

## The signaling pathways of atherosclerosis regulated by Taohong Siwu Decoction

Kaijie Yan, Sihe Gong, Yanni Li, Zhonghong Shi, Yimin Bao, Jing Leng, Ke Ning

**Citation:** Kaijie Yan, Sihe Gong, Yanni Li, Zhonghong Shi, Yimin Bao, Jing Leng, Ke Ning, The signaling pathways of atherosclerosis regulated by Taohong Siwu Decoction, *Chinese Journal of Natural Medicines*, 2026, 24(3), 279–288. doi: [10.1016/S1875-5364\(26\)61103-X](https://doi.org/10.1016/S1875-5364(26)61103-X).

View online: [https://doi.org/10.1016/S1875-5364\(26\)61103-X](https://doi.org/10.1016/S1875-5364(26)61103-X)

## Related articles that may interest you

Taohong Siwu Decoction: a classical Chinese prescription for treatment of orthopedic diseases

*Chinese Journal of Natural Medicines*. 2024, 22(8), 711–723 [https://doi.org/10.1016/S1875-5364\(24\)60581-9](https://doi.org/10.1016/S1875-5364(24)60581-9)

Effects of Bunao–Fuyuan decoction serum on proliferation and migration of vascular smooth muscle cells in atherosclerotic

*Chinese Journal of Natural Medicines*. 2021, 19(1), 36–45 [https://doi.org/10.1016/S1875-5364\(21\)60004-3](https://doi.org/10.1016/S1875-5364(21)60004-3)

Paeonol reduces microbial metabolite  $\alpha$ -hydroxyisobutyric acid to alleviate the ROS/TXNIP/NLRP3 pathway-mediated endothelial inflammation in atherosclerosis mice

*Chinese Journal of Natural Medicines*. 2023, 21(10), 759–774 [https://doi.org/10.1016/S1875-5364\(23\)60506-0](https://doi.org/10.1016/S1875-5364(23)60506-0)

Natural products: potential therapeutic agents for atherosclerosis

*Chinese Journal of Natural Medicines*. 2022, 20(11), 830–845 [https://doi.org/10.1016/S1875-5364\(22\)60219-X](https://doi.org/10.1016/S1875-5364(22)60219-X)

Si–Miao–Yong–An Decoction alleviates thromboangiitis obliterans by regulating miR–548j–5p/IL–17A signaling pathway

*Chinese Journal of Natural Medicines*. 2024, 22(6), 541–553 [https://doi.org/10.1016/S1875-5364\(24\)60626-6](https://doi.org/10.1016/S1875-5364(24)60626-6)

*Jiedu Sangen* decoction inhibits chemoresistance to 5–fluorouracil of colorectal cancer cells by suppressing glycolysis via PI3K/AKT/HIF–1 $\alpha$  signaling pathway

*Chinese Journal of Natural Medicines*. 2021, 19(2), 143–152 [https://doi.org/10.1016/S1875-5364\(21\)60015-8](https://doi.org/10.1016/S1875-5364(21)60015-8)



Wechat



Contents lists available at ScienceDirect

## Chinese Journal of Natural Medicines

journal homepage: [www.cjnmcpu.com/](http://www.cjnmcpu.com/)

## Review

## The signaling pathways of atherosclerosis regulated by Taohong Siwu Decoction

Kaijie Yan<sup>a,b</sup>, Sihe Gong<sup>a,b</sup>, Yanni Li<sup>a,b</sup>, Zhonghong Shi<sup>a,b</sup>, Yimin Bao<sup>a,b</sup>, Jing Leng<sup>c,\*</sup>, Ke Ning<sup>a,b,\*</sup><sup>a</sup> School of Integrative Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China<sup>b</sup> School of Traditional Chinese Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China<sup>c</sup> Preclinical Department, Shanghai Municipal Hospital of Traditional Chinese Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai 200071, China

## ARTICLE INFO

## Article history:

Received 4 March 2025

Revised 11 June 2025

Accepted 13 June 2025

Available online 20 March 2026

## Keywords:

Atherosclerosis

Traditional Chinese medicine

Taohong Siwu Decoction

Signaling pathways

## ABSTRACT

Taohong Siwu Decoction (THSWD), a traditional Chinese medicinal formulation, has been demonstrated to significantly modulate key signaling pathways implicated in atherosclerosis (AS). This review examines the complex mechanisms through which THSWD influences critical pathways, including nuclear factor kappa-B (NF-κB), phosphatidylinositol 3-kinase (PI3K)/serine-threonine kinase (AKT), Toll-like receptor 4 (TLR4), mitogen-activated protein kinase (MAPK), and mammalian target of rapamycin (mTOR), that play pivotal roles in AS pathogenesis. By synthesizing experimental evidence and existing literature, the review summarizes how THSWD and its bioactive constituents regulate these signaling cascades to ameliorate AS. Furthermore, it highlights the distinctive therapeutic advantages of traditional Chinese medicine (TCM) compounds in managing chronic diseases driven by multi-target and multifactorial mechanisms. Analyzing disease targets from the perspective of signaling pathways enhances the scientific validation of clinical efficacy for such formulations, thereby offering novel insights for future research.

## 1. Introduction

Atherosclerosis (AS) is considered one of the major causes of cardiovascular disease, primarily initiated by arterial endothelial cell damage, followed by a cascade of events including lipid accumulation, fibrous plaque formation, and plaque rupture. The inflammatory response is involved in nearly all stages of AS. During AS progression, endothelial dysfunction, lipid metabolism disorders, inflammation, and platelet activation are interrelated processes that collectively influence plaque development and destabilization. The vascular damage caused by AS can affect virtually all organs and has become a fundamental contributor to many common diseases in middle-aged and elderly individuals, posing a serious threat to human health and longevity. Taohong Siwu Decoction (THSWD), a classical formula for regulating menstruation, was first documented in the 'Golden Mirror of Medicine' compiled by Qian Wu, a renowned traditional Chinese medical practitioner during the Qing Dynasty. THSWD consists of *Angelica Sinensis Radix*, *Paeoniae Radix Alba*, *Chuanxiong Rhizoma*, *Rehmanniae Radix Preparata*, *Persicae Semen*, and *Carthami Flos*, and exerts dual effects of promoting blood circulation to resolve stasis and nourishing and replenishing blood. Modern pharmacological studies indicate that THSWD possesses anti-inflammatory<sup>1</sup>, anti-platelet<sup>2</sup>, anti-aggregation<sup>2</sup>, lipid-lowering<sup>3</sup>, antioxidant<sup>4</sup>, vascular endothelial protective<sup>5</sup>, hemorheological-improving properties<sup>6</sup>. These actions directly target key pathological stages of AS, exerting a positive regulatory effect on its pro-

gression. Consequently, THSWD has been clinically applied in the treatment of AS, and numerous studies have demonstrated its efficacy<sup>7-9</sup>, offering a safe and effective traditional Chinese medicine (TCM)-based approach for the prevention and management of AS.

A study<sup>10</sup> employed UPLC-Q-TOF-MS to qualitatively analyze and identify the components of THSWD, revealing that aromatic acids (e.g., ferulic acid, caffeic acid, coumaric acid, gallic acid), glycosides (e.g., paeoniflorin, albiflorin, rehmannioside, amygdalin), and phthalides (e.g., ligustilide) present in THSWD inhibit ADP-induced platelet aggregation and thrombin activity *in vitro*. These findings provide a scientific basis for THSWD's role in promoting blood circulation and resolving stasis. In TCM, *Persicae Semen* is believed to activate blood and dispel stasis. Experimental studies<sup>11</sup> have confirmed that *Persicae Semen* and its extracts reduce blood viscosity, enhance local blood flow, and improve hemorheological parameters. Its main active constituents include amygdalin and peach kernel oil. *Carthami Flos* promotes blood circulation, removes stasis, and relieves pain. A review<sup>12</sup> summarized recent findings on the chemical composition of *Carthami Flos*, identifying safflor yellow A, safflor yellow B, hydroxysafflor yellow A, quercetin, luteolin, baicalin, and kaempferol as primary components. *Chuanxiong Rhizoma* is recognized as a key herb with cardiovascular protective effects in clinical practice. Its principal bioactive compounds include ligustilide, senkyunolide, ferulic acid, and gallic acid<sup>13</sup>. *Angelica Sinensis Radix* invigorates blood and nourishes blood. As a member of the *Apiaceae* family, it shares considerable chemical similarity with *Chuanxiong Rhizoma*<sup>14</sup>. *Paeoniae Radix Alba* is traditionally used to nourish blood and regulate menstruation. Its bioactive constituents<sup>15</sup> include paeoniflorin, gallic acid, and albiflorin, with paeoniflorin

\* Corresponding author.

E-mail addresses: [lengjing2022szy@163.com](mailto:lengjing2022szy@163.com) (J. Leng); [ke.ning@vumc.org](mailto:ke.ning@vumc.org) (K. Ning)

being the most abundant compound and specific to the genus *Paeonia*. *Rehmanniae Radix Preparata* tonifies blood and nourishes yin. Research<sup>16</sup> has identified its chemical components as rehmannioside, catalpol, acteoside, sitosterol, and stigmasterol (Table 1).

In this review, we summarize the signaling pathways through which THSWD primarily exerts its therapeutic effects on AS (Table 2), discuss the roles and mechanisms of these pathways in AS pathogenesis, and outline how THSWD and its bioactive constituents modulate these signaling cascades. This synthesis is based on extensive experimental evidence and existing research, aiming to support future investigations in this field.

## 2. Signaling pathways in AS regulated by THSWD

### 2.1. Nuclear factor kappa-B (NF-κB) signaling pathway

#### 2.1.1. Overview of NF-κB signaling pathway

NF-κB is a family of inducible transcription factors involved in regulating diverse cellular processes such as proliferation, apoptosis, and inflammatory responses. It plays a critical role in innate and adaptive immunity by controlling the expression of cytokines and chemokines, and is widely recognized as a central regulator of inflammation.

The NF-κB family comprises five subunits: NF-κB1 p50, NF-κB2 p52, RelA (also known as p65), RelB, and c-Rel, which typically form hetero- or homodimers. These dimers are sequestered in the cytoplasm by inhibitory proteins known as inhibitor of NF-κB (IκB). Upon stimulation, they dissociate from IκB and translocate

to the nucleus to regulate target gene transcription<sup>17</sup>. Additionally, the precursor proteins p105 (NF-κB1) and p100 (NF-κB2) exhibit IκB-like functions due to their C-terminal structural similarity to IκB<sup>18</sup>.

This pathway operates via two distinct branches: the canonical and non-canonical pathways. The canonical pathway is activated by various receptors, including Toll-like receptors (TLRs)<sup>19</sup>, cytokine receptors, pattern recognition receptors (PRRs), tumor necrosis factor receptors (TNFRs), T cell receptors (TCRs), and B cell receptors (BCRs)<sup>20</sup>. Activation leads to transforming growth factor β (TGF-β)-activated kinase 1 (TAK1)-mediated phosphorylation of IκB kinase (IKK), resulting in IκBα ubiquitination and degradation. This releases p50/RelA and p50/c-Rel dimers, allowing nuclear translocation and initiation of gene transcription<sup>17</sup>. In contrast, the non-canonical pathway depends on p100 processing, primarily triggered by ligands of specific TNFR superfamily members such as LTβR, BAFF-R, and cluster of differentiation 40 (CD40). This process involves NF-κB-inducing kinase (NIK)-mediated phosphorylation of p100, leading to its partial proteolysis into p52 and subsequent nuclear translocation of p52/RelB dimers<sup>18</sup>.

