136653	Phase 1/2	Relapsed multiple myeloma
		NCT03418532	Phase 1/2	EGFR positive lung cancer
		NCT02194426	Phase 1/2	Neoplasms
ABL-001, NOV-1501, TR-009	VEGF × DLL4	NCT02857868	Phase 1	Neoplasms
		NCT03595917	Phase 1	Chronic myeloid leukemia, acute lymphoblastic leukemia
		NCT03106779	Phase 3	Chronic myelogenous leukemia
		NCT04216563	Phase 2	Philadelphia chromosome negative, BCR-ABL1 positive chronic myelogenous leukemia
		NCT03292783	Phase 1	Advanced solid tumors
		NCT02081378	Phase 1	Chronic myelogenous leukemia, Philadelphia chromosome-positive acute lymphoblastic leukemia
		NCT03605277	Phase 1	Neoplasms
Vanucizumab, RG-7221	ANGPT2 × VEGF	NCT03906292	Phase 2	Chronic myeloid leukemia
		NCT03578367	Phase 2	Chronic myelogenous leukemia
		NCT01688206	Phase 1	Neoplasms
		NCT02141295	Phase 2	Colorectal cancer
		NCT02665416	Phase 1	Advanced or metastatic solid tumors
BI-836880	ANGPT2 × VEGF	NCT02689505	Phase 1	Neoplasms
		NCT02674152	Phase 1	Neoplasms
		NCT03972150	Phase 1	Neoplasms
		NCT03861234	Phase 1	Neoplasms
		NCT03468426	Phase 1	Non-squamous, non-small-cell lung cancer, neoplasms
Navicixizumab, OMP-305B83	VEGF × DLL4	NCT03035253	Phase 1	Metastatic colorectal cancer
		NCT03030287	Phase 1	Ovaries cancer, fallopian tube cancer
		NCT02298387	Phase 1	Advanced solid tumor malignancies
ZW-25	HER2 × HER2	NCT04224272	Phase 2	Her2 or HR positive breast cancer
		NCT02892123	Phase 1	Her2 positive cancers
		NCT03929666	Phase 2	Her2 positive gastresophageal adenocarcinoma
		NCT04276493	Phase 1/2	Breast cancer, gastric cancer, gastresophageal junction cancer
MCLA-128	HER2 × HER3	NCT03321981	Phase 2	Metastatic breast cancer
		NCT02912949	Phase 1/2	Harboring NRG1 fusion solid tumors
BCD-147	HER2 × HER2	NCT03912441	Phase 1	Neoplasms
BI-905677	LRP5 × LRP6	NCT03604445	Phase 1	Neoplasms
MP0274	HER2 × HER2	NCT03084926	Phase 1	Neoplasms
DuoBody-PD-L1x4-1BB, GEN-1046	PD-L1 × 4-1BB	NCT03917381	Phase 1/2	Solid tumors
REGN-5678	CD28 × PSMA	NCT03972657	Phase 1/2	Metastatic castration-resistant prostate cancer
FS118 mAb2, FS-118, LAG-3/PD-L1 mAb2	PD-L1 × LAG-3	NCT03440437	Phase 1	Advanced cancer
LY-3434172	PD-1 × PD-L1	NCT03936959	Phase 1	Advanced cancer
XmAb-23104	PD-1 × ICOS	NCT03752398	Phase 1	Advanced solid tumors
ABBV-428	MSLN × CD40	NCT02955251	Phase 1	Advanced solid tumors
ADC-1015, ATOR-1015	OX40 × CTLA-4	NCT03782467	Phase 1	Solid tumor
MCLA-145	PD-L1 × 4-1BB	NCT03922204	Phase 1	Advanced solid tumor, B cell lymphoma
MEDI-5752	PD-1 × CTLA-4	NCT03530397	Phase 1	Selected advanced solid tumors
MGD-019	PD-1 × CTLA-4	NCT03761017	Phase 1	Advanced solid tumor
PRS-343	HER2 × 4-1BB	NCT03330561	Phase 1	Her2 positive solid tumor
		NCT03650348	Phase 1	Her2 positive solid tumor

(Continued)

Antibody name	Targets	Clinical studies	Phases	Cancer type
RG-7769, RO-7121661	PD-1 × TIM-3	NCT03708328	Phase 1	Solid tumors
XmAb-20717	PD-1 × CTLA-4	NCT03517488	Phase 1	Solid tumors
XmAb-22841	CTLA-4 × LAG-3	NCT03849469	Phase 1	Solid tumors
MP0310	FAP × CD40	NCT04049903	Phase 1	Advanced solid tumor
AK-112	VEGF × PD-1	NCT04047290	Phase 1	Neoplasms malignant
GEN-1042	CD40 × 4-1BB	NCT04083599	Phase 1/2	Solid tumor, non-small cell lung cancer, colorectal cancer, melanoma
AGEN-1423, GS-1423	CD73 × TGF-β	NCT03954704	Phase 1	Advanced solid tumors
Tebentafusp (IMCgp100)	gp100/HLA-A*0201 × CD3	NCT03070392	Phase 2	Uveal melanoma
		NCT02889861	Phase 2	Malignant melanoma
		NCT02535078	Phase 1/2	Malignant melanoma
		NCT01209676	Early phase 1	Melanoma, advanced tumors
		NCT02570308	Phase 1/2	Uveal melanoma
		NCT01211262	Phase 1	Malignant melanoma
OXS-1550, DT-2219	CD19 × CD22	NCT02370160	Phase 1/2	Refractory or relapsed B-lineage leukemia
		NCT00889408	Phase 1	Leukemia, lymphoma
AFM-13	CD16 × CD30	NCT02321592	Phase 2	Hodgkin lymphoma
		NCT01221571	Phase 1	Hodgkin lymphoma
		NCT03192202	Phase 1/2	T cell lymphoma
		NCT04074746	Phase 1	Recurrent anaplastic large cell lymphoma, recurrent B cell non-Hodgkin lymphoma, recurrent classic Hodgkin lymphoma
		NCT04101331	Phase 2	Peripheral T cell lymphoma
		NCT02665650	Phase 1	Hodgkin lymphoma
Odronextamab, REGN-1979	CD3 × CD20	NCT02651662	Phase 1	Lymphoma
		NCT03888105	Phase 2	B cell non-Hodgkin lymphoma
		NCT02290951	Phase 1	Non-Hodgkin lymphoma, chronic lymphocytic leukemia
IMC-C103C	MAGE-A4/HLA *A0201 × CD3	NCT03973333	Phase 1/2	Advanced solid tumors
IMCnyeso	NY-ESO-1/HLA *A0201 × CD3	NCT03515551	Phase 1/2	Advanced solid tumors
Mosunetuzumab, RG-7828	CD3 × CD20	NCT03671018	Phase 1/2	B cell non-Hodgkin lymphoma
		NCT04313608	Phase 1	B cell lymphoma
		NCT03677141	Phase 1/2	B cell non-Hodgkin lymphoma
		NCT04246086	Phase 1	Follicular lymphoma
		NCT03677154	Phase 1/2	Diffuse large B cell lymphoma
		NCT02500407	Phase 1	Lymphocytic leukemia
OXS-3550, CD161533 TriKE	CD16 × CD33	NCT03214666	Phase 1/2	Acute myelogenous leukemia, mast cell leukemia
GEN-3013	CD3 × CD20	NCT03625037	Phase 1/2	Lymphoma
MCLA-117	CD3 × CLEC12	NCT03038230	Phase 1	Acute myelogenous leukemia, acute myeloid leukemia
Flotetuzumab, MGD-006	CD3 × CD123	NCT03739606	Phase 2	Acute and chronic myelogenous leukemia
		NCT04158739	Phase 1	Recurrent or refractory acute myeloid leukemia
MGD-007	CD3 × GPA33	NCT03531632	Phase 1/2	Colorectal cancer metastatic
		NCT02248805	Phase 1	Colorectal carcinoma
REGN-4018	CD3 × MUC16	NCT03564340	Phase 1/2	Recurrent ovarian cancer, recurrent fallopian tube cancer, recurrent primary peritoneal cancer
Cibisatamab, RO-6958688, RG-7802	CD3 × CEA	NCT02650713	Phase 1	Solid tumors
		NCT02324257	Phase 1	Solid tumors
		NCT03337698	Phase 1/2	Carcinoma, non-small-cell lung
		NCT03866239	Phase 1	Colorectal cancer

(Continued)

