

Rhubarb in renal fibrosis: from traditional ethnopharmacology to mechanistic therapeutic development

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Review

Rhubarb in renal fibrosis: from traditional ethnopharmacology to mechanistic therapeutic development

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ABSTRACT

Rhubarb (*Rheum* spp.), a traditional herbal medicine, has attracted growing interest due to its anti-renal fibrosis effects in chronic kidney disease (CKD). This review systematically evaluates Rhubarb's botanical features, global distribution, and diverse processing methods, which influence its chemical composition and bioactivity. Major bioactive constituents, including anthraquinones, stilbenes, and polyphenols, are cataloged, and their potential roles in renal protection are elucidated. Traditional applications in nephropathy management are critically assessed alongside contemporary pharmacological evidence demonstrating Rhubarb's ability to attenuate renal fibrosis. Notably, this review highlights that multiple bioactive components in Rhubarb exert potent anti-fibrotic effects through complex, interactive modulation of multiple signaling pathways. Despite promising preclinical data, clinical translation remains limited by insufficient understanding of pharmacokinetics and potential herb-drug interactions. This synthesis identifies key research gaps, advocating for interdisciplinary studies to decipher multi-target mechanisms, refine pharmacokinetic profiles to enhance bioavailability, and translate preclinical findings into randomized controlled trials (RCTs). By integrating ethnopharmacological knowledge with modern drug discovery frameworks, this review underscores Rhubarb's potential as a multifaceted anti-fibrotic agent while calling for methodologically rigorous research to validate its therapeutic integration into CKD management protocols.

1. Introduction

Chronic kidney disease (CKD), characterized by progressive renal fibrosis, constitutes a significant global health burden with limited therapeutic options¹. Renal fibrosis, defined by excessive extracellular matrix (ECM) deposition, tubular atrophy, and inflammatory infiltration, represents the final common pathway of CKD progression, irrespective of etiology, such as diabetes, hypertension, or glomerulonephritis². Despite advances in renin-angiotensin system inhibitors and sodium-glucose cotransporter-2 inhibitors, these therapies often fail to halt fibrosis progression, highlighting the urgent need for novel anti-fibrotic agents³. In recent decades, natural products have re-emerged as promising candidates due to their multi-target mechanisms and long-standing use in traditional medicine⁴. Among these, Rhubarb (*Rheum* spp.), a perennial herb from the Polygonaceae family, has attracted increasing attention for its potential to mitigate renal injury and fibrosis through diverse bioactive constituents.

Rhubarb, native to temperate regions of Asia and Europe, has

been cultivated for millennia in China (*Da huang*), where its roots and rhizomes are processed into various formulations, such as raw, wine-fried, or carbonized forms, to modulate therapeutic effects. Traditionally, it has been prescribed in East Asian medicine for constipation, inflammation, and blood purification. Modern pharmacological studies attribute its bioactivity to a rich array of phytochemicals, including anthraquinones, flavonoids, stilbenes, and tannins, which collectively exhibit antioxidant, anti-inflammatory, and anti-fibrotic properties⁵. Notably, emerging evidence indicates that Rhubarb extracts and isolated compounds ameliorate renal fibrosis by targeting key signaling pathways, such as the transforming growth factor-beta (TGF-β)/Smad cascade, oxidative stress responses, and epithelial-mesenchymal transition (EMT). For instance, Emodin, a primary anthraquinone, improves renal dysfunction by activating autophagy and inhibiting EMT in rodent models of renal fibrosis, whereas Resveratrol derivatives attenuate mitochondrial dysfunction in glomerular mesangial cells⁶. These findings position Rhubarb as a multifaceted phytotherapeutic agent with substantial translational potential.

This review systematically consolidates current evidence on Rhubarb's anti-renal fibrotic properties, addressing its botanical background, functional constituents, and mechanistic insights. We further evaluate the roles of specific phytochemicals, such as

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anthraquinones and flavonoids, in modulating fibrotic pathways and propose strategies to bridge phytochemistry with translational nephrology. By elucidating Rhubarb's potential as a cost-effective adjuvant therapy, this work aims to inspire targeted research into its clinical validation and sustainable application in CKD management.

2. Botany and geographical distribution of Rhubarb

2.1. Botanical characteristics

Rheum palmatum L., commonly known as Chinese Rhubarb or Dahuang, belongs to the Polygonaceae family within the genus *Rheum*. This perennial herbaceous plant exhibits distinct morphological features that facilitate identification⁷. It develops a robust, fleshy rhizome system with characteristic yellow-orange transverse markings when cut longitudinally, a key diagnostic trait in traditional Chinese medicine (TCM) authentication. Mature plants typically reach 1–3 meters in height, bearing large palmately lobed leaves (15–40 cm in diameter) with 5–7 deeply divided segments and prominent venation. Reddish-green flowering stalks emerge in early summer, producing panicles of small hermaphroditic flowers, each with six tepals and nine stamens. Three primary medicinal species are recognized: *R. palmatum*, *R. officinale*, and *R. tanguticum*, distinguishable by leaf morphology and microscopic features of root cross-sections (Fig. 1). The bioactive compounds, particularly anthraquinones (e.g., Emodin, Chrysophanol) and tannins, accumulate in the parenchyma cells of the thickened roots and rhizomes, with optimal concentrations achieved after 3–5 years of growth.

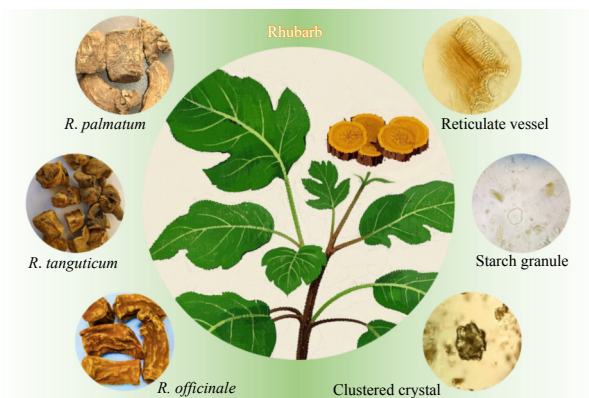


Fig. 1 Botanical characteristics of Rhubarb. Three primary medicinal varieties are recognized: *R. palmatum*, *R. officinale*, and *R. tanguticum* (left). The microstructure of Rhubarb is characterized by clustered crystals, reticulate vessels, and starch granules (right).

2.2. Geographical distribution

Native to temperate regions of East Asia, *Rheum palmatum* thrives at altitudes between 1500 and 4000 meters across specific biogeographical zones. Its natural habitat extends along the eastern Qinghai-Tibet Plateau, spanning western China (Gansu, Sichuan, Qinghai, Tibet), northern Nepal, and Bhutan. Commercial cultivation is concentrated in China's "Rhubarb Belt", a 1200 km corridor from southern Gansu to northwestern Yunnan, where well-drained sandy loam soils and diurnal temperature fluctuations (15–25 °C daytime; 5–12 °C nighttime) optimize secondary metabolite production. *R. palmatum* is primarily produced in Gansu, Qinghai, Tibet, Sichuan, and Yunnan provinces. *R. tanguticum* is mainly cultivated in Qinghai, Gansu, Tibet, and Sichuan provinces, while *R. officinale* is predominantly grown in Sichuan, Guizhou, Yunnan, Hubei, and Shaanxi provinces (Fig.

S2). Historical trade routes facilitated its spread to Central Asia, with wild populations persisting in Mongolia's Khangai Mountains and Russia's Altai region. Modern introductions to Europe (Germany, Poland) and North America have established limited cultivation, although these non-native populations exhibit reduced concentrations of bioactive compounds compared to indigenous Chinese varieties.

3. Processing methods of Rhubarb

The processing of medicine represents a critical pharmaceutical technique in TCM that directly influences its clinical applications⁸. Through distinct preparation methods, practitioners can modulate the herb's chemical composition and therapeutic properties to suit specific therapeutic needs⁹. The primary processed forms include raw Rhubarb (Sheng Dahuang), steamed Rhubarb (Shu Dahuang), wine-processed Rhubarb (Jiu Dahuang), and charred Rhubarb (Dahuang Tan), each demonstrating unique processing protocols and pharmacological outcomes (Fig. 2).

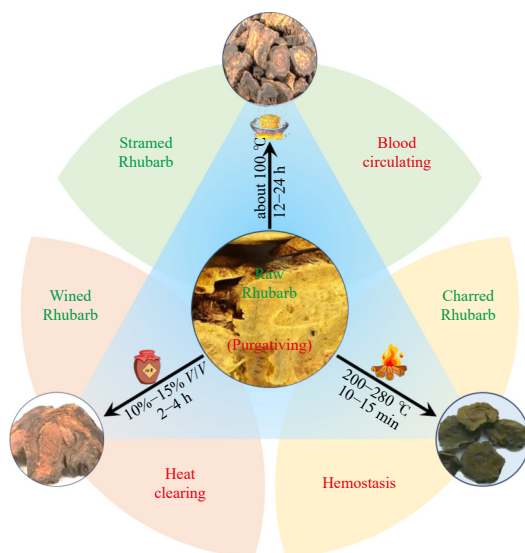


Fig. 2 Main processed products and effects of Rhubarb. The primary processed forms include raw Rhubarb (Sheng Dahuang), steamed Rhubarb (Shu Dahuang), wine-processed Rhubarb (Jiu Dahuang), and charred Rhubarb (Dahuang Tan), each demonstrating unique processing protocols and pharmacological outcomes.

3.1. Raw Rhubarb (Sheng Dahuang)

The simplest form involves cleaning fresh rhizomes with running water to remove soil impurities, followed by slicing into 2–3 mm thick pieces. These slices undergo natural sun-drying or low-temperature oven drying (40–50 °C) to preserve bioactive components¹⁰. Raw Rhubarb contains the highest concentration of anthraquinone glycosides, making it the most potent form for purgative effects¹¹. However, its strong cathartic properties and potential gastric irritation limit its use in patients with deficient spleen/stomach conditions.

3.2. Steamed Rhubarb (Shu Dahuang)

This method modifies the herb's action through prolonged moist-heat treatment¹². Sliced Rhubarb is layered with or without black bean soup (30%, W/W) in a bamboo steamer and processed for 12–24 hours at 100 °C. The extended steaming converts bitter naphthodianthrones into more palatable forms while preserving emodin and chrysophanol¹³. The processed herb demonstrates milder purgative effects with enhanced blood-nourishing capabilities, often prescribed for chronic constipation in debilitated patients¹⁴.

3.3. Wine-processed Rhubarb (*Jiu Dahuang*)

This preparation employs yellow rice wine (10%–15% W/W) as a processing medium. The cleaned Rhubarb slices are moistened with wine and allowed to marinate for 2–4 hours until complete absorption. The impregnated material is then stir-fried in a copper wok at 160–180 °C until achieving a dark brown color with slight scorch spots¹⁵. Wine processing enhances the herb's ability to clear upper-jiao heat while reducing its drastic purgative effects through partial decomposition of anthraquinones. The alcohol-soluble components improve blood-activating properties, making it suitable for treating blood stasis syndromes¹⁶.

3.4. Charred Rhubarb (*Dahuang Tan*)

Prepared through controlled carbonization, raw slices are stir-fried in an iron wok at 220–280 °C until surfaces turn charcoal-black while maintaining reddish-brown interiors¹⁷. This high-temperature processing (approximately 10–15 minutes) significantly increases tannin content through Maillard reactions while reducing anthraquinones by 40%–60%¹⁸. The resultant product exhibits enhanced hemostatic and astringent properties, particularly effective for treating hematemesis and hemorrhagic disorders¹⁹.

3.5. The Rhubarb processing and chemical compositions

The complex chemical components of TCM are the main material basis for its clinical therapeutic effects. Through various processing and preparation methods, TCM can induce changes in its chemical components, thereby enhancing therapeutic effects or reducing toxicity. Based on this, researchers^{15, 20} used different methods such as UHPLC-Q-TOF-MS to detect changes in the main chemical components of processed Rhubarb, revealing the internal logic of TCM processing (Table 1). It should be noted that numerous factors influence processing, including different excipients, processing time, and temperature, all of which can cause varying degrees of change in chemical composition. This variability constitutes both the complexity of TCM processing and a key challenge in its standardization.

3.6. The Rhubarb processing and pharmacokinetics

In addition, the pharmacokinetics of chemical components in different processed products also change, which is an important factor affecting pharmacodynamics. Prof. Xiao Yongqing's team established a rapid and efficient UPC-MS/MS method to determine pharmacokinetic changes of the main chemical components in rat plasma after oral administration of raw and steamed Rhubarb^{12, 14}. The results show that wine steaming alters the pharmacokinetic behavior of Rhubarb components, providing a theoretical basis for the differences in pharmacological activities among processed forms (Table 2).

3.7. The Rhubarb processing and pharmacodynamics

After processing, the efficacy of various Rhubarb preparations varies to some extent. For instance, the purgative effect of wine-processed and steamed Rhubarb is slightly milder. Wine-processed Rhubarb is particularly effective at clearing heat from the "upper jiao" and resolving "heat toxins," while steamed Rhubarb exhibits enhanced effects in promoting blood circulation and resolving blood stasis. Charred Rhubarb has a weak purgative effect but excels in cooling the blood and stopping bleeding. According to TCM theory, "blood stasis" is one of the key pathogenesises of CKD. Compared with other processed forms, steamed Rhubarb is advantageous due to its ability to "promote blood circulation and resolve blood stasis," a property supported by modern pharmacological evidence²¹. Additionally, anthraquinone

glycosides in raw Rhubarb can cause certain renal toxicity, whereas in steamed Rhubarb, these compounds decompose into free anthraquinones, thereby reducing nephrotoxicity²². Therefore, in the treatment of CKD, steamed Rhubarb is generally preferred. It should be noted that TCM emphasizes "syndrome differentiation and treatment." Drug selection is closely tied to the patient's constitution and syndrome type, and various TCM diagnostic frameworks exist. Thus, drug use is not fixed. For example, in chronic kidney failure, if a patient is highly sensitive to Rhubarb, steamed Rhubarb should be prioritized. If steamed Rhubarb proves ineffective, raw Rhubarb may be considered. Conversely, if the patient presents with an "excess syndrome (Shi Zheng)" requiring "purging the fu-organs and descending turbidity", raw Rhubarb is indicated. For physically weak patients, wine-processed Rhubarb may be selected to achieve both purgation and tonification.

4. The traditional applications of Rhubarb

Rhubarb's traditional applications reveal a multifaceted adaptogen-like profile, modulating diverse physiological pathways across cultures (Fig. 3). Its prominence in digestive, hepatic, inflammatory, gynecological, and topical therapies stems from empirical observations of its dose-dependent, preparation-specific actions. While modern pharmacology has clarified mechanisms, such as anthraquinones for constipation and rhein for inflammation, these historical uses underscore Rhubarb's enduring role as a versatile "regulator" of systemic homeostasis in pre-scientific medical systems.

4.1. Gastrointestinal disorders

Rhubarb has been a cornerstone in managing gastrointestinal ailments across TCM, Ayurveda, and European herbalism²³. Its primary application centers on constipation and dyspepsia, attributed to anthraquinone compounds (e.g., sennosides) that stimulate intestinal peristalsis²⁴. In TCM, raw Rhubarb root is prescribed for acute "excess heat" conditions, such as abdominal distention and fecal impaction, often combined with bitter herbs like Mangxiao (mirabilite) in formulations such as Dachengqi Tang²⁵. Conversely, processed Rhubarb (e.g., wine-fried or charcoal-baked) is used for chronic constipation in frail patients, reflecting TCM's emphasis on tailoring preparations to individual vitality. In addition to its laxative effects, Rhubarb has also been used in the treatment of dysentery. At low doses, its astringent tannins may help counteract diarrhea, highlighting its bidirectional regulatory role in gastrointestinal health²⁶.

4.2. Hepato-biliary system

Traditional systems have leveraged Rhubarb's purported detoxifying properties to address liver and gallbladder disorders²⁷. In TCM, it is integral to formulas targeting jaundice and hepatitis, believed to clear "damp-heat" pathogens obstructing bile flow. Classical texts such as Shanghan Lun describe Rhubarb paired with Yinchenhao (*Artemisia capillaris*) in Yinchenhao decoction to resolve yellowing of the skin and dark urine by relieving cholestasis^{28, 29}. Similarly, Tibetan medicine employs Rhubarb for liver congestion and gallstones, often blending it with gentian and chebulic myrobalan. Its choleric effects are thought to enhance bile secretion, while antioxidant components (e.g., rhein) may mitigate hepatic inflammation, a rationale later corroborated by modern studies^{30, 31}.

4.3. Inflammatory and febrile conditions

As a "cold" herb in TCM energetics, rhubarb is a key remedy for inflammatory and febrile diseases³². It is prescribed to clear

Table 1 The influence of different processing methods of Rhubarb on its main chemical components.

