

Nitrogen allocation to biochemical fractions shapes functional traits and N use efficiency in tobacco cultivars

Guiru ZHAO^{*1}, Shichen LI^{*1,2}, Xiaoyao DING¹, Xiaoci PENG^{1,3}, Waqar AHMED¹, Shaoming LI (✉)¹, Zhengxiong ZHAO (✉)¹

1 Key Laboratory for Improving Quality and Productivity of Arable Land of Yunnan Province, College of Resources and Environment, Yunnan Agricultural University, Kunming 650201, China.

2 Kunming Branch of Yunnan Tobacco Monopoly Bureau, Kunming 650051, China.

3 Qujing Branch of Yunnan Tobacco Monopoly Bureau, Qujing 655000, China.

*These authors contributed equally to the work.

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Correspondences: 248501858@qq.com, zhaozx0801@163.com

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SUPPLEMENTARY MATERIALS

S1 Morphological and functional partitioning of nitrogen

S1.1 Conceptual framework for nitrogen partitioning

Nitrogen (N) within plant tissues exists in diverse biochemical forms that fulfill distinct physiological roles. To facilitate interpretation of nitrogen use efficiency (NUE), plant nitrogen was categorized from two complementary perspectives: morphological nitrogen fractions (defined by solubility and molecular association) and functional nitrogen pools (defined by physiological roles in photosynthesis, respiration, structure, or storage)^[1].

Morphological nitrogen fractions included SDS-insoluble nitrogen (N_{in-SDS}), SDS-soluble nitrogen (N_s), water-soluble nitrogen (N_w), and non-protein nitrogen (N_{np}). Functional nitrogen pools were defined following the LUNA framework^[1], including photosynthetic nitrogen (N_{psn}), respiratory nitrogen (N_{resp}), structural nitrogen (N_{str}), and storage nitrogen (N_{store}).

S1.2 Determination of morphological nitrogen fractions

Morphological nitrogen fractions were determined in frozen root, stem, and leaf tissues following a modified protocol of Takashima et al.^[2]. Approximately 1.0 g of fresh tissue was flash-frozen in liquid nitrogen and homogenized in 1.0 mL of 100 mmol·L⁻¹ sodium phosphate buffer (pH 7.5) containing 2 mmol·L⁻¹ MgCl₂, 0.4 mol·L⁻¹ D-sorbitol, 5 mmol·L⁻¹ dithiothreitol, 5 mmol·L⁻¹ iodoacetate, 10 mmol·L⁻¹ NaCl, and 5 mmol·L⁻¹ phenylmethylsulfonyl fluoride.

The homogenate was centrifuged at $12,000 \times g$ for 10 min at 4 °C. The supernatant was collected as water-soluble nitrogen (N_w).

The remaining tissue was resuspended in phosphate buffer containing 3% sodium dodecyl sulfate (SDS), incubated at 90 °C for 5 min, and centrifuged at 5500 g for 8 min. The resulting supernatant was collected as SDS-soluble nitrogen (N_s), mainly composed of soluble functional proteins.

The remaining insoluble fraction was rinsed with anhydrous ethanol, filtered through quantitative filter paper, treated with 20% trichloroacetic acid, washed again with ethanol, air-dried, and digested using $H_2SO_4-H_2O_2$. Nitrogen in this fraction was designated as SDS-insoluble nitrogen (N_{in-SDS}), corresponding primarily to structural nitrogen.

Non-protein nitrogen (N_{np}) was calculated as the difference between total nitrogen and the sum of protein-associated nitrogen fractions ($N_w + N_s + N_{in-SDS}$), and includes nitrate, ammonium, free amino acids, and other low-molecular-weight nitrogenous compounds. Nitrogen concentrations were determined using continuous flow analysis (AA3, Seal Analytical Inc., Southampton, UK).

S1.3 Determination of Functional Nitrogen Fractions in Leaves

Functional nitrogen fractions were quantified only in leaves. All functional nitrogen components were initially calculated on a fresh weight basis. To ensure consistency with total nitrogen and morphological nitrogen fractions (measured on a dry weight basis), values were converted using sample-specific fresh-to-dry weight ratios.

Gas exchange measurements were conducted one day before harvest using a portable photosynthesis system (LI-6400XT; LI-COR Biosciences, Lincoln, NE, USA). Net photosynthetic rate (A_n) and A/C_i response curves were recorded to derive the maximum carboxylation rate ($V_{c,max}$) and maximum electron transport rate (J_{max})^[3].

