

Pharmacological effects of volatile oil from chrysanthemum and its associated mechanisms: a review

Jing Zhang¹, Weiqiang Su¹, Nina Filipczak², Ying Luo¹, Anping Wan³, Yao He¹, Shijuan Yan¹, Xiang Li^{1,3,*}, Ming Yang^{1,*}

¹Key Laboratory of Modern Preparation of TCM, Ministry of Education, Jiangxi University of Chinese Medicine, Nanchang, China; ²Center for Pharmaceutical Biotechnology and Nanomedicine, Northeastern University, Boston, USA; ³National Pharmaceutical Engineering Center for Solid Preparation in Chinese Herbal Medicine, Jiangxi University of Chinese Medicine, Jiangxi University of Chinese Medicine, Nanchang, China

Abstract

Volatile oil (VO) is the main chemical component of common plants in Chrysanthemum genus, and it possesses several beneficial pharmacological properties, including bacteriostatic, antioxidant, anti-tumor, anti-inflammatory, antipyretic, analgesic, anti-osteoporotic, antihypertensive, sedative, and hypnotic effects. To date, research on the effective components of Chrysanthemum extract has mainly focused on flavonoids, whereas limited data are available on the chemical constituents and underlying mechanisms of action of the VO components. In this review, the pharmacological activities and mechanisms of VO are comprehensively reviewed with the aim of providing a foundation for further development for medicinal, aromatherapy, and diet therapy applications.

Keywords: Action mechanisms, Chrysanthemum, Pharmacological activity, Volatile oil

Graphical abstract: <http://links.lww.com/AHM/A85>.

Introduction

Chrysanthemums, characterized by its unique flower color and structure, belong to the Compositae family and are annual or perennial herbs with an over 3,000-year history of cultivation^[1,2]. From the perspective of varieties, Chrysanthemum species worldwide are mainly divided into Chinese, Japanese, and European groups. The plant has been used as a food ingredient, additive, beverage, and medicine in several countries since ancient times, particularly in China and Korea^[3,4]. In China, Japan, Vietnam, and other Eastern countries, boiled dried Chrysanthemum has been used to prepare herbal teas to relieve fever and treat eye diseases for over 2,000 years^[5]. The medicinal efficacy of Chrysanthemum was first recorded in *Shennong Herbal Classic* (神农本草经) in China, with reported potency for the treatment of headaches and vertigo, among other symptoms^[6]. Chrysanthemum is included in the Pharmacopoeia of the People's Republic of China (2020 Edition) as a medicinal food plant and is classified into five medicinal varieties: Chrysanthemum morifolium (Bo), Chuzhou Chrysanthemum (Chu), Florist Chrysanthemum (Gong), Chrysanthemum morifolium (Hang), and Huaqing

Chrysanthemum (Huai), depending on the growing regions^[7].

Modern pharmacological studies have identified flavonoids, volatile oil (VO), organic acids, and triterpenoids as the major components of Chrysanthemum. To date, studies have predominantly focused on the VO from *Chrysanthemum morifolium* Ramat. and *Chrysanthemum indicum* L. in China, which are recorded in the Chinese Pharmacopoeia^[7]. The composition of Chrysanthemum VO is complex and includes monoterpenes, sesquiterpenes, and oxygen-containing derivatives^[8,9]. However, current research mainly involves the optimization of the VO extraction process, with limited attention to the pharmacological effects and mechanisms of action.

In this report, the pharmacological properties and underlying mechanisms of Chrysanthemum VO are comprehensively reviewed to provide a basis for exploring and developing its clinical applications. The word cloud describing the main pharmacological effects of Chrysanthemum VO is presented in Figure 1.

The following retrieval and summary methods were used: (1) initial search based on the main keywords,

Jing Zhang and Weiqiang Su contributed equally to this work.

*Corresponding author. Xiang Li, E-mail: xiang.li@jxutcm.edu.cn; Ming Yang, E-mail: 20050858@jxutcm.edu.cn.

Received 10 July 2023 / Accepted 12 January 2024

How to cite this article: Zhang J, Su WQ, Filipczak N, Luo Y, Wan AP, He Y, Yan SJ, Li X, Yang M. Pharmacological effects of volatile oil from chrysanthemum and its associated mechanisms: a review. *Acupunct Herb Med* 2024;4(1):79–91. doi: 10.1097/HM9.0000000000000090

Copyright © 2024 Tianjin University of Traditional Chinese Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

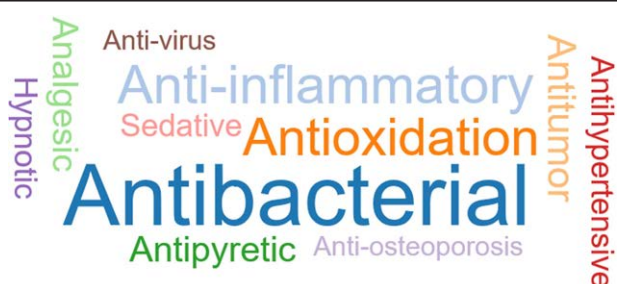


Figure 1. Pharmacological actions of Chrysanthemum VO. The common effects of Chrysanthemum VO are presented as a “word cloud,” whereby the size of each pharmacological activity is positively correlated with frequency of its reports in literature. VO: Volatile oil.

specifically, “medicinal plant” “volatile oil” “chrysanthemum” “volatile oil components” and “pharmacological effect” in PubMed, Web of Science, ScienceDirect, and other databases; (2) preliminary screening of the literature based on title, keywords, and guidelines; (3) addition of recent research progress and new references from the original literature; and (4) summary and collation of the available literature.

Pharmacological effects and underlying mechanisms

Antimicrobial effect

Chrysanthemum VO contains a mixture of terpenoids, phenols, alcohols, and other chemical components that exert significant antibacterial effects against several pathogenic microorganisms, such as *Escherichia coli*, *Staphylococcus aureus*, *Streptococcus mutans*, *Pseudomonas aeruginosa*, and *Listeria monocytogenes*^[9–11]. Common varieties of Chrysanthemum VO and their active antimicrobial components are presented in Table 1^[11–13].

The active components of Chrysanthemum VOs differ depending on the origin of the plant. The antibacterial effect is not attributed to individual components and mechanisms but may be the result of multi-component synergy, multiple pathways, and multi-target actions in cells^[14]. Figure 2 depicts the sites of action and mechanisms of VO components in bacterial cells. Notably, many of these mechanisms are affected by multiple pathways^[15,16].

Destruction of cellular structure integrity

The integrity of the cell structure is the premise for maintaining the normal physiological activities of microorganisms. Owing to their hydrophobic nature, VO and their components are proposed to be distributed in the microbial cell membrane, inducing destruction of the cellular structure, increased membrane permeability, and outflow of critical molecules from the cell, triggering internal disorder, disruption of the steady-state balance of bacterial cells, and ultimately, death of microorganisms^[17]. A previous study reported that Chrysanthemum VO inhibited the growth of *Phytophthora nicotianae* and exerted a bacteriostatic effect by increasing cell membrane permeability, destroying the structure of the mycelium, and causing cell rupture^[18]. Another report on the bacteriostatic activity of Chrysanthemum VO against

Listeria monocytogenes consistently demonstrated the destruction of the bacterial cell membrane and increased permeability, leading to the leakage of important physiological molecules, such as deoxyribonucleic acid (DNA), protein, and adenosine triphosphate (ATP). Subsequently, the intracellular content is significantly reduced and cell surface conductivity and adsorption rates increase, eventually leading to bacterial death^[10].

Effects on cellular nucleic acid metabolism and protein synthesis

VO inhibits the expression of nucleic acid metabolism genes and promotes cell death by blocking the biosynthesis of microbial nucleic acids, proteins, and peptidoglycans^[19]. DNA is a carrier of genetic information that is critical for the synthesis and genetic processes of microbial cells. DNA topoisomerases are essential enzymes involved in DNA replication, transcription, and translation, and damage to these enzymes affects the normal replication, growth, and reproduction of genetic material^[20]. The active compound thymol reduces bacterial adhesion and virulence by downregulating a gene encoding the outer membrane protein of *Acinetobacter baumannii*^[21]. The VO of Chu Chrysanthemum inhibits the activities of intracellular enzymes such as adenosine triphosphatase (ATPase) and alkaline phosphatase (ALP), which are crucial for bacterial growth. It also inhibits topoisomerases I and II, leading to a subsequent increase in the proportion of supercoiled DNA, which affects nucleic acid metabolism and eventually triggers bacterial death^[22].

Inhibition of microbial energy and respiratory metabolism

ATP is the direct source of energy for all life processes. Upon inhibition of ATP synthase, metabolism is significantly affected, leading to the death of microorganisms^[23]. Thymol and eugenol induced the acidification of the intracellular environment of *Candida* by inhibiting the activity of proton-translocating ATPases, resulting in bacterial cell death^[24]. Similarly, Chrysanthemum VO inhibited the respiratory metabolism of cells of *Listeria monocytogenes* through the inhibitory effects of iodoacetic acid, malonic acid, and trisodium phosphate dodecahydrate on the activity of enzymes in the glycolytic pathway^[10]. Collectively, these findings indicate that Chrysanthemum VO promotes intracellular ion loss by destroying microbial cell membranes, thereby disrupting the balance of intracellular and extracellular ATP and exerting a bacteriostatic effect.