Chemical composition	Raw Rhubarb	Steamed Rhubarb	Wine-Processed Rhubarb	Charred Rhubarb	References
Rhein		**		**	15, 20
Emodin		**	*	**	15, 20
Aloe-emodin		**		**	20
Chrysophanol		**	**	**	20
Physcion		**	*	**	20
Gallic acid		**	**	**	20
Catechin		*	**		15, 20
Polydatin		**	**	**	20
Frambione		**	**	**	20
Sennoside A		**	**	**	15
Sennoside B		*		**	15
Rhein-8- <i>O</i> - β - <i>D</i> -glucopyranoside		**	**	**	20
Chrysophanol-8- <i>O</i> - β - <i>D</i> -glucopyranoside				**	20
Emodin-8- <i>O</i> - β - <i>D</i> -glucopyranoside		*		/	15, 20
Physcion-8- <i>O</i> - β - <i>D</i> -glucopyranoside		**	**	**	20
Aloe-emodin-8- <i>O</i> - β - <i>D</i> -glucopyranoside		**	**	**	20
Dimer of catechin		/	**	/	15
Cassialoin		/	**	/	15
Sennoside C/D		/	*	/	15
Gallic acid 3- <i>O</i> - β - <i>D</i> -(6'- <i>O</i> -galloyl)-glucopyranoside		/	*	/	15
Epicatechin-(4 β -8)-epicatechin-(4 β -8)-catechin		/	*	/	15
(-)-Epicatechin-3- <i>O</i> -gallate		/	**	/	15
2- <i>O</i> - <i>p</i> -coumaroyl-1- <i>O</i> -galloyl- β - <i>D</i> -glucose		/	**	/	15
2- <i>O</i> -cinnamoyl-1- <i>O</i> -galloyl- β - <i>D</i> -glucose		/	**	/	15
Emodin-8- <i>O</i> -(6'- <i>O</i> -malonyl)-glucopyranoside		/	**	/	15
6-Methyl-aloe-emodin		/	**	/	15

Note: The red block indicates an increase in content compared with raw Rhubarb, the blue block indicates a decrease in content compared with raw Rhubarb, and the gray block indicates no significant difference in content compared with raw Rhubarb. * indicates $P < 0.05$ vs raw Rhubarb, ** indicates $P < 0.01$ vs raw Rhubarb. "/" indicates that the relevant data not available.

"heat-toxins" manifesting as high fever, sore throat, or skin eruptions, frequently combined with antipyretic herbs like Huangqin (*Scutellaria*) in Xiexin Tang. Topically, poultices of rhubarb paste treat burns, abscesses, and infected wounds, capitalizing on its anti-microbial and astringent properties³³. In Ayurveda, Rhubarb root (called Revandchini) is used for inflammatory joint disorders, though less prominently than in East Asian traditions. Notably, its dual role as both purgative and anti-inflammatory agent aligns with the ancient concept of "expelling pathogenic fire through the bowels."

4.4. Gynecological and circulatory disorders

Rhubarb's ability to "invigorate blood and resolve stagnation" makes it a unique agent for gynecological conditions³⁴. TCM practitioners prescribe it for amenorrhea, postpartum abdominal pain, and dysmenorrhea linked to blood stasis, often processing it with wine to enhance circulation. Formulas like TaoHeChengQi Tang combine Rhubarb with peach kernels to dissolve pelvic static blood. Additionally, its hemostatic effects are exploited for hematemesis or epistaxis caused by "heat in the blood," typically using charred Rhubarb. This paradoxical capacity, both promot-

ing blood flow and arresting bleeding, underscores its context-dependent applications guided by TCM diagnostics¹⁹.

4.5. Renal support

Rhubarb has a long history of clinical use in the treatment of kidney diseases, particularly³⁵. According to TCM theory, the kidneys and intestines are interconnected through the function of Qi transformation. CKD is often associated with damp-heat, blood stasis, and toxin accumulation³⁶. Its main pathogenesis is "internal accumulation of turbid toxins", so "unblocking the bowels and eliminating turbid toxins" is the treatment strategy. Rhubarb exerts purgative, heat-clearing, detoxifying, and accumulation-attacking effects, and is regarded as a key medicine for "unblocking the bowels and eliminating turbidity"^{37,38}. Thus, the pathogenesis of CKD aligns with the pharmacological mechanism of Rhubarb in TCM. A meta-analysis involving 34 studies and 2 786 patients indicated that Rhubarb use in chronic renal failure significantly reduces serum creatinine (Scr), blood urea nitrogen (BUN), and uric acid (UA), increases creatinine clearance rate (Ccr), and enhances the total effective rate of symptom and sign improvement³⁹. Research based on modern biotechnology shows that the

Table 2 The influence of steamed Rhubarb on the pharmacokinetics of its main chemical components

Pharmacokinetic parameters	$AUC_{(0-t)}$ ($\text{mg}\cdot\text{L}^{-1}\cdot\text{h}^{-1}$)	$AUC_{(0-\infty)}$ ($\text{mg}\cdot\text{L}^{-1}\cdot\text{h}^{-1}$)	$MRT_{(0-t)}$ (h)	$MRT_{(0-\infty)}$ (h)	$T_{1/2z}$ (h)	T_{\max} (h)	$V_z\cdot F$ ($\text{L}\cdot\text{kg}^{-1}$)	$CL_z\cdot F$ ($\text{L}\cdot\text{h}\cdot\text{kg}^{-1}$)	C_{\max} ($\mu\text{g}\cdot\text{L}^{-1}$)	References
Rhein	*	*	**	*	**	**	**	**	**	12, 14
Rhein glucuronide	**	**	**	*			*	**	*	14
Rhein-8-O- β -D-glucoside	**	**	**	**	*	*	**	**	**	12, 14
Emodin	**	**	*	**	**	**		*	*	12, 14
Emodin glucuronide	**	**	**	**	**			**	**	14
Aloe-emodin	**	*	*	**	**	**	**	**	*	12, 14
Aloe-emodin glucuronide	**	**					**	**	**	14
Chrysophanol	*	*					**	*	**	12, 14
Chrysophanol glucuronide	**	**					**	**	**	14
Physcion	**		**	**	**		**	**		12, 14

Note: The red block indicates an increase in value compared with raw Rhubarb, the blue block indicates a decrease in value compared with raw Rhubarb, and the gray block indicates no significant difference in value compared with raw Rhubarb. * represents $P < 0.05$ vs raw Rhubarb, ** represents $P < 0.01$ vs raw Rhubarb.

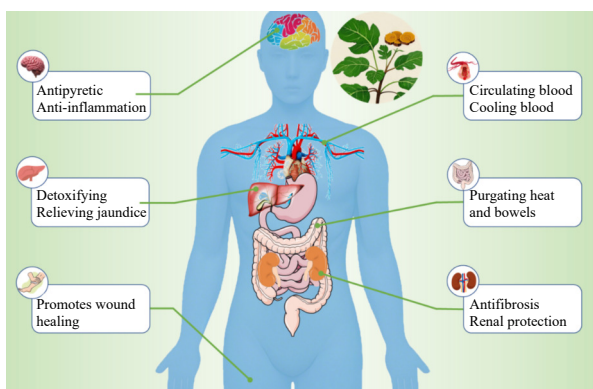


Fig. 3 Traditional applications of Rhubarb. Rhubarb has been widely used in traditional medicine. Its typical applications include promoting blood circulation, cooling the blood, purging heat and relieving constipation, exerting antifibrotic and renoprotective effects, providing antipyretic and anti-inflammatory activities, detoxifying and relieving jaundice, and promoting wound healing.

main active components of Rhubarb exert a synergistic anti-renal fibrosis effect by regulating abnormal ECM accumulation, controlling inflammatory factor release, and maintaining coagulation-fibrinolysis balance⁴⁰. Additionally, Rhubarb and Astragalus are often used together to treat various types of CKD⁴¹⁻⁴³. These reports further validate the traditional therapeutic value of Rhubarb in kidney diseases.

4.6. Topical applications

Externally, Rhubarb-based liniments address traumatic injuries and dermatitis; its anthraquinones inhibit microbial growth, while tannins reduce exudation in eczema^{44,45}. In medieval Europe, Rhubarb root was a prized ingredient in wound salves, paralleling its Eastern uses despite differing theoretical frameworks.

5. The main mechanism of Rhubarb against CKD

The multi-target therapeutic potential of Rhubarb in CKD stems from its synergistic modulation of inflammation, oxidative stress, autophagy, fibrosis, and gut-kidney interactions. These mechanisms collectively preserve renal architecture, delay functional deterioration, and interrupt the vicious cycle driving fibrosis. While clinical validation is ongoing, Rhubarb's pleiotropic actions position it as a promising adjunctive strategy for managing CKD progression.

5.1. Anti-inflammatory effects

Chronic inflammation is a hallmark of CKD progression, driving renal fibrosis and functional decline^{46,47}. Rhubarb contains bioactive compounds such as emodin and rhein that suppress pro-inflammatory signaling pathways, including nuclear factor κB (NF- κB) and mitogen-activated protein kinase (MAPK) cascades⁴⁸. By inhibiting the release of cytokines like TNF- α , interleukin-6 (IL-6), and IL-1 β , Rhubarb mitigates tubulointerstitial inflammation and reduces immune cell infiltration. This anti-inflammatory action alleviates endothelial dysfunction and glomerular damage, thereby slowing the transition from renal injury to fibrosis²². Its ability to modulate macrophage polarization toward an anti-inflammatory phenotype further underscores its role in disrupting the inflammatory microenvironment of diseased kidneys⁴⁹.

5.2. Antioxidant stress modulation

Oxidative stress exacerbates renal injury by promoting cellular apoptosis, mitochondrial dysfunction, and ECM deposition⁵⁰. Rhubarb-derived anthraquinones and polyphenols scavenge reactive oxygen species (ROS) while enhancing endogenous antioxidant defenses via nuclear factor erythroid 2-related factor 2 (Nrf2)/ARE pathway activation^{51,52}. This dual action reduces lipid peroxidation, deoxyribonucleic acid (DNA) damage, and protein carbonylation in renal tubular cells. By preserving redox homeostasis, Rhubarb attenuates oxidative stress-mediated podocyte loss and glomerulosclerosis, key contributors to CKD progression.

5.3. Autophagy regulation

Dysregulated autophagy in CKD leads to the accumulation of damaged organelles and protein aggregates, accelerating tubular atrophy and interstitial fibrosis^{53,54}. Rhubarb compounds modulate autophagy flux by balancing mammalian target of rapamycin (mTOR) and AMP-activated protein kinase (AMPK) signaling pathways⁵⁵. Enhanced autophagy clearance removes cytotoxic debris, mitigates endoplasmic reticulum stress, and inhibits apoptosis in renal parenchymal cells⁵⁶. This mechanism is particularly critical in diabetic nephropathy, where autophagic impairment drives hyperglycemia-induced renal injury.

5.4. Anti-fibrotic actions

Renal fibrosis, characterized by ECM deposition, represents the terminal pathway of CKD. Rhubarb targets TGF- β /Smad3 signaling, a central axis in fibroblast activation and ECM synthesis⁵⁷.

Emodin suppresses myofibroblast differentiation by downregulating α -smooth muscle actin (α -SMA) and collagen I/III expression, while rhein inhibits EMT in tubular cells⁵⁸. Additionally, Rhubarb disrupts crosstalk between angiotensin II and pro-fibrotic mediators, preventing glomerular basement membrane thickening and peritubular capillary rarefaction⁵⁹.

5.5. Gut-kidney axis modulation

The core view of the “gut-kidney axis” theory holds that during CKD progression, intestinal microecosystem disorders occur, which aggravate the accumulation of enterogenic uremic toxins in the blood, further decrease renal function, and eventually form a vicious cycle between the intestine and kidney⁶⁰. Emerging evidence highlights Rhubarb’s role in reshaping gut microbiota composition, which influences uremic toxin production and systemic inflammation in CKD⁶¹. By promoting the growth of short-

chain fatty acid (SCFA)-producing bacteria and reducing indoxyl sulfate and p-cresol levels, Rhubarb alleviates enteric dysbiosis-induced renal oxidative stress⁶². Its mild laxative properties also mitigate nitrogenous waste retention, indirectly lowering the metabolic burden on compromised kidneys⁶³.

6. Core signaling pathways involved in renal fibrosis

Renal fibrosis is a common mechanism leading to the progression of kidney disease from various etiologies to end-stage renal disease. In pathological conditions, the interaction between renal cells and matrix is disrupted, and inflammatory cell infiltration and intrinsic cell transformation lead to increased expression of pro-fibrotic cytokines⁶⁴. Abundant ECM deposition in the renal interstitium is the main cause of renal fibrosis. Multiple signaling pathways contribute to fibrogenesis, with the following being central players (Fig. 4).

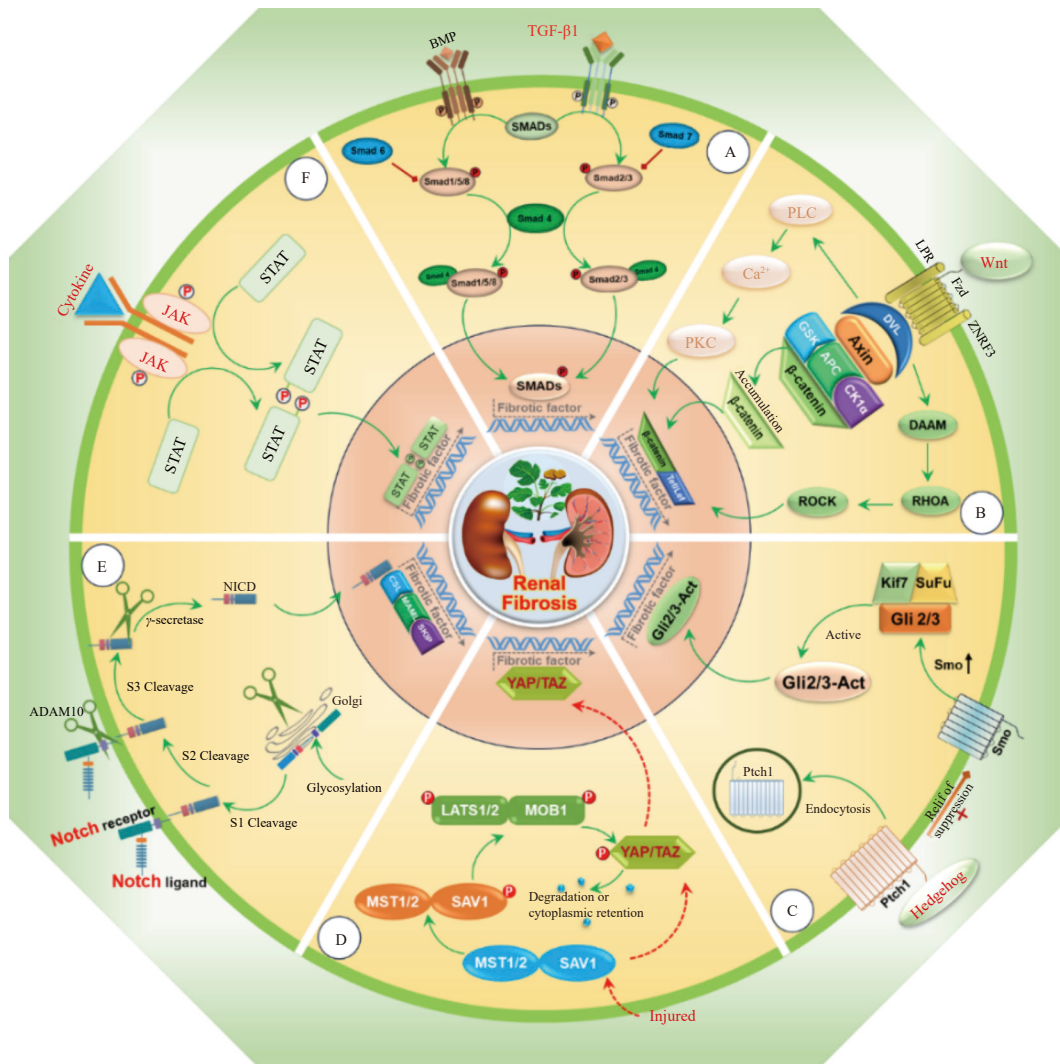


Fig. 4 Core signaling pathways involved in renal fibrosis. Multiple signaling pathways contribute to fibrogenesis, the central players including: TGF- β /Smad signaling (A), Wnt/ β -catenin signaling (B), Hedgehog signaling (C), Hippo/YAP signaling (D), Notch signaling (E), and JAK/STAT signaling (F).

6.1. TGF- β /Smad signaling

The TGF- β signaling pathway consists of TGF- β 1, TGF- β 2, and TGF- β 3, among which TGF- β 1 is the core molecule. The inactive form of TGF- β 1 is the TGF- β 1/leucine aminopeptidase (LAP) complex. Upon stimulation by injury, LAP dissociates, releasing active TGF- β 1⁶⁵. TGF- β ligands then bind to type II receptors, which phosphorylate type I receptors, activating downstream Smad2/3 proteins. Phosphorylated Smad2/3 complexes with

Smad4 translocate to the nucleus, driving transcription of pro-fibrotic genes (e.g., collagen, α -SMA)⁶⁶. Non-canonical pathways, including MAPK and phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt), further amplify ECM production. In renal fibrosis, TGF- β promotes EMT, fibroblast activation, and sustained inflammation, creating a vicious cycle of tissue scarring⁶⁷. Recent research has found that Taohe Chengqi Decoction, with Rhubarb as its core ingredient, significantly alleviates chronic renal failure in a 5/6 nephrectomy model by regulating the TGF- β

pathway. Further studies suggest that rhein is one of the main active components responsible for this effect⁶⁸.