Antibody name	Targets	Clinical studies	Phases	Cancer type
AMG-701	CD3 × BCMA	NCT03287908	Phase 1	Relapsed or refractory multiple myeloma
AMG-160	CD3 × PSMA	NCT03792841	Phase 1	Metastatic castration-resistant prostate cancer, prostate cancer
AMG-330, MT-114	CD3 × CD33	NCT02520427	Phase 1	Relapsed or refractory acute myeloid leukemia
AMG-424	CD3 × CD38	NCT03445663	Phase 1	Relapsed or refractory multiple myeloma
AMG-427	CD3 × FLT3	NCT03541369	Phase 1	Relapsed or refractory acute myeloid leukemia
AMG-562	CD3 × CD19	NCT03571828	Phase 1	Diffuse large B cell lymphoma, mantle cell lymphoma, follicular lymphoma
AMG-596	CD3 × EGFRvIII	NCT03296696	Phase 1	Glioblastoma or malignant glioma
AMG-673	CD3 × CD33	NCT03224819	Early Phase 1	Acute myeloid leukemia
AMG-757	CD3 × DLL3	NCT03319940	Phase 1	Small cell lung carcinoma
AMV-564, TandAb T564	CD3 × CD33	NCT03144245	Phase 1	Acute myeloid leukemia
		NCT04128423	Phase 1	Locally advanced or metastatic solid tumors
		NCT03516591	Phase 1	Myelodysplastic syndrome
APVO-436	CD3 × CD123	NCT03647800	Phase 1	Acute myeloid leukemia, myelodysplastic syndrome
BI-836909, AMG-420	CD3 × BCMA	NCT02514239	Phase 1	Multiple myeloma
		NCT03836053	Phase 1	Relapsed or refractory multiple myeloma
RG-6026, RO-7082859	CD3 × CD20	NCT03533283	Phase 1	Non-Hodgkin's lymphoma
		NCT03467373	Phase 1	B cell lymphoma, non-Hodgkin lymphoma
		NCT04313608	Phase 1	B cell lymphoma
		NCT04246086	Phase 1	Follicular lymphoma
		NCT03075696	Phase 1	Non-Hodgkin's lymphoma
		NCT04077723	Phase 1	Lymphoma, non-Hodgkin
EM-901, CC-93269	CD3 × BCMA	NCT03486067	Phase 1	Multiple myeloma
ERY-974	CD3 × GPC3	NCT02748837	Phase 1	Solid tumors
GBR-1302	CD3 × HER2	NCT02829372	Phase 1	Her2 positive solid tumors
		NCT03983395	Phase 1/2	Breast cancer
GBR-1342	CD3 × CD38	NCT03309111	Phase 1/2	Multiple myeloma
GEM-333	CD3 × CD33	NCT03516760	Phase 1	Acute myeloid leukemia
GEM-3PSCA, GEM3PSCA	CD3 × PSCA	NCT03927573	Phase 1	Non-small cell lung cancer, breast cancer, pancreatic cancer, urogenital cancer
IGM-2323	CD3 × CD20	NCT04082936	Phase 1	Non-Hodgkin lymphoma, follicular lymphoma, mantle cell lymphoma, marginal zone lymphoma
JNJ-67571244, JNJ-1244	CD3 × CD33	NCT03915379	Phase 1	Leukemia
JNJ-63709178, JNJ-9178	CD3 × CD123	NCT02715011	Phase 1	Leukemia
JNJ-64007957, JNJ-7957	CD3 × BCMA	NCT03145181	Phase 1	Hematological malignancies
JNJ-63898081, JNJ-8081	CD3 × PSMA	NCT03926013	Phase 1	Neoplasms
Orlotamab, MGD-009	CD3 × B7-H3	NCT03406949	Phase 1	Advanced solid tumors
		NCT02628535	Phase 1	Advanced solid tumors
Pasotuxizumab, AMG-212, (BAY2010112/MT112)	CD3 × PSMA	NCT01723475	Phase 1	Prostatic neoplasms
PF-06671008	CD3 × CDH3	NCT02659631	Phase 1	Neoplasms
PF-06863135, PF-3135	CD3 × BCMA	NCT03269136	Phase 1	Multiple myeloma
REGN-5458	CD3 × BCMA	NCT03761108	Phase 1/2	Multiple myeloma
RG-6194, BTRC-4017A	CD3 × HER2	NCT03448042	Phase 1	Solid tumors
TNB-383B	CD3 × BCMA	NCT03933735	Phase 1	Multiple myeloma
XmAb-13676, THG-338	CD3 × CD20	NCT02924402	Phase 1	B cell non-Hodgkins lymphoma, chronic lymphocytic leukemia
XmAb-14045, SQZ-622	CD3 × CD123	NCT02730312	Phase 1	Acute myelogenous leukemia, B cell acute lymphoblastic leukemia, blastic plasmacytoid dendritic cell neoplasm, chronic myeloid leukemia

(Continued)

Antibody name	Targets	Clinical studies	Phases	Cancer type
XmAb-18087, XENP-18087	CD3 × SSTR2	NCT03411915	Phase 1	Neuroendocrine tumor, gastrointestinal neoplasm
HPN-424	CD3 × PSMA	NCT03577028	Phase 1	Advanced prostate cancer
JNJ-64407564	CD3 × GPRC5D	NCT03399799	Phase 1	Hematological malignancies
		NCT04108195	Phase 1	Multiple myeloma
RG-6160 (BFCR4350A)	CD3 × FeRH5	NCT03275103	Phase 1	Multiple myeloma
NI-1701, TG-1801	CD19 × CD47	NCT03804996	Phase 1	B cell lymphoma
MCLA-158	EGFR × LGR5	NCT03526835	Phase 1	Advanced or metastatic solid tumors, colorectal cancer
ZW-49	HER2 × HER2	NCT03821233	Phase 1	Her2 positive cancers
SAR-440234	CD3 × CD123	NCT03594955	Phase 1/2	Leukemia
AFM-11	CD3 × CD19	NCT02848911	Phase 1	Leukemia
		NCT02106091	Phase 1	Relapsed or refractory B cell non-Hodgkin lymphoma
AFM-24	EGFR × CD16	NCT04259450	Phase 1/2	Advanced solid tumor
CCW-702	CD3 × PSMA	NCT04077021	Phase 1	Castration-resistant prostatic cancer
HPN-217	CD3 × BCMA	NCT04184050	Phase 1/2	Multiple myeloma in relapse
BI-905711	Cadherin-17 × TRAIL-R2	NCT04137289	Phase 1	Gastrointestinal neoplasms, cholangiocarcinoma, pancreatic neoplasms
MT110	CD3 × EpCAM	NCT00635596	Phase 1	Solid tumors

Abbreviations: VEGF, vascular endothelial growth factor; DLL4, delta like canonical notch ligand 4; NCT, clinicaltrials.gov identification number; HGF, hepatocyte growth factor; ANGPT2, angiopoietin 2; LRP, lipoprotein receptor related-protein; PSMA, prostate-specific membrane antigen; LAG-3, lymphocyte-activation gene 3; MSLN, mesothelin; TIM-3, T cell immunoglobulin domain and mucin domain-3; FAP, fibroblast activation protein; HLA, human leukocyte antigen; GPA33, glycoprotein A33; MUC16, mucin 16; BCMA, B cell maturation antigen; FLT3, FMS-liketyrosine kinase 3; EGFR, epidermal growth factor receptor; GPC3, glypican 3; PSCA, prostate stem cell antigen; CDH3, cadherin 3; SSTR2, somatostatin receptor 2; GPRC5D, G-protein coupled receptor family C group 5 member D; LGR5, leucine-rich repeat-containing G-protein coupled receptor 5; TRAIL-R2, TNF-related apoptosis-inducing ligand receptor 2; EpCAM, epithelial cell adhesion molecule.

FDA approval [114–117]. Clinical trials are also ongoing, and CAR T cells specific for CD30 (CD30-targeting CARs) have shown potential to treat Hodgkin’s lymphoma (HL) in two phase 1/2 clinical trials (NCT02690545, NCT02917083) [118]. A clinical trial of anti-CD7 universal CAR-T (U-CAR-T) cells indicated that patients with T cell lymphoma displayed robust CAR-T cell expansion (NCT04264078) [119]. In a phase 1/2 clinical study (NCT01869166), anti-EGFR CAR-T cells were found to be a feasible therapeutic strategy for EGFR-positive patients with NSCLC [120].

However, the success of CAR T cell therapy is yet to be applied clinically. Several impediments have been encountered, namely, poor availability of tumor specific antigens, immunosuppressive characteristics of the TME, and variability in manufacturing quality and high processing costs [121–123]. The use of “off-the-shelf” allogeneic CAR T cells from healthy donors could potentially overcome these issues. Allogeneic T cells are primarily derived from peripheral blood mononuclear cells, embryonic stem cells, and induced pluripotent stem cells. Allogeneic CAR T products can markedly decrease the costs owing to industrialized and scaled-up production, thereby rendering CAR T treatment immediately accessible to a large number of patients due to the batch manufacturing of cryopreserved T cells. The use of

allogeneic cells would also provide a high-quality product based on donor selection and allow for standardized dosing and re-dosing and a combination of CAR targets [122,123]. Other major issues must be addressed, including toxicities such as graft versus host disease (GVHD) and limited anti-tumor efficacy against solid tumors. Various safeguarding strategies, such as applying non- $\alpha\beta$ T cells including $\gamma\delta$ T cells [124], gene editing with $\alpha\beta$ T cell receptor (TCR) deletion [125], and using virus-specific T cells [126] or donor-derived allogeneic T cells [127], are needed to improve the clinical safety of CAR T cell therapy. All these techniques have been designed to specifically reduce GVHD toxicity.

Although CAR T cell therapies have shown unsatisfactory efficacy in solid tumors, many promising methods can be applied for optimization. Improving CAR T structures [128] and combining with different treatment strategies such as chemotherapy [129], local therapy [130], checkpoint blockades [131], bsAbs [132], epigenetic modulators [133], vaccines [134], and oncolytic viruses [135] have all been explored to enhance the persistence and antitumor activity of CAR T cell therapy.

Despite the bumpy road ahead, the future of CAR T cell therapy looks promising because of the continuous evolution of advanced gene editing techniques and novel solutions. These innovations will help “off-the-shelf”

allogeneic CAR T cell therapy to be effective, safe, and perhaps even revolutionize cancer treatment.

Therapeutic cancer vaccines

Cancer vaccines trigger immune responses against tumor cells by amplifying and broadening antigen-specific T cells [136]. Tumor antigens, immune adjuvants, delivery vehicles, and formulations are the four key components of therapeutic vaccines and are vital for efficacy. Tumor antigens can be delivered in the form of genetic vaccines (DNA/RNA/viral), protein/peptide vaccines, and cell vaccines. Delivery method is also a major factor influencing vaccine efficacy [136].

Antigens for tumor vaccines include TAAs and tumor-specific antigens (TSAs). Early cancer vaccines focused on TAAs, self-antigens that have elevated levels on tumor cells but may also be expressed on normal cells. However, TAAs lacking tumor specificity increase the risk of autoimmune toxicities and have been unsuccessful in generating effective antitumor immune responses due to immune tolerance [136,137].