Other effects

VO from medicinal plants can induce the coagulation of the microbial cytoplasm, resulting in a bacteriostatic effect. The cytoplasm is the hub for all physiological activities. Upon solidification, the metabolism and chemical reactions of cells cannot proceed normally, eventually causing microbial death^[25]. Carvacrol inhibited bacterial quorum sensing to decrease bacterial virulence and induces bacteriostasis by inhibiting LasI synthase activity, with a concomitant reduction in LasR protein expression, biofilm formation, swarming motility, and a

Table 1**Main active antibacterial components of VO from different varieties of Chrysanthemum**

Main antibacterial active ingredients	Chrysanthemum species			
	<i>Chrysanthemum morifolium</i> Ramat.	<i>Chrysanthemum indicum</i> L.	<i>Chrysanthemum boreale</i> Makino	<i>Chrysanthemum zawadskii</i> Herbich
Monoterpene hydrocarbons				
Pinene	√	√	√	√
Terpinene	√	√	√	√
Camphene		√	√	√
Thujene		√	√	√
Phellandrene				
Myrcene			√	√
Limonene	√	√	√	
Oxygenated monoterpenes				
Camphor	√	√	√	√
Borneol	√	√	√	√
Thujone	√	√	√	√
Linalool		√	√	
Carvacrol		√	√	
Terpinen-4-ol		√	√	√
Terpineol	√	√	√	
Myrtenol	√	√	√	√
Thymol		√	√	√
Cineole	√	√	√	√
Bornyl acetate	√	√	√	√
Sesquiterpenes hydrocarbons				
Caryophyllene	√	√	√	√
Curcumene	√	√	√	
Guaiene	√		√	
Elemene	√			√
α-Sesquiphellandrene			√	√
Oxygenated sesquiterpenes				
Nerolidol	√		√	√
Eudesmol	√	√		
Caryophyllene oxide	√	√	√	

VO: Volatile oil.

The overall composition and VO content in the same variety of Chrysanthemum can vary depending on the origin, processing method, and detection method.

subsequent decrease in acyl-homoserine lactone production in *Pseudomonas aeruginosa*^[26].

Anti-inflammatory effect

Inflammation is an essential biological mechanism that represents the defensive response of an organism to stimulation by various inflammatory factors^[27]. Luteolin, apigenin, linarin, and other flavonoids present in Chrysanthemum extracts exhibit anti-inflammatory properties. The significant anti-inflammatory activities of terpenoids, esters, phenols, and other compounds in VO have also been confirmed, which mainly regulate the synthesis and release of cellular inflammatory factors and block signaling pathways^[27], as depicted in Figure 3.

Inhibition of pro-inflammatory factor production

Cytokines are low-molecular-weight soluble proteins that are synthesized and secreted by cells such as lymphocytes, macrophages, and epithelial cells. They have diverse biological functions and exert both anti-inflammatory and pro-inflammatory effects^[28,29]. Pro-inflammatory cytokines, including tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), interleukin-6 (IL-6), and interleukin-8 (IL-8), enhance the inflammatory reaction and immunity^[28]. VO inhibits the production of inflammatory factors and the expression of inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2) to block the production of related mediators, thus exerting an anti-inflammatory effect. In addition, iNOS and COX-2 are modulated through nuclear factor-κB (NF-κB) activation to regulate

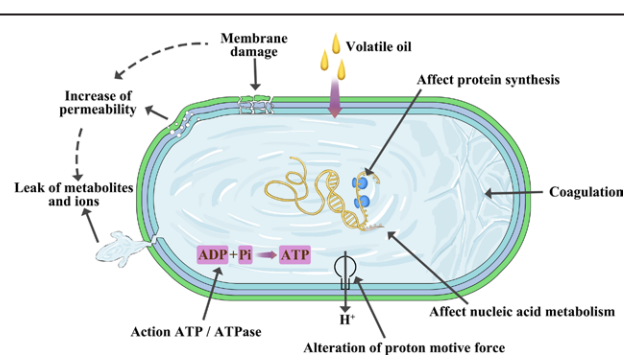


Figure 2. Mechanisms underlying the antibacterial effects of Chrysanthemum VO. ADP: Adenosine diphosphate; ATP: Adenosine triphosphate; ATPase: Adenosine triphosphatase; Pi: Inorganic phosphate; VO: Volatile oil.

the production of prostaglandin E_2 (PGE_2) and nitric oxide (NO)^[30]. Previous mechanistic studies have shown that specific sesquiterpenoids in Chrysanthemum exhibited significant anti-inflammatory activity by inhibiting NO production in RAW 264.7 cells stimulated with lipopolysaccharide (LPS)^[31,32]. Parthenolide (PTL) was reported to inhibit the expression of inflammatory cytokines (IL-1 β , IL-6, IL-8, IL-12p40, IL-18, TNF- α , and NO) in LPS-induced THP-1 cells in a dose-dependent manner, indicating activity against LPS-mediated pro-inflammatory responses in cells^[33]. In LPS-stimulated RAW264.7 macrophages *in vitro*, handelin from *Chrysanthemum boreale* not only inhibited the induction of the pro-inflammatory cytokines (IL-1 β and TNF- α), but also inhibited PGE_2 and NO produced by the cells in a concentration-dependent manner (20 μ M handelin inhibited NO production by >84% while the inhibition rate at 40 μ M was 98%). Moreover, this inhibitory effect was related to the down-regulation of mRNA and protein expression of COX-2 and iNOS^[34].

Blockage of signaling pathways

VO of medicinal plants exerts anti-inflammatory effects by the blockade of NF- κ B and mitogen-activated protein kinases (MAPK) signaling pathways.

Upon stimulation of cells, the stimulatory agent binds to the receptor protein on the membrane and activates the inhibitor of NF- κ B (I κ B) kinase (IKK). Subsequently, I κ B undergoes phosphorylation, thereby releasing NF- κ B that translocates to the nucleus and interacts with specific DNA sequences, inducing pro-inflammatory factors to enhance the expression of secondary inflammatory mediator enzymes and ultimately causing inflammation^[35,36]. VO can inhibit the release of NF- κ B by blocking specific pathways to reduce the release of inflammatory mediators and associated gene expression. Bornyl acetate, the main VO constituent in some traditional Chinese herbs, exerts anti-inflammatory effects *via* decreasing the transcriptional activity of NF- κ B in a concentration-dependent manner and blocking p65 nuclear translocation induced by the oxidized form of low-density lipoprotein (LDL)-cholesterol^[37].

MAPK is a key signaling pathway involved in the stress response. Upon stimulation of cells by the appropriate mediators, MAPK kinase and MAPK kinase are

activated, which in turn activates MAPK, which acts on downstream molecules to regulate specific genes, thereby modulating inflammation and other reactions^[38,39]. Chrysanthemum VO suppresses the activity of extracellular signal-regulated kinases (ERK), c-Jun N-terminal kinase (JNK), and p38 in the MAPK pathway. The anti-inflammatory mechanism of the PTL component involves inhibition of toll-like receptor 4 (TLR4)-mediated MAPK and NF- κ B pathways through dose-dependent suppression of LPS-induced upregulation of phosphorylated ERK 1/2, JNK, p38, NF- κ B, p65, I κ B α and expression of iNOS, TLR4, and TNF receptor-associated factor 6^[33]. Additionally, research has revealed that handelin ameliorated inflammation by downregulating NF- κ B and blocking the ERK and JNK signaling pathways in MAPK^[34]. In addition, the anti-inflammatory activity of the sesquiterpene compound chrysanthemulide A, a constituent of the herb *Chrysanthemum indicum*, appears to be mediated *via* the suppression of the LPS-induced NF- κ B pathway and downregulation of MAPK activation^[40].

Antioxidant activity

Oxidative stress is a known pathological mechanism in many diseases, and the inhibition of this process plays a crucial role in preventing and curing diseases. Chrysanthemum has been reported to protect against liver damage *via* antioxidant effects^[41]. Chrysanthemum VO are rich in phenols and other compounds with significant antioxidant activity, particularly thymol, carvacrol, and eugenol; therefore, they have potential utility as natural antioxidants^[42]. VO can be incorporated as a natural preservative into various food and pharmaceutical preparations. The antioxidant activity of VO is mainly exerted through the mechanisms described in the next sections.

Blockage of the free radical chain reaction

VO from medicinal plants can play an antioxidant role by blocking free radical chain reactions, including free radical scavenging and chelation with metal ions, as depicted in Figure 4^[43].

The phenol compounds in Chrysanthemum VO, such as eugenol, thymol, and carvacrol, can provide active H-atom to react with the free substrate (ROO) faster than substrate RH to generate stable substances of non-free substrate ROOH and antioxidant free radicals, thereby interrupting the free radical chain reaction to exert antioxidant effect^[44,45]. By preventing the generation of free radicals, thymol in VO reduced intracellular oxidative stress in a dose-dependent manner, exerted anti-cholinesterase activity, and improved the viability of neuronal PC-12 cells^[46].