6.2. Wnt/ β -catenin signaling

The Wnt signaling pathway is widely present in invertebrates and vertebrates and is highly conserved during species evolution. The canonical pathway begins when Wnt proteins bind to the cysteine-rich domain of frizzled (Fz) family receptors⁶⁹. This binding inhibits β -catenin degradation, leading to its accumulation⁷⁰. Stabilized β -catenin enters the nucleus and activates target genes such as fibronectin (FN) and matrix metalloproteinases (MMPs). In damaged kidneys, aberrant Wnt/ β -catenin signaling enhances fibroblast proliferation, disrupts tubular repair, and exacerbates ECM deposition⁷¹. Studies have shown that the Wnt/ β -catenin pathway is activated in the db/db mouse model of diabetic nephropathy. After 12 weeks of treatment with rhein, renal function is significantly improved through inhibition of this pathway⁷².

6.3. Notch signaling

The Notch signaling pathway comprises Notch receptors, ligands, CBF-1-suppressor of hairless-Lag (CSL) DNA-binding proteins, and regulatory molecules. Signaling is initiated by ligand-receptor interaction between adjacent cells. The Notch protein undergoes three proteolytic cleavages, releasing the Notch intracellular domain (NICD), which translocates to the nucleus and binds to the CSL transcription factor⁷³. In renal fibrosis, Notch hyperactivation in tubular cells induces EMT and inflammation while suppressing regeneration⁷⁴. Dysregulated Notch also promotes pericyte-to-myofibroblast differentiation, directly fueling interstitial fibrosis⁷⁵. Many compounds in Rhubarb, such as emodin and resveratrol, have been confirmed to regulate the Notch signaling pathway^{76, 77}, indicating Rhubarb's potential to ameliorate renal fibrosis *via* this pathway.

6.4. Hedgehog (Hh) signaling

There are three homologous Hh genes in mammals: Sonic Hh (SHH), Indian Hh (IHH), and Desert Hh (DHH), which encode SHH, IHH, and DHH proteins, respectively. Hh signaling is controlled by two receptors, Patched (Ptc) and Smoothed (Smo), on the target cell membrane. Hh ligands bind to Ptc, relieving its suppression on Smo⁷⁸. This activates Gli transcription factors, which regulate genes involved in fibroblast activation and ECM synthesis⁷⁹. In fibrotic kidneys, Hh signaling is upregulated in interstitial fibroblasts, driving myofibroblast differentiation and collagen accumulation⁸⁰. In unilateral ureteral obstruction (UUO) rats, treatment with resveratrol, an active component of Rhubarb, significantly antagonizes excessive activation of the Hh pathway, resulting in reduced cell proliferation, EMT, and ECM accumulation⁸¹.

6.5. Janus kinase (JAK)/signal transducer and activator of transcription (STAT) signaling

The JAK/STAT signaling pathway is a key cytokine-mediated signal transduction mechanism⁸². Its molecular conduction process includes four stages: ligand-receptor binding, JAK kinase activation, STAT phosphorylation and dimerization, and nuclear translocation for transcriptional regulation of target genes. When cytokines such as interferon and interleukin bind to receptors, receptor dimerization triggers self-phosphorylation of JAK family kinases⁸³. Activated JAK recruits and phosphorylates STATs, forming homodimers or heterodimers. These activated STATs translocate to the nucleus to regulate transcription of pro-inflammatory and ECM-related genes, ultimately accelerating fibrosis⁸⁴.

It has been reported that emodin-8- β -D-glucoside binds to JAK1/2, thereby inhibiting STAT activation and the associated inflammatory response⁸⁵.

6.6. Hippo/YAP signaling

The Hippo/YAP pathway is one of the core regulators of organ fibrosis. Under physiological conditions, mammalian sterile 20-like kinases 1/2 (MST1/2) and salvador homolog 1 (SAV1) form a complex that phosphorylates large tumor suppressors 1/2 (LATS1/2) and MOB kinase activator 1 (MOB1)⁸⁶. The latter further phosphorylates the downstream effectors YAP/TAZ. Phosphorylated YAP/TAZ are either ubiquitinated and degraded or bind to 14-3-3 proteins and remain in the cytoplasm⁸⁷. Conversely, under inflammatory or injurious stimuli, the upstream kinase cascade is disrupted, leading to YAP/TAZ dephosphorylation and nuclear translocation, which induces expression of fibrotic factors⁸⁸. Research suggests that oral administration of emodin activates YAP phosphorylation and promotes its intracellular degradation, thereby reducing oxidative stress damage mediated by nuclear entry⁸⁹. This provides evidence for Rhubarb's regulation of the Hippo/YAP pathway.

6.7. Other Pathways

Additional pathways include MAPK/ERK signaling (activated by oxidative stress or cytokines, promoting fibroblast proliferation) and peroxisome proliferator-activated receptor (PPAR) signaling (its downregulation exacerbates lipid metabolism dysfunction, linked to fibrosis progression). These interconnected pathways form a complex network driving renal fibrosis⁹⁰. Targeting key nodes or their crosstalk holds therapeutic potential. However, pathway-specific modulation remains challenging due to context-dependent roles in tissue repair versus fibrosis.

7. Major chemical constituents of Rhubarb and their effects on renal fibrosis

The chemical diversity of Rhubarb underpins its multifaceted pharmacological profile. Modern pharmacological studies have shown that Rhubarb contains various chemical components, including anthraquinones, flavonoids, anthrones, chromones, tannins, stilbenes, and others (Fig. 5). These components exhibit

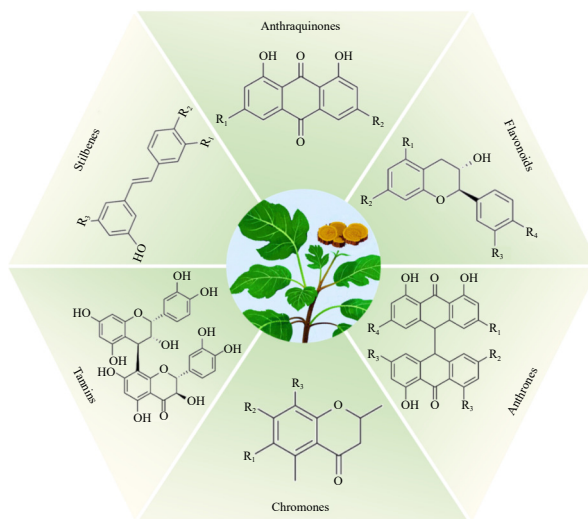


Fig. 5 Major chemical constituents of Rhubarb. The main chemical components of Rhubarb include: anthraquinones, flavonoids, anthrones, chromones, tannins, stilbenes, and so on. This figure lists the basic skeleton structure or representative compound of each class of compounds.

strong therapeutic effects in the treatment of CKD and renal fibrosis (Table 3).

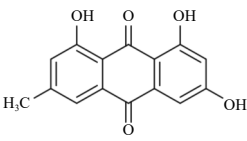
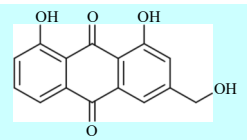
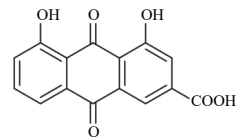
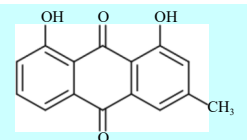
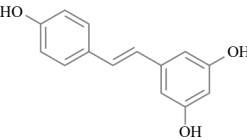
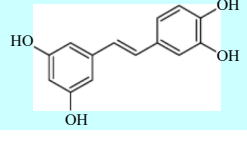
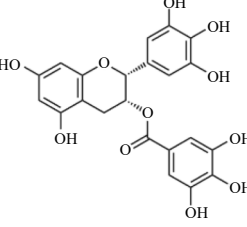
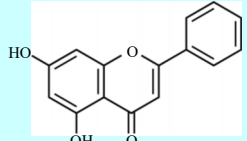
7.1. Anthraquinone derivatives

Anthraquinones are the most prominent and pharmacologically active compounds in Rhubarb, responsible for its characteristic laxative effects⁹¹. These compounds exist in both free forms (aglycones, e.g., emodin, rhein, aloe-emodin) and conjugated

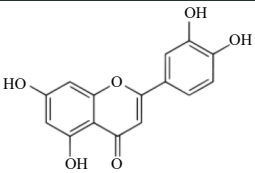
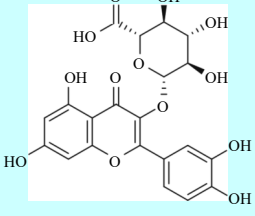
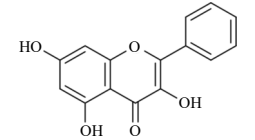
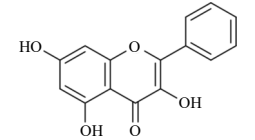
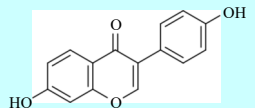
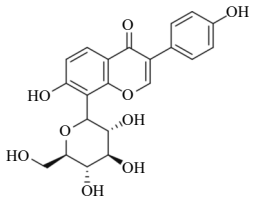
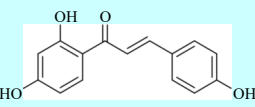
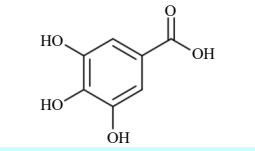
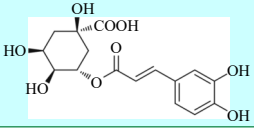
forms (glycosides, e.g., sennosides A–D). Structurally, they feature a 9,10-anthracenedione core with hydroxyl, methyl, or carboxyl substitutions⁹². Sennosides act as prodrugs metabolized by gut microbiota into active anthrones, stimulating colonic motility⁹³. Beyond their purgative properties, anthraquinones exhibit anti-microbial⁹⁴, anti-inflammatory⁹⁵, and anti-tumor activities⁹⁶.

Emodin (C₁₅H₁₀O₅, 1,3,8-trihydroxy-6-methylantraquinone) is a lipophilic anthraquinone compound isolated from Rhubarb,

Table 3 The chemical composition of Rhubarb and its effect on renal fibrosis.

Chemical structure	Name	Model	Mechanisms	Ref
	Emodin	<i>In vivo</i> : UUO rats	Regulating PGC-1 α -mediated mitochondria function and energy homeostasis	97
<i>In vitro</i> : TGF- β -induced NRK-52E				
<i>In vivo</i> : adriamycin-induced rat RF with unilateral nephrectomy		Alleviating EMT by actuating autophagy through BMP-7/TGF- β signaling	98	
<i>In vitro</i> : TGF- β -induced HK-2 cells				
<i>In vivo</i> : 5/6 renal mass reduction		Down-regulating expressions of TGF- β and Smurf 2 and up-regulating Smad7 expression	99	
<i>In vivo</i> : UUO mouse				
<i>In vitro</i> : TGF- β -induced HK-2 cells	Regulating HGF and TGF- β /Smad Signaling Pathway.	100		
	Aloe-emodin	<i>In vivo</i> : UUO mouse	Inhibiting PI3K/Akt/mTOR pathway	101
<i>In vitro</i> : TGF- β -induced NRK-49F cells				
	Rhein	<i>In vivo</i> : UIRI rats	Regulating the expression of MAPK and AKT in the NF- κ B and MAPK signaling pathways	102
		<i>In vitro</i> : TGF- β -induced HK-2 cells		
		<i>In vivo</i> : UIRI rats	Activating the PPAR α -CPT1A axis	103
		<i>In vitro</i> : TGF- β and oleic acid induced HK-2 and NRK-52E cells		
		<i>In vivo</i> : UUO rats	Regulating the SirT1/STAT3/Twist1 pathway	104
		<i>In vitro</i> : TGF- β -induced primary rat renal tubular epithelial (RTE) cells		
		<i>In vivo</i> : UUO rats	Regulating the SHH-Gli1-Snai1 signal pathway	105
<i>In vitro</i> : SHH-activated NRK-49F cells				
	Chrysophanol	<i>In vivo</i> : UUO rats	Inhibiting p-STAT3-mediated apoptosis	106
		<i>In vitro</i> : UUO mouse		
		<i>In vitro</i> : TGF- β induced HK2 and HEK293 cells	Regulating aberrant methyltransferase 1/3a expressions in preventing Klotho loss	107
		<i>In vivo</i> : UUO mouse		
		<i>In vitro</i> : TGF- β -induced HK-2 cells	Inhibiting the TGF- β /Smad signaling pathway	108
<i>In vitro</i> : TGF- β -induced HK-2 cells	Suppressing NKD2/NF- κ B pathway	109		
	Resveratrol	<i>In vivo</i> : UUO rats	Inactivating of Smad2/3 signaling	117
		<i>In vitro</i> : TGF- β -induced HK-2 cells		
		<i>In vivo</i> : 5/6Nx rats	Inhibiting TGF- β and CTGF	118
		<i>In vivo</i> : db/db mice		
		<i>In vitro</i> : high glucose-induced NRK-49F cells	Regulating of AMPK/NOX4/ROS signaling	119
		<i>In vivo</i> : UUO mouse		
		<i>In vitro</i> : aristolochic acid or TGF- β -induced HK-2 cells	Deacetylation of Smad4 and inhibition of MMP	120
<i>In vivo</i> : UUO mouse				
<i>In vitro</i> : TGF- β -induced HK-2 cells	Decreased EMT via Sirt1-dependent deacetylation of Smad3/Smad4	121		
	Picetannol	<i>In vivo</i> : UUO mouse	Inhibiting HDAC4/5 and the p38 MAPK signaling	122
	Epigallocatechin-3-gallate (EGCG)	<i>In vitro</i> : arsenic-induced Caki-1 and HK-2 cells	Antioxidant and epigenetic regulatory abilities	126
		<i>In vitro</i> : high glucose induced HPC	Inhibiting the NF- κ B pathway	127
		<i>In vitro</i> : pcDNA6.2-SNAI1 cells transfected with renal TEPEs	Inhibiting SNAI1 overexpression and promoting the process from MET to EMT	128
		<i>In vitro</i> : TGF- β induced HK-2 cells	Deactivating the GSK-3 β / β -catenin/Snai1 pathway and activating the Nrf2 pathway	129
		<i>In vitro</i> : TGF- β induced NRK-52E cells	Inhibiting EMT and fibrotic proteins expression by Nrf2 activation	130
		<i>In vivo</i> : UUO mouse	Inhibiting of TGF- β /Smad signaling pathway	131
	Chrysin (5,7-dihydroxyflavone)	<i>In vivo</i> : Cyclosporin A induced rats	Inhibiting the activation of TGF- β /Smad 3/Snai1 and Akt/GSK-3 β pathways	133
		<i>In vitro</i> : TGF- β or CsA induced LLC-PK1 cells		
		<i>In vivo</i> : db/db mice	Inhibiting renal tubular EMT	134
<i>In vitro</i> : High glucose induced RPTECs				

Continued

Chemical structure	Name	Model	Mechanisms	Ref
	Luteolin	<i>In vivo</i> : Aristolochic acid induced mice <i>In vitro</i> : Hypoxia/TGF- β or CsA induced NRK49F cells	Activating the SIRT1/FOXO3 pathway	136
	Quercetin	<i>In vivo</i> : UUO rats	Inhibiting the activation of AREG/EGFR pathway	137
		<i>In vitro</i> : TGF- β -induced HK-2 cells	Inhibiting the activation of NLRP3 inflammatory bodies	138
		<i>In vivo</i> : UUO rats		
		<i>In vitro</i> : TGF- β -induced HK-2 cells	Inhibiting IL-33/ST2 pathway	139
		<i>In vivo</i> : MRL/lpr mice		
<i>In vitro</i> : IL-33 induced HK-2 cells	Inhibiting FAO pathway by upregulating PPAR α	140		
<i>In vivo</i> : db/db mice				
<i>In vitro</i> : High glucose induced TECs	Inhibiting the PI3k/Akt pathway	142		
<i>In vivo</i> : adenine induced CRF rats				
	Rutin	<i>In vivo</i> : UUO rats	Activating the SIRT1/PINK1/Parkin-mediated mitophagy	143
		<i>In vitro</i> : AngII-treated RTECs		
	Rutin	<i>In vivo</i> : 5/6 Nx rats	Inhibiting of TGF- β /Smad signaling pathway	144
	Daidzein	<i>In vivo</i> : VOX + UUO rats	Regulating the expression of miR-33a and miR-27a	147
	Puerarin	<i>In vivo</i> : II DN rats	Regulating iron homeostasis and alleviating iron deprivation anemia	148
		<i>In vitro</i> : High glucose induced GMCs	Inhibiting TLR4/Nox4 pathway	149
		<i>In vivo</i> : renal I/R rats		
		<i>In vitro</i> : HR induced HK-2 cells	Inhibiting the cAMP/PKA/CREB pathway in podocytes activated via Gnai1	150
		<i>In vivo</i> : db/db mice		
<i>In vitro</i> : High glucose induced podocytes	Reducing renal injury and pyroptosis by targeting the miR-342-3p/TGF- β /SMAD axis	151		
<i>In vivo</i> : Adenine induced CRF rats				
<i>In vitro</i> : LPS induced HK-2 cells	Inhibiting the NF- κ B p65/STAT3 and TGF- β /Smads pathways	152		
<i>In vivo</i> : UUO mice				
	Isoliquiritigenin	<i>In vivo</i> : UUO mice	Inhibiting oxidative stress induced-epithelial cell apoptosis through MAPK signaling	153
		<i>In vitro</i> : H ₂ O ₂ induced HK-2 cells		
		<i>In vivo</i> : UUO mice	Inhibiting the Mincle/Syk/NF- κ B signaling pathway	154
		<i>In vitro</i> : LPS/TGF- β -induced bone marrow-derived macrophages (BMDM)		
		<i>In vivo</i> : Adenine induced CKD rats	Activating sGC, increasing cGMP and its downstream PKG	155
<i>In vitro</i> : glycation end-products (AGEs) induced HK-2 cells	Inhibiting the EMT and suppressing the TGF- β /STAT3 signaling	156		
<i>In vivo</i> : STZ-induced diabetes nephropathy rats	Inhibiting the JAK2/STAT3 signaling pathway	157		
<i>In vitro</i> : High glucose induced HK-2 cells				
<i>In vivo</i> : STZ-induced type 1 DN	Direct binding to SIRT1, the inhibition of MAPK activation, and the induction of Nrf-2 signaling	158		
<i>In vitro</i> : high glucose-induced NRK-52E cells				
	Gallic acid	<i>In vivo</i> : glyoxal induced renal fibrosis rats	ROS quenching and anti-glycation activity	162
	Chlorogenic acid	<i>In vivo</i> : Kidney ischemic/reperfusion (I/R) injury rats	Reducing inflammation, tubular injury, and myofibroblast formation	163
		<i>In vivo</i> : hyperuricemia nephropathy rats	Inhibiting the PI3K/AKT/mTOR signaling	164

which has been shown to alleviate fibrosis in multiple organs. Administration of emodin in renal fibrosis models significantly inhibits PGC-1 α -mediated lipid accumulation and mitochondrial biogenesis, indicating that emodin regulates mitochondrial function and energy homeostasis through PGC-1 α , thereby inhibiting renal fibrosis⁹⁷. Studies have also found that in renal fibrosis rats and cell models treated with emodin, upregulation of BMP-7 is observed, accompanied by activation of autophagy and inhibition of EMT. This suggests that emodin activates autophagy *via* the BMP-7 pathway, thereby reducing EMT in renal fibrosis⁹⁸. Emodin significantly improves pathological abnormalities and renal function in renal fibrosis, and its mechanism may involve upregu-

lation of Smad7 and inhibition of TGF- β /Smad signaling-related Smurf2 and ECM components⁹⁹. Additionally, another study found that the combination of emodin and HGF exhibits better anti-fibrotic effects than HGF alone, and this mechanism is related to the regulation of the TGF- β /Smad signaling pathway¹⁰⁰.