#### 2.1.2. Role of NF-κB in AS

Studies<sup>21</sup> have shown that the NF-κB signaling pathway is activated in human AS, with elevated DNA binding activity of NF-κB observed in nuclei within atherosclerotic plaques. This activity predominantly involves p50, RelA, and c-Rel, subunits of the canonical pathway, and lacks p52 and RelB, indicating that the canonical NF-κB pathway primarily mediates AS development.

This pathway contributes to plaque formation by upregulating pro-inflammatory mediators such as tissue factor (TF) and

**Table 1** The effects of active ingredients from THSWD on signaling pathways of AS.

Active ingredient	Herb	Related signaling pathway	Mechanism	Outcome	Refs.
Amygdalin	Persicae Semen	NF-κB	NF-κB p65↓	Anti-inflammation	28
		NF-κB	NF-κB p65↓, TNF-α↓	Anti-inflammation	29
		PI3K/AKT	PI3K↑, AKT↑	Inhibition of apoptosis	49
Quercetin	Persicae Semen, Carthami Flos	TLRs	TLR4↓	Reduce cellular calcification	65
		MAPK	p38↓	Inhibition of macrophage senescence in plaques	79
		mTOR	mTOR↓	Activate autophagy	97
Luteolin	Carthami Flos	NF-κB	NF-κB p65↓	Anti-inflammation	30
Baicalin	Carthami Flos	MAPK	ERK1/2, JNK, p38↓	Anti-inflammation, Anti-oxidation	80
		NF-κB	NF-κB p65↓	Anti-inflammation	31
Ferulic acid	Chuanxiong Rhizoma, Angelicae Sinensis Radix	PI3K/AKT	PI3K↑, AKT↑	Inhibition of apoptosis	48
		TLRs	TLR4↓	Anti-inflammation	63
Senkyunolide A	Chuanxiong Rhizoma	NF-κB	NF-κB p65↓	Anti-inflammation	32
Ligustilide	Chuanxiong Rhizoma, Angelicae Sinensis Radix	MAPK	ERK, JNK, p38↓	Inhibition of cell proliferation	81
Paeoniflorin	Paeoniae Radix Alba	NF-κB	NF-κB p65↓	Anti-inflammation	33
		TLRs	TLR4↓, MyD88↓	Anti-inflammation	64
		MAPK	p38↓	Anti-inflammation	83
Kaempferol	Carthami Flos, Paeoniae Radix Alba	PI3K/AKT	PI3K/AKT/mTOR↓	Activate autophagy	50
		TLRs	TLR2 6↓, miR-5a-4p↑	Inhibition of apoptosis	66
Gallic acid	Chuanxiong Rhizoma, Paeoniae Radix Alba	MAPK	p38↓	Anti-inflammation	82
Paeonol	Paeoniae Radix Alba	mTOR	AMPK↑, mTOR↓	Activate autophagy, Inhibition of vascular smooth muscle cell proliferation	98
Catalpol	Rehmanniae Radix Preparata	NF-κB	NF-κB p65↓	Anti-inflammation	34–35
Acteoside	Rehmanniae Radix Preparata	NF-κB	NF-κB p65↓	Anti-inflammation	34–35

matrix metalloproteinases (MMPs). NF-κB activation also enhances the expression<sup>19</sup> of cytokines (tumor necrosis factor α (TNF-α), interleukin-1β (IL-1β), IL-6), chemokines (monocyte chemoattractant protein-1 (MCP-1)), and adhesion molecules (intercellular adhesion molecule-1 (ICAM-1) and vascular adhesion molecule-1 (VCAM-1)). These molecules promote leukocyte and monocyte recruitment to endothelial cells, where they differentiate into macrophages<sup>22</sup>, internalize lipids to become foam cells, and initiate AS lesion formation.

However, evidence<sup>23</sup> suggests that suppressing NF-κB activation in macrophages may reduce anti-inflammatory IL-10 production and increase cell death, potentially exacerbating AS progression.

2.1.3. Regulation of NF-κB by THSWD

2.1.3.1 THSWD-mediated regulation of NF-κB signaling pathway

A recent study<sup>24</sup> demonstrated that in type 2 diabetic rats, THSWD treatment significantly suppressed NF-κB p65 expression, reduced TNF-α protein and messenger ribonucleic acid (mRNA) levels in myocardial tissue, and lowered serum concentrations of IL-1β and IL-6, thereby attenuating inflammatory factor expression. Another study<sup>25</sup> found that THSWD downregulates PTGS2 and NFKB1 target proteins in the NF-κB pathway, contributing to its anti-inflammatory effects. THSWD has also been shown to synergistically inhibit NF-κB when combined with other decoctions. For instance, Erchen Decoction and THSWD exhibit anti-atherosclerotic effects, possibly through inhibition of the Nox4/NF-κB/HIF-1α pathway<sup>26</sup>. Similarly, Baoyuan Decoction combined with THSWD reduces inflammation and vascular fibrosis<sup>27</sup>. Collectively, these findings demonstrate that THSWD mitigates AS progression by suppressing the activation of the NF-κB signaling pathway (Fig. 1).

2.1.3.2 Bioactive components of THSWD targeting NF-κB

Amygdalin, derived from Persicae Semen, inhibits the NF-κB/COX-2 pathway, thereby reducing inflammatory mediator expression, improving lipid profiles, and exerting anti-atherosclerotic effects<sup>28</sup>. Quercetin, an active constituent in both Carthami Flos and Persicae Semen, suppresses nuclear translocation of the NF-κB p65 subunit and TNF-α release, blocking downstream gene expression and attenuating inflammation in AS<sup>29</sup>. Luteolin, a bioactive compound in Carthami Flos, inhibits IκBα degradation and subsequent p65 nuclear translocation, thus suppressing NF-κB signaling<sup>30</sup>. Ferulic acid, present in Angelicae Sinensis Radix and Chuanxiong Rhizoma, restrains NF-κB activation, modulates NF-κB-dependent expression of VCAM-1 and ICAM-1, and reduces inflammation<sup>31</sup>. Senkyunolide A from Chuanxiong Rhizoma significantly inhibits TNF-α expression, thereby interfering with NF-κB pathway activation<sup>32</sup>. Paeoniflorin, the major component of Paeoniae Radix Alba, prevents inflammatory responses by inhibiting oxidized (ox) LDL-induced phosphorylation of the NF-κB p65 subunit, thereby delaying AS progression<sup>33</sup>. Catalpol and acetoside, key constituents of Rehmanniae Radix Preparata, have

been shown<sup>34,35</sup> to inhibit p65 nuclear translocation in the NF-κB pathway, markedly reducing downstream inflammatory factor synthesis and alleviating the inflammatory response.

2.2. Phosphatidylinositol 3-kinase (PI3K)/serine-threonine kinase (AKT) signaling pathway

2.2.1. Overview of PI3K/AKT signaling pathway

The PI3K/AKT (also known as protein kinase B or PKB) pathway is a key intracellular signal transduction cascade regulating essential cellular functions such as protein synthesis, metabolism, angiogenesis, proliferation, apoptosis, and autophagy. It is intricately linked to inflammation, growth, and survival processes<sup>36</sup>.

Central metabolites in this pathway include phosphatidylinositol-4,5-bisphosphate (PIP2) and phosphatidylinositol-3,4,5-trisphosphate (PIP3). Key regulatory genes are phosphatase and tensin homolog (PTEN) and phosphoinositide-dependent kinase-1 (PDK1). PI3K, a lipid kinase, is activated upon binding of growth factors, such as vascular endothelial growth factor (VEGF), epidermal growth factor (EGF), and hepatocyte growth factor (HGF), to their respective receptors<sup>36</sup>. Activated PI3K catalyzes the conversion of PIP2 to PIP3<sup>37</sup>. PTEN counteracts this process by dephosphorylating PIP3 at the 3' position, thereby limiting AKT activation and acting as a negative regulator<sup>38</sup>.

PIP3 serves as a second messenger, recruiting PH domain-containing proteins like phosphoinositide-dependent protein kinase-1 (PDK1) and AKT to the plasma membrane. PDK1 then phosphorylates AKT, leading to partial activation<sup>36</sup>. Fully activated AKT regulates downstream effectors: it activates FOXO and GSK3 to control proliferation, and Tsc, Rheb, and mammalian target of rapamycin (mTOR) to regulate protein synthesis<sup>39</sup>. AKT also activates IKK, establishing crosstalk with the NF-κB pathway<sup>40</sup>.

2.2.2. Role of PI3K/AKT in AS

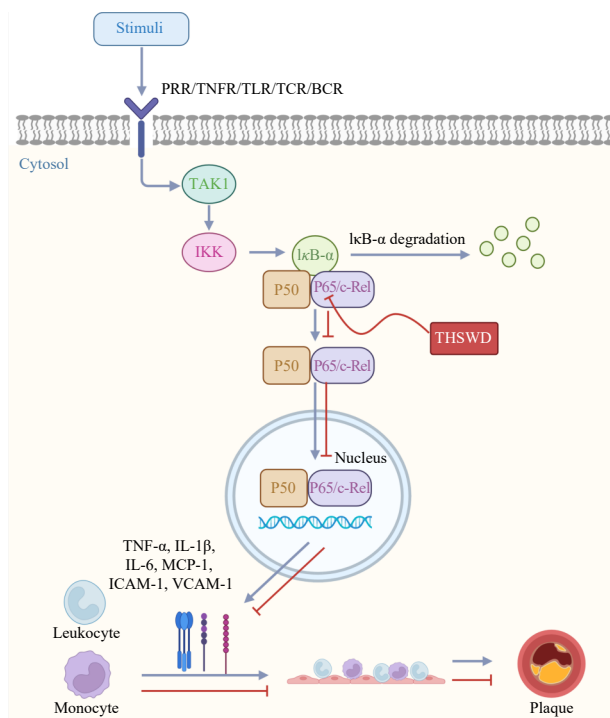
Substantial evidence indicates that PI3Kγ, a key immune regulator<sup>39</sup>, drives inflammatory responses in macrophages, neutrophils, and mast cells, thereby contributing to AS pathogenesis.

The PI3K/AKT pathway also influences macrophage polarization. Macrophages exist in two primary phenotypes: M1 and M2. Studies<sup>41</sup> show that M1 macrophages promote plaque development and instability, whereas M2 macrophages suppress inflammation<sup>42</sup> and are protective against AS progression. Deletion of the AKT1 isoform promotes M1 polarization and accelerates AS, while AKT2 deletion favors M2 differentiation and slows disease progression<sup>43</sup>.

Macrophage apoptosis has dual roles in AS. In early lesions, apoptosis reduces foam cell accumulation and limits plaque expansion. However, in advanced plaques, macrophage apoptosis enlarges the necrotic core and increases the risk of rupture<sup>44</sup>.

Table 2 Different mechanisms of signaling pathway regulation by THSWD.