TSAs comprise antigens expressed by neoantigens or oncoviruses and are found exclusively in cancer cells. Neoantigen-based cancer vaccines are tumor-specific, can enhance a tumor-specific T cell response, and prevent toxicities caused by “off-target” damage. Recent development on bioinformatics technologies has enabled the systematic identification of tumor neoantigens; several promising studies have explored neoantigen cancer vaccines [138]. In a phase 1 clinical trial, Ott *et al.* reported a neoantigen vaccine that was formulated with up to 20 personalized HLA-A/B-restricted peptides and has expanded neoantigen-specific T cells in patients with melanoma (NCT01970358) [139]. After a 4-year median follow-up of neoantigen vaccine therapy, a persistent T cell response was observed in patients with melanoma [140]. Neoantigen-specific T cells from peripheral blood also show the potential to migrate into intracranial tumors in glioblastoma after surgical resection cases in a phase 1b clinical trial (NCT02287428) [141]. These initial studies suggest that neoantigen-specific cancer vaccines are safe in patients with melanoma and glioblastoma. For further understanding on their therapeutic efficacy, in-depth studies must be conducted on the function of vaccine-induced T cells and the persistence of neoantigen-specific memory T cells.

Most therapeutic cancer vaccines are in ongoing trials, and their development can possibly enhance the efficacy of immunotherapy. In a phase 1b study of a neoantigen-based peptide vaccine NEO-PV-01 in combination with a PD-1 inhibitor, epitope spreading was detected post-vaccination and correlated with improved progression free survival in

patients (NCT02897765) [142]. Compared with sunitinib monotherapy, sunitinib in combination with ilixadencel, a cell-based allogeneic off-the-shelf product, exhibited a higher overall response rate in patients with synchronous metastatic renal cell carcinoma [143]. In a clinical trial of personalized tumor lysate-pulsed DCs for patients with recurrent ovarian cancer, a vaccine plus therapy seemed to improve the overall survival compared with a low-dose cyclophosphamide and bevacizumab combination therapy [144].

Although the above preliminary findings are encouraging, numerous challenges remain to be addressed. First, further discovery of personalized neoantigen targets is required to maximize their effects. Second, delivery strategies are an important factor affecting vaccine efficacy; the effectiveness of different delivery methods varies among tumor types. Finally, when a vaccine is being combined with existing treatment approaches, the timing, sequence, and dose of combination therapy must be further explored.

Challenges and future direction

New immunotherapeutic approaches provide opportunities for further drug development and bring benefits to patients. However, challenges persist during their development. Therefore, further basic and clinical research is needed.

Assessment of combination therapy

Given that anti-tumor immunity involves various steps, rational combinations to modulate different biological steps might strengthen anti-tumor responses. Effective transformation from basic discovery to clinical application could be achieved by exploring the molecular mechanisms and optimizing the strategies and timing of combination therapy to maximize its effects. The combination of four components (anti-PD1 therapy, tumor antigen-targeting antibody, interleukin-2, and a T cell vaccine) that engage in innate and adaptive immune responses was reported to eliminate large tumors in mouse model [145]. However, most drugs are in the early stages of clinical trials with complicated combinations and pose various challenges, specifically how to maximize their synergistic effects and how to avoid combinational toxicities. For a partial solution, MORPHEUS and FRACTION platforms were designed to evaluate the safety and effectiveness of combination immunotherapies in multiple phase 1b/2 trials [146,147]. A novel Quick efficacy seeking trial (QuEST1) was also designed to assess different immunotherapy combinations in patients with prostate cancer [148]. The rational selection of the combination and dosage based on known molecular mechanisms to maximize their synergistic effects is yet to be elucidated.

Validated biomarkers

Over 3000 interventional clinical trials of immunological drugs either alone or in combination are being conducted globally [149]. Nevertheless, the clinical benefits of many novel immunotherapies cannot be determined at this stage. Strategies for the identification of valid biomarkers are essential in identifying patients who will benefit the most. Many current clinical trials include the detection of serial sampling of peripheral blood or tumor specimens for the analyses of corresponding biomarkers (such as NCT01928576 and NCT03220477). In a phase 2 study of immunotherapy combinations of motolimod and doxorubicin for ovarian cancer, statistically significant differences in the overall survival of motolimod-treated patients were observed in a subgroup of patients who experienced injection site reactions; this investigation may provide biomarkers to evaluate the efficacy of combinational immunotherapies [150]. Owing to the complex interactions required for effective treatment, the development of actionable information and identifying feasible markers that can accurately classify patients is imperative.

Autoimmune toxicities

The mechanisms of immune-related toxicities must be understood to produce the best personalized treatment approach. Small changes in the molecular structures of small molecules may lead to tremendous variations in efficacy and toxicity. Diverse challenges have emerged during the exploration of bsAbs, such as reducing toxicity and immunogenicity induced by neo-epitopes, satisfying thresholds for sensitizing various molecular pathways, and assuring the quantity and quality of bsAbs.

The application of CAR T is also not without concerns. This treatment can lead to adverse effects, such as cytokine release syndrome and on-target off-tumor toxicity. Early recognition of cytokine release syndrome and aggressive steroid administration in CAR T treatment are important [151]. Moreover, drug–drug interactions must be considered for the toxicities of combination treatments. In a phase 2 study, the combination of pembrolizumab plus oral azacitidine CC-486 was associated with an increase in treatment-related adverse events compared with the pembrolizumab plus placebo group. This phenomenon can be attributed to the intestinal and hematological toxicities noted for the oral formulation of azacitidine [152].

Improving manufacturing practices

The production of bsAbs, CAR T cells, and neoantigen-based vaccines is expensive and time consuming. In the development of biological products, optimizing the structures and workflows according to the biological

mechanisms requires special attention. Designing a reasonable antibody structure according to different effect mechanisms is the focus of current bsAb research and development.

Complete CAR T cell therapy is complex compared with autologous products; however, allogeneic CAR T products offer the advantages of industrialized production and low costs [122]. Manufacturing some biological products is also time consuming, and the production of personalized vaccines is more expensive than off-the-shelf therapeutic agents. Vaccine preparation usually takes 3–5 months at best [139]. Technological developments, such as automated flow peptide production, might help promote peptide manufacturing and decrease the production time of personalized vaccines [153]. For these emerging immunotherapy drugs, their research, development, and production time and costs must be considered.

Conclusions

Immune checkpoint therapies, such as PD-1, PD-L1, and CTLA-4 antibodies, have made considerable headway in tumor treatments for the past decade. However, only a small number of cases respond to immunotherapy and are often accompanied by adverse reactions. Therefore, new treatment options are essential to enhance immunotherapy efficacy, overcome immunosuppression, and reduce toxicity. Understanding novel immuno-oncology therapeutic strategies allow us to provide additional opportunities for patients with advanced cancer. Small molecule drugs, bsAbs, CAR-T treatment, and cancer vaccines provide appealing avenues for immunotherapy. Related preliminary preclinical and clinical studies are already underway. Cost of treatment, lack of biomarker responses, and combination therapies targeting different immune mechanisms remain as challenges to be overcome. Nevertheless, these emerging strategies can bring about new opportunities for patients with cancer.

Compliance with ethics guidelines

Hongyun Zhao, Fan Luo, Jinhui Xue, Su Li, and Rui-Hua Xu declare that they have no conflicts of interest. This manuscript is a review article and does not involve a research protocol requiring approval from relevant institutional review board or ethics committee.

Electronic Supplementary Material Supplementary material is available in the online version of this article at <https://doi.org/10.1007/s11684-021-0886-x> and is accessible for authorized users.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format,

as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder.

To view a copy of this license, visit <https://creativecommons.org/licenses/by/4.0/>.