Aloe-emodin (C₁₅H₁₀O₅, 1,8-dihydroxy-3-hydroxymethylanthraquinone) is another common anthraquinone in Rhubarb, which alleviates histopathological and renal dysfunction in UUO mice by inhibiting the PI3K/Akt/mTOR pathway. Administration of the PI3K inhibitor wortmannin or siRNA-PI3K counteracts the renal protective effect of aloe-emodin¹⁰¹.

Rhein (C₁₅H₈O₆, 1,8-dihydroxy-3-carboxy anthraquinone) is

often used as a raw material for synthesizing the arthritis drug diclofenac. A metabolomics-based study found that rhein's alleviation of renal fibrosis is closely associated with sphingolipid, arachidonic acid, and glycerophospholipid metabolism pathways, and its molecular mechanism may involve modulation of NF- κ B and MAPK signaling pathways¹⁰². Xiao et al. developed a new integrated analysis method combining network pharmacology and metabolomics, and identified the PPAR- α -CPT1A-1-palmitoyl-carnitine axis as a potential target of rhein in anti-renal fibrosis. This result was validated in UIRI rats and TGF- β /oleic acid-induced NRK-52E and HK-2 cells¹⁰³. Based on combined transcriptomics and metabolomics analyses, rhein may exert its anti-renal fibrosis effect by regulating EMT. Cellular studies have confirmed that rhein promotes fatty acid oxidation (FAO) by regulating the Sirt1/STAT3/ Twist1 pathway, thereby inhibiting trans-differentiation of renal tubular epithelial cells (TECs)¹⁰⁴. In the UUO rat model, rhein treatment significantly alleviates renal fibrosis, possibly through inhibition of the SHH-Gli1-Snail signaling pathway, a finding confirmed in SHH-induced NRK-49F cells¹⁰⁵. Similar studies suggest that rhein mitigates renal fibrosis by restraining p-STAT3-mediated apoptosis¹⁰⁶. Additionally, in the UUO-induced renal fibrosis model, abnormal expression of methyltransferases 1 and 3a leads to hypermethylation of the promoter of the endogenous protective factor Klotho, and rhein can correct these epigenetic abnormalities, exerting a renal protective effect¹⁰⁷.

Chrysophanol (C₁₅H₁₀O₄, 1,8-dihydroxy-3-methyl-anthraquinone) is an anthraquinone isolated from Rheum, exhibiting various pharmacological activities. Treatment with chrysophanol in UUO mice reverses abnormal serum and urine biochemical parameters and reduces expression of fibrosis markers such as FN, collagen I/III, and α -SMA. Rhein also inhibits the secretion of TGF- β and p-Smad3 while promoting Smad7. Mechanistic studies suggest that chrysophanol alleviates fibrosis by blocking the crosstalk between Smad3 and TGF- β receptor I¹⁰⁸. In the same renal fibrosis models, chrysophanol significantly inhibits NF- κ B activation and NKD2 expression, and reduces inflammatory factors such as IL-1 β , IL-6, and TNF- α . Molecular docking and microscale thermal analysis indicate that NKD2 is a direct target of chrysophanol. Therefore, chrysophanol improves the fibrotic state by directly inhibiting NKD2¹⁰⁹. However, the poor water solubility and unstable physical properties of chrysophanol limit its efficacy. To address this, Wei et al. synthesized *N*-octyl-*O*-sulfate chitosan and developed nanoparticles loaded with chrysophanol. Their research shows that compared to free chrysophanol, the bioavailability of chrysophanol nanoparticles is 2.57 times higher, and its anti-renal fibrosis effect is enhanced¹¹⁰.

7.2. Stilbenes

Stilbenes, though less abundant than anthraquinones, are notable for their bioactive potential. The primary stilbene in Rhubarb is *trans*-resveratrol and its glycoside, polydatin (resveratrol-3-*O*- β -D-glucoside). These compounds feature a C6-C2-C6 structure with a central ethylene bond¹¹¹. Stilbenes are renowned for their cardioprotective, neuroprotective, and anti-aging properties¹¹², largely attributed to sirtuin pathway activation and suppression of pro-inflammatory cytokines¹¹³. Additionally, recent studies highlight their role in improving insulin sensitivity¹¹⁴. Their presence underscores Rhubarb's potential in managing chronic metabolic¹¹⁵ and degenerative diseases¹¹⁶.

Resveratrol (C₁₄H₁₂O₃, 3,5,4'-trihydroxy-*trans*-stilbene), a non-flavonoid polyphenolic compound, is an antitoxin produced by various plants under stress. Resveratrol suppresses myofibroblast phenotype and fibrosis formation in UUO kidneys by targeting EMT and fibroblast-to-myofibroblast transition (FMT), which is associated with inactivation of Smad2/3 signaling¹¹⁷. In 5/6 nephrectomy (5/6Nx) rats, resveratrol alleviates fibrosis by in-

hibiting TGF- β and CTGF, while also improving CKD-associated muscle atrophy¹¹⁸. Research has shown that resveratrol significantly inhibits high glucose-induced fibroblast proliferation and activation by reducing NOX4-derived ROS production. Further *in vivo* studies confirmed that resveratrol alleviates renal fibrosis in db/db mice by increasing p-AMPK and inhibiting NOX4¹¹⁹. Xiao et al.¹²⁰ found that resveratrol inhibits the EMT process to alleviate kidney injury and fibrosis, attributed to upregulation of SIRT1, leading to deacetylation of Smad4 and inhibition of MMP7 expression¹²⁰. Moreover, resveratrol's effect on renal fibrosis is dose-dependent. At low doses (≤ 25 mg·kg⁻¹), resveratrol reduces TGF- β -induced EMT *via* Sirt1-dependent Smad3/Smad4 deacetylation, exerting anti-fibrotic effects. However, at high doses (≥ 50 mg·kg⁻¹), resveratrol induces mitochondrial oxidative stress and ROCK1-mediated cytoskeletal disruption, promoting EMT in HK-2 cells and exacerbating fibrosis. Therefore, the risk of resveratrol use in CKD patients should be carefully considered¹²¹.

Picetannol (C₁₄H₁₂O₄, 3,3',4,5'-tetrahydroxy-*trans*-stilbene) is a metabolite of resveratrol, found in rhubarb, red wine, and grapes. In UUO mice, picetannol significantly inhibits ECM deposition. Mechanistic research indicates that picetannol may exert its effect by inhibiting HDAC4/5 expression and p38 MAPK signaling activation, rather than through the TGF- β /Smad2/3 pathway¹²².

7.3. Flavonoids

Rhubarb contains diverse flavonoids, including flavonols (e.g., quercetin, kaempferol) and flavan-3-ols (e.g., catechin, epicatechin). These compounds are characterized by a C6-C3-C6 skeleton with hydroxyl, glycosyl, or methoxy substitutions¹²³. Flavonoids contribute to Rhubarb's antioxidant capacity by scavenging free radicals and chelating metal ions. They also modulate inflammatory pathways by inhibiting cyclooxygenase (COX) and lipoxygenase (LOX) enzymes¹²⁴. Additionally, certain flavonoids enhance vascular integrity and exhibit cardiovascular protective effects, making them valuable in treating oxidative stress-related disorders¹²⁵. Their synergistic interactions with anthraquinones may mitigate oxidative damage caused by the latter.

7.3.1. Flavonoids (flavan-3-ols)

Epigallocatechin-3-gallate (EGCG, C₂₂H₁₈O₁₁, (2R,3R)-5,7-dihydroxy-2-(3,4,5-trihydroxyphenyl)-3,4-dihydro-2H-chromen-3-yl 3,4,5-trihydroxybenzoate), a green tea polyphenol, is known for its strong antioxidant activity. Iheanacho et al. investigated whether EGCG alleviates arsenic-induced renal fibrosis in TECs. The results indicate that EGCG significantly alleviates arsenic-induced cytotoxicity and abnormal expression of fibrogenic growth factors through its antioxidant and epigenetic regulatory abilities¹²⁶. In the high glucose-treated human podocyte cell line HPC, the actin $\alpha 4$ gene promoter is highly methylated. EGCG treatment reverses this hypermethylation, upregulates ACTN4 levels, downregulates DNA methyltransferase 1 (DNMT1), and inhibits the NF- κ B pathway and its mediated fibrotic pathology¹²⁷. The dynamic trans-differentiation of epithelial cells from EMT to mesenchymal-epithelial transition (MET) has attracted widespread attention in fibrosis treatment. Research has shown that EGCG inhibits SNAI1 overexpression and promotes the MET process, providing a novel strategy for preventing renal fibrosis¹²⁸. In TGF- β -induced HK-2 cells, EGCG pretreatment significantly eliminates the EMT process and reduces fibroblast and ECM deposition. This mechanism is related to EGCG deactivating the GSK-3 β / β -catenin/Snail1 pathway and activating the Nrf2 pathway^{129, 130}. Additionally, studies suggest that EGCG's anti-renal fibrosis effect is highly correlated with inhibition of inflammatory response¹³¹ and cell apoptosis¹³².

Chrysin ($C_{15}H_{10}O_4$, 5,7-dihydroxyflavone) is a raw material for synthesizing drugs with anti-cancer, lipid-lowering, cardio-cerebrovascular disease prevention, anti-bacterial, and anti-inflammatory properties. Chrysin treatment alleviates cyclosporin A-induced renal dysfunction in rats and reduces renal tubular injury and collagen deposition by inhibiting the TGF- β /Smad3/Snail and Akt/GSK-3 β pathways¹³³. In diabetic renal tubulointerstitial fibrosis, chrysin inhibits high glucose-induced renal EMT by suppressing vimentin, α -SMA, and FSP-1 expression in human proximal TECs and db/db mice¹³⁴. To address chrysin's poor water solubility, Kang et al. synthesized 5,7-di-*O*-acetylchrysin, which has higher solubility than salicylic acid. In db/db mice, this derivative improved renal injury caused by glomerulosclerosis and tubulointerstitial fibrosis¹³⁵.

Luteolin ($C_{15}H_{10}O_6$, 3',4',5,7-tetrahydroxyflavon) is mostly present in glycosidic form in various plants, including whole leaf orchids, chili peppers, wild chrysanthemums, honeysuckle, and perilla. After treatment with luteolin in mice with renal fibrosis, renal function and fibrosis were significantly improved. Transcriptomic analysis revealed significant increases in SIRT1 and FOXO3, validated both *in vivo* and *in vitro*. Molecular docking confirmed strong direct binding between luteolin and SIRT1. Therefore, luteolin may alleviate renal fibrosis *via* the SIRT1/FOXO3 pathway¹³⁶.

7.3.2. Flavonoids (flavonols)

Quercetin ($C_{15}H_{10}O_7$, 3,5,7,3',4'-pentahydroxyflavonoid dihydrate) is a widely distributed and biologically active flavonoid that shows potential to inhibit fibrosis in various diseases. Recent research confirms that quercetin improves the fibrotic area in UUO rats and alleviates TGF- β -induced EMT in HK-2 cells. Mechanistic studies show that quercetin exerts anti-renal fibrosis effects by inhibiting the AREG/EGFR signaling pathway¹³⁷. Similar studies indicate that quercetin inhibits NLRP3 inflammasome activation in renal TECs both *in vivo* and *in vitro*, thereby reducing EMT and ECM accumulation¹³⁸. Tubulointerstitial fibrosis is also a key pathological feature in lupus nephritis progression. Quercetin administration significantly improves renal function in MRL/lpr mice and inhibits fibrosis and inflammatory responses. In the IL-33-induced HK-2 cell model, quercetin improves fibrosis and cell pyroptosis *via* the IL-33/ST2 pathway, suggesting its potential in treating lupus nephritis-associated renal fibrosis and inflammation¹³⁹. The combination of quercetin and dasatinib significantly improves fibrosis in db/db mice and reduces lipid deposition. Further investigation found that quercetin inhibits the FAO pathway by upregulating PPAR α . Knocking out PPAR α reverses this inhibition and weakens quercetin's renal protective effect. Molecular docking and dynamic simulation analysis indicate direct binding between quercetin and PPAR α ¹⁴⁰. To improve quercetin's water solubility, absorption, and bioavailability, Sánchez Jaramillo et al. developed a quercetin-nanoparticle formulation. Results showed that low-dose nanoparticles produced the same beneficial effect as high-dose quercetin in preventing kidney injury¹⁴¹. Additionally, quercetin reduces renal fibrosis in chronic renal failure rats by targeting the PI3K/Akt pathway and reduces water retention and toxin accumulation by regulating AQP1 and AQP2 expression¹⁴². Quercetin may also reduce renal TEC (RTEC) aging by activating SIRT1/PINK1/parkin-mediated mitophagy, thereby alleviating renal fibrosis¹⁴³.

Rutin ($C_{27}H_{30}O_{16}$, 3,3',4',5,7-pentahydroxyflavon-3-(*O*-rhamnosylglucoside)) is a natural antioxidant that scavenges ROS by donating hydrogen atoms to peroxides, superoxide anions, singlet oxygen, and hydroxyl radicals. After inducing chronic renal failure in adult Wistar rats *via* 5/6 nephrectomy, oral rutin treatment significantly improved renal function indicators, alleviated tubulointerstitial injury, and reduced glomerulosclerosis scores. This mechanism may involve rutin's antioxidant activity and in-

hibition of the Smad pathway¹⁴⁴. After four weeks of treatment in alloxan-induced diabetic nephropathy rats, rutin significantly reduced serum triglyceride and cholesterol levels, and histopathological results showed marked improvement in renal fibrosis¹⁴⁵.

7.3.3. Flavonoids (Isoflavones)

Daidzein ($C_{15}H_{10}O_4$, 7,4'-dihydroxyisoflavone) is a major isoflavone found in leguminous plants and grains such as soybeans, kudzu, clover, and alfalfa. Daidzein has two hydroxyl groups at the 7 and 4' positions, which are key to its antioxidant properties. Oophorectomized (OVX) rats subjected to UUO were used to simulate pathological damage in postmenopausal CKD. Daidzein treatment alleviates renal dysfunction, fibrosis, and cell apoptosis, especially when combined with losartan. The mechanism involves regulation of miR-33a and miR-27a expression and their interaction with angiotensin AT1 and Mas receptors^{146,147}.

Puerarin ($C_{21}H_{20}O_9$, 8-(β -*D*-glucopyranosyl)-4',7-dihydroxyisoflavone) is a flavonoid derivative originally isolated from Pueraria lobata, known for its coronary vasodilatory effect. It has been reported that puerarin inhibits ECM production by regulating iron homeostasis and alleviating ferroptosis, thereby reducing high glucose-induced cell damage, ROS production, and excessive collagen fiber accumulation, ultimately reducing renal fibrosis in diabetic nephropathy rats¹⁴⁸. Jian et al. also suggested that puerarin alleviates ferroptosis during renal fibrosis in a renal ischemia-reperfusion model by inhibiting the TLR4/NOX4 pathway¹⁴⁹. In the db/db mouse model of type 2 diabetic nephropathy, puerarin plays a renal protective role by inhibiting the cAMP/PKA/CREB pathway in podocytes activated *via* Gna11¹⁵⁰. In a rat model of chronic renal failure induced by adenine, puerarin treatment significantly alleviated renal injury and fibrosis while increasing miR-342-3p levels. Elevated miR-342-3p negatively regulates the TGF- β /Smad axis, inhibiting IL-1 β and IL-18 secretion and pyroptosis. This was confirmed in LPS-induced HK-2 cells¹⁵¹. Additionally, puerarin inhibits inflammatory factor recruitment and ECM deposition by regulating the NF- κ B/STAT3 and TGF- β /Smads pathways, thereby alleviating UUO-induced inflammation and fibrosis¹⁵². Zhou et al. found that puerarin improves renal fibrosis by inhibiting oxidative stress-induced epithelial cell apoptosis *via* the MAPK pathway¹⁵³.