Research has also shown that Chrysanthemum VO had a strong antioxidant capacity in 2,2-diphenyl-1-picrylhydrazyl (DPPH), hydroxyl radical scavenging, and superoxide radical scavenging experiments, with 50% inhibitory concentration (IC_{50}) values of 2.59, 2.89, and 5.92 mg/mL, respectively. An explanation for this activity, in addition to its role as a pro-oxidant, can be found in the synergistic effect between all VO components^[12]. Compounds

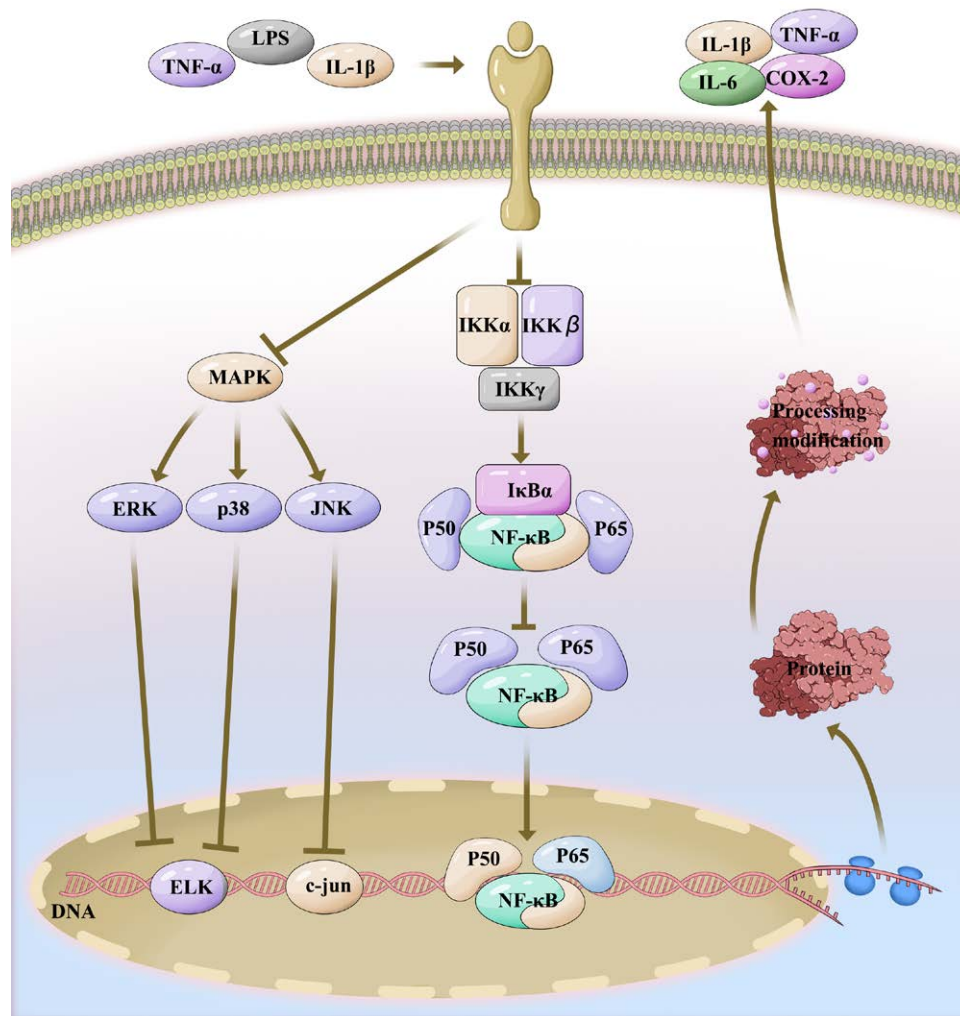


Figure 3. Signaling pathways underlying the anti-inflammatory effects of Chrysanthemum VO. COX: Cyclooxygenase-2; ELK: Transcription factor ELK; ERK: Extracellular signal-regulated kinases; IKK: NF-κB (IκB) kinase; IL: Interleukin; c-jun JNK: c-Jun N-terminal kinase; LPS: Lipopolysaccharide; MAPK: Mitogen-activated protein kinases; NF-κB: Nuclear factor-κB; P50: NF-κB p50 subunit; P65: NF-κB p65 subunit; TNF-α: Tumor necrosis factor-α; VO: Volatile oil.

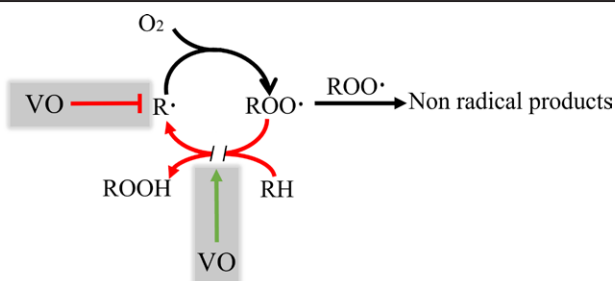


Figure 4. Effect of antioxidants in Chrysanthemum VO on free radical chain reaction. VO: Volatile oil.

including α -terpinene, β -terpinene, β -terpinolene, 1,8-cineole, and terpinen-4-ol are considered to contribute to the antioxidant effect of VO^[47]. Moreover, VO obtained at different growth stages (vegetative, pre-flowering, and full-flowering stages) of *Chrysanthemum boreale* Makino displayed varying degrees of antioxidant activity, which promoted the DPPH and azinobis-(3-ethylvensothiazoline-6-sulfonic acid) scavenging of cells. Eugenol from *Chrysanthemum boreale* Makino VO exhibited the highest scavenging efficiency, compared with that of other components, at concentration from 0.1 to 20 $\mu\text{g/mL}$ ^[48].

Involvement in other antioxidant pathways

Various bioactive components in Chrysanthemum VO can improve the antioxidant defense ability of the body by inhibiting lipid peroxidation or regulating antioxidant enzyme levels by participating in other related antioxidant stress signaling pathways^[43], such as the activation of nuclear factor erythroid 2-related factor 2 (Nrf2)-related antioxidant pathways^[49]. According to a recent study, germacrene-type sesquiterpenoids in Chrysanthemum can promote the dissociation and translocation of Nrf2 from Kelch-like ECH-associated protein 1 (Keap1) to the nucleus by anchoring to the Kelch domain-binding site of Keap1, thereby reducing the accumulation of reactive oxygen species (ROS) and increasing glutathione and heme oxygenase-1 levels and superoxide dismutase activity, thereby protecting the liver against oxidative damage^[50].

Anti-tumor effect

Chrysanthemum has a long history of clinical applications for therapeutic purposes, such as killing pathogenic microorganisms, improving immunity, and treating malignant tumors^[51,52]. Herbal medicines with cold and

cool in nature are generally used for tumor management^[53,54]. Recent research has identified a variety of effective chemical components in *Chrysanthemum VO*, such as PTL, β -elemene, and 1,8-cineole, that effectively exert anti-tumor effects against several cancer cell types. The therapeutic effects and mechanisms of action of the active ingredients of VO in different cancer types are shown in Table 2.

Parthenolide

PTL is a natural sesquiterpene lactone bioactive component extracted from the traditional medicinal plants of Asteraceae, such as *Chrysanthemum parthenium* L^[81]. It exerts anti-proliferative and pro-apoptotic effects in various malignant tumor types, including leukemia, prostate cancer, and osteosarcoma, both *in vitro* and in animal models, supporting its potential as a first-line clinical anticancer drug^[80–82]. Moreover, the synergistic effects of the interactions of several drugs with their targets, combined with the actions of PTL, have been reported to effectively inhibit the malignant proliferation of cancer cells^[78]. Data from the available literature suggest that PTL regulates related signaling pathways, such as signal transducer and activator of transcription 3 (STAT3), phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt), mitogen extracellular signal-regulated kinase (MEK)/ERK, Wnt (wingless)/ β -catenin, pyruvate kinase M2 (PKM2)-STAT3, and NF- κ B, to block the cancer cell cycle and induce apoptosis^[79]. Subcutaneous injection or oral administration of PTL suppressed the production of IL-8 and vascular endothelial growth factor an OUR-10 nude mouse xenograft model and reduced nuclear localization of NF- κ B and its phosphorylated form, causing decreased expression of metalloproteinase-9, anti-apoptotic factor Bcl-xL, and COX-2^[83]. Collectively, these findings indicate that PTL inhibits the growth of renal cancer cells by blocking NF- κ B activity.

β -Elemene

β -Elemene, a non-cytotoxic anti-tumor drug developed by China with broad-spectrum anti-tumor properties, is also abundant in *Chrysanthemum VO*. Elemene Injection (approval number H20110114), Elemene Emulsion Injection (approval number H10960114), and Elemene Oral Emulsion (approval number H20010337) have been approved by the National Medical Products Administration in China for the treatment of liver, lung, esophageal, and other cancer types. As a non-cytotoxic drug, β -elemene exerts effects against different tumor cell types *via* multiple mechanisms. In general, the molecular mechanisms of β -elemene include blockage of the cell proliferation cycle, apoptosis induction, inhibition of tumor growth and metastasis, enhancement of tumor cell immunogenicity, and reversal of multidrug resistance^[69]. In addition, multiple signaling pathways and enzymes or proteins (including NF- κ B, MAPK, PI3K-Akt-mammalian target of rapamycin (mTOR), B-cell lymphoma 2 (Bcl-2) protein family and caspases, and Wnt/ β -catenin) are involved^[70]. Research on the combination of β -elemene and 5-fluorouracil showed

that β -elemene reversed the resistance of HCT116p53 to 5-fluorouracil by inducing apoptosis, autophagy, and cyclin D3-dependent cell cycle arrest^[84].