References

- Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD, Carvajal RD, Sosman JA, Atkins MB, Leming PD, Spigel DR, Antonia SJ, Horn L, Drake CG, Pardoll DM, Chen L, Sharfman WH, Anders RA, Taube JM, McMiller TL, Xu H, Korman AJ, Jure-Kunkel M, Agrawal S, McDonald D, Kollia GD, Gupta A, Wigginton JM, Sznol M. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med* 2012; 366(26): 2443–2454
- Skoulidis F, Goldberg ME, Greenawalt DM, Hellmann MD, Awad MM, Gainor JF, Schrock AB, Hartmaier RJ, Trabucco SE, Gay L, Ali SM, Elvin JA, Singal G, Ross JS, Fabrizio D, Szabo PM, Chang H, Saxon A, Srinivasan S, Kirov S, Szustakowski J, Vitazka P, Edwards R, Bufill JA, Sharma N, Ou SI, Peled N, Spigel DR, Rizvi H, Aguilar EJ, Carter BW, Erasmus J, Halpenny DF, Plodkowski AJ, Long NM, Nishino M, Denning WL, Galan-Cobo A, Hamdi H, Hirz T, Tong P, Wang J, Rodriguez-Canales J, Villalobos PA, Parra ER, Kalhor N, Sholl LM, Sauter JL, Jungbluth AA, Mino-Kenudson M, Azimi R, Elamin YY, Zhang J, Leonardi GC, Jiang F, Wong KK, Lee JJ, Papadimitrakopoulou VA, Wistuba II, Miller VA, Frampton GM, Wolchok JD, Shaw AT, Jänne PA, Stephens PJ, Rudin CM, Geese WJ, Albacker LA, Heymach JV. STK11/LKB1 mutations and PD-1 inhibitor resistance in *KRAS*-mutant lung adenocarcinoma. *Cancer Discov* 2018; 8(7): 822–835
- Postow MA, Sidlow R, Hellmann MD. Immune-related adverse events associated with immune checkpoint blockade. *N Engl J Med* 2018; 378(2): 158–168
- van der Zanden SY, Luimstra JJ, Neeffes J, Borst J, Ovaas H. Opportunities for small molecules in cancer immunotherapy. *Trends Immunol* 2020; 41(6): 493–511
- Skalniak L, Zak KM, Guzik K, Magiera K, Musielak B, Pachota M, Szelazek B, Kocik J, Grudnik P, Tomala M, Krzanik S, Pyrc K, Dömling A, Dubin G, Holak TA. Small-molecule inhibitors of PD-1/PD-L1 immune checkpoint alleviate the PD-L1-induced exhaustion of T-cells. *Oncotarget* 2017; 8(42): 72167–72181
- Ganesan A, Ahmed M, Okoye I, Arutyunova E, Babu D, Turnbull WL, Kundu JK, Shields J, Agopsowicz KC, Xu L, Tabana Y, Srivastava N, Zhang G, Moon TC, Belovodskiy A, Hena M, Kandadai AS, Hosseini SN, Hitt M, Walker J, Smylie M, West FG, Siraki AG, Lemieux MJ, Elahi S, Nieman JA, Tyrrell DL, Houghton M, Barakat K. Comprehensive *in vitro* characterization of PD-L1 small molecule inhibitors. *Sci Rep* 2019; 9(1): 12392
- Chen FF, Li Z, Ma D, Yu Q. Small-molecule PD-L1 inhibitor BMS1166 abrogates the function of PD-L1 by blocking its ER export. *OncImmunity* 2020; 9(1): 1831153
- Guzik K, Zak KM, Grudnik P, Magiera K, Musielak B, Törner R, Skalniak L, Dömling A, Dubin G, Holak TA. Small-molecule inhibitors of the programmed cell death-1/programmed death-ligand 1 (PD-1/PD-L1) interaction via transiently induced protein states and dimerization of PD-L1. *J Med Chem* 2017; 60(13): 5857–5867
- Zak KM, Grudnik P, Guzik K, Zieba BJ, Musielak B, Dömling A, Dubin G, Holak TA. Structural basis for small molecule targeting of the programmed death ligand 1 (PD-L1). *Oncotarget* 2016; 7(21): 30323–30335
- Sasikumar P, Sudarshan N, Ramachandra R, Gowda N, Samiulla D, Bilugudi P, Adurthi S, Mani J, Nair R, Ramachandra M. Pre-clinical efficacy in multiple syngeneic models with oral immune checkpoint antagonists targeting PD-L1 and TIM-3. *Eur J Cancer* 2016; 1(69): S98
- Powderly J, Patel M, Lee J, Brody J, Meric-Bernstam F, Hamilton E, Aix SP, Garcia-Corbacho J, Bang Y, Ahn M. CA-170, a first in class oral small molecule dual inhibitor of immune checkpoints PD-L1 and VISTA, demonstrates tumor growth inhibition in pre-clinical models and promotes T cell activation in phase 1 study. *Ann Oncol* 2017; 28: v405–v406
- Radhakrishnan V, Banavali S, Gupta S, Kumar A, Deshmukh C, Nag S, Beniwal S, Gopichand M, Naik R, Lakshmaiah K, Mandavia D, Ramchandra M, Prabhash K. Excellent CBR and prolonged PFS in non-squamous NSCLC with oral CA-170, an inhibitor of VISTA and PD-L1. *Ann Oncol* 2019; 30: v494
- Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat Immunol* 2010; 11(5): 373–384
- Work Group; Invited Reviewers, Kim JYS, Kozlow JH, Mittal B, Moyer J, Olencki T, Rodgers P. Guidelines of care for the management of basal cell carcinoma. *J Am Acad Dermatol* 2018; 78(3): 540–559
- Donin NM, Chamie K, Lenis AT, Pantuck AJ, Reddy M, Kivlin D, Holldack J, Pozzi R, Hakim G, Karsh LI, Lamm DL, Belkoff LH, Beldegrun AS, Holden S, Shore N. A phase 2 study of TMX-101, intravesical imiquimod, for the treatment of carcinoma *in situ* bladder cancer. *Urol Oncol* 2017; 35(2): 39.e1–39.e7
- Dietsch GN, Lu H, Yang Y, Morishima C, Chow LQ, Disis ML, Hershberg RM. Coordinated activation of Toll-like receptor 8 (TLR8) and NLRP3 by the TLR8 agonist, VTX-2337, ignites tumoricidal natural killer cell activity. *PLoS One* 2016; 11(2): e0148764
- Rodell CB, Arlauckas SP, Cuccarese MF, Garris CS, Li R, Ahmed MS, Kohler RH, Pittet MJ, Weissleder R. TLR7/8-agonist-loaded nanoparticles promote the polarization of tumour-associated macrophages to enhance cancer immunotherapy. *Nat Biomed Eng* 2018; 2(8): 578–588
- Woo SR, Fuertes MB, Corrales L, Spranger S, Furdyna MJ, Leung MY, Duggan R, Wang Y, Barber GN, Fitzgerald KA, Alegre ML, Gajewski TF. STING-dependent cytosolic DNA sensing mediates innate immune recognition of immunogenic tumors. *Immunity* 2014; 41(5): 830–842
- Corrales L, Glickman LH, McWhirter SM, Kanne DB, Sivick KE,

- Katibah GE, Woo SR, Lemmens E, Banda T, Leong JJ, Metchette K, Dubensky TW Jr, Gajewski TF. Direct activation of STING in the tumor microenvironment leads to potent and systemic tumor regression and immunity. *Cell Rep* 2015; 11(7): 1018–1030
20. Sivick KE, Desbrien AL, Glickman LH, Reiner GL, Corrales L, Surh NH, Hudson TE, Vu UT, Francica BJ, Banda T, Katibah GE, Kanne DB, Leong JJ, Metchette K, Brumbl JR, Ndubaku CO, McKenna JM, Feng Y, Zheng L, Bender SL, Cho CY, Leong ML, van Elsland A, Dubensky TW Jr, McWhirter SM. Magnitude of therapeutic STING activation determines CD8⁺ T cell-mediated anti-tumor immunity. *Cell Rep* 2018; 25(11): 3074–3085.e5
 21. Meric-Bernstam F, Sandhu S K, Hamid O, Spreafico A, Kasper S, Dummer R, Shimizu T, Steeghs N, Lewis N, Talluto C. Phase Ib study of MIW815 (ADU-S100) in combination with spartalizumab (PDR001) in patients (pts) with advanced/metastatic solid tumors or lymphomas. *J Clin Oncol* 2019; 37 (15 suppl): 2507
 22. Tye H, Kennedy CL, Najdovska M, McLeod L, McCormack W, Hughes N, Dev A, Sievert W, Ooi CH, Ishikawa TO, Oshima H, Bhathal PS, Parker AE, Oshima M, Tan P, Jenkins BJ. STAT3-driven upregulation of TLR2 promotes gastric tumorigenesis independent of tumor inflammation. *Cancer Cell* 2012; 22(4): 466–478
 23. Ochi A, Graffeo CS, Zambirinis CP, Rehman A, Hackman M, Fallon N, Barilla RM, Henning JR, Jamal M, Rao R, Greco S, Deutsch M, Medina-Zea MV, Bin Saeed U, Ego-Osuala MO, Hajdu C, Miller G. Toll-like receptor 7 regulates pancreatic carcinogenesis in mice and humans. *J Clin Invest* 2012; 122(11): 4118–4129
 24. Li X, Wenes M, Romero P, Huang SC, Fendt SM, Ho PC. Navigating metabolic pathways to enhance antitumor immunity and immunotherapy. *Nat Rev Clin Oncol* 2019; 16(7): 425–441
 25. Liu X, Shin N, Koblisch HK, Yang G, Wang Q, Wang K, Leffert L, Hansbury MJ, Thomas B, Rupal M, Waeltz P, Bowman KJ, Polam P, Sparks RB, Yue EW, Li Y, Wynn R, Fridman JS, Burn TC, Combs AP, Newton RC, Scherle PA. Selective inhibition of IDO1 effectively regulates mediators of antitumor immunity. *Blood* 2010; 115(17): 3520–3530
 26. Luke J, Tabernero J, Joshua A, Desai J, Varga A, Moreno V, Gomez-Roca C, Markman B, Braud F, Patel S, Carlino M, Siu L, Curigliano G, Liu Z, Ishii Y, Wind-Rotolo M, Basciano P, Azrilevich A, Gelmon K. BMS-986205, an indoleamine 2, 3-dioxygenase 1 inhibitor (IDO1i), in combination with nivolumab (nivo): updated safety across all tumor cohorts and efficacy in advanced bladder cancer (advBC). *J Clin Oncol* 2019; 37(7 suppl): 358
 27. Jung KH, LoRusso P, Burris H, Gordon M, Bang YJ, Hellmann MD, Cervantes A, Ochoa de Olza M, Marabelle A, Hodi FS, Ahn MJ, Emens LA, Barlesi F, Hamid O, Calvo E, McDermott D, Soliman H, Rhee I, Lin R, Pourmohamad T, Suchomel J, Tshako A, Morrissey K, Mahrus S, Morley R, Pirzkall A, Davis SL. Phase I study of the indoleamine 2,3-dioxygenase 1 (IDO1) inhibitor navoximod (GDC-0919) administered with PD-L1 inhibitor (atezolizumab) in advanced solid tumors. *Clin Cancer Res* 2019; 25(11): 3220–3228
 28. Long GV, Dummer R, Hamid O, Gajewski TF, Caglevic C, Dalle S, Arance A, Carlino MS, Grob JJ, Kim TM, Demidov L, Robert C, Larkin J, Anderson JR, Maleski J, Jones M, Diede SJ, Mitchell TC. Epacadostat plus pembrolizumab versus placebo plus pembrolizumab in patients with unresectable or metastatic melanoma (ECHO-301/KEYNOTE-252): a phase 3, randomised, double-blind study. *Lancet Oncol* 2019; 20(8): 1083–1097
 29. Adams JL, Smothers J, Srinivasan R, Hoos A. Big opportunities for small molecules in immuno-oncology. *Nat Rev Drug Discov* 2015; 14(9): 603–622
 30. Steggerda SM, Bennett MK, Chen J, Emberley E, Huang T, Janes JR, Li W, MacKinnon AL, Makkouk A, Marguier G, Murray PJ, Neou S, Pan A, Parlati F, Rodriguez MLM, Van de Velde LA, Wang T, Works M, Zhang J, Zhang W, Gross MI. Inhibition of arginase by CB-1158 blocks myeloid cell-mediated immune suppression in the tumor microenvironment. *J Immunother Cancer* 2017; 5(1): 101
 31. Johnson M O, Wolf M, Madden M Z, Andrejeva G, Sugiura A, Contreras D C, Maseda D, Liberti M V, Paz K, Kishton R J, Johnson M E, de Cubas A, Wu P, Li G, Zhang Y, Newcomb D C, Wells A D, Restifo N P, Rathmell W K, Locasale J W, Davila M L, Blazar B R, Rathmell J C. Distinct regulation of Th17 and Th1 cell differentiation by glutaminase-dependent metabolism. *Cell* 2018; 175(7): 1780–1795.e1719
 32. Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, Chen JF, Enjyoji K, Linden J, Oukka M, Kuchroo VK, Strom TB, Robson SC. Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med* 2007; 204(6): 1257–1265
 33. Beavis PA, Milenkovski N, Henderson MA, John LB, Allard B, Loi S, Kershaw MH, Stagg J, Darcy PK. Adenosine receptor 2A blockade increases the efficacy of anti-PD-1 through enhanced antitumor T-cell responses. *Cancer Immunol Res* 2015; 3(5): 506–517
 34. Emens L, Powderly J, Fong L, Brody J, Forde P, Hellmann M, Hughes B, Kummar S, Loi S, Luke J. CPI-444, an oral adenosine A2a receptor (A2aR) antagonist, demonstrates clinical activity in patients with advanced solid tumors. *Cancer Res* 2017; 77(13 suppl): CT119
 35. Ivanov II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, Cua DJ, Littman DR. The orphan nuclear receptor ROR γ t directs the differentiation program of proinflammatory IL-17⁺ T helper cells. *Cell* 2006; 126(6): 1121–1133
 36. Hu X, Liu X, Moisan J, Wang Y, Lesch CA, Spooner C, Morgan RW, Zawadzka EM, Mertz D, Bousley D, Majchrzak K, Kryczek I, Taylor C, Van Huis C, Skalizky D, Hurd A, Aicher TD, Toogood PL, Glick GD, Paulos CM, Zou W, Carter LL. Synthetic ROR γ agonists regulate multiple pathways to enhance antitumor immunity. *OncoImmunology* 2016; 5(12): e1254854
 37. Herberich S, Sawyer JS, Stauber AJ, Gueorguieva I, Driscoll KE, Estrem ST, Cleverly AL, Desai D, Guba SC, Benhadji KA, Slapak CA, Lahn MM. Clinical development of galunisertib (LY2157299 monohydrate), a small molecule inhibitor of transforming growth factor-beta signaling pathway. *Drug Des Devel Ther* 2015; 9: 4479–4499
 38. Holmgaard RB, Schaer DA, Li Y, Castaneda SP, Murphy MY, Xu X, Inigo I, Dobkin J, Manro JR, Iversen PW, Surguladze D, Hall GE, Novosiadly RD, Benhadji KA, Plowman GD, Kalos M, Driscoll KE. Targeting the TGF β pathway with galunisertib, a TGF β RI small molecule inhibitor, promotes anti-tumor immunity leading to durable, complete responses, as monotherapy and in combination with checkpoint blockade. *J Immunother Cancer*