7.3.4. Flavonoids (chalcones)

Isoliquiritigenin ($C_{15}H_{12}O_4$, 2',4,4'-trihydroxy-trans-chalcone) is a flavonoid initially isolated from licorice, reported to inhibit inflammation and fibrosis. In LPS/TGF- β -induced bone marrow-derived macrophages and UUO mice, isoliquiritigenin exerts renal protective effects by inhibiting the Mincle/Syk/NF- κ B signaling pathway, reducing α -SMA and Col III expression¹⁵⁴. Long-term treatment improves adenine-induced renal fibrosis by activating sGC and increasing cGMP and its downstream PKG¹⁵⁵. In diabetic patients, hyperglycemia-related advanced glycation end products (AGEs) can lead to renal fibrosis by increasing renal tubular EMT and ECM synthesis. Isoliquiritigenin treatment significantly inhibits the TGF- β /STAT3 pathway and its mediated collagen secretion in AGEs-induced HK-2 cells, alleviating EMT and fibrosis progression¹⁵⁶. In the STZ-induced diabetic nephropathy model, isoliquiritigenin improves renal fibrosis in rats by inhibiting the JAK2/STAT3 signaling pathway¹⁵⁷. Isoliquiritigenin also exerts anti-diabetic nephropathy and anti-fibrotic effects by directly binding to SIRT1, inhibiting MAPK activation, and inducing Nrf2 activation¹⁵⁸.

7.4. Other phytochemicals

Rhubarb contains hydrolyzable tannins, primarily gallotannins and galloyl glucoses, which are polymers of gallic acid bound to a glucose core. These compounds contribute to the astringent

taste of Rhubarb and possess astringent and anti-diarrheal properties, counterbalancing the laxative effects of anthraquinones¹⁵⁹. Tannins also exhibit anti-microbial activity by disrupting bacterial cell membranes and inhibiting enzyme function¹⁶⁰. Moreover, their antioxidant and anti-inflammatory effects make them useful in gastrointestinal protection and wound healing¹⁶¹. Interestingly, tannins may reduce the bioavailability of anthraquinones via complexation, highlighting the plant's inherent chemical equilibrium. Rhubarb also produces minor constituents such as chromones (e.g., rheumone), dianthrones, and polysaccharides. Chromones exhibit anti-allergic and vasodilatory effects, while dianthrones (dimers of anthrone units) display laxative activity similar to anthraquinones. Polysaccharides, including pectins and glucomannans, contribute to Rhubarb's immunomodulatory and prebiotic properties by promoting gut microbiota diversity. Additionally, organic acids (e.g., citric acid, malic acid) enhance the solubility and absorption of other bioactive compounds. These components collectively broaden Rhubarb's therapeutic spectrum beyond its classical applications.

Gallic acid (C₇H₆O₅, 3,4,5-trihydroxybenzoic acid) is a polyphenolic compound widely found in plants such as *Rheum palmatum*, *Eucalyptus grandiflorus*, and *Cornus officinalis*. It is widely used in the food, biological, pharmaceutical, and chemical industries. Yousuf et al. investigated the protective effect of gallic acid on experimental rat renal fibrosis induced by glyoxal. The results indicated that glyoxal induces renal fibrosis by enhancing ROS production via AGE receptors, while gallic acid significantly counteracts this effect through its ROS quenching and anti-glycation activity¹⁶².

Chlorogenic acid (CGA, C₁₆H₁₈O₉, 1,4,5-trihydroxycyclohexanecarboxylic acid 3-(3,4-dihydroxycinnamic acid)) is an organic compound commonly used in anti-bacterial and anti-viral treatments. In renal ischemia-reperfusion injury (IRI) mice, renal tubular injury was observed, with increased inflammatory mediators, macrophage count, and myofibroblast expansion. After treatment with CGA, the number of macrophages and myofibroblasts significantly decreased. These findings are related to the inhibition of inflammatory factors such as TLR-4, TNF- α , NF- κ B, and MCP-1 by CGA¹⁶³. In hyperuricemic nephropathy, CGA treatment promotes renal function recovery and reduces renal fibrosis damage. Gut microbiota studies showed that CGA acts by regulating trimethylamine N-oxide (TMAO)-associated microbiota and inhibiting the PI3K/Akt/mTOR pathway cascade¹⁶⁴.

7.5. The multi-dimensional interaction of Rhubarb against renal fibrosis

TCM is characterized by multiple components acting on multiple targets, and Rhubarb exemplifies this complexity. Based on a synthesis of existing literature, we analyzed the anti-fibrotic effects of Rhubarb's chemical constituents and illustrated their complex interactions with molecular targets using Sankey plots (Fig. 6). This multi-dimensional pharmacological profile operates at three levels. First, individual components exert single-target effects, for example, Aloe-emodin modulates the PI3K/Akt pathway; Emodin, for instance, regulates NF- κ B, MAPK, and PPAR α signaling. Third, multiple constituents converge on shared targets: Emodin, Resveratrol, and Rutin all influence the TGF- β /Smad pathway. This multi-level regulatory network reflects Rhubarb's therapeutic advantages in anti-renal fibrosis through a holistic mechanism, consistent with TCM's systemic approach, while also highlighting the intricate pharmacology of herbal medicine. As previously noted, the TGF- β /Smad pathway drives fibrotic factor expression and ECM deposition, serving as a central and downstream mediator of fibrosis. Consequently, it remains the most extensively studied target, aligning with our analytical findings.

Notably, current research on Rhubarb's anti-renal fibrosis effects highlights Rhein as the most frequently investigated constituent, suggesting its potential significance in CKD and renal fibrosis. Network pharmacology analyses further support this: one study identifies Catechin, Aloe-emodin, Rhein, and Emodin as core components in preventing diabetic nephropathy¹⁶⁵, while another implicates Rhein, β -sitosterol, and Aloe-emodin as key agents against diabetic nephropathy¹⁶⁶. Collectively, these findings indicate that Rhein may represent the most promising Rhubarb-derived compound for combating renal fibrosis. However, this conclusion is based on current literature, which may contain publication bias, and the efficacy of Rhubarb's constituents can vary across different pathological contexts. Comparative studies among major components are therefore still required.

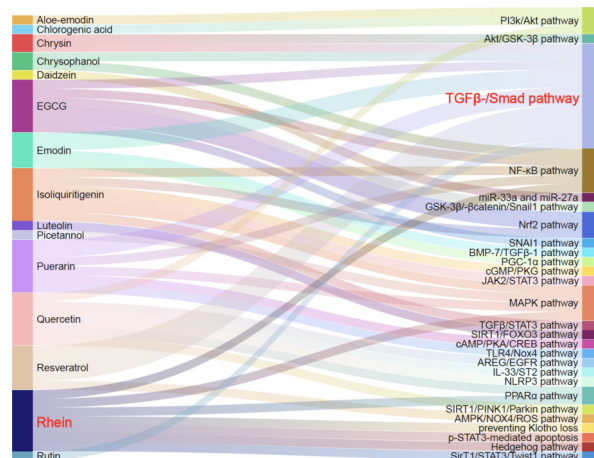


Fig. 6 The multi-dimensional interaction between the chemical components of Rhubarb and the targets in renal fibrosis.

8. Perspectives

8.1. Integrating multi-disciplinary approaches to decipher mechanism synergies

Despite advances in identifying Rhubarb's anti-fibrotic constituents, most studies focus on isolated pathways, such as TGF- β /Smad or NF- κ B inhibition, without elucidating how its complex phytochemical composition produces synergistic effects. For example, while emodin suppresses fibroblast activation and resveratrol mitigates oxidative stress, their combined actions within the renal microenvironment remain uncharacterized. Future research should employ multi-omics technologies, including single-cell sequencing and spatial transcriptomics¹⁶⁷, to map the dynamic interactions between rhubarb's constituents and renal cellular networks. Advanced computational models, such as network pharmacology¹⁶⁸ and artificial intelligence¹⁶⁹, could predict system-level targets and optimize compound ratios for enhanced efficacy. Moreover, *in vivo* validation using genetically engineered models, such as cell-specific knockout mice, is essential to confirm these interactions. By integrating reductionist and holistic methodologies, such approaches will clarify how rhubarb's multi-component system achieves therapeutic specificity while minimizing off-target effects, ultimately guiding the development of standardized polyherbal formulations for CKD.

8.2. Optimizing processing methods and pharmacokinetic profiling

The pharmacological effects of rhubarb's various processing methods, such as raw, wine-fried, or carbonized forms, on renal fibrosis remain poorly characterized. Traditional processing tech-

niques alter the composition of bioactive compounds; for instance, heating reduces anthraquinone glycosides while increasing free aglycones, potentially influencing bioavailability and renal tissue targeting¹⁷⁰. However, systematic studies correlating these processing methods with *in vivo* pharmacokinetics or tissue distribution are limited^{171, 172}, particularly across different stages and etiologies of kidney disease. Future research should employ advanced analytical tools, such as HPLC-QTOF-MS and MALDI imaging, to quantify how processing influences the spatiotemporal release of key constituents in renal tissues¹⁷³. Concurrently, *in vitro-in vivo correlation (IVIVC)* models could help predict dose-response relationships and guide the development of standardized processing protocols that maximize anti-fibrotic efficacy while minimizing gastrointestinal toxicity. Furthermore, novel delivery systems, such as nanoparticles¹⁷⁴ or phytosomes¹⁷⁵, may enhance the stability and renal accumulation of labile compounds like resveratrol. Addressing these knowledge gaps will facilitate evidence-based standardization of Rhubarb preparations, thereby ensuring reproducibility in clinical applications.

8.3. Translating preclinical findings into clinical validation

Although preclinical studies highlight Rhubarb's anti-fibrotic potential, clinical evidence remains sparse and fragmented. While clinical trials on Rhubarb-containing formulations, such as Niaoduoqing Granules, have been conducted¹⁷⁶⁻¹⁸⁰, randomized

controlled trials (RCTs) evaluating Rhubarb as a single agent are scarce³⁵ (Table 4). Moreover, a substantial number of clinical studies on rhubarb and its formulations have been carried out in China, but many suffer from methodological limitations, including small sample sizes, lack of rigorous design, incomplete data reporting, absence of subgroup analyses, and potential ethnic or regional biases^{43, 181}. Rigorous, large-scale, multicenter phase II/III RCTs are therefore urgently needed to evaluate rhubarb's efficacy as an adjunct therapy in CKD populations stratified by fibrosis severity and underlying etiology (e.g., diabetic nephropathy versus hypertensive nephrosclerosis). Additionally, biomarker-driven approaches, such as monitoring urinary TGF- β or collagen IV levels, could provide mechanistic insights alongside clinical endpoints like eGFR decline. Long-term safety assessments are equally critical, particularly regarding anthraquinone-associated risks such as electrolyte imbalances or tubular toxicity¹⁸². Complementary models, including patient-derived organoids^{183, 184} and microphysiological systems^{185, 186}, may enable personalized rhubarb regimens based on individual genetic and microbiome profiles. Finally, interdisciplinary collaboration among phytochemists, nephrologists, and regulatory agencies is essential to address challenges in quality control, intellectual property, and global regulatory acceptance. By prioritizing translational rigor, rhubarb may transition from empirical use to a clinically validated, cost-effective intervention in the growing CKD pandemic.

Table 4 Clinical research on Rhubarb and its formulations for the treatment of chronic kidney disease.

No.	Research type	Research subject	Research drugs	Dosage	Grouping and sample size	Primary outcome indicators	Pharmacodynamic results	Ref
1	Randomized, prospective, double-blinded, and parallel group study	CKD patients (stages 3 & 4)	Rhubarb Capsule	350 mg, thrice daily for 12 weeks	Control: (conservative management treatment, $n = 71$) Rhubarb: (conservative management treatment + Rhubarb Capsule, $n = 74$)	B. urea, Scr, TUP, TUV, GFR	Compared to control group, blood glucose, B. urea, Scr, TUP decreased; Hb, TUV increased	35
2	Randomized, single-blind, and parallel-control study	CKD patients staged G1-G3b with hyperuricemia	Dahuang Mudan Tang	8 weeks	Control group: (Allopurinol Tablets, $n = 40$) Treated group (allopurinol + Dahuang Mudan Tang, $n = 40$)	Kidney function and proteinuria indicators	Treated group showed a better recovery in kidney function, proteinuria, oxidative stress, a higher efficiency rate	176
3	Randomized, double-blind, placebo-controlled, multicenter clinical study	CKD patients staged 3b-4	Niaoduoqing Particles	5 g thrice daily and 10 g before bedtime for 24 weeks	Control group: (placebo, $n = 146$) Test group: (Niaoduoqing Particles, $n = 146$)	Scr, eGFR	Compared to control group, significantly reduced Scr and increased eGFR	177
4	Post-trial, open-label, follow-up study	CKD patients staged 3b-4	Niaoduoqing Particles	daily and 10 g before bedtime for 48 weeks	Control group: (placebo, $n = 146$) Test group: (Niaoduoqing Particles, $n = 146$)	Scr, eGFR	The early use of Niaoduoqing Particles seems to ameliorate the worsening of renal function	178
5	Retrospective cohort study	CKD patients staged 3-5	Niaoduoqing Particles	NA	Control group: (never taken Niaoduoqing Particles, $n = 232$) Niaoduoqing group: (Niaoduoqing prescription duration/follow-up duration > 80%, $n = 1039$)	All-cause mortality, ESRD, eGFR, Scr.	The long-term use of Niaoduoqing granules improved Scr variation and lowered the risk of CKD progression	179
6	Retrospective study	non-diabetic patients with stage IV CKD	Niaoduoqing Particles	5 g thrice daily and 10 g before bedtime for 24 weeks	Control group: (only Western medicine, $n = 220$) Test group: (Niaoduoqing Particles, $n = 252$)	Deaths, doubling of Scr, ESRD	Niaoduoqing Particles reduced Scr and BUN, delayed dialysis time, improved the incidence of compound outcome	180

Note: BUN: blood urea nitrogen, B. urea: blood urea, CKD: chronic kidney disease, eGFR: estimated glomerular filtration rate, ESRD: end-stage renal disease, GFR: glomerular filtration rate, Hb: haemoglobin, Scr: serum creatinine, TUP: 24-hour total urine protein, TUV: 24-hour total urine volume. NA: not available.

9. Conclusion

Rhubarb exhibits significant potential as a phytotherapeutic agent for mitigating renal fibrosis, supported by its diverse bioactive constituents, including anthraquinones, stilbenes, and tannins, and multi-target mechanisms involving anti-inflammatory and antioxidant activities, as well as modulation of the TGF- β /Smad signaling pathway. Preclinical evidence indicates differential efficacy among processed forms and isolated compounds; however, clinical validation remains limited. Future research should prioritize standardized quality control of Rhubarb-derived preparations, comprehensive pharmacokinetic profiling of bioactive metabolites, and rigorously designed human trials to

establish dose-response relationships and long-term safety profiles. Integrating modern pharmacological approaches with traditional knowledge of processing methods may enhance the therapeutic utility of Rhubarb in CKD management.

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Declaration of competing interests

The authors have no conflict of interest to declare.