1,8-Cineole

1,8-Cineole is a highly selective cyclic monoterpene compound with potential anti-tumor activity against human colon, skin, liver, ovarian, osteosarcoma, and breast cancer cells *in vitro* and *in vivo*^[74]. Numerous preclinical efficacy studies have provided extensive evidence that 1,8-cineole regulates related signaling pathways and enzymes or proteins, such as tumor suppressor protein p53, PI3K/Akt, aryl hydrocarbon receptor (AHR)/cellular Src (c-Src)/epidermal growth factor receptor (EGFR), rapidly accelerated fibrosarcoma (RAF)-MEK1/2-ERK1/2, Bcl-2 protein family, and adenine monophosphate-activated protein kinase (AMPK), to destroy mitochondrial integrity, inhibit tumor cell proliferation and induce apoptosis^[85–87].

1,8-Cineole has also been shown to promote G0/G1 cycle arrest in HepG2 cells by inhibiting cell proliferation. Furthermore, when cells were incubated with 1,8-cineole at a concentration of 8 mM, the level of cyclin-dependent kinase 4/6 decreased, ROS production significantly increased, and ERK phosphorylation (p-ERK) and p-p38 increased by 2- and 5.7-fold, respectively. Although the p-Akt was higher, the p-p70S6K (p70 ribosomal protein S6 kinase, mTOR downstream target) was decreased by 57%, and AMPK was activated, which induced the senescence of HepG2 cells to anti-senescence compounds. Therefore, the combination of 1,8-cineole and these compounds synergistically inhibited tumor cell viability and induced tumor cell apoptosis^[88].

Other pharmacological effects and mechanisms

Anti-pyretic and analgesic activity

Fever is usually caused by the interaction between pyrogens and the organum vasculosum lamina terminalis, which leads to the upregulation of body temperature by the hypothalamic thermoregulation center, resulting in fever^[89]. Pain is triggered by the formation and release of pain-inducing substances, such as histamine, dopamine (DA), 5-hydroxytryptamine (5-HT), and prostaglandins (PG), at the site of inflammation or injury, which act on pain receptors^[90,91]. The mechanism underlying antipyretic and analgesic activity is similar to that of anti-inflammatory action, which involves blockage or reduction of NO, PGE₂, and other related mediators through inhibiting expression of COX-2 and iNOS or regulation of related signaling pathways, in particular NF- κ B^[92,93]. In studies on the antipyretic effect and underlying mechanisms of VO from *Chrysanthemum morifolium* using a New Zealand rabbit model of endotoxin-induced fever, the high-dose group (0.096 mL/kg VO) showed reduced norepinephrine (NE) and DA contents from 0.17 and 0.09 μ g/mg to 0.11 and 0.05 μ g/mg, respectively, compared with the model group, while the 5-HT content increased from 1.59 to 2.40 ng/mg, inferring that the antipyretic effect of VO is related to alterations in the 5-HT, DA, and NE contents^[94].

Table 2

Main anti-tumor components and mechanisms of Chrysanthemum VO

Active component	Cancer type	Mechanism										Reference
		Induces cell autophagy	Induces oxidative stress	Inhibits cell proliferation	Blocks cell cycle	Induces apoptosis	Induces cell DNA damage	Inhibits migration and invasion	Induces mitochondrial structural change or apoptosis			
Borneol	Liver cancer, glioma	✓	✓	✓	✓	✓	✓	✓	✓	✓	[55,56]	
Geraniol	Breast cancer, colon cancer, kidney cancer, liver cancer, lung cancer, oral cancer, pancreatic carcinoma, prostate cancer, skin cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[57,58]	
Thujone	Human placental choriocarcinoma, malignant melanoma, ovarian cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[59-61]	
Limonene	Bladder cancer, breast cancer, colon cancer, gastric cancer, leukemia, liver cancer, lung cancer, lymphoma, neuroblastoma, pancreatic cancer, prostate cancer, skin cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[62,63]	
Carvacrol	Breast cancer, cervical cancer, colon cancer, gastric cancer, leukemia, liver cancer, lung cancer, melanoma, neuroblastoma, oral cancer, prostate cancer, human glioblastoma, human tongue squamous cell carcinoma	✓	✓	✓	✓	✓	✓	✓	✓	✓	[64,65]	
α-Pinene	Cervical cancer, gastric cancer, liver cancer, ovarian cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[66,67]	
Thymol	Bladder cancer, bone cancer, breast cancer, cervical cancer, colon cancer, colorectal adenocarcinoma, gastric cancer, glioblastoma, liver cancer, lung cancer, neuroblastoma, prostate cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[64-68]	
β-Elemene	Bladder cancer, bone metastasis, brain cancer, breast cancer, cervical cancer, esophageal cancer, gastric cancer, liver cancer, lung cancer, malignant glioma, nasopharyngeal cancer, ovarian cancer, prostate cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[69,70]	
β-Caryophyllene	Breast cancer, cervical cancer, colon cancer, gastric cancer, leukemia, liver cancer, lung cancer, ovarian cancer, prostate cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[71-73]	
1,8-Cineole	Breast cancer, colon cancer, leukemia, liver cancer, oral cancer, ovarian cancer, skin cancer,	✓	✓	✓	✓	✓	✓	✓	✓	✓	[74]	
α-Bisabolol	Breast cancer, endometrial cancer, glioblastoma cells, hepatocellular carcinoma, non-small cell lung cancer, leukemia cells, liver cancer, pancreatic cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[75-77]	
PTL	Breast cancer, colorectal cancer, gastric cancer, leukemia, liver cancer, lung cancer, malignant glioma, melanoma, prostate cancer, thyroid cancer	✓	✓	✓	✓	✓	✓	✓	✓	✓	[78-80]	

PTL: Parthenolide.

Anti-hypertensive activity

Herbal medicine protects against hypertension mainly through the inhibition of sympathetic nerve activity, blockage of the calcium channel, diuresis, hypotension, suppression of the pathological processes in the renin-angiotensin system, improvement of vascular endothelial function and insulin resistance, and effects on cardiac hemodynamics and hemorheology or synergistic effects of multiple pathways of hypotension^[95,96]. 1,8-Cineole from VO inhibited calcium ion influx in cells with concomitant relaxation of isolated arteries and reduction of aortic pressure^[97], while carvacrol increased cyclic adenosine monophosphate (cAMP) and cyclic guanosine monophosphate (cGMP) levels by inhibiting various phosphodiesterases, exerting an endothelium-independent vasodilatory effect^[98]. The systolic blood pressure and heart rate of subjects decreased after inhalation of VO; in addition, brain waves θ and α increased in the relaxed brain state while β and γ waves decreased during activity, indicating that VO from *Chrysanthemum indicum* helps to reduce blood pressure and relax the body and mental state^[99]. Chrysanthemum extract, rich in polyphenols, reduces renovascular hypertension. Through inhibiting the expression of hypoxia-inducible factor-1 α and regulating carnitine palmitoyltransferase-1a, pyruvate dehydrogenase kinase-4, and glucose transporter-4, Chrysanthemum extract improved myocardial energy metabolism, leading to the attenuation of hypertension-induced myocardial hypertrophy in rats^[100].

Sedative and hypnotic effects

Insomnia is a disorder that leads to difficulty falling asleep or maintaining sleep, poor sleep quality, and shortened total sleep time as the main manifestations. The pathogenesis of insomnia is mainly associated with central neurotransmitters, inflammatory factors, and the hypothalamus-pituitary-adrenal axis^[101,102]. Sleep deficiency reduces the quality of life and leads to mental health problems, especially anxiety and depression^[103]. A number of clinical and pharmacological studies have validated the utility of VO in medicinal plants administered *via* inhalation, oral administration, or aromatherapy for sedation and hypnosis, and relief of anxiety and depression^[104]. Studies have indicated that the ethanolic extract of *Chrysanthemum morifolium* can regulate the expression of glutamic acid decarboxylase and enhance pentobarbital-induced sleep behavior, which may be caused by the activation of Cl⁻ channels^[105,106]. Chrysanthemum VO may participate in sleep regulation mechanisms to improve sleep by regulating the levels of 5-HT, NE, or TNF- α to exhibit sedative and hypnotic effects^[107]. However, the underlying mechanisms have not yet been elucidated.

Anti-osteoporotic activity

Osteoporosis is a systemic metabolic bone disease characterized by decreased bone mass, destruction of bone tissue microstructure, and an increased risk of bone fragility and fracture^[108]. Herbal medicines exert anti-osteoporotic effects through regulation of Wnt/ β -catenin,

bone morphogenetic protein (BMP)/Smad, MAPK pathways and the receptor activator of NF- κ B ligand (RANKL)/osteoprotegerin system (OPG)^[109]. VOs are herbal components that effectively regulate bone metabolism and prevent osteoporosis^[110]. Monoterpenes (borneol, camphor, menthol, and thymol) isolated from VOs inhibit bone resorption *in vivo* by directly affecting osteoclast formation in hematopoietic cells^[111]. The growth level of osteoblasts treated with 10 μ g/mL VO of *Chrysanthemum indicum* L. increased to 111%, alkaline phosphatase activity increased to 109%, collagen synthesis level reached 114%, and mineralization increased to 118% compared with the control group, indicating that VO promotes collagen synthesis, activates alkaline phosphatase activity, and accelerates calcium deposition of MC3T3-E1 cells, supporting its utility as a pharmacological agent for osteoporosis^[112].