- 2018; 6(1): 47
39. Yokosuka T, Takamatsu M, Kobayashi-Imanishi W, Hashimoto-Tane A, Azuma M, Saito T. Programmed cell death 1 forms negative costimulatory microclusters that directly inhibit T cell receptor signaling by recruiting phosphatase SHP2. *J Exp Med* 2012; 209(6): 1201–1217
 40. Zhao M, Guo W, Wu Y, Yang C, Zhong L, Deng G, Zhu Y, Liu W, Gu Y, Lu Y, Kong L, Meng X, Xu Q, Sun Y. SHP2 inhibition triggers anti-tumor immunity and synergizes with PD-1 blockade. *Acta Pharm Sin B* 2019; 9(2): 304–315
 41. Dammeyer F, Lievens LA, Kaijen-Lambers ME, van Nimwegen M, Bezemer K, Hegmans JP, van Hall T, Hendriks RW, Aerts JG. Depletion of tumor-associated macrophages with a CSF-1R kinase inhibitor enhances antitumor immunity and survival induced by DC immunotherapy. *Cancer Immunol Res* 2017; 5(7): 535–546
 42. Gumbleton M, Sudan R, Fernandes S, Engelman RW, Russo CM, Chisholm JD, Kerr WG. Dual enhancement of T and NK cell function by pulsatile inhibition of SHIP1 improves antitumor immunity and survival. *Sci Signal* 2017; 10(500): eaam5353
 43. Evans CA, Liu T, Lescarbeau A, Nair SJ, Grenier L, Pradeilles JA, Glenadel Q, Tibbitts T, Rowley AM, DiNitto JP, Brophy EE, O’Hearn EL, Ali JA, Winkler DG, Goldstein SI, O’Hearn P, Martin CM, Hoyt JG, Soglia JR, Cheung C, Pink MM, Proctor JL, Palombella VJ, Tremblay MR, Castro AC. Discovery of a selective phosphoinositide-3-kinase (PI3K)- γ inhibitor (IPI-549) as an immuno-oncology clinical candidate. *ACS Med Chem Lett* 2016; 7(9): 862–867
 44. Dwyer CJ, Arhontoulis DC, Rangel Rivera GO, Knochelmann HM, Smith AS, Wyatt MM, Rubinstein MP, Atkinson C, Thaxton JE, Neskey DM, Paulos CM. *Ex vivo* blockade of PI3K gamma or delta signaling enhances the antitumor potency of adoptively transferred CD8⁺ T cells. *Eur J Immunol* 2020; 50(9): 1386–1399
 45. Long M, Beckwith K, Do P, Mundy BL, Gordon A, Lehman AM, Maddocks KJ, Cheney C, Jones JA, Flynn JM, Andritsos LA, Awan F, Fraietta JA, June CH, Maus MV, Woyach JA, Caligiuri MA, Johnson AJ, Muthusamy N, Byrd JC. Ibrutinib treatment improves T cell number and function in CLL patients. *J Clin Invest* 2017; 127(8): 3052–3064
 46. Zlotnik A, Yoshie O. The chemokine superfamily revisited. *Immunity* 2012; 36(5): 705–716
 47. Sanford DE, Belt BA, Panni RZ, Mayer A, Deshpande AD, Carpenter D, Mitchem JB, Plambeck-Suess SM, Worley LA, Goetz BD, Wang-Gillam A, Eberlein TJ, Denardo DG, Goedegebuure SP, Linehan DC. Inflammatory monocyte mobilization decreases patient survival in pancreatic cancer: a role for targeting the CCL2/CCR2 axis. *Clin Cancer Res* 2013; 19(13): 3404–3415
 48. Chen Y, Ramjiawan RR, Reiberger T, Ng MR, Hato T, Huang Y, Ochiai H, Kitahara S, Unan EC, Reddy TP, Fan C, Huang P, Bardeesy N, Zhu AX, Jain RK, Duda DG. CXCR4 inhibition in tumor microenvironment facilitates anti-programmed death receptor-1 immunotherapy in sorafenib-treated hepatocellular carcinoma in mice. *Hepatology* 2015; 61(5): 1591–1602
 49. Uy GL, Rettig MP, Motabi IH, McFarland K, Trinkaus KM, Hladnik LM, Kulkarni S, Abboud CN, Cashen AF, Stockerl-Goldstein KE, Vij R, Westervelt P, DiPersio JF. A phase 1/2 study of chemosensitization with the CXCR4 antagonist plerixafor in relapsed or refractory acute myeloid leukemia. *Blood* 2012; 119(17): 3917–3924
 50. Nisonoff A, Wissler FC, Lipman LN. Properties of the major component of a peptic digest of rabbit antibody. *Science* 1960; 132(3441): 1770–1771
 51. Labrijn AF, Janmaat ML, Reichert JM, Parren PWHI. Bispecific antibodies: a mechanistic review of the pipeline. *Nat Rev Drug Discov* 2019; 18(8): 585–608
 52. Chelius D, Ruf P, Gruber P, Plösch M, Liedtke R, Gansberger E, Hess J, Wasiliu M, Lindhofer H. Structural and functional characterization of the trifunctional antibody catumaxomab. *MAbs* 2010; 2(3): 309–319
 53. Klinger M, Brandl C, Zugmaier G, Hijazi Y, Bargou RC, Topp MS, Gökbuget N, Neumann S, Goebeler M, Viardot A, Stelljes M, Brüggemann M, Hoelzer D, Degenhard E, Nagorsen D, Baeuerle PA, Wolf A, Kufer P. Immunopharmacologic response of patients with B-lineage acute lymphoblastic leukemia to continuous infusion of T cell-engaging CD19/CD3-bispecific BiTE antibody blinatumomab. *Blood* 2012; 119(26): 6226–6233
 54. Kitazawa T, Esaki K, Tachibana T, Ishii S, Soeda T, Muto A, Kawabe Y, Igawa T, Tsunoda H, Nogami K, Shima M, Hattori K. Factor VIIIa-mimetic cofactor activity of a bispecific antibody to factors IX/IXa and X/Xa, emicizumab, depends on its ability to bridge the antigens. *Thromb Haemost* 2017; 117(7): 1348–1357
 55. Dickopf S, Georges GJ, Brinkmann U. Format and geometries matter: structure-based design defines the functionality of bispecific antibodies. *Comput Struct Biotechnol J* 2020; 18: 1221–1227
 56. Mazor Y, Hansen A, Yang C, Chowdhury PS, Wang J, Stephens G, Wu H, Dall’Acqua WF. Insights into the molecular basis of a bispecific antibody’s target selectivity. *MAbs* 2015; 7(3): 461–469
 57. Mazor Y, Sachsenmeier KF, Yang C, Hansen A, Filderman J, Mulgrew K, Wu H, Dall’Acqua WF. Enhanced tumor-targeting selectivity by modulating bispecific antibody binding affinity and format valence. *Sci Rep* 2017; 7(1): 40098
 58. Lopez-Albaitero A, Xu H, Guo H, Wang L, Wu Z, Tran H, Chandralapaty S, Scaltriti M, Janjigian Y, de Stanchina E, Cheung NK. Overcoming resistance to HER2-targeted therapy with a novel HER2/CD3 bispecific antibody. *OncoImmunology* 2017; 6(3): e1267891
 59. Moores SL, Chiu ML, Bushey BS, Chevalier K, Luistro L, Dorn K, Brezski RJ, Haytko P, Kelly T, Wu SJ, Martin PL, Neijssen J, Parren PW, Schuurman J, Attar RM, Laquerre S, Lorenzi MV, Anderson GM. A novel bispecific antibody targeting EGFR and cMet is effective against EGFR inhibitor-resistant lung tumors. *Cancer Res* 2016; 76(13): 3942–3953
 60. Thakur A, Huang M, Lum LG. Bispecific antibody based therapeutics: strengths and challenges. *Blood Rev* 2018; 32(4): 339–347
 61. Zhang MY, Lu JJ, Wang L, Gao ZC, Hu H, Ung CO, Wang YT. Development of monoclonal antibodies in China: overview and prospects. *BioMed Res Int* 2015; 2015: 168935
 62. Staerz UD, Kanagawa O, Bevan MJ. Hybrid antibodies can target sites for attack by T cells. *Nature* 1985; 314(6012): 628–631
 63. Staerz UD, Bevan MJ. Hybrid hybridoma producing a bispecific monoclonal antibody that can focus effector T-cell activity. *Proc Natl Acad Sci USA* 1986; 83(5): 1453–1457
 64. Kontermann RE. Dual targeting strategies with bispecific anti-