Signaling pathway	Mechanism	Outcome	Refs.
NF-κB	NF-κB p65↓, TNF-α, IL-1β, IL-6↓		24
NF-κB	PTGS2, NFKB1↓	THSWD plays an anti-inflammatory role by regulating the NF-κB signaling pathway.	25
NF-κB	NF-κB p65↓, HIF-1α↓		26
PI3K/AKT	PI3K, AKT↑, eNOS↑, NO↑	THSWD plays an antioxidant and anti-inflammatory role by regulating the PI3K/AKT signaling pathway.	46, 47
TLRs	TLR4↓, NF-κB↓	THSWD plays an anti-inflammatory role by regulating the TLR signaling pathway.	60, 61, 62
MAPK	JNK, p38↓	THSWD inhibits pyroptosis by regulating the MAPK signaling pathway.	60, 78
mTOR	PI3K/AKT↓, mTOR↓		96
mTOR	AMPK↑, mTOR↓	THSWD activates autophagy and reduces inflammatory response by regulating the mTOR signaling pathway.	97



**Fig. 1** Mechanism of THSWD regulating NF- $\kappa$ B signaling pathway in the treatment of AS. THSWD can inhibit the nuclear translocation of NF- $\kappa$ B p65 subunit, thus blocking the expression of related inflammatory factors, inhibiting leukocytes, monocytes, etc. from accumulating in the endothelial cells and differentiating into macrophages, which plays a role in inhibiting the formation of atherosclerotic plaques.

Since AKT signaling inhibits apoptosis<sup>38</sup>, its activation may exert opposing effects depending on disease stage<sup>44</sup>.

The activated AKT can catalyze the phosphorylation of eNOS at Ser-1177 and promote the release of NO<sup>45</sup>. NO can suppress the release and expression of inflammatory factors, the oxidation of LDL, the expression of cell adhesion protein, the migration and proliferation of vascular smooth muscle cells<sup>6</sup>, which acts as an antioxidant and anti-inflammatory agent and inhibits the aggravation of AS. AKT signaling in macrophages functions as an inhibitor of apoptosis<sup>38</sup>, which may affect different processes of AS differently.

Among the downstream molecules activated by this signaling pathway, it is worth mentioning eNOS and mTOR (the specific effects of this pathway will be described in detail in the mTOR signaling pathway). Activated AKT can catalyze the phosphorylation of eNOS at Ser-1177 and promote the release of NO<sup>45</sup>, which can inhibit the release and expression of inflammatory factors, LDL oxidation, the expression of cell adhesion molecules (CAMs) expression, and the migration and proliferation of vascular smooth muscle cells (VSMCs)<sup>6</sup>, and play an antioxidant and anti-inflammatory role and inhibit the aggravation of AS.

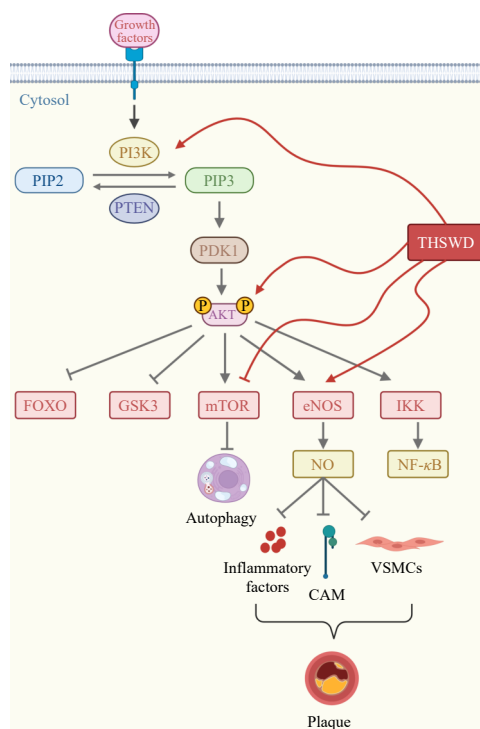
Collectively, the role of the PI3K/AKT pathway in AS is complex and context-dependent, with no single mechanism clearly dominating its overall impact.

### 2.2.3. Regulation of PI3K/AKT by THSWD

#### 2.2.3.1 THSWD-mediated regulation of PI3K/AKT signaling pathway

A study<sup>46</sup> using Erchen Decoction combined with THSWD to treat ox-LDL-induced endothelial injury found significantly increased expression of PI3K and eNOS proteins and enhanced AKT phosphorylation in the intervention group compared to controls, along with reduced apoptosis. When a PI3K inhibitor (LY294002) was introduced, these effects were reversed, confirming the involvement of the PI3K/AKT pathway. Another study<sup>47</sup> showed that THSWD activates eNOS and elevates serum NO levels via

PI3K/AKT signaling, suggesting that THSWD's therapeutic effect in AS may largely depend on eNOS activation and NO release through this pathway (Fig. 2).



**Fig. 2** Mechanism of THSWD regulating PI3K/AKT signaling pathway in the treatment of AS. In the downstream signaling, it inhibits the aggravation of AS by promoting the phosphorylation of eNOS and stimulating its release of NO, thereby inhibiting the release of inflammatory factors, the expression of CAM, and the migration and proliferation of VSMCs. THSWD can also enhance cellular autophagy by inhibiting the PI3K/AKT/mTOR signaling pathway, thereby stabilizing plaque.

#### 2.2.3.2 Bioactive components of THSWD targeting PI3K/AKT

Ferulic acid has been shown<sup>48</sup> to enhance PI3K and AKT expression in cardiomyocytes, activating the pathway to prevent apoptosis and inhibit early plaque formation. Quercetin similarly enhances AKT activity and suppresses apoptosis<sup>49</sup>. Kaempferol, present in *Paoniae Radix Alba* and *Carthami Flos*, has been found<sup>50</sup> to inhibit the PI3K/AKT/mTOR pathway, thereby enhancing autophagy and stabilizing atherosclerotic plaques.

### 2.3. Toll-like receptor (TLR) signaling pathway

#### 2.3.1. Overview of the TLR signaling pathway

TLRs are a class of PRRs belonging to the type I transmembrane protein family. Their extracellular domains contain leucine-rich repeats (LRRs) and a Toll-interleukin-1 receptor (TIR) domain<sup>51</sup>, enabling recognition of pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs)<sup>52</sup>. This recognition initiates downstream signaling cascades critical for both innate and adaptive immunity. TLR expression profoundly influences inflammatory responses and macrophage differentiation.

TLR signaling proceeds via two main pathways: myeloid differentiation primary response gene 88 (MyD88)-dependent and TRIF-dependent. Except for TLR3, all TLRs utilize the adaptor protein MyD88 via Toll-IL-1 receptor (TIR) domain containing adaptor protein (TIRAP)<sup>52</sup>, leading to activation of NF- $\kappa$ B and mitogen-activated protein kinases (MAPKs), which stimulate pro-inflammatory cytokine production<sup>53</sup>. TLR3 and TLR4 can use the TRIF-dependent pathway: TLR3 directly recruits TRIF, while TLR4 requires TRAM as a bridging adaptor<sup>54</sup>. TRIF activates

TRAF6 and TRAF3, which in turn induce NF- $\kappa$ B and MAPKs or activate IRF3/5 to promote interferon- $\beta$  expression<sup>51</sup>.

### 2.3.2. Role of TLRs in AS

Among TLRs, TLR4 is most closely associated with AS development. Research<sup>55</sup> indicates that increased lipid accumulation elevates TLR4 levels and enhances MyD88/TRIF signaling, accelerating AS progression and inflammation. Recent studies<sup>56</sup> show that ox-LDL binds to myeloid differentiation factor 2 (MD2) on macrophages, facilitating TLR4 activation and triggering downstream inflammatory cascades.

TLR4 mediates inflammation in multiple cell types. Monocyte accumulation at damaged sites is a hallmark of inflammation. Studies<sup>57</sup> report elevated TLR4 expression on circulating mononuclear cells in AS patients. TLR4 also mediates ox-LDL-induced macrophage activation, with Sub1 identified as a key regulator in macrophage-driven AS, likely through upstream regulation of IRF1 and promotion of M1 polarization, leading to plaque expansion<sup>58</sup>. Dendritic cell maturation is crucial in inflammatory cascades involving lipid accumulation, foam cell formation, and antigen presentation. Upregulation of TLR4 on dendritic cells activates endogenous responses and worsens AS<sup>59</sup>.

### 2.3.3. Regulation of TLRs by THSWD

#### 2.3.3.1 THSWD-mediated regulation of the TLR signaling pathway

A study<sup>60</sup> demonstrated that THSWD ameliorates ischemic stroke injury by inhibiting pyroptosis, and further revealed that THSWD suppresses TLR4 activation and downstream signals NF- $\kappa$ B and MAPK, thereby reducing inflammatory factor expression. In a rat model of rheumatoid arthritis<sup>61</sup>, THSWD reduced joint swelling by downregulating the TLR4/NF- $\kappa$ B pathway. Another study<sup>62</sup> confirmed that THSWD intervention decreased mRNA and protein levels of TLR4, MyD88, and NF- $\kappa$ B in the model group. Thus, THSWD primarily attenuates inflammation by suppressing TLR4 and its downstream signaling components, slowing AS progression (Fig. 3).

#### 2.3.3.2 Bioactive components of THSWD targeting TLRs

Network pharmacology analysis<sup>63</sup> indicated that ferulic acid's anti-AS mechanism is closely linked to the TLR4 pathway, with TLR4 being a core target. Paeoniflorin reduces expression of

TLR4 and MyD88, and inhibits I $\kappa$ B and NF- $\kappa$ B p65 phosphorylation<sup>64</sup>, thereby interrupting the inflammatory cascade. Quercetin attenuates ox-LDL-induced oxidative stress by dampening TLR4 signaling and reducing calcification<sup>65</sup>. Kaempferol not only modulates TLR4 but also alleviates ox-LDL-induced endothelial apoptosis by inhibiting the TLR2/6/NF- $\kappa$ B signaling pathway and up-regulating miR-5a-4p, thereby exerting anti-atherosclerotic effects<sup>66</sup> to alleviate ox-LDL-induced endothelial apoptosis by suppressing the TLR2/6/NF- $\kappa$ B pathway and upregulating miR-5a-4p, thus inhibiting AS.