Anti-viral effects

Terpenoids, phenols, aldehydes, and their derivatives in plant VO exert inhibitory effects during different stages of the viral replication cycle by inactivating the virus, protecting host cells, inhibiting viral adsorption by host cells, controlling the proliferation and spread of the virus, and regulating immunity^[113,114]. 1,8-Cineole directly combines with and inactivates free viruses by binding to viral proteins involved in the entry and penetration of host cells, thus effectively preventing viral infections^[115]. Carvacrol attenuated the excessive immune response induced by the influenza A virus by suppressing viral replication and recognizing TLR/retinoic acid-induced gene I-like receptor (RLR) patterns^[116]. β -Caryophyllene improved damage induced by toxic Newcastle disease virus in birds by suppressing viral replication and regulating immunity to increase the level of interferon α (a signaling cytokine), exerting optimal therapeutic effects before or during infection^[117]. Certain sesquiterpenoids in Chrysanthemum VO exhibit selective anti-viral activities. Three sesquiterpenoids (chrysanthemum B, 6,8-cycloeuodesm-4(15)-en-1-ol, and 1 β -hydroxy-4(15),5E,10(14)-germacatriene) isolated from *Chrysanthemum indicum* inhibited the replication of porcine epidemic diarrhea virus in a dose-dependent manner (concentration range: 20–90 μ M). Moreover, chrysanthemum B had an inhibitory effect on the synthesis of viral nucleocapsid in a dose-dependent manner and spike protein at 80, 40, 20, and 10 μ M^[118].

Prospects

Novel VO delivery systems

Chrysanthemum VO exhibits a variety of biological activities and pharmacological effects and has been widely investigated and used in the fields of medicine, food, agriculture, and skin care. Notably, because of the volatility, instability, and hydrophobicity of VO and its susceptibility to enzymatic reactions, its utility is limited^[119]. The use of novel delivery systems (NDS) for encapsulating VOs can overcome these drawbacks. This is a promising strategy for the development and innovation of Chrysanthemum VO application.

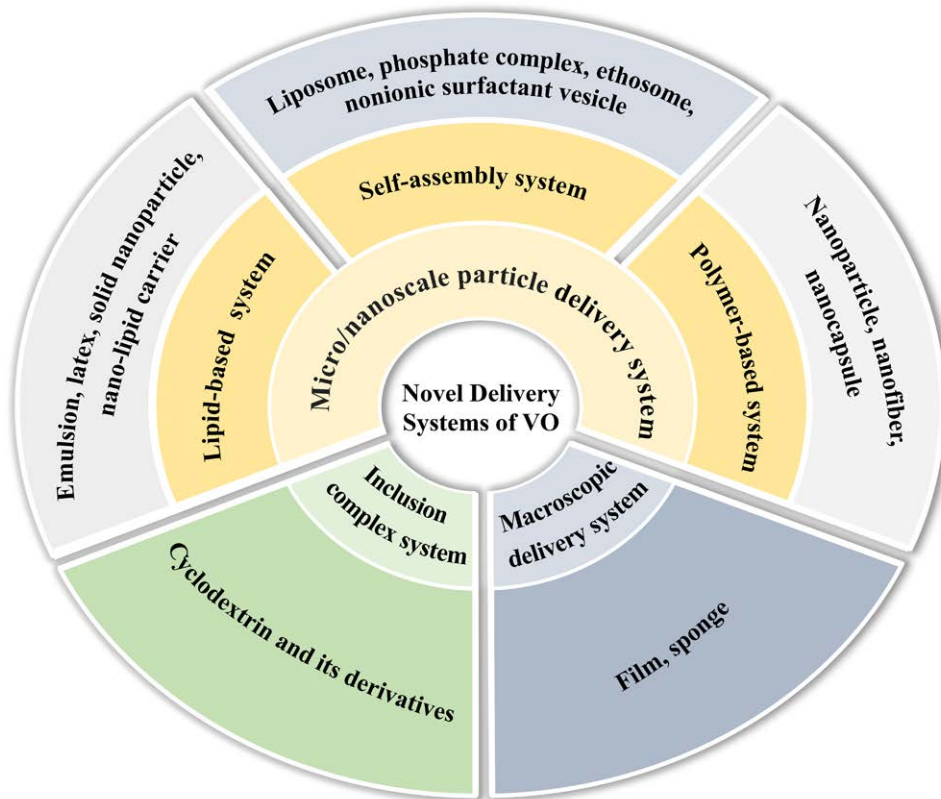


Figure 5. Novel delivery systems designed for VO. VO: Volatile oil.

The encapsulation of VO is commonly defined as a technical means and method strategy to encapsulate VO in “inert” materials with special structure to improve the stability and bioavailability of VO^[120]. Based on the different encapsulation materials and techniques, novel delivery systems can be mainly divided into inclusion complexes, micro/nanoscale particles, and macroscopic delivery systems^[121]. Figure 5 shows the NDS used to improve the stability and bioavailability of the VO.

Among them, cyclodextrins, liposomes, and nanoparticles have been widely explored as potential delivery systems for VO. Cyclodextrin inclusion complexes have polar hydroxyl groups on the outside of the ring and nonpolar hydrogen and etheral oxygen on the inside of the ring, forming hydrophobic cavities that can encapsulate VO within the cavity and improve its stability^[122]. Qin et al.^[123] have demonstrated that the combination of Chrysanthemum VO with β -cyclodextrin can reduce the instability and photosensitivity of VO, and effectively disguise the bad odor of VO, whereas the properties and main components of VO remain unchanged. Moreover, researchers have used chitosan and pectin to modify Chrysanthemum VO liposomes to form more stable Chrysanthemum VO triple-layer liposomes, which showed better antibacterial activity against *Campylobacter jejuni* in chickens without affecting their quality^[124]. Moreover, nanotechnology for VO delivery, which is characterized by improved cellular uptake, controlled release, and precise targeting, is a current focus of interest^[125,126]. Compared to traditional emulsions, nanoemulsions are stabler because of their higher surface-to-volume ratio, which can be used as a promising

antibacterial delivery method to enhance the stability of VO, organ and cellular targeting, and antibacterial efficacy^[127].

Pharmacological effects of VO can be further enhanced by (1) achieving targeted drug delivery and controlled drug release to enhance bioavailability and efficiency, (2) preventing hydrolysis and oxidation to improve the chemical stability, and (3) reducing toxicity and volatility^[128]. However, studies on the NDS of Chrysanthemum VO are still in the preliminary exploratory stage, particularly for nanodelivery systems and clinical availability may take some time.

VO mechanisms of action

Since Chrysanthemum VO has complex components that act on pathological tissues through multiple pathways and targets, study of the mechanisms of the pharmacological effects of Chrysanthemum VO is challenging. The pharmacological mechanisms of the sedative, hypnotic, hypotensive, and analgesic effects remain the focus.

To develop more feasible patented products for use in the medical, food, and agricultural fields, the mechanisms of action and pharmacological toxicity of VO should be comprehensively investigated, along with further *in vivo* studies and innovations in the formula encapsulation of VO, with the aim of providing reliable data to ensure safe and effective usage.

Conclusions

Chrysanthemum is a medicinal and edible herb containing abundant VO. Over the years, the utility of

Chrysanthemum VO as a therapeutic agent with great potential has been gradually recognized by researchers in various fields, not only in clinical and cosmetic skin care as a potential natural bacteriostatic or antioxidant agent and in other therapeutic drugs^[129,130], but also as a potential natural antiseptic additive and biodegradable food packaging in the food industry^[131]. The VO extract is also a natural biological insecticide that is commonly used in agricultural pest control^[132].

This review focuses on the pharmacological effects of Chrysanthemum VO and their mechanisms of action. Recent advances in effective delivery by NDS are also summarized to shed light on further applications of VO as an important resource for medicinal treatment, aromatherapy, and dietary therapeutic applications.

Conflict of interest statement

The authors declare no conflict of interest.

Funding

This research was funded by the National Natural Science Foundation of China (82260695), the Jiangxi Provincial Natural Science Foundation (20232ACB206062, 20212ACB206004), Young Jinggang Scholar of Jiangxi Province and New Century Talents Project of Jiangxi Province (2017082, 2020028), the Science and Technology Innovation Team of Jiangxi University of Chinese Medicine (CXTD22001 and CXTD22006), and Project of College Students' Innovation and Entrepreneurship Training Program of Jiangxi University of Chinese Medicine.

Author contributions

Jing Zhang, Xiang Li, and Ming Yang conceived and designed the review. Weiqiang Su, Yao He, Ying Luo, Nina Filipczak, Shijuan Yan, Ming Yang, and Anping Wan performed the research. Jing Zhang and Weiqiang Su drafted the manuscript. Xiang Li and Nina Filipczak revised the manuscript. All authors read and approved the final manuscript.

Ethical approval of studies and informed consent

Not applicable.

Acknowledgments

None.

Data availability

All data generated or analyzed during this study are included in this published article.