- bodies. *MAbs* 2012; 4(2): 182–197
65. Nie S, Wang Z, Moscoso-Castro M, D'Souza P, Lei C, Xu J, Gu J. Biology drives the discovery of bispecific antibodies as innovative therapeutics. *Antib Ther* 2020; 3(1): 18–62
 66. Grugan KD, Dorn K, Jarantow SW, Bushey BS, Pardinias JR, Laquerre S, Moores SL, Chiu ML. Fc-mediated activity of EGFR x c-Met bispecific antibody JNJ-61186372 enhanced killing of lung cancer cells. *MAbs* 2017; 9(1): 114–126
 67. Wang Q, Chung CY, Chough S, Betenbaugh MJ. Antibody glycoengineering strategies in mammalian cells. *Biotechnol Bioeng* 2018; 115(6): 1378–1393
 68. Kontermann RE, Brinkmann U. Bispecific antibodies. *Drug Discov Today* 2015; 20(7): 838–847
 69. Gupta A, Kumar Y. Bispecific antibodies: a novel approach for targeting prominent biomarkers. *Hum Vaccin Immunother* 2020; 16(11): 2831–2839
 70. Kontermann RE, Brinkmann U. Bispecific antibodies. *Drug Discov Today* 2015; 20(7): 838–847
 71. Xu T, Tau X, Wang X, Li Q, Minjie P, Zhang H, Han L, Zhang Q. Patent US 10808043 (B2); PCT/CN2016/070447. 2020
 72. Li F, Zhang B, Ye P, Zhao J, Huang S, Jin C. Patent US 9745382 (B1); PCT/CN2017/093816, 2017
 73. Liu J, Song N, Yang Y, Jin M. Patent WO2018177324 (A1); PCT/CN2018/080858. 2018
 74. Liu J, Song N, Yang Y. Patent WO2018090950 (A1); PCT/CN2017/111310. 2018
 75. Xu T, Dong Y, Wang P. Patent US 20180291103 (A1); PCT/CN2016/092679. 2017
 76. Wu C. Patent US 10266608 (B2); PCT/US2014/072336. 2019
 77. Eckelman B, Timmer JC, Hata C, Jones KS, Hussain A, Razai AS, Becklund B, Pandit R, Kaplan M, Rason L, Deveraux Q, Eckelman BP, Timmer JC, Hata C, Jones KS, Hussain A, Razai AS, Becklund B, Pandit R, Kaplan M, Rascon L, Deveraux Q. Patent US 20170198050 (A1); PCT/US2017/013040. 2017
 78. Kong J, Ye Y, Zhou P, Huang Y, Kong Q, Yang S, Xu L, Zhang K, Zhang K, Wang S. Patent US 20190284279 (A1); PCT/CN2018/085397. 2019
 79. Li B, Xia Y, Wang ZM, Zhang P. Patent US 20190185569 (A1); PCT/CN2017/098466. 2019
 80. Gao Z, TAN P, Kovacevich B, Renshaw B, Adamo J, Mak SA, Zhuo S, Chen L. Patent WO2016106157 (A1); PCT/US2015/066951. 2015
 81. LaMotte-Mohs R, Shah K, Smith D, Gorlatov S, Ciccarone V, Tamura J, Li H, Rillema J, Licea M. MGD013, a bispecific PD-1 X LAG-3 dual affinity re-targeting (DART[®]) protein with T-cell immunomodulatory activity for cancer treatment. *Cancer Res* 2016; 76 (14 Supplement): 3217–3217
 82. Gu J, Luo X, Tao W. Patent CN 201880004344.6A; PCT/CN2018/086451. 2018
 83. Tian W, Li S. Patent WO201816650; PCT/CN2018/079187. 2018
 84. Huang Y, Zhang F, Xi G. Patent WO2019109357; PCT/CN2017/115323. 2019
 85. Hinner MJ, Aiba RSB, Wiedenmann A, Schlosser C, Allersdorfer A, Matschiner G, Rothe C, Moebius U, Kohrt HE, Olwill SA. Costimulatory T cell engagement via a novel bispecific anti-CD137/anti-HER2 protein. *J Immunother Cancer* 2015; 3(Suppl 2): 187
 86. Chames P, Baty D. Bispecific antibodies for cancer therapy. *Curr Opin Drug Discov Devel* 2009; 12(2): 276–283
 87. Poole RM. Pembrolizumab: first global approval. *Drugs* 2014; 74 (16): 1973–1981
 88. Markham A. Atezolizumab: first global approval. *Drugs* 2016; 76 (12): 1227–1232
 89. Kim ES. Avelumab: first global approval. *Drugs* 2017; 77(8): 929–937
 90. Syed YY. Durvalumab: first global approval. *Drugs* 2017; 77(12): 1369–1376
 91. Osipov A, Zaidi N, Laheru DA. Dual checkpoint inhibition in pancreatic cancer: revealing the limitations of synergy and the potential of novel combinations. *JAMA Oncol* 2019; 5(10): 1438–1439
 92. Reck M, Borghaei H, O'Byrne KJ. Nivolumab plus ipilimumab in non-small-cell lung cancer. *Future Oncol* 2019; 15(19): 2287–2302
 93. Winer A, Ghatalia P, Bubes N, Anari F, Varshavsky A, Kasireddy V, Liu Y, El-Deiry WS. Dual checkpoint inhibition with ipilimumab plus nivolumab after progression on sequential PD-1/PDL-1 inhibitors pembrolizumab and atezolizumab in a patient with Lynch syndrome, metastatic colon, and localized urothelial cancer. *Oncologist* 2019; 24(11): 1416–1419
 94. Hassel JC, Heinzerling L, Aberle J, Bähr O, Eigentler TK, Grimm MO, Grünwald V, Leipe J, Reinmuth N, Tietze JK, Trojan J, Zimmer L, Gutzmer R. Combined immune checkpoint blockade (anti-PD-1/anti-CTLA-4): evaluation and management of adverse drug reactions. *Cancer Treat Rev* 2017; 57: 36–49
 95. Topalian SL, Drake CG, Pardoll DM. Targeting the PD-1/B7-H1 (PD-L1) pathway to activate anti-tumor immunity. *Curr Opin Immunol* 2012; 24(2): 207–212
 96. Hugo W, Zaretsky JM, Sun L, Song C, Moreno BH, Hu-Lieskovan S, Berent-Maoz B, Pang J, Chmielowski B, Cherry G, Seja E, Lomeli S, Kong X, Kelley MC, Sosman JA, Johnson DB, Ribas A, Lo RS. Genomic and transcriptomic features of response to anti-PD-1 therapy in metastatic melanoma. *Cell* 2016; 165(1): 35–44
 97. Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, Lindeman N, Gale CM, Zhao X, Christensen J, Kosaka T, Holmes AJ, Rogers AM, Cappuzzo F, Mok T, Lee C, Johnson BE, Cantley LC, Jänne PA. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science* 2007; 316(5827): 1039–1043
 98. Turke AB, Zejnullahu K, Wu YL, Song Y, Dias-Santagata D, Lifshits E, Toschi L, Rogers A, Mok T, Sequist L, Lindeman NI, Murphy C, Akhavanfard S, Yeap BY, Xiao Y, Capelletti M, Iafrate AJ, Lee C, Christensen JG, Engelman JA, Jänne PA. Preexistence and clonal selection of MET amplification in EGFR mutant NSCLC. *Cancer Cell* 2010; 17(1): 77–88
 99. Yano S, Yamada T, Takeuchi S, Tachibana K, Minami Y, Yatabe Y, Mitsudomi T, Tanaka H, Kimura T, Kudoh S, Nokihara H, Ohe Y, Yokota J, Uramoto H, Yasumoto K, Kiura K, Higashiyama M, Oda M, Saito H, Yoshida J, Kondoh K, Noguchi M. Hepatocyte growth factor expression in EGFR mutant lung cancer with intrinsic and acquired resistance to tyrosine kinase inhibitors in a Japanese cohort. *J Thorac Oncol* 2011; 6(12): 2011–2017
 100. van Lengerich B, Agnew C, Puchner EM, Huang B, Jura N. EGF and NRG induce phosphorylation of HER3/ERBB3 by EGFR using distinct oligomeric mechanisms. *Proc Natl Acad Sci USA*