### 2.4. MAPK signaling pathway

#### 2.4.1. Overview of the MAPK signaling pathway

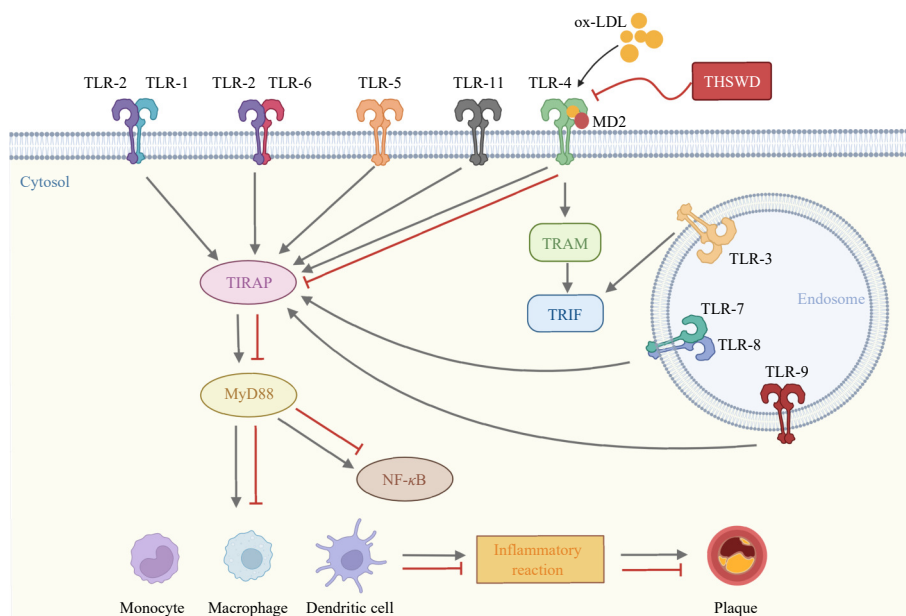
MAPKs are AKTs activated by extracellular stimuli such as cytokines, neurotransmitters, and hormones. They regulate vital cellular processes, including growth, differentiation, development, and apoptosis. The MAPK cascade involves three sequential phosphorylation events: MAPK kinase kinase (MKKK), MAPK kinase (MKK), and finally MAPK<sup>67</sup>, transmitting signals to the nucleus to initiate gene transcription. The best-characterized MAPK families are extracellular signal-regulated kinases 1/2 (ERK1/2), c-jun N-terminal kinases/stress-activated protein kinases (JNK/SAPK), and p38 (with  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$  isoforms).

In the ERK1/2 pathway, Ras-GTP binds Raf (the MKKK), which activates MEK1/2, subsequently phosphorylating ERK1/2 to execute its function<sup>68</sup>. The JNK/SAPK pathway is stimulated by ionizing radiation, heat shock, or inflammatory cytokines, with JNK activation mediated by mainly affected by MEK4 (MKK4) and MEK7 (MKK7)<sup>67</sup>; this upstream signaling overlaps with the p38 pathway. Similarly, the p38 pathway is activated by UV radiation, cytokines, or lipopolysaccharide. Signal transduction leads to phosphorylation of p38 primarily by MKK3 and MKK6, with MKK4 also contributing partially<sup>69</sup>.

The ERK pathway is generally associated with cell proliferation and differentiation<sup>70,71</sup>, whereas JNK and p38 pathways regulate not only proliferation but also inflammatory responses<sup>72</sup>.

#### 2.4.2. Role of MAPK in AS

The p38 MAPK pathway is particularly implicated in AS. It in-



**Fig. 3** Mechanism of THSWD regulating the TLR signaling pathway in the treatment of AS. THSWD is able to reduce the expression of TLR4 and MyD88, thus inhibiting the transduction of the TLR4/NF- $\kappa$ B signaling pathway. Moreover, THSWD can decrease the occurrence of inflammatory responses in different cells, such as monocytes, macrophages, and dendritic cells, by inhibiting the signaling of the MyD88-dependent pathway, which ultimately suppresses the formation of atherosclerotic plaques.

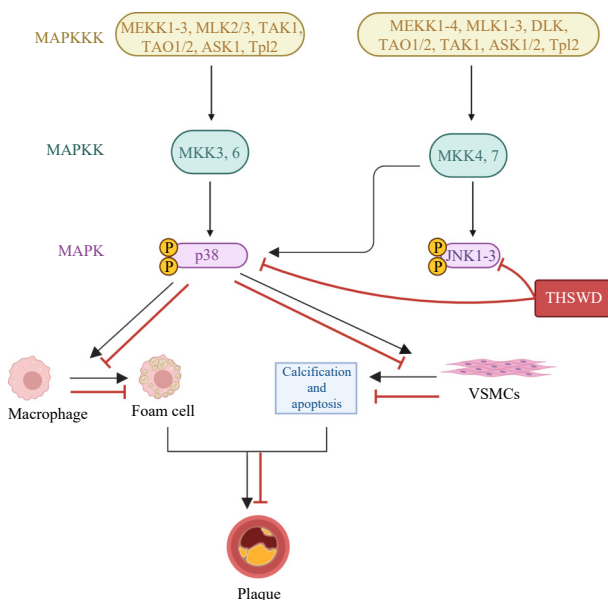
duces expression of inflammatory mediators such as VCAM-1<sup>73</sup> and MCP-1<sup>74</sup>, promoting inflammatory cell infiltration and accelerating AS. Beyond inflammation, vascular calcification contributes to AS progression. Ceramide has been shown<sup>75</sup> to promote ox-LDL-induced VSMC calcification and apoptosis via p38 MAPK activation.

In addition to endothelial and VSMCs, the MAPK pathway is activated in macrophages. Ox-LDL induces p38-mediated macrophage proliferation, leading to foam cell formation and accelerating plaque development and rupture<sup>76</sup>. CD137 signaling has also been shown<sup>77</sup> to trigger mitochondrial fission via p38 MAPK, generating reactive oxygen species from dysfunctional mitochondria and promoting macrophage apoptosis, thereby expanding unstable plaques.

#### 2.4.3. Regulation of MAPK by THSWD

##### 2.4.3.1 THSWD-mediated regulation of MAPK signaling pathway

A study<sup>60</sup> reported that THSWD treatment significantly reduced JNK and p38 MAPK levels in rats with middle cerebral artery occlusion/reperfusion (MCAO/R), accompanied by decreased levels of inflammatory cytokines (TNF- $\alpha$ , IL-2, IL-6, TGF- $\beta$ , IL-1 $\beta$ , IL-18). This suggests that THSWD may inhibit pyroptosis by suppressing the MAPK pathway. Another study<sup>78</sup>, integrating network pharmacology, molecular docking, and experimental validation, confirmed that THSWD mitigates pyroptosis in ischemia-reperfusion injury by blocking MAPK signaling (Fig. 4).



**Fig. 4** Mechanism of THSWD regulating the MAPK signaling pathway in the treatment of AS. THSWD mainly inhibits the p38 MAPK signaling pathway, thus preventing macrophages from phagocytosis of lipids into foam cells as well as preventing VSMCs from undergoing calcification and apoptosis, and ultimately slowing down the production and rupture of atherosclerotic plaque. On the other hand, THSWD can also affect the JNK pathway, which may be related to the inhibition of inflammatory responses.

##### 2.4.3.2 Bioactive components of THSWD targeting MAPK

Quercetin has been shown<sup>79</sup> to alleviate ox-LDL-induced senescence in plaque macrophages by inhibiting p38 MAPK/p16 phosphorylation, thereby slowing AS progression. Baicalin, a component of Carthami Flos, reduces activation of ERK1/2, JNK, and p38, indicating suppression of the MAPK pathway and conferring anti-inflammatory and antioxidant effects<sup>80</sup>. Ligustilide, present in Chuanxiong Rhizoma and Angelicae Sinensis Radix, dose-dependently suppresses ERK, JNK, and p38 activation, suggesting its role in modulating cell proliferation via MAPK inhibition<sup>81</sup>. Gallic acid, found in Chuanxiong Rhizoma and Paeoniae Radix Alba, attenuates inflammation by blocking p38 MAPK activation<sup>82</sup>. Paeoniflorin improves cardiac function in chronic heart failure by inhibiting the p38 MAPK pathway<sup>83</sup>.

ation<sup>82</sup>. Paeoniflorin improves cardiac function in chronic heart failure by inhibiting the p38 MAPK pathway<sup>83</sup>.

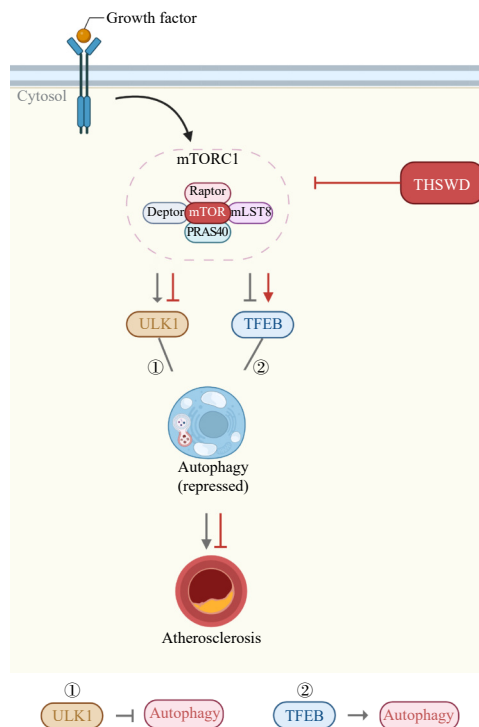
#### 2.5. mTOR signaling pathway

##### 2.5.1. Overview of the mTOR signaling pathway

mTOR is an atypical serine/threonine protein kinase belonging to the phosphatidylinositol kinase-related kinase (PIKK) family<sup>84</sup>. It integrates diverse extracellular signals to regulate cell proliferation, autophagy, and apoptosis, processes critically involved in AS plaque stability (Fig. 5).

mTOR forms two distinct complexes: mTOR complex 1 (mTORC1) and mTORC2. mTORC1 consists of five components<sup>84</sup>: mTOR (catalytic subunit), mLST8 (G $\beta$ L), Raptor (substrate-recruiting protein via TOS motif<sup>85</sup>), PRAS40, and Deptor. The first three are core components; the latter two are inhibitory. mTORC2 contains six subunits<sup>86</sup>: mTOR, mLST8, Rictor, mSIN1, Protor1/2, and Deptor. The first four constitute the core. Notably, mLST8 is dispensable for mTORC1 activity but essential for mTORC2 function<sup>87</sup>.

mTORC1 is regulated by growth factors, amino acids, oxygen, and energy status<sup>88</sup>, and controls downstream processes such as protein, lipid, and nucleotide synthesis, as well as autophagy and catabolism. Key effectors include S6K1, 4EBP, sterol response element binding protein (SREBP), ULK1, and transcription factor EB (TFEB)<sup>84</sup>. mTORC2 is less well understood but is associated with cell survival, proliferation, and cytoskeletal remodeling. Its upstream signals originate mainly from insulin/PI3K signaling, and its downstream targets belong to the AGC kinase family. Notably, AKT and mTORC2 engage in a positive feedback loop: partial AKT activation promotes mTORC2 activation<sup>89</sup>, which in turn fully activates AKT via phosphorylation.



**Fig. 5** Mechanism of THSWD regulating the mTOR signaling pathway in the treatment of AS. Cellular autophagy prevents endothelial cells from being damaged by exogenous lipid accumulation in the vascular wall and vascular stress, while ULK1 in the mTORC1 signaling pathway can inhibit autophagy, and TFEB can promote autophagy. THSWD can influence the mTORC1 signaling pathway to inhibit ULK1 phosphorylation, thus promoting the formation of autophagosomes. At the same time, THSWD can also stimulate the nuclear translocation of TFEB to promote cellular autophagy and ultimately inhibit the formation of AS.

2.5.2. Role of mTOR in AS

Autophagy protects against exogenous lipid accumulation and vascular stress in endothelial cells<sup>90</sup>, thereby preventing AS. mTORC1 negatively regulates autophagy: adenosine 5'-monophosphate-activated protein kinase (AMPK)-mediated ULK1 activation is essential for autophagosome formation, but mTORC1 directly phosphorylates ULK1 to inhibit this process<sup>91-92</sup>. mTORC1 also suppresses autophagy<sup>93</sup> by blocking TFEB nuclear translocation. Foam cell formation, a hallmark of AS, is promoted by mTOR via inhibition of autophagy<sup>92</sup>.