References

- Wu D, Zhuang F, Wang J, et al. Metabolomics and transcriptomics revealed a comprehensive understanding of the biochemical and genetic mechanisms underlying the color variations in Chrysanthemums. *Metabolites* 2023;13(6):742.
- Liang X, Wu H, Su W. A rapid UPLC-PAD fingerprint analysis of Chrysanthemum morifolium Ramat combined with chemometrics methods. *Food Anal Methods* 2014;7(1):197–204.
- Lee M, Shim SY. Inhibitory effects of eriodictyol-7-O- β -d-glucuronide and 5,7-dihydroxy-4-chromene isolated from Chrysanthemum zawadskii var. latilobum in Fc ϵ RI-mediated human basophilic KU812F cell activation. *Molecules* 2020;25(4):994.
- Park S, Lee JB, Kang S. Topical application of Chrysanthemum indicum L. attenuates the development of atopic dermatitis-like skin lesions by suppressing serum IgE levels, IFN- γ , and IL-4 in Nc/Nga mice. *Evid Based Complement Alternat Med* 2012;2012:821967.
- Hanieh H, Leila S, Abolfazl S. Chrysanthemum, an ornamental genus with considerable medicinal value: a comprehensive review. *S Afr J Bot* 2022;144:23–43.
- Yuan H, Jiang S, Liu Y, et al. The flower head of Chrysanthemum morifolium Ramat. (Juhua): a paradigm of flowers serving as Chinese dietary herbal medicine. *J Ethnopharmacol* 2020;261:113043.
- Chinese Pharmacopoeia Commission. *Pharmacopoeia of the People's Republic of China*. Beijing: China Medical Science Press; 2020:323, 328.
- Zhu S, Yang Y, Yu H, et al. Chemical composition and antimicrobial activity of the essential oils of Chrysanthemum indicum. *J Ethnopharmacol* 2005;96(1-2):151–158.
- Kuang CL, Lv D, Shen GH, et al. Chemical composition and antimicrobial activities of volatile oil extracted from Chrysanthemum morifolium Ramat. *J Food Sci Technol* 2018;55(7):2786–2794.
- Lin L, Mao X, Sun Y, et al. Antibacterial properties of nanofibers containing chrysanthemum essential oil and their application as beef packaging. *Int J Food Microbiol* 2018;292:21–30.
- Kim BS, Park SJ, Kim MK, et al. Inhibitory effects of Chrysanthemum boreale essential oil on biofilm formation and virulence factor expression of Streptococcus mutans. *Evid Based Complement Alternat Med* 2015;2015:616309.
- Youssef FS, Eid SY, Alshammari E, et al. Chrysanthemum indicum and Chrysanthemum morifolium: chemical composition of their essential oils and their potential use as natural preservatives with antimicrobial and antioxidant activities. *Foods* 2020;9(10):1460.
- Chang KM, Kim GH. Volatiles of Chrysanthemum zawadskii var. latilobum K. *Prev Nutr Food Sci* 2012;17(3):234–238.
- Carson CF, Mee BJ, Riley TV. Mechanism of action of Melaleuca alternifolia (tea tree) oil on Staphylococcus aureus determined by time-kill, lysis, leakage, and salt tolerance assays and electron microscopy. *Antimicrob Agents Chemother* 2002;46(6):1914–1920.
- Angane M, Swift S, Huang K, et al. Essential oils and their major components: an updated review on antimicrobial activities, mechanism of action and their potential application in the food industry. *Foods* 2022;11(3):464.
- Nazzaro F, Fratianni F, De Martino L, et al. Effect of essential oils on pathogenic bacteria. *Pharmaceuticals (Basel)* 2013;6(12):1451–1474.
- Burt S. Essential oils: their antibacterial properties and potential applications in foods—a review. *Int J Food Microbiol* 2004;94(3):223–253.
- Han XB, Zhao J, Cao JM, et al. Essential oil of Chrysanthemum indicum L.: potential biocontrol agent against plant pathogen Phytophthora nicotianae. *Environ Sci Pollut Res Int* 2019;26(7):7013–7023.
- Kalemba D, Kunicka A. Antibacterial and antifungal properties of essential oils. *Curr Med Chem* 2003;10(10):813–829.
- Labbozzetta M, Poma P, Occhipinti C, et al. Antitumor effect of Glandora rosmarinifolia (Boraginaceae) essential oil through inhibition of the activity of the Topo II enzyme in acute myeloid leukemia. *Molecules* 2022;27(13):4203.
- Hassannejad N, Bahador A, Rudbari NH, et al. In vivo antibacterial activity of Zataria multiflora Boiss extract and its components, carvacrol, and thymol, against colistin-resistant Acinetobacter baumannii in a pneumonic BALB/c mouse model. *J Cell Biochem* 2019;120(11):18640–18649.
- Cui H, Bai M, Sun Y, et al. Antibacterial activity and mechanism of Chuzhou chrysanthemum essential oil. *J Funct Foods* 2018;48:159–166.
- Vasconcelos NG, Croda J, Simionatto S. Antibacterial mechanisms of cinnamon and its constituents: a review. *Microb Pathog* 2018;120:198–203.
- Ahmad A, Khan A, Yousuf S, et al. Proton translocating ATPase mediated fungicidal activity of eugenol and thymol. *Fitoterapia* 2010;81(8):1157–1162.
- da Silva BD, Bernardes PC, Pinheiro PF, et al. Chemical composition, extraction sources and action mechanisms of essential oils: natural preservative and limitations of use in meat products. *Meat Sci* 2021;176:108463.