- 2017; 114(14): E2836–E2845
101. Mujoo K, Choi BK, Huang Z, Zhang N, An Z. Regulation of ERBB3/HER3 signaling in cancer. *Oncotarget* 2014; 5(21): 10222–10236
 102. Tian W, Li S, Chen D, Liang G, Zhang L, Zhang W, Tu X, Peng L, Weng J, Zhao G. Preclinical development of a bispecific antibody-trap selectively targeting CD47 and CD20 for the treatment of B cell lineage cancer. *Cancer Res* 2019; 79(13 Suppl): Abstract nr 545
 103. Robert B, Dorvillius M, Buchegger F, Garambois V, Mani JC, Pugnères M, Mach JP, Pèlerin A. Tumor targeting with newly designed biparatopic antibodies directed against two different epitopes of the carcinoembryonic antigen (CEA). *Int J Cancer* 1999; 81(2): 285–291
 104. Wei H, Cai H, Jin Y, Wang P, Zhang Q, Lin Y, Wang W, Cheng J, Zeng N, Xu T, Zhou A. Structural basis of a novel heterodimeric Fc for bispecific antibody production. *Oncotarget* 2017; 8(31): 51037–51049
 105. Li F, Zhang B, Ye P, Zhao J, Huang S, Jin C. Bispecific anti-HER2 antibody. Patent US 9745382 (B1); PCT/CN2017/093816. 2017
 106. Center for Drug Evaluation of the National Medical Products Authority. <http://www.cde.org.cn/> (accessed August 31, 2020)
 107. Li B, Xia Y, Wang Z M, Zhang P. Patent MX2019002254 (A). 2019
 108. Du X, Liu M, Su J, Zhang P, Tang F, Ye P, Devenport M, Wang X, Zhang Y, Liu Y, Zheng P. Uncoupling therapeutic from immunotherapy-related adverse effects for safer and effective anti-CTLA-4 antibodies in CTLA4 humanized mice. *Cell Res* 2018; 28(4): 433–447
 109. Du X, Tang F, Liu M, Su J, Zhang Y, Wu W, Devenport M, Lazarski CA, Zhang P, Wang X, Ye P, Wang C, Hwang E, Zhu T, Xu T, Zheng P, Liu Y. A reappraisal of CTLA-4 checkpoint blockade in cancer immunotherapy. *Cell Res* 2018; 28(4): 416–432
 110. Liu Y, Zheng P. Preserving the CTLA-4 checkpoint for safer and more effective cancer immunotherapy. *Trends Pharmacol Sci* 2020; 41(1): 4–12
 111. Duell J, Lurati S, Dittrich M, Bedke T, Pule M, Einsele H, Rossig C, Topp M S. First generation chimeric antigen receptor display functional defects in key signal pathways upon antigen stimulation. *Blood* 2010; 116(21): 2088
 112. Carpenito C, Milone MC, Hassan R, Simonet JC, Lakhai M, Suhoski MM, Varela-Rohena A, Haines KM, Heitjan DF, Albelda SM, Carroll RG, Riley JL, Pastan I, June CH. Control of large, established tumor xenografts with genetically retargeted human T cells containing CD28 and CD137 domains. *Proc Natl Acad Sci USA* 2009; 106(9): 3360–3365
 113. June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science* 2018; 359(6382): 1361–1365
 114. Prasad V. Tisagenlecleucel—the first approved CAR-T-cell therapy: implications for payers and policy makers. *Nat Rev Clin Oncol* 2018; 15(1): 11–12
 115. Bouchkouj N, Kasamon YL, de Claro RA, George B, Lin X, Lee S, Blumenthal GM, Bryan W, McKee AE, Pazdur R. FDA approval summary: axicabtagene ciloleucel for relapsed or refractory large B-cell lymphoma. *Clin Cancer Res* 2019; 25(6): 1702–1708
 116. Voelker R. CAR-T therapy is approved for mantle cell lymphoma. *JAMA* 2020; 324(9): 832
 117. Mullard A. FDA approves fourth CAR-T cell therapy. *Nat Rev Drug Discov* 2021; 20(3): 166
 118. Ramos CA, Grover NS, Beaven AW, Lulla PD, Wu MF, Ivanova A, Wang T, Shea TC, Rooney CM, Dittus C, Park SI, Gee AP, Eldridge PW, McKay KL, Mehta B, Cheng CJ, Buchanan FB, Grilley BJ, Morrison K, Brenner MK, Serody JS, Dotti G, Heslop HE, Savoldo B. Anti-CD30 CAR-T cell therapy in relapsed and refractory Hodgkin lymphoma. *J Clin Oncol* 2020; 38(32): 3794–3804
 119. Huang R, Li X, He Y, Zhu W, Gao L, Liu Y, Gao L, Wen Q, Zhong JF, Zhang C, Zhang X. Recent advances in CAR-T cell engineering. *J Hematol Oncol* 2020; 13(1): 86
 120. Liu Y, Guo Y, Wu Z, Feng K, Tong C, Wang Y, Dai H, Shi F, Yang Q, Han W. Anti-EGFR chimeric antigen receptor-modified T cells in metastatic pancreatic carcinoma: a phase I clinical trial. *Cytotherapy* 2020; 22(10): 573–580
 121. Cutmore LC, Brown NF, Raj D, Chaudhuri S, Wang P, Maher J, Wang Y, Lemoine NR, Marshall JF. Pancreatic Cancer UK Grand Challenge: developments and challenges for effective CAR T cell therapy for pancreatic ductal adenocarcinoma. *Pancreatolgy* 2020; 20(3): 394–408
 122. Depil S, Duchateau P, Grupp SA, Mufti G, Poirot L. ‘Off-the-shelf’ allogeneic CAR T cells: development and challenges. *Nat Rev Drug Discov* 2020; 19(3): 185–199
 123. Cutmore LC, Marshall JF. Current perspectives on the use of off the shelf CAR-T/NK cells for the treatment of cancer. *Cancers (Basel)* 2021; 13(8): 1926
 124. Capsomidis A, Benthall G, Van Acker HH, Fisher J, Kramer AM, Abeln Z, Majani Y, Gileadi T, Wallace R, Gustafsson K, Flutter B, Anderson J. Chimeric antigen receptor-engineered human gamma delta T cells: enhanced cytotoxicity with retention of cross presentation. *Mol Ther* 2018; 26(2): 354–365
 125. Torikai H, Reik A, Liu PQ, Zhou Y, Zhang L, Maiti S, Huls H, Miller JC, Kebriaei P, Rabinovich B, Lee DA, Champlin RE, Bonini C, Naldini L, Rebar EJ, Gregory PD, Holmes MC, Cooper LJ. A foundation for universal T-cell based immunotherapy: T cells engineered to express a CD19-specific chimeric-antigen-receptor and eliminate expression of endogenous TCR. *Blood* 2012; 119(24): 5697–5705
 126. Melenhorst JJ, Leen AM, Bollard CM, Quigley MF, Price DA, Rooney CM, Brenner MK, Barrett AJ, Heslop HE. Allogeneic virus-specific T cells with HLA alloreactivity do not produce GVHD in human subjects. *Blood* 2010; 116(22): 4700–4702
 127. Kochenderfer JN, Dudley ME, Carpenter RO, Kassim SH, Rose JJ, Telford WG, Hakim FT, Halverson DC, Fowler DH, Hardy NM, Mato AR, Hickstein DD, Gea-Banacloche JC, Pavletic SZ, Sportes C, Maric I, Feldman SA, Hansen BG, Wilder JS, Blacklock-Schuber B, Jena B, Bishop MR, Gress RE, Rosenberg SA. Donor-derived CD19-targeted T cells cause regression of malignancy persisting after allogeneic hematopoietic stem cell transplantation. *Blood* 2013; 122(25): 4129–4139
 128. Guo F, Cui J. CAR-T in cancer treatment: develop in self-optimization, win-win in cooperation. *Cancers (Basel)* 2021; 13(8): 1955
 129. Hu J, Sun C, Bernatchez C, Xia X, Hwu P, Dotti G, Li S. T-cell homing therapy for reducing regulatory T cells and preserving