AS is a chronic inflammatory condition involving immune cells. In T cells, mTOR inhibition promotes a shift from effector to memory phenotype and suppresses CD8<sup>+</sup> T cell inflammation, contributing to plaque stabilization<sup>94</sup>. Additionally, mTOR modulates specific leukocyte subsets and inhibits immune cell proliferation, thereby dampening inflammation and slowing AS progression<sup>95</sup>.

2.5.3. Regulation of mTOR by THSWD

2.5.3.1 THSWD-mediated regulation of the mTOR signaling pathway

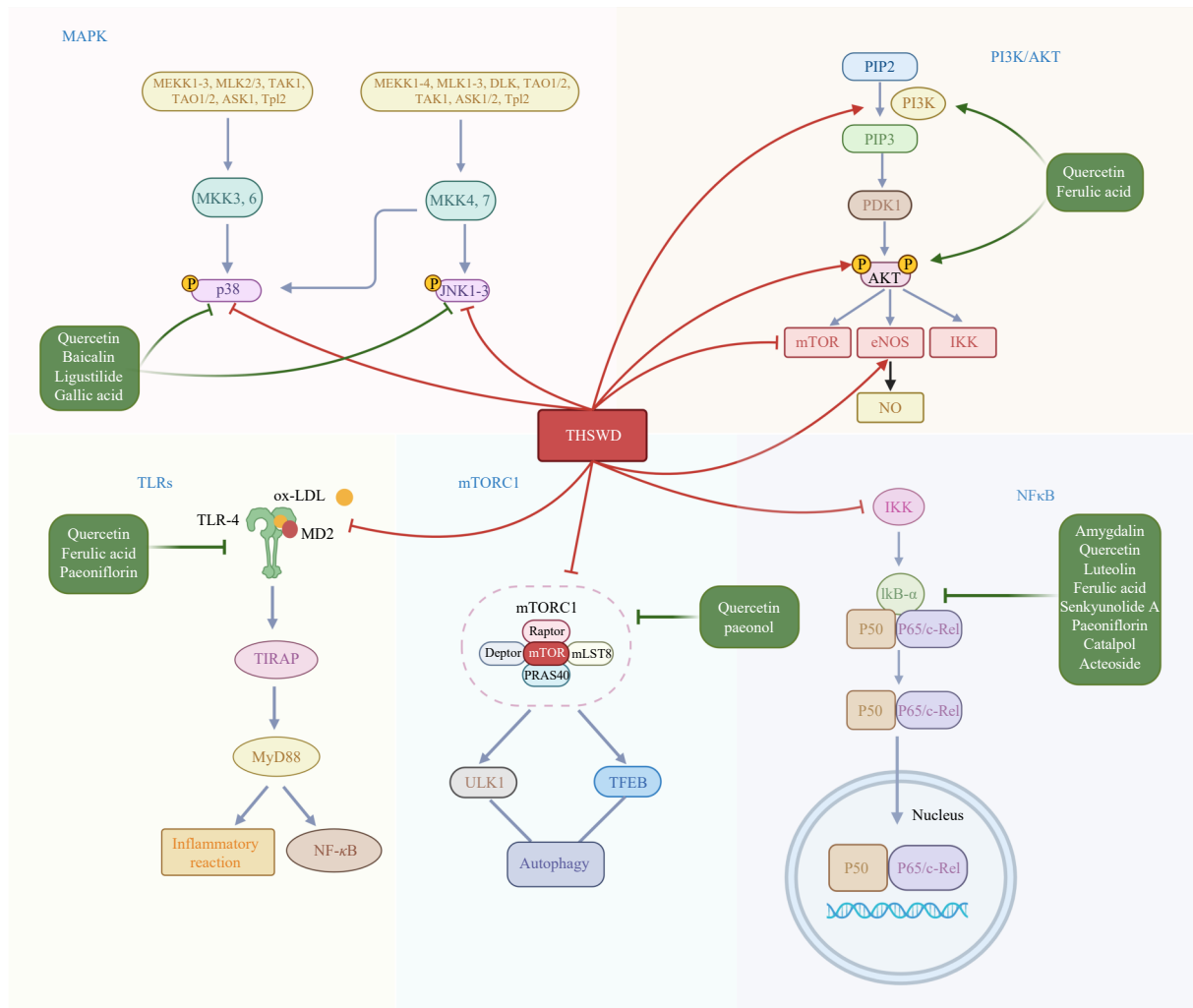
A study<sup>96</sup> found that medium and high doses of THSWD significantly reduced mTOR mRNA and protein levels in rabbit tendon-bone interface tissues, indicating that THSWD suppresses mTOR expression, possibly via the PI3K/AKT/mTOR pathway (Fig. 6).

2.5.3.2 Bioactive components of THSWD targeting mTOR

Quercetin<sup>97</sup> suppresses mTOR expression, enhancing autophagy and reducing aortic root lipid accumulation in mice, thereby exerting protective effects against AS. Paoniflorin activates AMPK and inhibits mTOR phosphorylation, promoting autophagy and suppressing VSMC proliferation<sup>98</sup>.

3. Conclusion and prospect

In summary, the signaling pathways regulating AS described in this article have a close relationship with the treatment of THSWD, which can reduce the release of inflammatory factors, inhibit macrophage proliferation, reduce cellular calcification, and enhance cellular autophagy by promoting or inhibiting the above signaling pathways (Fig. 6). These pathways are likely to be the potential therapeutic targets of THSWD for AS. However, a large number of studies have found that there are numerous signaling pathways that can regulate the process of AS, such as the Wnt signaling pathway<sup>99-100</sup>, the NLRP3 inflammasome<sup>101-102</sup>, the RhoA/ROCK signaling pathway<sup>103-104</sup>, the JAK/STAT signaling pathway<sup>105</sup>, the Notch signaling pathway<sup>106-107</sup>, and the VEGF signaling pathway<sup>108-109</sup>. However, few studies have correlated these pathways with the treatment of AS through THSWD. Several key questions remain to be addressed, including whether THSWD exerts therapeutic effects on atherosclerosis (AS) via the aforementioned pathways, and what specific molecular targets



**Fig. 6** The comprehensive connection between the signaling pathways of AS regulated by THSWD. THSWD can have different effects on NF-κB, PI3K/AKT, TLRs, MAPK, mTOR signaling pathways, and their downstream signaling molecules, and ultimately play a role in inhibiting the formation and development of atherosclerotic plaques. Among them, NF-κB, PI3K/AKT, TLR, and mTOR signaling pathways are interconnected and interact with each other, and are not a single independent signaling pathway.

and active components of THSWD are involved in these mechanisms. Further in-depth investigation is required to clarify these issues. Based on the potential cross-regulatory mechanisms between pathways that have not been thoroughly studied (e.g., Wnt, NLRP3, etc.) and those with well-elucidated mechanisms (e.g., NF- $\kappa$ B, TLR4 pathway, etc.), it provides the possibility of in-depth investigation and elaboration of the relationship between them and the treatment of AS with THSWD. For instance, in the Wnt signaling pathway, it has been pointed out<sup>110</sup> that Wnt signaling transduction is inextricably linked to the NF- $\kappa$ B and the TLR4 pathway during the regulation of inflammatory responses. Additionally, research<sup>111</sup> has also demonstrated that Wnts are able to activate p38 MAPK, and that the two interact with each other. In the NLRP3 inflammasome, study<sup>112</sup> concluded that MyD88- and TRIF-induced caspase-11 synthesis contributes to the activation of the NLRP3 inflammasome, suggesting a connection between the TLR4 signaling pathway and the NLRP3 inflammasome. In the RhoA/ROCK signaling pathway, Rho-kinase was shown<sup>113</sup> to inhibit the PI3K/AKT signaling pathway, thereby affecting downstream eNOS phosphorylation and NO release. It has been demonstrated<sup>114</sup> that the JAK/STAT signaling pathway exhibits crosstalk with the MAPK, PI3K/AKT, and NF- $\kappa$ B signaling pathways. Regarding the Notch signaling pathway, study<sup>115</sup> has revealed a positive feedback loop between Notch signaling and STAT3 after TLR activation, which involves several signaling pathways, including the TLR4, NF- $\kappa$ B, and JAK/STAT pathways. Additionally, research<sup>108</sup> has also demonstrated that VEGF activates STAT-3, NF- $\kappa$ B, and Akt pathways, creating crosstalk between the vascular and the inflammatory response.

A growing body of evidence indicates that TCM formulations offer significant advantages in managing complex, multifactorial chronic diseases. Unlike Western medicine's single-target approach, TCM treatment is guided by a holistic philosophy, enabling multi-target, multi-level, multi-pathway, and systemic regulation. The conventional drug development model in Western medicine typically follows a linear sequence: identification of disease-related targets, discovery of lead compounds, optimization of candidates, and subsequent pharmacodynamic, pharmacological, and clinical evaluation. While systematic, this approach may fail to capture the dynamic, staged, and multifactorial nature of chronic conditions such as AS<sup>116</sup>. In contrast, under the principles of "holistic regulation" and "treatment based on syndrome differentiation," TCM employs the classical formulation theory of "sovereign, minister, assistant, and courier" to rationally combine herbs. This enables TCM formulas to comprehensively regulate physiological functions, restore homeostasis, strengthen host defense, eliminate pathogenic factors<sup>117</sup>, and simultaneously address both the symptoms and root causes of disease.

Nevertheless, it has been argued that the clinical application of TCM formulations lacks reproducibility and objective mechanistic validation when based solely on empirical observation and anecdotal efficacy. Therefore, elucidating the precise roles of TCM compounds through the lens of signaling pathways can provide robust scientific evidence for their therapeutic mechanisms. Furthermore, it is anticipated that future research will build upon this review to identify the primary bioactive components in THSWD. By screening and optimizing these constituents according to the principles of enhancing efficacy and minimizing toxicity, it may be possible to advance from crude herb compatibility to rational combinations of purified active ingredients. Such secondary development could yield formulations with superior pharmacodynamic profiles compared to the original prescription, while ensuring greater clarity and control over the key pharmacologically active substances.

The in-depth investigation of TCM formulations is not only essential for ensuring their safety and efficacy, promoting accurate understanding of herbal medicines, and objectively interpret-

ing their disease-modifying mechanisms<sup>118</sup>, but also holds broad implications for modern drug development and the global promotion of integrative preventive medicine. It is imperative to recognize that single-target interventions are often insufficient for preventing or treating increasingly prevalent complex chronic diseases. A paradigm shift is needed, one that emphasizes the value of comprehensive, systems-level approaches and fosters greater appreciation for the integrative effects of TCM formulations.

## Funding

This work was supported by the National Natural Science Foundation of China (Nos. 82104430 and 82274133), the Shanghai Sailing Program (No. 21YF1447600), and the Future Plan for Traditional Chinese Medicine Development of Science and Technology of Shanghai Municipal Hospital of Traditional Chinese Medicine (No. WL-HBQN-2022002K).

## Declaration of competing interest

These authors have no conflict of interest to declare.

## Acknowledgements

We gratefully acknowledge the use of BioRender for the creation of the scientific illustrations in this study.