References

- Levin A, Ahmed SB, Carrero JJ, et al. Executive summary of the KDIGO Clinical Practice Guideline for the evaluation and management of chronic kidney disease: known knowns and known unknowns. *Kidney Int.* 2024; 105(4):684-701. <https://doi.org/10.1016/j.kint.2023.10.016>.
- Roccatello D, Lan H, Sciascia S, et al. From inflammation to renal fibrosis: is a one-way road in autoimmunity? *Autoimmun Rev.* 2024;23(4):103466. <https://doi.org/10.1016/j.autrev.2023.103466>.
- Lahane GP, Dhar A, Bhat A. Therapeutic approaches and novel antifibrotic agents in renal fibrosis: a comprehensive review. *J Biochem Mol Toxicol.* 2024;38(8):e23795. <https://doi.org/10.1002/jbt.23795>.
- Liang L, Mi Y, Zhou S, et al. Advances in the study of key cells and signaling pathways in renal fibrosis and the interventional role of Chinese medicines. *Front Pharmacol.* 2024;15:1403227. <https://doi.org/10.3389/fphar.2024.1403227>.
- Xiang H, Zuo J, Guo F, et al. What we already know about rhubarb: a comprehensive review. *Chin Med.* 2020;15:88. <https://doi.org/10.1186/s13020-020-00363-1>.
- Zhu X, Deng Z, Cao Y, et al. Resveratrol prevents Drp1-mediated mitochondrial fission in the diabetic kidney through the PDE4D/PKA pathway. *Phytother Res.* 2023;37(12):5916-5931. <https://doi.org/10.1002/ptr.8004>.
- Kumar A, Jugran AK, Bargali SS, et al. Ethno-medicinal uses, ecology, phytochemistry, biological activities, and conservation approaches for Himalayan Rhubarb species. *Nat Prod Res.* 2025;1-14. <https://doi.org/10.1080/14786419.2024.2441171>.
- Yang B, Zhang Z, Song J, et al. Interpreting the efficacy enhancement mechanism of Chinese medicine processing from a biopharmaceutic perspective. *Chin Med.* 2024;19(1):14. <https://doi.org/10.1186/s13020-024-00887-0>.
- Wu X, Wang S, Lu J, et al. Seeing the unseen of Chinese herbal medicine processing (Paozhi): advances in new perspectives. *Chin Med.* 2018;13(1):4. <https://doi.org/10.1186/s13020-018-0163-3>.
- Sun J, Wu Y, Dong S, et al. Influence of the drying method on the bioactive compounds and pharmacological activities of rhubarb. *J Sci Food Agric.* 2018;98(9):3551-3562. <https://doi.org/10.1002/jsfa.8871>.
- Zhang J, Zhou P, Xu Y, et al. Metabolic profile and dynamic characteristic of rhubarb during the *in vitro* biotransformation by human gut microbiota. *Food Chem.* 2022;397:133840. <https://doi.org/10.1016/j.foodchem.2022.133840>.
- Zhang J, Fu Y, Li L, et al. Pharmacokinetic comparisons of major bioactive components after oral administration of raw and steamed rhubarb by UPLC-MS/MS. *J Pharm Biomed Anal.* 2019;171:43-51. <https://doi.org/10.1016/j.jpba.2019.04.002>.
- Liu T, Yu M, Dai Y, et al. Traditional method of rhubarb processing optimized by combining flavor analysis with anthraquinone content determination. *Front Nutr.* 2024;11:1406430. <https://doi.org/10.3389/fnut.2024.1406430>.
- Zhou P, Zhang J, Xu Y, et al. Simultaneous quantification of anthraquinone glycosides, aglycones, and glucuronic acid metabolites in rat plasma and tissues after oral administration of raw and steamed rhubarb in blood stasis rats by UHPLC-MS/MS. *J Sep Sci.* 2022;45(2):529-541. <https://doi.org/10.1002/jssc.2021000623>.
- Zhu T, Liu X, Wang X, et al. Profiling and analysis of multiple compounds in rhubarb decoction after processing by wine steaming using UHPLC-Q-TOF-MS coupled with multiple statistical strategies. *J Sep Sci.* 2016;39(15):3081-3090. <https://doi.org/10.1002/jssc.201600256>.
- Wang M, Fu J, Lv M, et al. Effect of wine processing and acute blood stasis on the serum pharmacokinetics of rhubarb: a possible explanation for processing mechanism. *J Sep Sci.* 2014;37(18):2499-2503. <https://doi.org/10.1002/jssc.201400473>.
- Ma J, Meng X, Guo X, et al. Thermal analysis during partial carbonizing process of rhubarb, moutan and burnet. *PLoS One.* 2017;12(3):e0173946. <https://doi.org/10.1371/journal.pone.0173946>.
- Yang L, Yang L, Pei W, et al. Color-reflected chemical regulations of the scorched rhubarb (*Rhei Radix et Rhizoma*) revealed by the integration analysis of visible spectrophotometry, fourier transform infrared spectroscopy and high performance liquid chromatography. *Food Chem.* 2022;367:130730. <https://doi.org/10.1016/j.foodchem.2021.130730>.
- Zhong L, Sun J, Li S, et al. Scorch processing of rhubarb (*Rheum tanguticum* Maxim. ex Balf.) pyrolyzed anthraquinone glycosides into aglycones and improved the therapeutic effects on thromboinflammation via regulating the complement and coagulation cascades pathway. *J Ethnopharmacol.* 2024;333:118475. <https://doi.org/10.1016/j.jep.2024.118475>.
- Zhang Q, Chen Y, Yue S, et al. Study on the content changes of 16 chemical components in *Radix et Rhizoma Rhei* and its different processed products. *China J Tradit Chin Med Pharm.* 2022;37(2):1036-1044. <https://doi.org/10.4266/cissn.1672-2779.2022.0204>.
- Xin L, Jing X, Li H, et al. Efficacy and safety of activating blood circulation and removing blood stasis of Traditional Chinese Medicine for managing renal fibrosis in patients with chronic kidney disease: a systematic review and meta-analysis. *J Tradit Chin Med.* 2023;43(3):429-440. <https://doi.org/10.1016/j.jtcm.2023.03.003>.
- Zhang F, Wu R, Liu Y, et al. Nephroprotective and nephrotoxic effects of rhubarb and their molecular mechanisms. *Biomed Pharmacother.* 2023;160:114297. <https://doi.org/10.1016/j.biopha.2023.114297>.
- Gao H, He C, Xin S, et al. Rhubarb extract rebuilding the mucus homeostasis and regulating mucin-associated flora to relieve constipation. *Exp Biol Med (Maywood).* 2023;248(23):2449-2463. <https://doi.org/10.1177/15353702231218594>.
- Neyrinck AM, Rodriguez J, Taminiau B, et al. Constipation mitigation by rhubarb extract in middle-aged adults is linked to gut microbiome modulation: a double-blind randomized placebo-controlled trial. *Int J Mol Sci.* 2022;23(23):14685. <https://doi.org/10.3390/ijms232314685>.
- Luo Y, Feng X, Wu D, et al. A randomized controlled trial of Chinese traditional medicine Dachengqi Decoction in the treatment of postoperative intestinal function recovery. *Transl Cancer Res.* 2020;9(8):4498-4506. <https://doi.org/10.21037/tcr-19-2671>.
- Khivah A, Hashempour MH, Shakiba M, et al. Effects of rhubarb (*Rheum ribes* L.) syrup on dysenteric diarrhea in children: a randomized, double-blind, placebo-controlled trial. *J Integr Med.* 2017;15(5):365-372. [https://doi.org/10.1016/S2095-4964\(17\)60344-3](https://doi.org/10.1016/S2095-4964(17)60344-3).
- Zhuang T, Gu X, Zhou N, et al. Hepatoprotection and hepatotoxicity of Chinese herb rhubarb (Dahuang): how to properly control the "General (Jiang Jun)" in Chinese medical herb. *Biomed Pharmacother.* 2020;127:110224. <https://doi.org/10.1016/j.biopha.2020.110224>.
- Liu W, Tu Z, Liu J, et al. Therapeutic effect of Yinchenhao Decoction on cholelithiasis via mucin in the gallbladder and intestine. *Fitoterapia.* 2024;172:105746. <https://doi.org/10.1016/j.fitote.2023.105746>.
- Luo S, Huang M, Lu X, et al. Optimized therapeutic potential of Yinchenhao Decoction for cholestatic hepatitis by combined network meta-analysis and network pharmacology. *Phytotherapy.* 2024;129:155573. <https://doi.org/10.1016/j.phymed.2024.155573>.
- Wang L, Wang H, Niu J, et al. Cholesterol-lowering effects of rhubarb free anthraquinones and their mechanism of action. *Eur J Pharmacol.* 2024;966:176348. <https://doi.org/10.1016/j.ejphar.2024.176348>.
- Zhang F, Wu R, Liu Y, et al. Integration of pharmacodynamics and metabolomics to reveal rhubarb anthraquinone protection against nonalcoholic fatty liver disease rat model. *J Pharm Pharmacol.* 2024;76(4):381-390. <https://doi.org/10.1093/jpp/rgae014>.
- Wan Y, Sun S, Fu H, et al. Adjuvant rhubarb alleviates organs dysfunction and inhibits inflammation in heat stroke. *Exp Ther Med.* 2018;16(2):1493-1498. <https://doi.org/10.3892/etm.2018.6344>.
- Huang QJ, Wang MY, Wang M, et al. *Scutellaria baicalensis*: a promising natural source of antiviral compounds for the treatment of viral diseases. *Chin J Nat Med.* 2023;21(8):563-575. [https://doi.org/10.1016/S1875-5364\(23\)60401-7](https://doi.org/10.1016/S1875-5364(23)60401-7).
- Cai Z, Li S, Wang Y. Integrated serum pharmacokinetics, network pharmacology, and molecular docking to study the mechanism of rhubarb against atherosclerosis. *Curr Pharm Biotechnol.* 2025;26(4):564-575. <https://doi.org/10.2174/0113892010296117240531071301>.
- Khan IA, Nasiruddin M, Haque SF, et al. Evaluation of rhubarb supplementation in stages 3 and 4 of chronic kidney disease: a randomized clinical trial. *Int J Chron Dis.* 2014;2014:485960. <https://doi.org/10.1155/2014/485960>.
- Li S, Du S, Du Z, et al. Distribution of Traditional Chinese Medicine syndromes in diabetic kidney disease chronic kidney disease 1-5: a correlation study. *J Tradit Chin Med.* 2024;44(3):572-580. <https://doi.org/10.1016/j.jtcm.2024.02.002>.
- Cao Y, Pu Z, Tang Y, et al. Advances in bio-active constituents, pharmacology and clinical applications of rhubarb. *Chin Med.* 2017;12:36. <https://doi.org/10.1186/s13020-017-0144-6>.
- Xu H, Wang W, Li X, et al. Botany, traditional use, phytochemistry, pharmacology and clinical applications of rhubarb (*Rhei Radix et Rhizoma*): a systematic review. *Am J Chin Med.* 2024;52(7):1925-1967. <https://doi.org/10.1142/S0192415X24500757>.
- Huang W, Rao Y, Li L, et al. Clinical effect of rhubarb on the treatment of chronic renal failure: a meta-analysis. *Front Pharmacol.* 2023;14:1108861. <https://doi.org/10.3389/fphar.2023.1108861>.
- Xiang Z, Sun H, Cai X, et al. The study on the material basis and the mechanism for anti-renal interstitial fibrosis efficacy of rhubarb through integration of metabolomics and network pharmacology. *Mol Biosyst.* 2015;11(4):1067-1078. <https://doi.org/10.1039/C4MB00573B>.
- Qin MY, Huang SQ, Zou XQ, et al. Drug-containing serum of rhubarb-astragalus capsule inhibits the epithelial-mesenchymal transformation of HK-2 by downregulating TGF- β 1/p38MAPK/Smad2/3 pathway. *J Ethnopharmacol.* 2021;280:114414. <https://doi.org/10.1016/j.jep.2021.114414>.
- Zeng X, Cai G, Liang T, et al. Rhubarb and astragalus capsule attenuates renal interstitial fibrosis in rats with unilateral ureteral obstruction by alleviating apoptosis through regulating transforming growth factor beta1 (TGF- β 1)/p38 mitogen-activated protein kinases (p38 MAPK) pathway. *Med Sci Monit.* 2020;26:e920720. <https://doi.org/10.12659/MSM.920720>.
- Zhang Q, Ye X, Zhu L, et al. Spatiotemporal delivery of multiple components of rhubarb-astragalus formula for the synergistic treatment of renal fibrosis. *Front Pharmacol.* 2024;15:1456721. <https://doi.org/10.3389/fphar.2024.1456721>.
- Haq SM, Khoja AA, Lone FA, et al. Keeping healthy in your skin—plants and fungi used by indigenous Himalayan communities to treat dermatological ailments. *Plants.* 2023;12(7):1575. <https://doi.org/10.3390/plants12071575>.
- Tang T, Yin L, Yang J, et al. Emodin, an anthraquinone derivative from *Rheum officinale* Baill, enhances cutaneous wound healing in rats. *Eur J*