- effector T-cell function in large solid tumors. *Clin Cancer Res* 2018; 24(12): 2920–2934
130. Murty S, Haile ST, Beinart C, Aalipour A, Alam IS, Murty T, Shaffer TM, Patel CB, Graves EE, Mackall CL, Gambhir SS. Intravital imaging reveals synergistic effect of CAR T-cells and radiation therapy in a preclinical immunocompetent glioblastoma model. *OncoImmunology* 2020; 9(1): 1757360
131. Gresser R, Cherkassky L, Chintala N, Adusumilli PS. Combination immunotherapy with CAR T cells and checkpoint blockade for the treatment of solid tumors. *Cancer Cell* 2019; 36(5): 471–482
132. Lee YG, Marks I, Srinivasarao M, Kanduluru AK, Mahalingam SM, Liu X, Chu H, Low PS. Use of a single CAR T cell and several bispecific adapters facilitates eradication of multiple antigenically different solid tumors. *Cancer Res* 2019; 79(2): 387–396
133. Driouk L, Gicobi JK, Kamihara Y, Rutherford K, Dranoff G, Ritz J, Baumeister SHC. Chimeric antigen receptor T cells targeting NKG2D-ligands show robust efficacy against acute myeloid leukemia and T-cell acute lymphoblastic leukemia. *Front Immunol* 2020; 11: 580328
134. Caruana I, Weber G, Ballard BC, Wood MS, Savoldo B, Dotti G. K562-derived whole-cell vaccine enhances antitumor responses of CAR-redirection virus-specific cytotoxic T lymphocytes *in vivo*. *Clin Cancer Res* 2015; 21(13): 2952–2962
135. Bommareddy PK, Shettigar M, Kaufman HL. Integrating oncolytic viruses in combination cancer immunotherapy. *Nat Rev Immunol* 2018; 18(8): 498–513
136. Hu Z, Ott PA, Wu CJ. Towards personalized, tumour-specific, therapeutic vaccines for cancer. *Nat Rev Immunol* 2018; 18(3): 168–182
137. Parkhurst MR, Yang JC, Langan RC, Dudley ME, Nathan DA, Feldman SA, Davis JL, Morgan RA, Merino MJ, Sherry RM, Hughes MS, Kammula US, Phan GQ, Lim RM, Wank SA, Restifo NP, Robbins PF, Laurencot CM, Rosenberg SA. T cells targeting carcinoembryonic antigen can mediate regression of metastatic colorectal cancer but induce severe transient colitis. *Mol Ther* 2011; 19(3): 620–626
138. Blass E, Ott PA. Advances in the development of personalized neoantigen-based therapeutic cancer vaccines. *Nat Rev Clin Oncol* 2021; 18(4): 215–229
139. Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, Zhang W, Luoma A, Giobbie-Hurder A, Peter L, Chen C, Olive O, Carter TA, Li S, Lieb DJ, Eisenhaure T, Gjini E, Stevens J, Lane WJ, Javeri I, Nellaippan K, Salazar AM, Daley H, Seaman M, Buchbinder EI, Yoon CH, Harden M, Lennon N, Gabriel S, Rodig SJ, Barouch DH, Aster JC, Getz G, Wucherpfennig K, Neuberg D, Ritz J, Lander ES, Fritsch EF, Hacohen N, Wu CJ. An immunogenic personal neoantigen vaccine for patients with melanoma. *Nature* 2017; 547(7662): 217–221
140. Hu Z, Leet DE, Allesøe RL, Oliveira G, Li S, Luoma AM, Liu J, Forman J, Huang T, Iorgulescu JB, Holden R, Sarkizova S, Gohil SH, Redd RA, Sun J, Elagina L, Giobbie-Hurder A, Zhang W, Peter L, Ciantra Z, Rodig S, Olive O, Shetty K, Pyrdol J, Uduman M, Lee PC, Bachireddy P, Buchbinder EI, Yoon CH, Neuberg D, Pentelute BL, Hacohen N, Livak KJ, Shukla SA, Olsen LR, Barouch DH, Wucherpfennig KW, Fritsch EF, Keskin DB, Wu CJ, Ott PA. Personal neoantigen vaccines induce persistent memory T cell responses and epitope spreading in patients with melanoma. *Nat Med* 2021; 27(3): 515–525
141. Keskin DB, Anandappa AJ, Sun J, Tirosh I, Mathewson ND, Li S, Oliveira G, Giobbie-Hurder A, Felt K, Gjini E, Shukla SA, Hu Z, Li L, Le PM, Allesøe RL, Richman AR, Kowalczyk MS, Abdelrahman S, Geduldig JE, Charbonneau S, Pelton K, Iorgulescu JB, Elagina L, Zhang W, Olive O, McCluskey C, Olsen LR, Stevens J, Lane WJ, Salazar AM, Daley H, Wen PY, Chiocca EA, Harden M, Lennon NJ, Gabriel S, Getz G, Lander ES, Regev A, Ritz J, Neuberg D, Rodig SJ, Ligon KL, Suvà ML, Wucherpfennig KW, Hacohen N, Fritsch EF, Livak KJ, Ott PA, Wu CJ, Reardon DA. Neoantigen vaccine generates intratumoral T cell responses in phase Ib glioblastoma trial. *Nature* 2019; 565(7738): 234–239
142. Ott PA, Hu-Lieskovan S, Chmielowski B, Govindan R, Naing A, Bhardwaj N, Margolin K, Awad MM, Hellmann MD, Lin JJ, Friedlander T, Bushway ME, Balogh KN, Sciuto TE, Kohler V, Turnbull SJ, Besada R, Curran RR, Trapp B, Scherer J, Poran A, Harjanto D, Barthelme D, Ting YS, Dong JZ, Ware Y, Huang Y, Huang Z, Wanamaker A, Cleary LD, Moles MA, Manson K, Greshock J, Khondker ZS, Fritsch E, Rooney MS, DeMario M, Gaynor RB, Srinivasan L. A phase Ib trial of personalized neoantigen therapy plus anti-PD-1 in patients with advanced melanoma, non-small cell lung cancer, or bladder cancer. *Cell* 2020; 183(2): 347–362.e24
143. Lindskog M, Laurell A, Kjellman A, Melichar B, Niezabitowski J, Maroto P, Zieliński H, Villacampa F, Bigot P, Bajory ZA. A randomized phase II study with ilixadencel, a cell-based immune primer, plus sunitinib versus sunitinib alone in synchronous metastatic renal cell carcinoma. *J Clin Oncol* 2020; 38(5_suppl): 11
144. Tanyi JL, Bobisse S, Ophir E, Tuytaerts S, Roberti A, Genolet R, Baumgartner P, Stevenson BJ, Iseli C, Dangaj D, Czerniecki B, Semiletov A, Racle J, Michel A, Xenarios I, Chiang C, Monos DS, Torigian DA, Nisenbaum HL, Michielin O, June CH, Levine BL, Powell DJ Jr, Gfeller D, Mick R, Dafni U, Zoete V, Harari A, Coukos G, Kandalaf LE. Personalized cancer vaccine effectively mobilizes antitumor T cell immunity in ovarian cancer. *Sci Transl Med* 2018; 10(436): ea05931
145. Moynihan KD, Opel CF, Szeto GL, Tzeng A, Zhu EF, Engreitz JM, Williams RT, Rakhra K, Zhang MH, Rothschilds AM, Kumari S, Kelly RL, Kwan BH, Abraham W, Hu K, Mehta NK, Kauke MJ, Suh H, Cochran JR, Lauffenburger DA, Wittrup KD, Irvine DJ. Eradication of large established tumors in mice by combination immunotherapy that engages innate and adaptive immune responses. *Nat Med* 2016; 22(12): 1402–1410
146. Chau I, Haag G, Rahma O, Macarulla T, McCune S, Yardley D, Solomon B, Johnson M, Vidal G, Schmid P, Argiles G, Dimick K, Mahrus S, Abdullah H, He X, Sayyed P, Barak H, Bleul C, Cha E, Drakaki A. MORPHEUS: a phase Ib/II umbrella study platform evaluating the safety and efficacy of multiple cancer immunotherapy (CIT)-based combinations in different tumour types. *Ann Oncol* 2018; 29(suppl_8): 439–440
147. Simonsen KL, Fracasso PM, Bernstein SH, Wind-Rotolo M, Gupta M, Comprelli A, Reilly TP, Cassidy J. The Fast Real-time Assessment of Combination Therapies in Immuno-ONcology (FRACTION) program: innovative, high-throughput clinical screening of immunotherapies. *Eur J Cancer* 2018; 103: 259–266
148. Redman JM, Steinberg SM, Gulley JL. Quick efficacy seeking trial (QuEST1): a novel combination immunotherapy study designed

- for rapid clinical signal assessment metastatic castration-resistant prostate cancer. *J Immunother Cancer* 2018; 6(1): 91
149. Tang J, Shalabi A, Hubbard-Lucey VM. Comprehensive analysis of the clinical immuno-oncology landscape. *Ann Oncol* 2018; 29(1): 84–91
150. Monk BJ, Brady MF, Aghajanian C, Lankes HA, Rizack T, Leach J, Fowler JM, Higgins R, Hanjani P, Morgan M, Edwards R, Bradley W, Kolevska T, Foukas P, Swisher EM, Anderson KS, Gottardo R, Bryan JK, Newkirk M, Manjarrez KL, Mannel RS, Hershberg RM, Coukos G. A phase 2, randomized, double-blind, placebo-controlled study of chemo-immunotherapy combination using motolimod with pegylated liposomal doxorubicin in recurrent or persistent ovarian cancer: a Gynecologic Oncology Group partners study. *Ann Oncol* 2017; 28(5): 996–1004
151. Yu S, Yi M, Qin S, Wu K. Next generation chimeric antigen receptor T cells: safety strategies to overcome toxicity. *Mol Cancer* 2019; 18(1): 125
152. Levy BP, Giaccone G, Besse B, Felip E, Garassino MC, Domine Gomez M, Garrido P, Piperdi B, Ponce-Aix S, Menezes D, MacBeth KJ, Risueño A, Slepets R, Wu X, Fandi A, Paz-Ares L. Randomised phase 2 study of pembrolizumab plus CC-486 versus pembrolizumab plus placebo in patients with previously treated advanced non-small cell lung cancer. *Eur J Cancer* 2019; 108: 120–128
153. Mijalis AJ, Thomas DA 3rd, Simon MD, Adamo A, Beaumont R, Jensen KF, Pentelute BL. A fully automated flow-based approach for accelerated peptide synthesis. *Nat Chem Biol* 2017; 13(5): 464–466