- [26] Tapia-Rodriguez MR, Bernal-Mercado AT, Gutierrez-Pacheco MM, et al. Virulence of *Pseudomonas aeruginosa* exposed to carvacrol: alterations of the quorum sensing at enzymatic and gene levels. *J Cell Commun Signaling* 2019;13(4):531–537.
- [27] Tasneem S, Liu B, Li B, et al. Molecular pharmacology of inflammation: medicinal plants as anti-inflammatory agents. *Pharmacol Res* 2019;139:126–140.
- [28] Liu C, Chu D, Kalantar-Zadeh K, et al. Cytokines: from clinical significance to quantification. *Adv Sci (Weinb)* 2021;8(15):e2004433.
- [29] Tayal V, Kalra BS. Cytokines and anti-cytokines as therapeutics—an update. *Eur J Pharmacol* 2008;579(1–3):1–12.
- [30] Cheon MS, Yoon T, Lee DY, et al. Chrysanthemum indicum Linné extract inhibits the inflammatory response by suppressing NF-kappaB and MAPKs activation in lipopolysaccharide-induced RAW 264.7 macrophages. *J Ethnopharmacol* 2009;122(3):473–477.
- [31] Jiang S, Wang M, Jiang Z, et al. Chemistry and pharmacological activity of sesquiterpenoids from the Chrysanthemum genus. *Molecules* 2021;26(10):3038.
- [32] Kim JG, Lee JW, Le TPL, et al. Sesquiterpenoids from Chrysanthemum indicum with inhibitory effects on NO production. *J Nat Prod* 2021;84(3):562–569.
- [33] Li S, Gao X, Wu X, et al. Parthenolide inhibits LPS-induced inflammatory cytokines through the toll-like receptor 4 signal pathway in THP-1 cells. *Acta Biochim Biophys Sin (Shanghai)* 2015;47(5):368–375.
- [34] Pyee Y, Chung H-J, Choi TJ, et al. Suppression of inflammatory responses by handelin, a guaianolide dimer from Chrysanthemum boreale, via downregulation of NF- κ B signaling and pro-inflammatory cytokine production. *J Nat Prod* 2014;77(4):917–924.
- [35] Yoon WJ, Moon JY, Song G, et al. Artemisia fukudo essential oil attenuates LPS-induced inflammation by suppressing NF-kappaB and MAPK activation in RAW 264.7 macrophages. *Food Chem Toxicol* 2010;48(5):1222–1229.
- [36] Hoesel B, Schmid JA. The complexity of NF- κ B signaling in inflammation and cancer. *Mol Cancer* 2013;12:86.
- [37] Yang L, Liu J, Li Y, et al. Bornyl acetate suppresses ox-LDL-induced attachment of THP-1 monocytes to endothelial cells. *Biomed Pharmacother* 2018;103:234–239.
- [38] Krishna M, Narang H. The complexity of mitogen-activated protein kinases (MAPKs) made simple. *Cell Mol Life Sci* 2008;65(22):3525–3544.
- [39] Raman M, Chen W, Cobb MH. Differential regulation and properties of MAPKs. *Oncogene* 2007;26(22):3100–3112.
- [40] Zhuo F-F, Zhang C, Zhang H, et al. Chrysanthemulide A induces apoptosis through DR5 upregulation via JNK-mediated autophagosome accumulation in human osteosarcoma cells. *J Cell Physiol* 2019;234(8):13191–13208.
- [41] Chen L, Liu Y, Huang X, et al. Comparison of chemical constituents and pharmacological effects of different varieties of Chrysanthemum Flos in China. *Chem Biodivers* 2021;18(8):e2100206.
- [42] Yang L, Nuerbiye A, Cheng P, et al. Analysis of floral volatile components and antioxidant activity of different varieties of Chrysanthemum morifolium. *Molecules* 2017;22(10):1790.
- [43] Amorati R, Foti MC, Valgimigli L. Antioxidant activity of essential oils. *J Agric Food Chem* 2013;61(46):10835–10847.
- [44] Zeb A. Concept, mechanism, and applications of phenolic antioxidants in foods. *J Food Biochem* 2020;44(9):e13394.
- [45] Kim S-J. Chapter 21—Herbal Chrysanthemi Flos, oxidative damage and protection against diabetic complications. In: Preedy VR, ed. *Diabetes: Oxidative Stress and Dietary Antioxidants*. Pittsburgh: Academic Press; 2014:201–211.
- [46] Lee BH, Nam TG, Park WJ, et al. Antioxidative and neuroprotective effects of volatile components in essential oils from Chrysanthemum indicum Linn, flowers. *Food Sci Biotechnol* 2015;24(2):717–723.
- [47] Bardaweel SK, Tawaha KA, Hudaib MM. Antioxidant, antimicrobial and antiproliferative activities of Anthemis palestina essential oil. *BMC Complement Altern Med* 2014;14:297.
- [48] Kim DY, Won KJ, Hwang DI, et al. Chemical composition, antioxidant and anti-melanogenic activities of essential oils from Chrysanthemum boreale Makino at different harvesting stages. *Chem Biodivers* 2018;15(2):e1700506.
- [49] Zhan G, Long M, Shan K, et al. Antioxidant effect of Chrysanthemum morifolium (Chujin) extract on H₂O₂-treated L-O₂ cells as revealed by LC/MS-based metabolic profiling. *Antioxidants (Basel)* 2022;11(6):1068.
- [50] Liu Y, Zhou F, Shu HZ, et al. Germacrane-type sesquiterpenoids from the flowers of Chrysanthemum indicum with hepatoprotective activity. *Food Chem Toxicol* 2023;177:113850.
- [51] Ma A, Zou F, Zhang R, et al. The effects and underlying mechanisms of medicine and food homologous flowers on the prevention and treatment of related diseases. *J Food Biochem* 2022;46(12):e14430.
- [52] Yang HM, Sun CY, Liang JL, et al. Supercritical-carbon dioxide fluid extract from Chrysanthemum indicum enhances anti-tumor effect and reduces toxicity of bleomycin in tumor-bearing mice. *Int J Mol Sci* 2017;18(3):465.
- [53] Tang T, Liao ZG, Dong W, et al. Correlation between four properties of traditional Chinese medicine and function of reversing multidrug resistance of tumor cells. *Zhongguo Zhong Yao Za Zhi* 2017;42(4):795–799.
- [54] Liu LL, Chen J, Shi YP. Advance in studies on antitumor of Chinese materia medica with heat-clearing and toxin-resolving functions. *Chin Tradit Herb Drugs* 2012;43(6):1203–1212.
- [55] Su J, Lai H, Chen J, et al. Natural borneol, a monoterpene compound, potentiates selenocystine-induced apoptosis in human hepatocellular carcinoma cells by enhancement of cellular uptake and activation of ROS-mediated DNA damage. *PLoS One* 2013;8(5):e63502.
- [56] Cao WQ, Zhai XQ, Ma JW, et al. Natural borneol sensitizes human glioma cells to cisplatin-induced apoptosis by triggering ROS-mediated oxidative damage and regulation of MAPKs and PI3K/AKT pathway. *Pharm Biol* 2020;58(1):72–79.
- [57] Cho M, So I, Chun JN, et al. The antitumor effects of geraniol: modulation of cancer hallmark pathways (review). *Int J Oncol* 2016;48(5):1772–1782.
- [58] Silva G, Marques JNJ, Linhares EPM, et al. Review of anticancer activity of monoterpenoids: geraniol, nerol, geranial and neral. *Chem Biol Interact* 2022;362:109994.
- [59] Lee JY, Park H, Lim W, et al. Therapeutic potential of α , β -thujone through metabolic reprogramming and caspase-dependent apoptosis in ovarian cancer cells. *J Cell Physiol* 2021;236(2):1545–1558.
- [60] Biswas R, Mandal SK, Dutta S, et al. Thujone-rich fraction of Thuja occidentalis demonstrates major anti-cancer potentials: evidences from in vitro studies on A375 cells. *Evid Based Complement Alternat Med* 2011;2011:568148.
- [61] Lee JY, Park H, Lim W, et al. α , β -Thujone suppresses human placental choriocarcinoma cells via metabolic disruption. *Reproduction* 2020;159(6):745–756.
- [62] Mukhtar YM, Adu-Frimpong M, Xu X, et al. Biochemical significance of limonene and its metabolites: future prospects for designing and developing highly potent anticancer drugs. *Biosci Rep* 2018;38(6):BSR20181253.
- [63] Araújo-Filho HG, Dos Santos JF, Carvalho MTB, et al. Anticancer activity of limonene: a systematic review of target signaling pathways. *Phytother Res* 2021;35(9):4957–4970.
- [64] Sampaio LA, Pina LTS, Serafini MR, et al. Antitumor effects of carvacrol and thymol: a systematic review. *Front Pharmacol* 2021;12:702487.
- [65] Suntres ZE, Coccimiglio J, Alipour M. The bioactivity and toxicological actions of carvacrol. *Crit Rev Food Sci Nutr* 2015;55(3):304–318.
- [66] Xu Q, Li M, Yang M, et al. α -Pinene regulates miR-221 and induces G(2)/M phase cell cycle arrest in human hepatocellular carcinoma cells. *Biosci Rep* 2018;38(6):BSR20180980.
- [67] Chen W, Liu Y, Li M, et al. Anti-tumor effect of α -pinene on human hepatoma cell lines through inducing G2/M cell cycle arrest. *J Pharmacol Sci* 2015;127(3):332–338.
- [68] Islam MT, Khalipha ABR, Bagchi R, et al. Anticancer activity of thymol: a literature-based review and docking study with emphasis on its anticancer mechanisms. *IUBMB Life* 2019;71(1):9–19.
- [69] Bai Z, Yao C, Zhu J, et al. Anti-tumor drug discovery based on natural product β -elemene: anti-tumor mechanisms and structural modification. *Molecules* 2021;26(6):1499.
- [70] Zhai B, Zhang N, Han X, et al. Molecular targets of β -elemene, a herbal extract used in traditional Chinese medicine, and its potential role in cancer therapy: a review. *Biomed Pharmacother* 2019;114:108812.
- [71] Chung KS, Hong JY, Lee JH, et al. β -Caryophyllene in the essential oil from Chrysanthemum boreale induces G(1) phase cell cycle arrest in human lung cancer cells. *Molecules* 2019;24(20):3754.
- [72] Fidyk K, Fiedorowicz A, Strzadala L, et al. β -Caryophyllene and β -caryophyllene oxide-natural compounds of anticancer and analgesic properties. *Cancer Med* 2016;5(10):3007–3017.
- [73] Park KR, Nam D, Yun HM, et al. β -Caryophyllene oxide inhibits growth and induces apoptosis through the suppression of PI3K/AKT/mTOR/S6K1 pathways and ROS-mediated MAPKs activation. *Cancer Lett* 2011;312(2):178–188.