- Pharmacol. 2007;567(3):177-185. <https://doi.org/10.1016/j.ejphar.2007.02.033>.
- 46 Ebert T, Pawelzik S, Witasz A, et al. Inflammation and premature ageing in chronic kidney disease. *Toxins*. 2020;12(4):227. <https://doi.org/10.3390/toxins12040227>.
- 47 Zoccali C, Mallamaci F, Kanbay M, et al. The autonomic nervous system and inflammation in chronic kidney disease. *Nephrol Dial Transplant*. 2025;40(8):1470-1482. <https://doi.org/10.1093/ndt/gfaf020>.
- 48 Zhu Y, Yang S, Lv L, et al. Research progress on the positive and negative regulatory effects of rhein on the kidney: a review of its molecular targets. *Molecules*. 2022;27(19):6572. <https://doi.org/10.3390/molecules27196572>.
- 49 Xiong W, Tang J, Yu H, et al. Emodin inhibits M1 macrophage activation that related to acute and chronic kidney injury through EGFR/MAPK pathway. *Funct Integr Genomics*. 2024;24(4):131. <https://doi.org/10.1007/s10142-024-01327-8>.
- 50 Sun L, Han Y, Shen C, et al. Emodin alleviates high glucose-induced oxidative stress, inflammation and extracellular matrix accumulation of mesangial cells by the circ_0000064/miR-30c-5p/Lmp7 axis. *J Recept Signal Transduct Res*. 2022;42(3):302-312. <https://doi.org/10.1080/10799893.2021.1933028>.
- 51 Ji J, Tao P, Wang Q, et al. Emodin attenuate diabetic kidney disease by inhibiting ferroptosis via up regulating Nrf2 expression. *Aging (Albany NY)*. 2023;15(15):7673-7688. <https://doi.org/10.18632/aging.205984>.
- 52 Zhu Y, Jin H, Huo X, et al. Protective effect of rhein against vancomycin-induced nephrotoxicity through regulating renal transporters and Nrf2 pathway. *Phytother Res*. 2022;36(11):4244-4262. <https://doi.org/10.1002/ptr.7559>.
- 53 Njeim R, Merscher S, Fornoni A. Mechanisms and implications of podocyte autophagy in chronic kidney disease. *Am J Physiol Renal Physiol*. 2024;326(6):F877-F893. <https://doi.org/10.1152/ajprenal.00415.2023>.
- 54 Ruby M, Gifford CC, Pandey R, et al. Autophagy as a therapeutic target for chronic kidney disease and the roles of TGF- β 1 in autophagy and kidney fibrosis. *Cells*. 2023;12(3):412. <https://doi.org/10.3390/cells12030412>.
- 55 Tu Y, Gu L, Chen D, et al. Rhein inhibits autophagy in rat renal tubular cells by regulation of AMPK/mTOR signaling. *Sci Rep*. 2017;7(1):43790. <https://doi.org/10.1038/srep43790>.
- 56 Katuri A, Bryant J, Patel D, et al. HIVAN associated tubular pathology with reference to ER stress, mitochondrial changes, and autophagy. *Exp Mol Pathol*. 2019;106:139-148. <https://doi.org/10.1016/j.yexmp.2018.12.009>.
- 57 Li J, Qin X, Xu W, et al. Herb pair of rhubarb-astagalus mitigates renal interstitial fibrosis through downregulation of autophagy via p38-MAPK/TGF- β 1 and p38-MAPK/smad2/3 pathways. *Int J Biochem Cell Biol*. 2024;169:106549. <https://doi.org/10.1016/j.biocel.2024.106549>.
- 58 Xiong D, Hu W, Han X, et al. Rhein inhibited ferroptosis and EMT to attenuate diabetic nephropathy by regulating the Rac1/NOX1/ β -catenin axis. *Front Biosci (Landmark Ed)*. 2023;28(5):100. <https://doi.org/10.31083/j.fbl2805100>.
- 59 HaoShang, Jia X, Liu H, et al. A comprehensive review of emodin in fibrosis treatment. *FitoTerapia*. 2023;165:105358. <https://doi.org/10.1016/j.fitote.2022.105358>.
- 60 Tsuji K, Uchida N, Nakanoh H, et al. The gut-kidney axis in chronic kidney diseases. *Diagnostics*. 2025;15(1):21. <https://doi.org/10.3390/diagnostics15010021>.
- 61 Ji C, Li Y, Mo Y, et al. Rhubarb enema decreases circulating trimethylamine N-oxide level and improves renal fibrosis accompanied with gut microbiota change in chronic kidney disease rats. *Front Pharmacol*. 2021;12:780924. <https://doi.org/10.3389/fphar.2021.780924>.
- 62 Ji C, Lu F, Wu Y, et al. Rhubarb enema increasing short-chain fatty acids that improves the intestinal barrier disruption in CKD may be related to the regulation of gut dysbiosis. *Biomed Res Int*. 2022;2022:1896781. <https://doi.org/10.1155/2022/1896781>.
- 63 Li XJ, Shan QY, Wu X, et al. Gut microbiota regulates oxidative stress and inflammation: a double-edged sword in renal fibrosis. *Cell Mol Life Sci*. 2024;81(1):480. <https://doi.org/10.1007/s00018-024-05532-5>.
- 64 Reiss AB, Jacob B, Zubair A, et al. Fibrosis in chronic kidney disease: pathophysiology and therapeutic targets. *J Clin Med*. 2024;13(7):1881. <https://doi.org/10.3390/jcm13071881>.
- 65 Deng Z, Fan T, Xiao C, et al. TGF- β signaling in health, disease, and therapeutics. *Signal Transduct Target Ther*. 2024;9(1):61. <https://doi.org/10.1038/s41392-024-01764-w>.
- 66 Lathan R. Exploring unconventional targets in myofibroblast transdifferentiation outside classical TGF- β signaling in renal fibrosis. *Front Physiol*. 2024;15:1296504. <https://doi.org/10.3389/fphys.2024.1296504>.
- 67 Soudeh G, Arian A, Hamed S, et al. Antioxidant therapy against TGF- β /SMAD pathway involved in organ fibrosis. *J Cell Mol Med*. 2023;28(2):e18052. <https://doi.org/10.1111/jcmm.18052>.
- 68 Liu W, Hu C, Qian X, et al. TaoHeChengQi Decoction alleviate chronic renal failure via regulation of PHD2/UCP1 and RPK3/AKT/TGF- β pathway. *Phytomedicine*. 2025;141:156548. <https://doi.org/10.1016/j.phymed.2025.156548>.
- 69 Liu J, Xiao Q, Xiao J, et al. Wnt/ β -catenin signalling: function, biological mechanisms, and therapeutic opportunities. *Signal Transduct Target Ther*. 2022;7(1):3. <https://doi.org/10.1038/s41392-021-00762-6>.
- 70 Qin K, Yu M, Fan J, et al. Canonical and noncanonical Wnt signaling: multilayered mediators, signaling mechanisms and major signaling crosstalk. *Genes Dis*. 2023;11(1):103-134. <https://doi.org/10.1016/j.gendis.2022.11.013>.
- 71 Somanander DVN, Zhao P, Widdop RE, et al. The involvement of the Wnt/ β -catenin signaling cascade in fibrosis progression and its therapeutic targeting by relaxin. *Biochem Pharmacol*. 2024;223:116130. <https://doi.org/10.1016/j.bcp.2024.116130>.
- 72 Duan S, Wu Y, Zhao C, et al. The Wnt/ β -catenin signaling pathway participates in rhein ameliorating kidney injury in DN mice. *Mol Cell Biochem*. 2016;411(1-2):73-82. <https://doi.org/10.1007/s11010-015-2569-x>.
- 73 Lv Y, Pang X, Cao Z, et al. Evolution and function of the Notch signaling pathway: an invertebrate perspective. *Int J Mol Sci*. 2024;25(6):3322. <https://doi.org/10.3390/ijms25063322>.
- 74 Mukherjee M, Fogarty E, Janga M, et al. Notch signaling in kidney development, maintenance, and disease. *Biomolecules*. 2019;9(11):692. <https://doi.org/10.3390/biom9110692>.
- 75 Asanuma K, Trejo JO, Tanaka E. The role of Notch signaling in kidney podocytes. *Clin Exp Nephrol*. 2017;21(1):1-6. <https://doi.org/10.1007/s10157-016-1247-y>.
- 76 Xia Z, Wang J, Yang S, et al. Emodin alleviates hypertrophic scar formation by suppressing macrophage polarization and inhibiting the Notch and TGF- β pathways in macrophages. *Braz J Med Biol Res*. 2021;54(8):e11184. <https://doi.org/10.1590/1414-431x2021e11184>.
- 77 Hu Y, Peng L, Zhuo X, et al. Resveratrol promotes diabetic wound healing by inhibiting Notch pathway. *J Surg Res*. 2024;297:63-70. <https://doi.org/10.1016/j.jss.2024.02.004>.
- 78 Hu Y, Peng L, Zhuo X, et al. Hedgehog signaling pathway in fibrosis and targeted therapies. *Biomolecules*. 2024;14(12):1485. <https://doi.org/10.3390/biom14121485>.
- 79 Greenberg D, D'Cruz R, Lacanale JL, et al. Hedgehog-Gli mediated control of renal formation and malformation. *Front Nephrol*. 2023;3:1176347. <https://doi.org/10.3389/fnep.2023.1176347>.
- 80 Bai Y, Lu H, Lin C, et al. Sonic hedgehog-mediated epithelial-mesenchymal transition in renal tubulointerstitial fibrosis. *Int J Mol Med*. 2016;37(5):1317-1327. <https://doi.org/10.3892/ijmm.2016.2546>.
- 81 Bai Y, Lu H, Wu C, et al. Resveratrol inhibits epithelial-mesenchymal transition and renal fibrosis by antagonizing the hedgehog signaling pathway. *Biochem Pharmacol*. 2014;92(3):484-493. <https://doi.org/10.1016/j.bcp.2014.09.002>.
- 82 Wang T, Liu B, Huang J, et al. IFN- γ -mediated inhibition of JAK/STAT signaling via nano-scutellarin treatment is an efficient strategy for ameliorating liver fibrosis. *J Transl Med*. 2025;23(1):195. <https://doi.org/10.1186/s12967-025-06155-5>.
- 83 Liu J, Wang F, Luo F. The role of JAK/STAT pathway in fibrotic diseases: molecular and cellular mechanisms. *Biomolecules*. 2023;13(1):119. <https://doi.org/10.3390/biom13010119>.
- 84 Chuang PY, He JC. JAK/STAT signaling in renal diseases. *Kidney Int*. 2010;78(3):231-234. <https://doi.org/10.1038/ki.2010.158>.
- 85 Shim K, Song HK, Park M, et al. *Reynoutria japonica* consisted of emodin-8- β -D-glucoside ameliorates *Dermatophagoides farinae* extract-induced atopic dermatitis-like skin inflammation in mice by inhibiting JAK/STAT signaling. *Biomed Pharmacother*. 2024;176:116765. <https://doi.org/10.1016/j.biopha.2024.116765>.
- 86 Lao Z, Chen X, Pan B, et al. Pharmacological regulators of Hippo pathway: advances and challenges of drug development. *FASEB J*. 2025;39(6):e70438. <https://doi.org/10.1096/fj.2024018953R>.
- 87 Sun Y, Jin D, Zhang Z, et al. The critical role of the Hippo signaling pathway in kidney diseases. *Front Pharmacol*. 2022;13:988175. <https://doi.org/10.3389/fphar.2022.988175>.
- 88 Zhao C, Wang H, Xu C, et al. The critical role of the Hippo signaling pathway in renal fibrosis. *Cell Signal*. 2025;130:111661. <https://doi.org/10.1016/j.cellsig.2025.111661>.
- 89 Lee EH, Baek SY, Park JY, et al. Emodin in *Rheum undulatum* inhibits oxidative stress in the liver via AMPK with Hippo/Yap signalling pathway. *Pharm Biol*. 2020;58(1):333-341. <https://doi.org/10.1080/13880209.2020.1750658>.
- 90 Niculae A, Gherghina M, Peride I, et al. Pathway from acute kidney injury to chronic kidney disease: molecules involved in renal fibrosis. *Int J Mol Sci*. 2023;24(18):14019. <https://doi.org/10.3390/ijms241814019>.
- 91 Wang Z, Cheng Y, Su W, et al. Organ specific differences in alteration of aquaporin expression in rats treated with sennoside A, senna anthraquinones and rhubarb anthraquinones. *Int J Mol Sci*. 2021;22(15):8026. <https://doi.org/10.3390/ijms22158026>.
- 92 Khatoun M, Dubey A, Km VJ. Unveiling anthraquinones: diverse health benefits of an essential secondary metabolite. *Recent Pat Biotechnol*. 2025;19(3):179-197. <https://doi.org/10.2174/0118722083301761240628083511>.
- 93 Zhang M, Gong Z, Zhao Q, et al. Time-dependent laxative effect of sennoside A, the core functional component of rhubarb, is attributed to gut microbiota and aquaporins. *J Ethnopharmacol*. 2023;311:116431. <https://doi.org/10.1016/j.jep.2023.116431>.
- 94 Chan YL, Tang SN, Osman CP, et al. Exploring naphthoquinone and anthraquinone derivatives as antibiotic adjuvants against *Staphylococcus aureus* biofilms: synergistic effects of menadiene. *Microb Pathog*. 2024;195:106886. <https://doi.org/10.1016/j.micpath.2024.106886>.
- 95 Lin C, Chuang S, Huang T, et al. A systematic comparison of the effect of topically applied anthraquinone aglycones to relieve psoriasisiform lesion: the evaluation of percutaneous absorption and anti-inflammatory potency. *Biomed Pharmacother*. 2022;145:112482. <https://doi.org/10.1016/j.biopha.2021.112482>.
- 96 Chen Z, Xu Z, Chen X, et al. Characterization of five natural anthraquinone compounds as potent inhibitors against CYP1B1: implications for cancer treatment. *Curr Drug Metab*. 2024;25(9):677-684. <https://doi.org/10.2174/0113892002329282250108163208>.
- 97 Feng L, Lin Z, Tang Z, et al. Emodin improves renal fibrosis in chronic kidney disease by regulating mitochondrial homeostasis through the mediation of peroxisome proliferator-activated receptor-gamma coactivator-1 alpha (PGC-1 α). *Eur J Histochem*. 2024;68(2):3917. <https://doi.org/10.4081/ejh.2024.3917>.

- 98 Liu W, Gu R, Lou Y, et al. Emodin-induced autophagic cell death hinders epithelial-mesenchymal transition via regulation of BMP-7/TGF- β 1 in renal fibrosis. *J Pharmacol Sci.* 2021;146(4):216-225. <https://doi.org/10.1016/j.jphs.2021.03.009>.
- 99 Ma L, Li H, Zhang S, et al. Emodin ameliorates renal fibrosis in rats via TGF- β 1/Smad signaling pathway and function study of Smurf2. *Int Urol Nephrol.* 2018;50(2):373-382. <https://doi.org/10.1007/s11255-017-1757-x>.
- 100 Yang F, Deng L, Li J, et al. Emodin retarded renal fibrosis through regulating HGF and TGF β -Smad signaling pathway. *Drug Des Devel Ther.* 2020;14:3567-3575. <https://doi.org/10.2147/DDDT.S245847>.
- 101 Dou F, Liu Y, Liu L, et al. Aloe-emodin ameliorates renal fibrosis via inhibiting PI3K/Akt/mTOR signaling pathway in vivo and in vitro. *Rejuvenation Res.* 2019;22(3):218-229. <https://doi.org/10.1089/rej.2018.2104>.
- 102 Wang L, Yu X, Li H, et al. Cell and rat serum, urine and tissue metabolomics analysis elucidates the key pathway changes associated with chronic nephropathy and reveals the mechanism of action of rhein. *Chin Med.* 2023;18(1):158. <https://doi.org/10.1186/s13020-023-00862-1>.
- 103 Xiao Q, Yu X, Yu X, et al. An integrated network pharmacology and cell metabolomics approach to reveal the role of rhein, a novel PPAR α agonist, against renal fibrosis by activating the PPAR α -CPT1A axis. *Phytomedicine.* 2022;102:154147. <https://doi.org/10.1016/j.phymed.2022.154147>.
- 104 Song X, Du Z, Yao Z, et al. Rhein improves renal fibrosis by restoring Cpt1a-mediated fatty acid oxidation through SirT1/STAT3/twist1 pathway. *Molecules.* 2022;27(7):2344. <https://doi.org/10.3390/molecules27072344>.
- 105 Feng HY, Wang YQ, Yang J, et al. Anthraquinones from *Rheum officinale* ameliorate renal fibrosis in acute kidney injury and chronic kidney disease. *Drug Des Devel Ther.* 2025;19:5739-5760. <https://doi.org/10.2147/DDDT.S521265>.
- 106 Chen Y, Mu L, Xing L, et al. Rhein alleviates renal interstitial fibrosis by inhibiting tubular cell apoptosis in rats. *Biol Res.* 2019;52(1):50. <https://doi.org/10.1186/s40659-019-0257-0>.
- 107 Zhang Q, Yin S, Liu L, et al. Rhein reversal of DNA hypermethylation-associated Klotho suppression ameliorates renal fibrosis in mice. *Sci Rep.* 2016;6(1):34597. <https://doi.org/10.1038/srep34597>.
- 108 Dou F, Ding Y, Wang C, et al. Chrysophanol ameliorates renal interstitial fibrosis by inhibiting the TGF- β /Smad signaling pathway. *Biochem Pharmacol.* 2020;180:114079. <https://doi.org/10.1016/j.bcp.2020.114079>.
- 109 Gu M, Zhou Y, Liao N, et al. Chrysophanol, a main anthraquinone from *Rheum palmatum* L. (rhubarb), protects against renal fibrosis by suppressing NKG2D/NF- κ B pathway. *Phytomedicine.* 2022;105:154381. <https://doi.org/10.1016/j.phymed.2022.154381>.
- 110 Wei Q, Gao F, Gao L, et al. Construction of chrysophanol loaded nanoparticles with *N*-octyl-*O*-sulfate chitosan for enhanced nephroprotective effect. *Eur J Pharm Sci.* 2023;193:106685. <https://doi.org/10.1016/j.ejps.2023.106685>.
- 111 Zhou L, Cai X, Wang Y, et al. Chemistry and biology of natural stilbenes: an update. *Nat Prod Rep.* 2025;19:42(2):359-405. <https://doi.org/10.1039/d4np00033a>.
- 112 Tekka T, Zhang L, Ge X, et al. Stilbenes: source plants, chemistry, biosynthesis, pharmacology, application and problems related to their clinical application—a comprehensive review. *Phytochemistry.* 2022;197:113128. <https://doi.org/10.1016/j.phytochem.2022.113128>.
- 113 Wiciński M, Erdmann J, Nowacka A, et al. Natural phytochemicals as SIRT activators—focus on potential biochemical mechanisms. *Nutrients.* 2023;15(16):3578. <https://doi.org/10.3390/nu15163578>.
- 114 Shazmeen, Haq IU, Rajoka MSR, et al. Role of stilbenes against insulin resistance: a review. *Food Sci Nutr.* 2021;9(11):6389-6405. <https://doi.org/10.1002/fsn3.2620>.
- 115 Regnier M, Van Hul M, Roumain M, et al. Inulin increases the beneficial effects of rhubarb supplementation on high-fat high-sugar diet-induced metabolic disorders in mice: impact on energy expenditure, brown adipose tissue activity, and microbiota. *Gut Microbes.* 2023;15(1):2178796. <https://doi.org/10.1080/19490976.2023.2178796>.
- 116 Begh MZA, Khan J, Zehravi M, et al. Targeting neurological disorders with stilbenes: bridging the preclinical-clinical gap. *Int J Biol Sci.* 2024;20(14):5474-5494. <https://doi.org/10.7150/ijbs.102032>.
- 117 Zhang X, Lu H, Xie S, et al. Resveratrol suppresses the myofibroblastic phenotype and fibrosis formation in kidneys via proliferation-related signalling pathways. *Br J Pharmacol.* 2019;176(24):4745-4759. <https://doi.org/10.1111/bph.14842>.
- 118 Wang R, Yuan W, Li L, et al. Resveratrol ameliorates muscle atrophy in chronic kidney disease via the axis of SIRT1/FoxO1. *Phytother Res.* 2022;36(8):3265-3275. <https://doi.org/10.1002/ptr.7499>.
- 119 He T, Xiong J, Nie L, et al. Resveratrol inhibits renal interstitial fibrosis in diabetic nephropathy by regulating AMPK/NOX4/ROS pathway. *J Mol Med (Berl).* 2016;94(12):1359-1371. <https://doi.org/10.1007/s00109-016-1451-y>.
- 120 Xiao Z, Chen C, Meng T, et al. Resveratrol attenuates renal injury and fibrosis by inhibiting transforming growth factor- β pathway on matrix metalloproteinase 7. *Exp Biol Med (Maywood).* 2016;241(2):140-146. <https://doi.org/10.1177/1535370215598401>.
- 121 Liu S, Zhao M, Zhou Y, et al. Resveratrol exerts dose-dependent anti-fibrotic or pro-fibrotic effects in kidneys: a potential risk to individuals with impaired kidney function. *Phytomedicine.* 2019;57:223-235. <https://doi.org/10.1016/j.phymed.2018.12.024>.
- 122 Choi SY, Piao ZH, Jin L, et al. Piceatannol attenuates renal fibrosis induced by unilateral ureteral obstruction via downregulation of histone deacetylase 4/5 or p38-MAPK signaling. *PLoS One.* 2016;11(11):e0167340. <https://doi.org/10.1371/journal.pone.0167340>.
- 123 Shen N, Wang T, Gan Q, et al. Plant flavonoids: classification, distribution, biosynthesis, and antioxidant activity. *Food Chem.* 2022;383:132531. <https://doi.org/10.1016/j.foodchem.2022.132531>.
- 124 Cao X, Shang Y, Kong W, et al. Flavonoids derived from *Anemarrhena* Rhizoma ameliorate inflammation of benign prostatic hyperplasia via modulating COX/LOX pathways. *J Ethnopharmacol.* 2022;284:114740. <https://doi.org/10.1016/j.jep.2021.114740>.
- 125 Huang M, Xie X, Yuan R, et al. The multifaceted anti-atherosclerotic properties of herbal flavonoids: a comprehensive review. *Pharmacol Res.* 2025;211:107551. <https://doi.org/10.1016/j.phrs.2024.107551>.
- 126 Iheanacho MS, Kandel R, Roy P, et al. Epigallocatechin-3-gallate attenuates arsenic-induced fibrogenic changes in human kidney epithelial cells through reversal of epigenetic aberrations and antioxidant activities. *Biofactors.* 2023;50(3):542-557. <https://doi.org/10.1002/biof.1883>.
- 127 He C, Wang D, Wang R, et al. Epigallocatechin gallate induces the demethylation of actinin alpha 4 to inhibit diabetic nephropathy renal fibrosis via the NF- κ B signaling pathway in vitro. *Dose Response.* 2022;20(2):155932582211057. <https://doi.org/10.1177/155932582211057>.
- 128 Kanlaya R, Kapincharanon C, Fong-Ngern K, et al. Induction of mesenchymal-epithelial transition (MET) by epigallocatechin-3-gallate to reverse epithelial-mesenchymal transition (EMT) in SNAI1-overexpressed renal cells: a potential anti-fibrotic strategy. *J Nutr Biochem.* 2022;107:109066. <https://doi.org/10.1016/j.jnutbio.2022.109066>.
- 129 Kanlaya R, Peerapen P, Nilnumkhum A, et al. Epigallocatechin-3-gallate prevents TGF- β 1-induced epithelial-mesenchymal transition and fibrotic changes of renal cells via GSK-3 β / β -catenin/Snai1 and Nrf2 pathways. *J Nutr Biochem.* 2020;76:108266. <https://doi.org/10.1016/j.jnutbio.2019.108266>.
- 130 Wang Y, Liu N, Su X, et al. Epigallocatechin-3-gallate attenuates transforming growth factor- β 1 induced epithelial-mesenchymal transition via Nrf2 regulation in renal tubular epithelial cells. *Biomed Pharmacother.* 2015;70:260-267. <https://doi.org/10.1016/j.biopha.2015.02.004>.
- 131 Wang Y, Wang B, Du F, et al. Epigallocatechin-3-gallate attenuates unilateral ureteral obstruction-induced renal interstitial fibrosis in mice. *J Histochem Cytochem.* 2015;63(4):270-279. <https://doi.org/10.1369/0022155414568294>.
- 132 Wang Y, Liu N, Bian X, et al. Epigallocatechin-3-gallate reduces tubular cell apoptosis in mice with ureteral obstruction. *J Surg Res.* 2015;197(1):145-154. <https://doi.org/10.1016/j.jss.2015.03.034>.
- 133 Nagavally R, Sunilkumar S, Akhtar M, et al. Chrysin ameliorates cyclosporine-A-induced renal fibrosis by inhibiting TGF- β -induced epithelial-mesenchymal transition. *Int J Mol Sci.* 2021;22(19):10252. <https://doi.org/10.3390/ijms221910252>.
- 134 Kang M, Park S, Choi Y, et al. Chrysin inhibits diabetic renal tubulointerstitial fibrosis through blocking epithelial to mesenchymal transition. *J Mol Med (Berl).* 2015;93(7):759-772. <https://doi.org/10.1007/s00109-015-1301-3>.
- 135 Kang Y, Park S, Sim YE, et al. Highly water-soluble diacetyl chrysin ameliorates diabetes-associated renal fibrosis and retinal microvascular abnormality in db/db mice. *Nutr Res Pract.* 2023;17(3):421. <https://doi.org/10.1007/nrp.2023.17.3.421>.
- 136 Li F, Wei R, Huang M, et al. Luteolin can ameliorate renal interstitial fibrosis-induced renal anaemia through the SIRT1/FOXO3 pathway. *Food Funct.* 2022;13(22):11896-11914. <https://doi.org/10.1039/D2FO02477B>.
- 137 Wang Q, Wang F, Li X, et al. Quercetin inhibits the amphiregulin/EGFR signaling-mediated renal tubular epithelial-mesenchymal transition and renal fibrosis in obstructive nephropathy. *Phytother Res.* 2023;37(1):111-123. <https://doi.org/10.1002/ptr.7670>.
- 138 Ma Z, Wang F, Wang H, et al. Quercetin ameliorates renal tubulointerstitial transformation and renal fibrosis by regulating NLRP3 in obstructive nephropathy. *Minerva Med.* 2023;114(4):530-532. <https://doi.org/10.23736/S0026-4806.22.08373-8>.
- 139 Chen H, Chiang Y, Hong Y, et al. Quercetin ameliorates renal injury and pyroptosis in lupus nephritis through inhibiting IL-33/ST2 pathway in vitro and in vivo. *Antioxidants.* 2022;11(11):2238. <https://doi.org/10.3390/antiox11112238>.
- 140 Guo X, Wen S, Wang J, et al. Senolytic combination of dasatinib and quercetin attenuates renal damage in diabetic kidney disease. *Phytomedicine.* 2024;130:155705. <https://doi.org/10.1016/j.phymed.2024.155705>.
- 141 Sánchez-Jaramillo EA, Gasca-Lozano LE, Vera-Cruz JM, et al. Nanoparticles formulation improves the antifibrotic effect of quercetin on an adenine-induced model of chronic kidney disease. *Int J Mol Sci.* 2022;23(10):5392. <https://doi.org/10.3390/ijms23105392>.
- 142 Tu H, Ma D, Luo Y, et al. Quercetin alleviates chronic renal failure by targeting the PI3k/Akt pathway. *Bioengineered.* 2021;12(1):6538-6558. <https://doi.org/10.1080/21655979.2021.1973877>.
- 143 Liu T, Yang Q, Zhang X, et al. Quercetin alleviates kidney fibrosis by reducing renal tubular epithelial cell senescence through the SIRT1/PINK1/mitophagy axis. *Life Sci.* 2020;257:118116. <https://doi.org/10.1016/j.lfs.2020.118116>.
- 144 Han Y, Lu J, Xu Y, et al. Rutin ameliorates renal fibrosis and proteinuria in 5/6-nephrectomized rats by anti-oxidation and inhibiting activation of TGF β -smad signaling. *Int J Clin Exp Pathol.* 2015;8(5):4725-4734.
- 145 Ganesan D, Albert A, Paul E, et al. Rutin ameliorates metabolic acidosis and fibrosis in alloxan induced diabetic nephropathy and cardiomyopathy in experimental rats. *Mol Cell Biochem.* 2020;471(1-2):41-50. <https://doi.org/10.1007/s11010-020-03758-y>.
- 146 Majid A, Hamid N, Shadan S, et al. Daidzein mitigates oxidative stress and inflammation in the injured kidney of ovariectomized rats: AT1 and Mas receptor functions. *Iran J Kidney Dis.* 2022;16(1):32-43. <https://doi.org/10.15271/ijkd.16.1.32>.
- 147 Majid A, Hamid N, Shadan S, et al. Isoflavone daidzein ameliorates renal dysfunction and fibrosis in a postmenopausal rat model: intermediation of