## References

- Zhang B. Effect of medicated serum prepared with Taohong Siwu Decoction on expressions of TNF- $\alpha$ , MCP-1 and IL-1 $\beta$  of human umbilical vein endothelial cells treated with lipopolysaccharide. *Liaoning J Tradit Chin Med.* 2014;41:2280-2283. <https://doi.org/10.13192/j.issn.1000-1719.2014.11.004>.
- Han L, Peng DY, Xu F, et al. Studies on anti-platelet activation and partial mechanism of Taohong Siwu Decoction. *China J Chin Mater Med.* 2010;35(19):2609-2612. <https://doi.org/10.4268/cjcm20101924>.
- Lai LN, Zhou XQ. Discussion on the mechanism of Taohong Siwu Decoction in treating dyslipidemia based on network pharmacology. *J Liaoning Univ Tradit Chin Med.* 2021;23(4):72-78. <https://doi.org/10.13194/j.issn.1673-842x.2021.04.017>.
- Luo ZR, Li H, Xiao ZX, et al. Taohong Siwu Decoction exerts a beneficial effect on cardiac function by possibly improving the microenvironment and decreasing mitochondrial fission after myocardial infarction. *Cardiol Res Pract.* 2019;2019:5198278. <https://doi.org/10.1155/2019/5198278>.
- Li RS, Li DY, Chen WN, et al. Taohong Siwu Decoction regulated functions of endothelial cells and treated arteriosclerosis obliterans: an experimental study. *Chin J Integr Med.* 2014;34(2):191-196. <https://doi.org/10.7661/CJIM.2014.02.0191>.
- Kattoor AJ, Pothineni NVK, Palagiri D, et al. Oxidative stress in atherosclerosis. *Curr Atheroscler Rep.* 2017;19(11):42. <https://doi.org/10.1007/s11883-017-0678-6>.
- Wei SG. Observation on the clinical efficacy of Taohong Siwu Decoction combined with conventional drugs in the treatment of coronary atherosclerotic heart disease and angina pectoris. *Cardiovasc Dis J Integr Tradit Chin West Med.* 2018;6(29):111-112. <https://doi.org/10.16282/j.cnki.cn11-9336/r.2018.29.084>.
- Wang CB, Hu XH, Li L. Clinical observation of Taohong Siwu Decoction combined with cilostazol in treating ASO of lower extremity. *Acta Chin Med Pharmacol.* 2020;48(11):63-67. <https://doi.org/10.19664/j.cnki.1002-2392.200199>.
- Yang QH. 47 Cases of coronary artery disease and angina treated in modified Peach Seed and Safflower Decoction of Four Drugs. *J Henan Univ Chin Med.* 2008;4:77-78. <https://doi.org/10.16368/j.issn.1674-8999.2008.04.021>.
- Liu L, Duan JA, Su SL, et al. Effect of different fractions of Taohong Siwu Decoction on ADP-induced platelet aggregation and thrombin activity. *China J Chin Mater Med.* 2016;41(4):716-721. <https://doi.org/10.4268/cjcm20160429>.
- Liu L, Duan JA, Tang Y, et al. Taoren-Honghua herb pair and its main components promoting blood circulation through influencing on hemorheology, plasma coagulation and platelet aggregation. *J Ethnopharmacol.* 2012;139(2):381-387. <https://doi.org/10.1016/j.jep.2011.11.016>.
- Yang Y, Huang XL, Jiang ZM, et al. New research progress for chemical compositions and pharmacological effect of Honghua (Carthami Flos). *Chin Arch Tradit Chin Med.* 2023;41(10):119-126. <https://doi.org/10.13193/j.issn.1673-7717.2023.10.024>.
- Li WX, Tang YP, Chen YY, et al. Advances in the chemical analysis and biological activities of chuanxiang. *Molecules.* 2012;17(9):10614-10651.