- [74] Cai ZM, Peng JQ, Chen Y, et al. 1,8-Cineole: a review of source, biological activities, and application. *J Asian Nat Prod Res* 2021;23(10):938–954.
- [75] Eddin LB, Jha NK, Goyal SN, et al. Health benefits, pharmacological effects, molecular mechanisms, and therapeutic potential of α -bisabolol. *Nutrients* 2022;14(7):1370.
- [76] Jin M, Xiao Z, Zhang S, et al. Possible involvement of Fas/FasL-dependent apoptotic pathway in α -bisabolol induced cardiotoxicity in zebrafish embryos. *Chemosphere* 2018;219:557–566.
- [77] Rigo A, Vinante F. The antineoplastic agent α -bisabolol promotes cell death by inducing pores in mitochondria and lysosomes. *Apoptosis* 2016;21(8):917–927.
- [78] Sztiller-Sikorska M, Czyn M. Parthenolide as cooperating agent for anti-cancer treatment of various malignancies. *Pharmaceuticals (Basel)* 2020;13(8):194.
- [79] Liu X, Wang X. Recent advances on the structural modification of parthenolide and its derivatives as anticancer agents. *Chin J Nat Med* 2022;20(11):814–829.
- [80] Freund RRA, Gobrecht P, Fischer D, et al. Advances in chemistry and bioactivity of parthenolide. *Nat Prod Rep* 2020;37(4):541–565.
- [81] Cui ZY, Wang G, Zhang J, et al. Parthenolide, bioactive compound of *Chrysanthemum parthenium* L., ameliorates fibrogenesis and inflammation in hepatic fibrosis via regulating the crosstalk of TLR4 and STAT3 signaling pathway. *Phytother Res* 2021;35(10):5680–5693.
- [82] Czyn M, Lesiak-Mieczkowska K, Koprowska K, et al. Cell context-dependent activities of parthenolide in primary and metastatic melanoma cells. *Br J Pharmacol* 2010;160(5):1144–1157.
- [83] Oka D, Nishimura K, Shiba M, et al. Sesquiterpene lactone parthenolide suppresses tumor growth in a xenograft model of renal cell carcinoma by inhibiting the activation of NF- κ B. *Int J Cancer* 2007;120(12):2576–2581.
- [84] Zhang R, Pan T, Xiang Y, et al. β -Elemene reverses the resistance of p53-deficient colorectal cancer cells to 5-fluorouracil by inducing pro-death autophagy and cyclin D3-dependent cycle arrest. *Front Bioeng Biotechnol* 2020;8:378.
- [85] Sampath S, Subramani S, Janardhanan S, et al. Bioactive compound 1,8-cineole selectively induces G2/M arrest in A431 cells through the upregulation of the p53 signaling pathway and molecular docking studies. *Phytomedicine* 2018;46:57–68.
- [86] Murata S, Shiragami R, Kosugi C, et al. Antitumor effect of 1, 8-cineole against colon cancer. *Oncol Rep* 2013;30(6):2647–2652.
- [87] Lee J, Ha SJ, Park J, et al. 1,8-Cineole prevents UVB-induced skin carcinogenesis by targeting the aryl hydrocarbon receptor. *Oncotarget* 2017;8(62):105995–106008.
- [88] Rodenak-Kladniew B, Castro A, Stärkel P, et al. 1,8-Cineole promotes G0/G1 cell cycle arrest and oxidative stress-induced senescence in HepG2 cells and sensitizes cells to anti-senescence drugs. *Life Sci* 2020;243:117271.
- [89] Walter EJ, Hanna-Jumma S, Carraretto M, et al. The pathophysiological basis and consequences of fever. *Crit Care* 2016;20(1):200.
- [90] Basbaum AI, Bautista DM, Scherrer G, et al. Cellular and molecular mechanisms of pain. *Cell* 2009;139(2):267–284.
- [91] Woolf CJ. What is this thing called pain? *J Clin Invest* 2010;120(11):3742–3744.
- [92] Wang R, Han L, Gao Q, et al. Progress on active analgesic components and mechanisms of commonly used traditional Chinese medicines: a comprehensive review. *J Pharm Pharm Sci* 2018;21(1):437–480.
- [93] Inaba H, Yoshigai E, Okuyama T, et al. Antipyretic analgesic drugs have different mechanisms for regulation of the expression of inducible nitric oxide synthase in hepatocytes and macrophages. *Nitric Oxide* 2015;44:61–70.
- [94] Zhao-dan W, Ji-han S, Tian-qi P, et al. Antipyretic mechanism of *Chrysanthemum morifolium* essential oil on fever New Zealand rabbits model induced by endotoxin. *Shizhen Guoyi Guoyao* 2018;29(9):2053–2056.
- [95] Dong Z, Dai H, Feng Z, et al. Mechanism of herbal medicine on hypertensive nephropathy (Review). *Mol Med Rep* 2021;23(4):234.
- [96] Tang F, Yan HL, Wang LX, et al. Review of natural resources with vasodilation: traditional medicinal plants, natural products, and their mechanism and clinical efficacy. *Front Pharmacol* 2021;12:627458.
- [97] Pinto NV, Assreuy AMS, Coelho-de-Souza AN, et al. Endothelium-dependent vasorelaxant effects of the essential oil from aerial parts of *Alpinia zerumbet* and its main constituent 1,8-cineole in rats. *Phytomedicine* 2009;16(12):1151–1155.
- [98] Alamgeer, Auger C, Chabert P, et al. Mechanisms underlying vasorelaxation induced in the porcine coronary arteries by Thymus linearis, Benth. *J Ethnopharmacol* 2018;225:211–219.
- [99] Kim D-S, Goo Y-M, Cho J, et al. Effect of volatile organic chemicals in *Chrysanthemum indicum* Linné on blood pressure and electroencephalogram. *Molecules* 2018;23(8):2063.
- [100] Gao T, Zhu Z-Y, Zhou X, et al. *Chrysanthemum morifolium* extract improves hypertension-induced cardiac hypertrophy in rats by reduction of blood pressure and inhibition of myocardial hypoxia inducible factor-1 α expression. *Pharm Biol* 2016;54(12):2895–2900.
- [101] Vgontzas AN, Puzino K, Fernandez-Mendoza J, et al. Effects of trazodone versus cognitive behavioral therapy in the insomnia with short sleep duration phenotype: a preliminary study. *J Clin Sleep Med* 2020;16(12):2009–2019.
- [102] Shi MM, Piao JH, Xu XL, et al. Chinese medicines with sedative-hypnotic effects and their active components. *Sleep Med Rev* 2016;29:108–118.
- [103] Riemann D, Krone LB, Wulff K, et al. Sleep, insomnia, and depression. *Neuropsychopharmacology* 2020;45(1):74–89.
- [104] Cui J, Li M, Wei Y, et al. Inhalation aromatherapy via brain-targeted nasal delivery: natural volatiles or essential oils on mood disorders. *Front Pharmacol* 2022;13:860043.
- [105] Kim JW, Han JY, Hong JT, et al. Ethanol extract of the flower *Chrysanthemum morifolium* augments pentobarbital-induced sleep behaviors: involvement of Cl⁻ channel activation. *Evid Based Complement Alternat Med* 2011;2011:109164.
- [106] Kim M, Kim Y, Lee HW, et al. *Chrysanthemum morifolium* and its bioactive substance enhanced the sleep quality in rodent models via Cl⁻ channel activation. *Nutrients* 2023;15(6):1309.
- [107] Wang W. *The study on sedative and hypnotic effects of Chrysanthemum extract*. Master. Henan University. 2020.
- [108] Yang TL, Shen H, Liu A, et al. A road map for understanding molecular and genetic determinants of osteoporosis. *Nat Rev Endocrinol* 2020;16(2):91–103.
- [109] Zhang ND, Han T, Huang BK, et al. Traditional Chinese medicine formulas for the treatment of osteoporosis: implication for antiosteoporotic drug discovery. *J Ethnopharmacol* 2016;189:61–80.
- [110] Mühlbauer RC, Lozano A, Palacio S, et al. Common herbs, essential oils, and monoterpenes potently modulate bone metabolism. *Bone* 2003;32(4):372–380.
- [111] Dolder S, Hofstetter W, Wetterwald A, et al. Effect of monoterpenes on the formation and activation of osteoclasts *in vitro*. *J Bone Miner Res* 2006;21(4):647–655.
- [112] Mi CK, Mi CE, Hee KG. Chemical constituents of *Chrysanthemum indicum* L. flower oil and effect on osteoblastic MC3T3-E1 cells. *Food Sci Biotechnol* 2010;19(3):815–819.
- [113] Liao CH, Xiao HT, Zheng K, et al. Natural products: the master regulators of antiviral cytokines. *Curr Org Chem* 2017;21(18):1805–1823.
- [114] Parvez MK, Arbab AH, Al-Dosari MS, et al. Antiviral natural products against chronic hepatitis B: recent developments. *Curr Pharm Des* 2016;22(3):286–293.
- [115] Mieres-Castro D, Ahmar S, Shabbir R, et al. Antiviral activities of eucalyptus essential oils: their effectiveness as therapeutic targets against human viruses. *Pharmaceuticals (Basel)* 2021;14(12):1210.
- [116] Zheng K, Wu SZ, Lv YW, et al. Carvacrol inhibits the excessive immune response induced by influenza virus A via suppressing viral replication and TLR/RLR pattern recognition. *J Ethnopharmacol* 2021;268:113555.
- [117] Hassanin O, Abdallah F, Galal AAA. *In vitro* and *in vivo* experimental trials to assess the modulatory influence of β -caryophyllene on NDV replication and immunopathogenesis. *Comp Immunol Microbiol Infect Dis* 2020;73:101547.
- [118] Liu LL, Ha TK, Ha W, et al. Sesquiterpenoids with various carbocyclic skeletons from the flowers of *Chrysanthemum indicum*. *J Nat Prod* 2017;80(2):298–307.
- [119] Sherry M, Charcosset C, Fessi H, et al. Essential oils encapsulated in liposomes: a review. *J Liposome Res* 2013;23(4):268–275.
- [120] Sedaghat Doost A, Nikbakht Nasrabadi M, Kassozi V, et al. Recent advances in food colloidal delivery systems for essential oils and their main components. *Trends Food Sci Technol* 2020;99:474–486.
- [121] Wu Y, Wan N, Liu Y, et al. Stability improvement of Chinese medicinal essential oils based on delivery systems and their application in medical field. *Zhongguo Zhong Yao Za Zhi* 2022;47(3):603–610.

- [122] Arora D, Saneja A, Jaglan S. Cyclodextrin-based delivery systems for dietary pharmaceuticals. *Environ Chem Lett* 2019;17(3):1263–1270.
- [123] Qin D, Meng X, Li L. Synthesis and characterization of Jia-Xiang chrysanthemum volatile oil/ β -cyclodextrin inclusion complex. *Proc Natl Acad Sci India Phys Sci* 2014;84(3):381–385.
- [124] Lin L, Gu Y, Sun Y, et al. Characterization of chrysanthemum essential oil triple-layer liposomes and its application against *Campylobacter jejuni* on chicken. *LWT* 2019;107:16–24.
- [125] Mun H, Townley HE. Nanoencapsulation of plant volatile organic compounds to improve their biological activities. *Planta Med* 2021;87(3):236–251.
- [126] Sana SS, Li H, Zhang Z, et al. Recent advances in essential oils-based metal nanoparticles: a review on recent developments and biopharmaceutical applications. *J Mol Liq* 2021;333:115951.
- [127] Garcia CR, Malik MH, Biswas S, et al. Nanoemulsion delivery systems for enhanced efficacy of antimicrobials and essential oils. *Biomater Sci* 2022;10(3):633–653.
- [128] Cimino C, Maurel OM, Musumeci T, et al. Essential oils: pharmaceutical applications and encapsulation strategies into lipid-based delivery systems. *Pharmaceutics* 2021;13(3):327.
- [129] Kim DY, Won KJ, Yoon MS, et al. Chrysanthemum boreale Makino essential oil induces keratinocyte proliferation and skin regeneration. *Nat Prod Res* 2015;29(6):562–564.
- [130] Tran KN, Nguyen NPK, Nguyen LTH, et al. Screening for neuroprotective and rapid antidepressant-like effects of 20 essential oils. *Biomedicines* 2023;11(5):1248.
- [131] Tan LF, Elaine E, Pui LP, et al. Development of chitosan edible film incorporated with Chrysanthemum morifolium essential oil. *Acta Sci Pol Technol Aliment* 2021;20(1):55–66.
- [132] Alvarez-Castellanos PP, Bishop CD, Pascual-Villalobos MJ. Antifungal activity of the essential oil of flowerheads of garland chrysanthemum (*Chrysanthemum coronarium*) against agricultural pathogens. *Phytochem* 2001;57(1):99–102.