The photosynthesis model equations were:

$$A_n = \frac{V_{c,max}(C_i - \Gamma^*)}{C_i + K_c(1 + O/K_o)} - R_d \quad (1)$$

$$A_n = \frac{J_{max}(C_i - \Gamma^*)}{4C_i + 8\Gamma^*} - R_d \quad (2)$$

where C_i is intercellular CO₂ concentration ($\mu\text{mol}\cdot\text{mol}^{-1}$), Γ^* is the CO₂ compensation point ($\mu\text{mol}\cdot\text{mol}^{-1}$), K_c and K_o are Michaelis constants for carboxylation and oxygenation, respectively, O is the O₂ concentration ($210,000 \mu\text{mol}\cdot\text{mol}^{-1}$), and R_d is mitochondrial respiration in the light ($\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1} \text{CO}_2$).

Chlorophyll a and b were extracted from frozen leaf disks using 80% acetone for 48 h and quantified spectrophotometrically (UV2102 PCS, Unico, China)^[4].

Calculation of Functional Nitrogen Pools

Photosynthetic nitrogen (N_{psn}) was calculated as the sum of nitrogen allocated to the carboxylation system (N_{cb}), electron transport system (N_{et}), and light-harvesting system (N_{lc}):

$$N_{psn} = N_{cb} + N_{et} + N_{lc} \quad (3)$$

N_{cb} was estimated as:

$$N_{cb} = \frac{V_{c,max}}{6.25 \times V_{cr} \times f_{V_{c,max}}} \quad (4)$$

where 6.25 is the Rubisco-to-nitrogen conversion factor, V_{cr} is the Rubisco carboxylation rate (20.78 $\mu\text{mol CO}_2 \text{ g}^{-1} \text{ Rubisco s}^{-1}$ at 25 °C), and $f_{V_{c,max}}$ is the temperature correction factor (0.359 at 15 °C).

N_{et} was calculated as:

$$N_{et} = \frac{J_{max}}{8.06 \times J_{mc} \times f_{J_{max}}} \quad (5)$$

where 8.06 is the cytochrome f nitrogen coefficient, J_{mc} is the electron transport capacity of cytochrome f (155.65 $\mu\text{mol} \cdot \text{s}^{-1}$ at 25 °C), and $f_{J_{max}}$ is the temperature correction factor (0.714 at 15 °C).

N_{lc} was calculated from chlorophyll content:

$$N_{lc} = \frac{C_c}{C_B} \quad (6)$$

where C_c is chlorophyll concentration ($\text{mmol} \cdot \text{m}^{-2}$) and C_B is the chlorophyll-to-nitrogen ratio (2.15 $\text{mmol} \cdot \text{g}^{-1}$).

Respiratory, Structural, and Storage Nitrogen

Respiratory nitrogen (N_{resp}) was estimated as:

$$N_{resp} = \frac{R_t}{33.69 \times f_r} \quad (7)$$

where $R_t = 0.015 \times V_{c,max}$, 33.69 represents respiratory nitrogen use efficiency at 25 °C, and f_r is the temperature correction factor (0.522 at 15 °C).

Total leaf nitrogen (N_a) was partitioned as:

$$N_a = N_{psn} + N_{resp} + N_{str} + N_{store} \quad (8)$$

Structural nitrogen (N_{str}) was defined as N_{in-SDS} .

Storage nitrogen (N_{store}) comprised three components:

- Water-soluble storage nitrogen (N_{ow}):

$$N_{ow} = N_w - N_{cb} - N_{resp} \quad (9)$$

- SDS-soluble storage nitrogen (N_{os}):

$$N_{os} = N_s - N_{lc} - N_{et} \quad (10)$$

- Non-protein storage nitrogen (N_{np}), representing residual nitrogen not included in protein-associated fractions.

REFERENCES

1. Liu T, Ren T, White P J, Cong R, Lu J. Storage nitrogen co-ordinates leaf expansion and photosynthetic capacity in winter oilseed rape. *Journal of Experimental Botany*, 2018, **69**(12): 2995–3007
2. Takashima T, Hikosaka K, Hirose T. Photosynthesis or persistence: nitrogen allocation in leaves of evergreen and deciduous *Quercus* species. *Plant, Cell & Environment*, 2004, **27**(8): 1047–1054
3. Song G, Wang Q, Jin J. Exploring the instability of the relationship between maximum potential electron transport rate and maximum carboxylation rate in cool-temperate deciduous forests. *Agricultural and Forest Meteorology*, 2021, **308–309**: 108614
4. Li M, Zhang Y, Xu X, Chen Y, Chu J, Yao X. The combined treatments of brassinolide and zeaxanthin better alleviate oxidative damage and improve hypocotyl length, biomass, and the quality of radish sprouts stored at low temperature. *Food Chemistry*: 2022, **15**: 100394