- <https://doi.org/10.3390/molecules170910614>.
- 14 Lyu CL, Li HH, Shi YJ, et al. Research progress of *Angelicae Sinensis Radix* and predictive analysis on its quality markers. *China J Chin Mater Med*. 2022;47(19):5140-5157. <https://doi.org/10.19540/j.cnki.cjcm.20220225.203>.
  - 15 Yan BJ, Shen ML, Fang JY, et al. Advancement in the chemical analysis of *Paeoniae Radix* (Shaoyao). *J Pharm Biomed Anal*. 2018;160:276-288. <https://doi.org/10.1016/j.jpba.2018.08.009>.
  - 16 Ge N, Yan GL, Sun H, et al. Research progress on effective constituents in *Radix Rehmanniae Praeparata*. *Chin Tradit Herb Drugs*. 2023;54(1):292-302. <https://doi.org/10.7501/j.issn.0253-2670.2023.01.031>.
  - 17 Liu T, Zhang LY, Joo DH, et al. NF- $\kappa$ B signaling in inflammation. *Signal Transduct Targeted Ther*. 2017;2:17023. <https://doi.org/10.1038/sigtrans.2017.23>.
  - 18 Sun SC. The non-canonical NF- $\kappa$ B pathway in immunity and inflammation. *Nat Rev Immunol*. 2017;17(9):545-558. <https://doi.org/10.1038/nri.2017.52>.
  - 19 Baker RG, Hayden MS, Ghosh S. NF- $\kappa$ B, inflammation, and metabolic disease. *Cell Metab*. 2011;13(1):11-22. <https://doi.org/10.1016/j.cmet.2010.12.008>.
  - 20 Zhang H, Sun SC. NF- $\kappa$ B in inflammation and renal diseases. *Cell Biosci*. 2015;5:63. <https://doi.org/10.1186/s13578-015-0056-4>.
  - 21 Monaco C, Andreaskos E, Kiriakidis S, et al. Canonical pathway of nuclear factor  $\kappa$ B activation selectively regulates proinflammatory and prothrombotic responses in human atherosclerosis. *Proc Natl Acad Sci USA*. 2004;101(15):5634-5639. <https://doi.org/10.1073/pnas.0401060101>.
  - 22 Zhou P, Luo Y, Xing N, et al. Research progress of pathogenesis of atherosclerosis induced by tumor necrosis factor- $\alpha$ . *World Chin Med*. 2015;10(8):1163-1168. <https://doi.org/10.3969/j.issn.1673-7202.2015.08.008>.
  - 23 Kanters E, Pasparakis M, Gijbels MJ, et al. Inhibition of NF- $\kappa$ B activation in macrophages increases atherosclerosis in LDL receptor-deficient mice. *J Clin Invest*. 2003;112(8):1176-1185. <https://doi.org/10.1172/jci18580>.
  - 24 Shen AL, Shi H, Peng DY, et al. Regulation of NF- $\kappa$ B signal pathway and protection of cardiac structure and function of type 2 diabetes mellitus rat by treating with Taohong Siwu Decoction. *China J Tradit Chin Med Pharm*. 2019;34(4):1359-1362.
  - 25 Wang WJ, Deng XY, Wang W. Mechanism of Taohong Siwu Decoction in treating soft tissue injury based on UPLC-Q-TOF-MS, network pharmacology and experimental verification. *China J Chin Mater Med*. 2021;46(12):3043-3051. <https://doi.org/10.19540/j.cnki.cjcm.20210311.403>.
  - 26 Wang JY, Chen WN, Jia LQ, et al. Effects of Erchen Decoction and Taohong Siwu Decoction on Nox4/NF- $\kappa$ B/HIF-1 $\alpha$  signaling pathway in aorta of ApoE<sup>-/-</sup> atherosclerosis mice. *China J Tradit Chin Med Pharm*. 2019;34(6):2417-2420. <https://doi.org/10.37155/2717-5693-0102-2>.
  - 27 Yuan GZ, Han AB, Wu J, et al. Bao Yuan Decoction and Tao Hong Si Wu Decoction improve lung structural remodeling in a rat model of myocardial infarction: possible involvement of suppression of inflammation and fibrosis and regulation of the TGF- $\beta$ 1/Smad3 and NF- $\kappa$ B pathways. *Biosci Trends*. 2018;12(5):491-501. <https://doi.org/10.5582/bst.2018.01242>.
  - 28 Wu FF, Zhang XW. Protective effect of peach kernel extract on coronary heart disease model rats through nuclear factor- $\kappa$ B/cyclooxygenase-2 pathway. *Med J Wuhan Univ*. 2020;41(5):725-731. <https://doi.org/10.14188/j.1671-8852.2019.0996>.
  - 29 Bhaskar S, Helen A. Quercetin modulates toll-like receptor-mediated protein kinase signaling pathways in oxLDL-challenged human PBMCs and regulates TLR-activated atherosclerotic inflammation in hypercholesterolemic rats. *Mol Cell Biochem*. 2016;423(1-2):53-65. <https://doi.org/10.1007/s11010-016-2824-9>.
  - 30 Jia ZQ, Nallasamy P, Liu DM, et al. Luteolin protects against vascular inflammation in mice and TNF- $\alpha$ -induced monocyte adhesion to endothelial cells via suppressing I $\kappa$ B $\alpha$ /NF- $\kappa$ B signaling pathway. *J Nutr Biochem*. 2015;26(3):293-302. <https://doi.org/10.1016/j.jnutbio.2014.11.008>.
  - 31 Jung KJ, Go EK, Kim JY, et al. Suppression of age-related renal changes in NF- $\kappa$ B and its target gene expression by dietary ferulate. *J Nutr Biochem*. 2009;20(5):378-388. <https://doi.org/10.1016/j.jnutbio.2008.04.008>.
  - 32 Liu L, Ning ZQ, Shan S, et al. Phthalide lactones from *Ligusticum chuanxiong* inhibit lipopolysaccharide-induced TNF- $\alpha$  production and TNF- $\alpha$ -mediated NF- $\kappa$ B activation. *Planta Med*. 2005;71(9):808-813. <https://doi.org/10.1055/s-2005-871231>.
  - 33 Li WF, Zhi WB, Liu F, et al. Paeoniflorin inhibits VSMCs proliferation and migration by arresting cell cycle and activating HO-1 through MAPKs and NF- $\kappa$ B pathway. *Int Immunopharmacol*. 2018;54:103-111. <https://doi.org/10.1016/j.intimp.2017.10.017>.
  - 34 Gao F, He QF, Wu SH, et al. Catalpol ameliorates LPS-induced inflammatory response by activating AMPK/mTOR signaling pathway in rat intestinal epithelial cells. *Eur J Pharmacol*. 2023;960:176125. <https://doi.org/10.1016/j.ejphar.2023.176125>.
  - 35 Chen SS, Liu HH, Wang SM, et al. The neuroprotection of verbasoside in Alzheimer's disease mediated through mitigation of neuroinflammation via blocking NF- $\kappa$ B-p65 signaling. *Nutrients*. 2022;14(7):1417. <https://doi.org/10.3390/nu14071417>.
  - 36 Zhang JX, Wang XL, Vikash V, et al. ROS and ROS-mediated cellular signaling. *Oxid Med Cell Longev*. 2016;2016:4350965. <https://doi.org/10.1155/2016/4350965>.
  - 37 Sarbassov DD, Guertin DA, Ali SM, et al. Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science*. 2005;307(5712):1098-1101. <https://doi.org/10.1126/science.1106148>.
  - 38 Linton MF, Mosleh JJ, Babaev VR. Akt signaling in macrophage polarization, survival, and atherosclerosis. *Int J Mol Sci*. 2019;20(11):2703. <https://doi.org/10.3390/ijms20112703>.
  - 39 Hirsch E, Ciruolo E, Ghigo A, et al. Taming the PI3K team to hold inflammation and cancer at bay. *Pharmacol Ther*. 2008;118(2):192-205. <https://doi.org/10.1016/j.pharmthera.2008.02.004>.
  - 40 Romashkova JA, Makarov SS. NF- $\kappa$ B is a target of AKT in anti-apoptotic PDGF signalling. *Nature*. 1999;401(6748):86-90. <https://doi.org/10.1038/43474>.
  - 41 Mantovani A, Garlanda C, Locati M. Macrophage diversity and polarization in atherosclerosis: a question of balance. *Arterioscler Thromb Vasc Biol*. 2009;29(10):1419-1423. <https://doi.org/10.1161/atvbaha.108.180497>.
  - 42 Williams HJ, Fisher EA, Greaves DR. Macrophage differentiation and function in atherosclerosis: opportunities for therapeutic intervention? *J Innate Immun*. 2012;4(5-6):498-508. <https://doi.org/10.1159/000336618>.
  - 43 Arranz A, Doxaki C, Vergadi E, et al. Akt1 and Akt2 protein kinases differentially contribute to macrophage polarization. *Proc Natl Acad Sci U S A*. 2012;109(24):9517-9522. <https://doi.org/10.1073/pnas.1119038109>.
  - 44 Seimon T, Tabas I. Mechanisms and consequences of macrophage apoptosis in atherosclerosis. *J Lipid Res*. 2009;50(Suppl):S382-S387. <https://doi.org/10.1194/jlr.R800032-JLR200>.
  - 45 Michell BJ, Griffiths JE, Mitchell KI, et al. The Akt kinase signals directly to endothelial nitric oxide synthase. *Curr Biol*. 1999;9(15):845-848. [https://doi.org/10.1016/s0960-9822\(99\)80371-6](https://doi.org/10.1016/s0960-9822(99)80371-6).
  - 46 Wang JY, Qu NN, Jia LQ, et al. Effect and mechanism of serum containing of Erchen Decoction and Taohong Siwu Decoction on ox-LDL induced endothelial cell damage. *Chin J Arterioscler*. 2018;26(10):987-992. <https://doi.org/10.3969/j.issn.1007-3949.2018.10.004>.
  - 47 Han L, Liang J, Zhang YY, et al. Effects of Taohong Siwu Decoction on the VEGF, NO expression and PI3K/Akt pathway alterations in uterine tissue of postpartum rats with blood stasis syndrome. *China J Tradit Chin Med Pharm*. 2016;31(5):1625-1629.
  - 48 Porta C, Paglino C, Mosca A. Targeting PI3K/Akt/mTOR signaling in cancer. *Front Oncol*. 2014;4:64. <https://doi.org/10.3389/fonc.2014.00064>.
  - 49 Lu XL, Zhao CH, Yao XL, et al. Quercetin attenuates high fructose feeding-induced atherosclerosis by suppressing inflammation and apoptosis via ROS-regulated PI3K/AKT signaling pathway. *Biomed Pharmacother*. 2017;85:658-671. <https://doi.org/10.1016/j.biopha.2016.11.077>.
  - 50 Che JB, Liang B, Zhang Y, et al. Kaempferol alleviates ox-LDL-induced apoptosis by up-regulation of autophagy via inhibiting PI3K/Akt/mTOR pathway in human endothelial cells. *Cardiovasc Pathol*. 2017;31:57-62. <https://doi.org/10.1016/j.carpath.2017.08.001>.
  - 51 Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat Immunol*. 2010;11(5):373-384. <https://doi.org/10.1038/ni.1863>.
  - 52 Gouloupoulou S, McCarthy CG, Webb RC. Toll-like receptors in the vascular system: sensing the dangers within. *Pharmacol Rev*. 2016;68(1):142-167. <https://doi.org/10.1124/pr.114.010090>.
  - 53 Kawasaki T, Kawai T. Toll-like receptor signaling pathways. *Front Immunol*. 2014;5:461. <https://doi.org/10.3389/fimmu.2014.00461>.
  - 54 Yamamoto M, Sato S, Hemmi H, et al. TRAM is specifically involved in the Toll-like receptor 4-mediated MyD88-independent signaling pathway. *Nat Immunol*. 2003;4(11):1144-1150. <https://doi.org/10.1038/ni986>.
  - 55 Laurent YC, Carrie W, Tamara AP, et al. Increased inflammatory gene expression in ABC transporter-deficient macrophages: free cholesterol accumulation, increased signaling via toll-like receptors, and neutrophil infiltration of atherosclerotic lesions. *Circulation*. 2008;118(18):1837-1847. <https://doi.org/10.1161/circulationaha.108.793869>.
  - 56 Chen TW, Huang WJ, Qian JF, et al. Macrophage-derived myeloid differentiation protein 2 plays an essential role in ox-LDL-induced inflammation and atherosclerosis. *EBioMedicine*. 2020;53:102706. <https://doi.org/10.1016/j.ebiom.2020.102706>.
  - 57 Schoneveld AH, Hoefler I, Sluiter JPG, et al. Atherosclerotic lesion development and Toll like receptor 2 and 4 responsiveness. *Atherosclerosis*. 2008;197(1):95-104. <https://doi.org/10.1016/j.atherosclerosis.2007.08.004>.
  - 58 Huang RZ, Hu ZC, Chen XR, et al. The transcription factor SUB1 is a master regulator of the macrophage TLR response in atherosclerosis. *Adv Sci (Weinheim)*. 2021;8(19):e2004162. <https://doi.org/10.1002/adv.202004162>.
  - 59 Ye ZS, Jin MY, Wang SJ, et al. Subcutaneous injection of dendritic cells aggravates atherosclerosis in ApoE-knockout mice by activation of TLR4. *Mol Med Rep*. 2017;16(5):6041-6049. <https://doi.org/10.3892/mmr.2017.7339>.
  - 60 Wang MM, Liu ZQ, Hu SS, et al. Taohong Siwu Decoction ameliorates ischemic stroke injury via suppressing pyroptosis. *Front Pharmacol*. 2020;11:590453. <https://doi.org/10.3389/fphar.2020.590453>.
  - 61 Fan WC, Lyu LT, Bai DY, et al. Study on improvement effect of Taohong Siwu Decoction on joint swelling in rat models with rheumatoid arthritis and its mechanism. *New J Tradit Chin Med*. 2023;55(1):53-57. <https://doi.org/10.13457/j.cnki.jncm.2023.01.011>.
  - 62 Chang FJ, Zhou P, Li GY, et al. Taohong Siwu Decoction ameliorates atherosclerosis in rats possibly through toll-like receptor 4/myeloid differentiation primary response protein 88/nuclear factor- $\kappa$ B signal pathway. *J Tradit Chin Med*. 2024;44(1):103-112. <https://doi.org/10.19852/j.cnki.jtcm.20231215.003>.
  - 63 Yang LJ, Yang KL, Zhong WL, et al. Study on anti-atherosclerosis mechanism of ligustrazine and ferulic acid based on network pharmacology. *Drug Eval Res*. 2021;44(12):2555-2562. <https://doi.org/10.7501/j.issn.1674-6376.2021.12.005>.
  - 64 Li H, Jiao Y, Xie M. Paeoniflorin ameliorates atherosclerosis by suppressing TLR4-mediated NF- $\kappa$ B activation. *Inflammation*. 2017;40(6):2042-2051. <https://doi.org/10.1007/s10753-017-0644-z>.
  - 65 Liang QC, Chen YT, Li CX, et al. Quercetin attenuates Ox-LDL-induced calcification in vascular smooth muscle cells by regulating ROS-TLR4 signaling pathway. *J South Med Univ*. 2018;38(8):980-985. <https://doi.org/10.3969/j.issn.1673-4254.2018.08.13>.
  - 66 Zhong XM, Zhang L, Li YM, et al. Kaempferol alleviates ox-LDL-induced