- angiotensin AT1 and Mas receptors and microRNAs 33a and 27a. *Iran J Basic Med Sci.* 2022;25(11):1317-1325. <https://doi.org/10.22038/IJBMS.2022.66619.14270>.
- 148 Biyu H, Ma P, Yang X, et al. *In silico* prediction and experimental validation to reveal the protective mechanism of puerarin against excessive extracellular matrix accumulation through inhibiting ferroptosis in diabetic nephropathy. *J Ethnopharmacol.* 2023;319:117281. <https://doi.org/10.1016/j.jep.2023.117281>.
- 149 Jian J, Wang D, Xiong Y, et al. Puerarin alleviated oxidative stress and ferroptosis during renal fibrosis induced by ischemia/reperfusion injury via TLR4/Nox4 pathway in rats. *Acta Cir Bras.* 2023;38:e382523. <https://doi.org/10.1590/acb382523>.
- 150 Zhu Q, Yang S, Wei C, et al. Puerarin attenuates diabetic kidney injury through interaction with guanidine nucleotide-binding protein G subunit alpha-1 (Gnai1) subunit. *J Cell Mol Med.* 2022;26(14):3816-3827. <https://doi.org/10.1111/jcmm.17414>.
- 151 Yang J, Li B, Wang J, et al. Puerarin alleviates chronic renal failure-induced pyroptosis in renal tubular epithelial cells by targeting miR-342-3p/TGF- β /SMAD axis. *Genes Genomics.* 2023;45(12):1563-1573. <https://doi.org/10.1007/s13258-023-01448-9>.
- 152 Wang J, Ge S, Wang Y, et al. Puerarin alleviates UUO-induced inflammation and fibrosis by regulating the NF- κ B P65/STAT3 and TGF β 1/Smads signaling pathways. *Drug Des Devel Ther.* 2021;15:3697-3708. <https://doi.org/10.2147/DDDT.S321879>.
- 153 Zhou X, Bai C, Sun X, et al. Puerarin attenuates renal fibrosis by reducing oxidative stress induced-epithelial cell apoptosis via MAPK signal pathways *in vivo* and *in vitro*. *Ren Fail.* 2017;39(1):423-431. <https://doi.org/10.1080/0886022X.2017.1305409>.
- 154 Liao Y, Tan R, Li J, et al. Isoliquiritigenin attenuates UUO-induced renal inflammation and fibrosis by inhibiting Mincle/Syk/NF- κ B signaling pathway. *Drug Des Devel Ther.* 2020;14:1455-1468. <https://doi.org/10.2147/DDDT.S243420>.
- 155 Atteia HH, Alamri ES, Sirag N, et al. Soluble guanylate cyclase agonist, isoliquiritigenin attenuates renal damage and aortic calcification in a rat model of chronic kidney failure. *Life Sci.* 2023;317:121460. <https://doi.org/10.1016/j.lfs.2023.121460>.
- 156 Lin C, Lin Y, Paul CR, et al. Isoliquiritigenin ameliorates advanced glycation end-products toxicity on renal proximal tubular epithelial cells. *Environ Toxicol.* 2022;37(8):2096-2102. <https://doi.org/10.1002/tox.23553>.
- 157 Sun L, Yang Z, Zhang J, et al. Isoliquiritigenin attenuates acute renal injury through suppressing oxidative stress, fibrosis and JAK2/STAT3 pathway in streptozotocin-induced diabetic rats. *Bioengineered.* 2021;12(2):1188-1200. <https://doi.org/10.1080/21655979.2021.2006978>.
- 158 Huang X, Shi Y, Chen H, et al. Isoliquiritigenin prevents hyperglycemia-induced renal injuries by inhibiting inflammation and oxidative stress via SIRT1-dependent mechanism. *Cell Death Dis.* 2020;11(12):1040. <https://doi.org/10.1038/s41419-020-03242-5>.
- 159 Maugeri A, Lombardo GE, Cirmi S, et al. Pharmacology and toxicology of tannins. *Arch Toxicol.* 2022;96(5):1257-1277. <https://doi.org/10.1007/s00204-022-03250-0>.
- 160 Huang J, Zaynab M, Sharif Y, et al. Tannins as antimicrobial agents: understanding toxic effects on pathogens. *Toxicol.* 2024;247:107812. <https://doi.org/10.1016/j.toxicol.2024.107812>.
- 161 Su X, Liu X, Wang S, et al. Wound-healing promoting effect of total tannins from *Entada phaseoloides* (L.) Merr. in rats. *Burns.* 2017;43(4):830-838. <https://doi.org/10.1016/j.burns.2016.10.010>.
- 162 Yousuf MJ, Vellaichamy E. Protective activity of gallic acid against glyoxal-induced renal fibrosis in experimental rats. *Toxicol Rep.* 2015;2:1246-1254. <https://doi.org/10.1016/j.toxrep.2015.07.007>.
- 163 Arfian N, Wahyudi DAP, Zulfatma IB, et al. Chlorogenic acid attenuates kidney ischemic/reperfusion injury via reducing inflammation, tubular injury, and myofibroblast formation. *Biomed Res Int.* 2019;2019:1-10. <https://doi.org/10.1155/2019/3834802>.
- 164 Zhou X, Zhang B, Zhao X, et al. Chlorogenic acid prevents hyperuricemia nephropathy via regulating TMAO-related gut microbes and inhibiting the PI3K/AKT/mTOR pathway. *J Agric Food Chem.* 2022;70(33):10182-10193. <https://doi.org/10.1021/acs.jafc.2c03099>.
- 165 Gao Y, Nan Z. Mechanistic insights into the use of rhubarb in diabetic kidney disease treatment using network pharmacology. *Medicine.* 2022;101(1):e28465. <https://doi.org/10.1097/MD.00000000000028465>.
- 166 Fu S, Zhou Y, Hu C, et al. Network pharmacology and molecular docking technology-based predictive study of the active ingredients and potential targets of rhubarb for the treatment of diabetic nephropathy. *BMC Complement Med Ther.* 2022;22(1):210. <https://doi.org/10.1186/s12906-022-03662-6>.
- 167 Verma A, Wilson PC. Advances in single-cell sequencing and spatial profiling of kidney disease. *Am J Pathol.* 2025;195(1):5-6. <https://doi.org/10.1016/j.ajpath.2024.10.010>.
- 168 Shi X, Yin H, Shi X. Bibliometric analysis of literature on natural medicines against chronic kidney disease from 2001 to 2024. *Phytomedicine.* 2025;138:156410. <https://doi.org/10.1016/j.phymed.2025.156410>.
- 169 Simeri A, Pezzi G, Arena R, et al. Artificial intelligence in chronic kidney diseases: methodology and potential applications. *Int Urol Nephrol.* 2025;57(1):159-168. <https://doi.org/10.1007/s11225-024-04026-6>.
- 170 Wen Y, Yan P, Fan P, et al. The application of rhubarb concoctions in traditional Chinese medicine and its compounds, processing methods, pharmacology, toxicology and clinical research. *Front Pharmacol.* 2024;15:1376457. <https://doi.org/10.3389/fphar.2024.1376457>.
- 171 Zhao D, Feng S, Zhang H, et al. Pharmacokinetics, tissue distribution and excretion of five rhubarb anthraquinones in rats after oral administration of effective fraction of anthraquinones from *Rheum officinale*. *Xenobiotica.* 2021;51(8):916-925. <https://doi.org/10.1080/00498254.2021.1940353>.
- 172 Feng SX, Li JS, Qu LB, et al. Comparative pharmacokinetics of five rhubarb anthraquinones in normal and thrombotic focal cerebral ischemia-induced rats. *Phytother Res.* 2013;27(10):1489-1494. <https://doi.org/10.1002/ptr.4890>.
- 173 Islam A, Sakamoto T, Zhai Q, et al. Application of AP-MALDI imaging mass microscope for the rapid mapping of imipramine, chloroquine, and their metabolites in the kidney and brain of wild-type mice. *Pharmaceuticals.* 2022;15(11):1314. <https://doi.org/10.3390/ph15111314>.
- 174 Li H, Ouyang Y, Lv H, et al. Nanoparticle-mediated Klotho gene therapy prevents acute kidney injury to chronic kidney disease transition through regulating PPAR α signaling in renal tubular epithelial cells. *Biomaterials.* 2025;315:122926. <https://doi.org/10.1016/j.biomaterials.2024.122926>.
- 175 Zeng Y, Deng X, Shen L, et al. Advances in plant-derived extracellular vesicles: isolation, composition, and biological functions. *Food Funct.* 2024;15(23):11319-11341. <https://doi.org/10.1039/D4FO01871H>.
- 176 Fang C, Zhu X, Zhang Y, et al. Combination of allopurinol with Dahuang Mudan Tang significantly improve kidney function and alleviate oxidative stress and inflammation of chronic kidney disease stage I-III patients with hyperuricemia. *J Tradit Chin Med.* 2024;44(1):182-187. <https://doi.org/10.1016/j.jtc.2023.10.006>.
- 177 Zheng Y, Cai GY, He LQ, et al. Efficacy and safety of Niaoduqing Particles for delaying moderate-to-severe renal dysfunction: a randomized, double-blind, placebo-controlled, multicenter clinical study. *Chin Med J (Engl).* 2017;130(20):2402-2409. <https://doi.org/10.4103/0366-6999.216407>.
- 178 Zheng Y, Wang NS, Liu YN, et al. Effects of Niaoduqing Particles on delaying progression of renal dysfunction: a post-trial, open-label, follow-up study. *Chin J Integr Med.* 2019;25(3):168-174. <https://doi.org/10.1007/s11655-018-2998-y>.
- 179 Chen H, Lin Y, Han Y, et al. A Chinese patent medicine's long-term efficacy on non-dialysis patients with CKD stages 3-5: a retrospective cohort study. *Front Pharmacol.* 2024;15:1379338. <https://doi.org/10.3389/fphar.2024.1379338>.
- 180 Wei Z, Li Q, Han X, et al. Retrospective study on the effect of Niaoduqing Particles on outcome of non-diabetic patients with stage IV chronic kidney disease. *Medicine.* 2024;103(6):e37051. <https://doi.org/10.1097/MD.00000000000037051>.
- 181 Song Z, Qin T, Pan Y, et al. Shenkang Injection improves coagulation in patients with chronic kidney disease: a systematic review and meta-analysis. *J Tradit Chin Med.* 2019;39(4):451-458. <https://doi.org/10.1016/j.jtc.2019.06.003>.
- 182 Liu C, Liu J, Zheng Y, et al. Subchronic oral toxicity study of rhubarb extract in Sprague-Dawley rats. *Regul Toxicol Pharmacol.* 2021;123:104921. <https://doi.org/10.1016/j.yrtph.2021.104921>.
- 183 Liu Q, Yue L, Deng J, et al. Progress and breakthroughs in human kidney organoid research. *Biochem Biophys Res.* 2024;39:101736. <https://doi.org/10.1016/j.bbrep.2024.101736>.
- 184 Yang H, Li J, Wang Z, et al. Bridging the organoid translational gap: integrating standardization and micropatterning for drug screening in clinical and pharmaceutical medicine. *Life Med.* 2024;3(2):lnae016. <https://doi.org/10.1093/lifemedi/lnae016>.
- 185 Caetano-Pinto P, Nordell P, Nieskens T, et al. Amplifying the impact of kidney microphysiological systems: predicting renal drug clearance using mechanistic modelling based on reconstructed drug secretion. *ALTEX.* 2023;40(3):408-424. <https://doi.org/10.14573/altex.2209131>.
- 186 Mahadeo A, Yeung CK, Himmelfarb J, et al. Kidney microphysiological models for nephrotoxicity assessment. *Curr Opin Toxicol.* 2022;30:100341. <https://doi.org/10.1016/j.cotox.2022.03.002>.