- apoptosis by up-regulation of miR-26a-5p via inhibiting TLR4/NF- $\kappa$ B pathway in human endothelial cells. *Biomed Pharmacother.* 2018;108:1783-1789. <https://doi.org/10.1016/j.biopha.2018.09.175>.
- 67 Raman M, Chen W, Cobb MH. Differential regulation and properties of MAPKs. *Oncogene.* 2007;26(22):3100-3112. <https://doi.org/10.1038/sj.onc.1210392>.
- 68 Muslin AJ. MAPK signalling in cardiovascular health and disease: molecular mechanisms and therapeutic targets. *Clin Sci (Lond).* 2008;115(7):203-218. <https://doi.org/10.1042/cs20070430>.
- 69 Meier R, Rouse J, Cuenda A, et al. Cellular stresses and cytokines activate multiple mitogen-activated-protein kinase homologues in PC12 and KB cells. *Eur J Biochem.* 1996;236(3):796-805. <https://doi.org/10.1111/j.1432-1033.1996.00796.x>.
- 70 Carnello M, Roux PP. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol Mol Biol Rev.* 2011;75(1):50-83. <https://doi.org/10.1128/mmb.00031-10>.
- 71 Yoon S, Seger R. The extracellular signal-regulated kinase: multiple substrates regulate diverse cellular functions. *Growth Factors.* 2006;24(1):21-44. <https://doi.org/10.1080/02699050500284218>.
- 72 Cuadrado A, Nebreda AR. Mechanisms and functions of p38 MAPK signalling. *Biochem J.* 2010;429(3):403-417. <https://doi.org/10.1042/bj20100323>.
- 73 Pietersma A, Tilly BC, Gaestel M, et al. P38 mitogen activated protein kinase regulates endothelial VCAM-1 expression at the post-transcriptional level. *Biochem Biophys Res Commun.* 1997;230(1):44-48. <https://doi.org/10.1006/bbrc.1996.5886>.
- 74 Goebeler M, Kilian K, Gillitzer R, et al. The MKK6/p38 stress kinase cascade is critical for tumor necrosis factor-alpha-induced expression of monocyte-chemoattractant protein-1 in endothelial cells. *Blood.* 1999;93(3):857-865. [https://doi.org/10.1182/blood.V93.3.857.403k03.857\\_865](https://doi.org/10.1182/blood.V93.3.857.403k03.857_865).
- 75 Liao LZ, Zhou Q, Song Y, et al. Ceramide mediates Ox-LDL-induced human vascular smooth muscle cell calcification via p38 mitogen-activated protein kinase signaling. *PLoS One.* 2013;8(12):e82379. <https://doi.org/10.1371/journal.pone.0082379>.
- 76 Senokuchi T, Matsumura T, Sakai M, et al. Extracellular signal-regulated kinase and p38 mitogen-activated protein kinase mediate macrophage proliferation induced by oxidized low-density lipoprotein. *Atherosclerosis.* 2004;176(2):233-245. <https://doi.org/10.1016/j.atherosclerosis.2004.05.019>.
- 77 Xu Y, Zhang Y, Xu Y, et al. Activation of CD137 signaling promotes macrophage apoptosis dependent on p38 MAPK pathway-mediated mitochondrial fission. *Int J Biochem Cell Biol.* 2021;136:106003. <https://doi.org/10.1016/j.jbiocel.2021.106003>.
- 78 Tang LF, Chang H, Wang DD, et al. Active components and potential mechanism of Taohong Siwu Decoction in regulating ischemic stroke based on target cell trapping combined with network pharmacology, molecular docking, and experimental validation. *China J Chin Mater Med.* 2023;48(17):4761-4773. <https://doi.org/10.19540/j.cnki.cjcm.20230423.403>.
- 79 Luo G, Xiang L, Xiao L. Quercetin alleviates atherosclerosis by suppressing oxidized LDL-induced senescence in plaque macrophage via inhibiting the p38MAPK/p16 pathway. *J Nutr Biochem.* 2023;116:109314. <https://doi.org/10.1016/j.jnutbio.2023.109314>.
- 80 Wu YL, Wang F, Fan LH, et al. Baicalin alleviates atherosclerosis by relieving oxidative stress and inflammatory responses via inactivating the NF- $\kappa$ B and p38 MAPK signaling pathways. *Biomed Pharmacother.* 2018;97:1673-1679. <https://doi.org/10.1016/j.biopha.2017.12.024>.
- 81 Lu Q, Qiu TQ, Yang H. Ligustilide inhibits vascular smooth muscle cells proliferation. *Eur J Pharmacol.* 2006;542(1-3):136-140. <https://doi.org/10.1016/j.ejphar.2006.04.023>.
- 82 Bai JR, Zhang YS, Tang C, et al. Gallic acid: pharmacological activities and molecular mechanisms involved in inflammation-related diseases. *Biomed Pharmacother.* 2021;133:110985. <https://doi.org/10.1016/j.biopha.2020.110985>.
- 83 Liu M, Feng J, Du Q, et al. Paeoniflorin attenuates myocardial fibrosis in isoprenaline-induced chronic heart failure rats via inhibiting P38 MAPK pathway. *Curr Med Sci.* 2020;40(2):307-312. <https://doi.org/10.1007/s11596-020-2178-0>.
- 84 Saxton RA, Sabatini DM. mTOR signaling in growth, metabolism, and disease. *Cell.* 2017;168(6):960-976. <https://doi.org/10.1016/j.cell.2017.02.004>.
- 85 Schalm SS, Fingar DC, Sabatini DM, et al. TOS motif-mediated raptor binding regulates 4E-BP1 multisite phosphorylation and function. *Curr Biol.* 2003;13(10):797-806. [https://doi.org/10.1016/s0960-9822\(03\)00329-4](https://doi.org/10.1016/s0960-9822(03)00329-4).
- 86 Fu W, Hall MN. Regulation of mTORC2 signaling. *Genes(Basel).* 2020;11(9):1045. <https://doi.org/10.3390/genes11091045>.
- 87 Guertin DA, et al. Ablation in mice of the mTORC components raptor, rictor, or mLST8 reveals that mTORC2 is required for signaling to Akt-FOXO and PKC $\alpha$ , but not S6K1. *Dev Cell.* 2006;11(6):859-871. <https://doi.org/10.1016/j.devcel.2006.10.007>.
- 88 Laplante M, Sabatini DM. mTOR signaling at a glance. *J Cell Sci.* 2009;122(Pt 20):3589-3594. <https://doi.org/10.1242/jcs.051011>.
- 89 Yang G, Murashige DS, Humphrey SJ, et al. A positive feedback loop between Akt and mTORC2 via S1N1 phosphorylation. *Cell Rep.* 2015;12(6):937-943. <https://doi.org/10.1016/j.celrep.2015.07.016>.
- 90 Perrotta C, Cattaneo MG, Molteni R, et al. Autophagy in the regulation of tissue differentiation and homeostasis. *Front Cell Dev Biol.* 2020;8:602901. <https://doi.org/10.3389/fcell.2020.602901>.
- 91 Kim J, Kundu M, Viollet B, et al. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol.* 2011;13(2):132-141. <https://doi.org/10.1038/ncb2152>.
- 92 Wang XC, Li LX, Niu XL, et al. mTOR enhances foam cell formation by suppressing the autophagy pathway. *DNA Cell Biol.* 2014;33(4):198-204. <https://doi.org/10.1089/dna.2013.2164>.
- 93 Martina JA, Chen Y, Gucek M, et al. mTORC1 functions as a transcriptional regulator of autophagy by preventing nuclear transport of TFEB. *Autophagy.* 2012;8(6):903-914. <https://doi.org/10.4161/auto.19653>.
- 94 Araki K, Turner AP, Shaffer VO, et al. mTOR regulates memory CD8 T-cell differentiation. *Nature.* 2009;460(7251):108-112. <https://doi.org/10.1038/nature08155>.
- 95 Kaldirim M, Lang A, Pfeiler S, et al. Modulation of mTOR signaling in cardiovascular disease to target acute and chronic inflammation. *Front Cardiovasc Med.* 2022;9:907348. <https://doi.org/10.3389/fcvm.2022.907348>.
- 96 Xu ZH, Xu JB, Qian WW, et al. Repair effect and mechanism of Taohong Siwu Decoction on rotator cuff injury in rabbits. *China Pharmacy.* 2021;32(24):2975-2979. <https://doi.org/10.6039/j.issn.1001-0408.2021.24.06>.
- 97 Cao H, Jia QL, Shen DZ, et al. Quercetin has a protective effect on atherosclerosis via enhancement of autophagy in ApoE<sup>-/-</sup> mice. *Exp Ther Med.* 2019;18(4):2451-2458. <https://doi.org/10.3892/etm.2019.7851>.
- 98 Wu HF, Song AW, Hu WJ, et al. The anti-atherosclerotic effect of paeonol against vascular smooth muscle cell proliferation by up-regulation of autophagy via the AMPK/mTOR signaling pathway. *Front Pharmacol.* 2017;8:948. <https://doi.org/10.3389/fphar.2017.00948>.
- 99 Marinou K, Christodoulides C, Antoniadis C, et al. Wnt signaling in cardiovascular physiology. *Trends Endocrinol Metab.* 2012;23(12):628-636. <https://doi.org/10.1016/j.tem.2012.06.001>.
- 100 Catalano A, Bellone F, Morabito N, et al. Sclerostin and vascular pathophysiology. *Int J Mol Sci.* 2020;21(13):4779. <https://doi.org/10.3390/ijms21134779>.
- 101 Kong P, Cui ZY, Huang XF, et al. Inflammation and atherosclerosis: signaling pathways and therapeutic intervention. *Signal Transduct Target Ther.* 2022;7(1):131. <https://doi.org/10.1038/s41392-022-00955-7>.
- 102 Sharma BR, Kanneganti TD. NLRP3 inflammasome in cancer and metabolic diseases. *Nat Immunol.* 2021;22(5):550-559. <https://doi.org/10.1038/s41590-021-00886-5>.
- 103 Loirand G, Guérin P, Pacaud P. Rho kinases in cardiovascular physiology and pathophysiology. *Circ Res.* 2006;98(3):322-334. <https://doi.org/10.1161/01.RES.0000201960.04223.3c>.
- 104 Kloc M, Uosef A, Kubiak JZ, et al. Role of macrophages and RhoA pathway in atherosclerosis. *Int J Mol Sci.* 2020;22(1):216. <https://doi.org/10.3390/ijms22010216>.
- 105 Baldini C, Moriconi FR, Galimberti S, et al. The JAK-STAT pathway: an emerging target for cardiovascular disease in rheumatoid arthritis and myeloproliferative neoplasms. *Eur Heart J.* 2021;42(42):4389-4400. <https://doi.org/10.1093/eurheartj/ehab447>.
- 106 Segá FVD, Fortini F, Aquila G, et al. Notch signaling regulates immune responses in atherosclerosis. *Front Immunol.* 2019;10:1130. <https://doi.org/10.3389/fimmu.2019.01130>.
- 107 Martos-Rodríguez CJ, Albarrán-Juárez J, Morales-Cano D, et al. Fibrous caps in atherosclerosis form by Notch-dependent mechanisms common to arterial media development. *Arterioscler Thromb Vasc Biol.* 2021;41(9):e427-e439. <https://doi.org/10.1161/atvbaha.120.315627>.
- 108 Dabravolski SA, Khotina VA, Omelchenko AV, et al. The role of the VEGF family in atherosclerosis development and its potential as treatment targets. *Int J Mol Sci.* 2022;23(2):931. <https://doi.org/10.3390/ijms23020931>.
- 109 Li RJ, Dai YY, Qin C, et al. Application of traditional Chinese medicine in treatment of Helicobacter pylori infection. *World J Clin Cases.* 2021;9(35):10781-10791. <https://doi.org/10.12998/wjcc.v9.i35.10781>.
- 110 Kim J, Kim J, Kim DW, et al. Wnt5a induces endothelial inflammation via  $\beta$ -catenin-independent signaling. *J Immunol.* 2010;185(2):1274-1282. <https://doi.org/10.4049/jimmunol.1000181>.
- 111 Bikkavilli RK, Feigin ME, Malbon CC. P38 mitogen-activated protein kinase regulates canonical Wnt-beta-catenin signaling by inactivation of GSK3beta. *J Cell Sci.* 2008;121(Pt 21):3598-3607. <https://doi.org/10.1242/jcs.032854>.
- 112 Gurusu P, Malireddi RKS, Anand PK, et al. Toll or interleukin-1 receptor (TIR) domain-containing adaptor inducing interferon- $\beta$  (TRIF)-mediated caspase-11 protease production integrates Toll-like receptor 4 (TLR4) protein- and Nlrp3 inflammasome-mediated host defense against enteropathogens. *J Biol Chem.* 2012;287(41):34474-34483. <https://doi.org/10.1074/jbc.M112.401406>.
- 113 Wolfrum S, Dendorfer A, Rikitake Y, et al. Inhibition of Rho-kinase leads to rapid activation of phosphatidylinositol 3-kinase/protein kinase Akt and cardiovascular protection. *Arterioscler Thromb Vasc Biol.* 2004;24(10):1842-1847. <https://doi.org/10.1161/01.Atm.0000142813.33538.82>.
- 114 Hu X, Li J, Fu M, et al. The JAK/STAT signaling pathway: from bench to clinic. *Signal Transduct Target Ther.* 2021;6(1):402. <https://doi.org/10.1038/s41392-021-00791-1>.
- 115 Hildebrand D, Uhle F, Sahin D, et al. The interplay of Notch signaling and STAT3 in TLR-activated human primary monocytes. *Front Cell Infect Microbiol.* 2018;8:241. <https://doi.org/10.3389/fcimb.2018.00241>.
- 116 Luo GA, Wang YM, Fan XM, et al. Research strategy and practice from clinical reality, targeting at signaling pathways for the innovative compound drug—the sixth discussion on the proposal of holistic systems medicine. *WST-MTCM.* 2018;20(7):1047-1068. <https://doi.org/10.11842/wst.2018.07.001>.
- 117 Luo GA, Liang QL, Liu QF, et al. Chemomics-integrated global systems biology: a holistic methodology of study on compatibility and mechanism of formulas in traditional Chinese medicine. *World Sci Technol-Mod Tradit Chin Med.* 2007;1:10-15, 24. <https://doi.org/10.3969/j.issn.1674-3849.2007.01.005>.
- 118 Luo CY, Luo PZ, Wang GB, et al. Characteristics and advantages of traditional Chinese medicine based on complexity science. *Chin J Basic Med Tradit Chin Med.* 2018;24(10):1368-1372. <https://doi.org/10.19945/j.cnki.issn.1006-3250.2018.10.012>.