

# Version updates of strategies for drug discovery based on effective constituents of traditional Chinese medicine

Nan Ge<sup>1</sup>, Guangli Yan<sup>1</sup>, Hui Sun<sup>1,\*</sup>, Le Yang<sup>2</sup>, Ling Kong<sup>1</sup>, Ye Sun<sup>2</sup>, Ying Han<sup>1</sup>, Qiqi Zhao<sup>1</sup>, Shuyu Kang<sup>1</sup>, Xijun Wang<sup>1,2,3,\*</sup>

## Abstract

The discovery of effective constituents of traditional Chinese medicine (TCM) is an important approach in new drug development. Several well-known drugs, such as artemisinin, berberine, and ephedrine have been developed using this approach. However, the efficacy and safety of TCM, two key issues for drug development based on TCM clinical experience, remain unclear worldwide. The discovery strategy of relevant constituents is the most important step for determining efficacy and safety, which still a key scientific problem that restricts the development of new drugs. Furthermore, TCM formulas used as clinical drugs address a specific TCM syndrome (*Zheng*), and the complexity of the formula and vagueness of the syndrome make the identification of the effective constituents related to clinical effectiveness challenging. Over decades, researchers have developed transdisciplinary technologies and research methodologies to identify effective constituents *in vivo*. In this paper, the history of strategy development for identifying the effective constituents related to the clinical efficacy of TCM is reviewed and summarized. The main approaches include the phytochemical method, which involves the classical systematic separation and screening (extraction, separation, purification, structure identification, and activity test); bioactivity-guided separation; serum pharmacology of TCM *in vivo*; and Chinmedomics, which connects *in vivo* constituents with the biomarkers of the relevant TCM syndrome. Chinmedomics is a promising strategy that conforms to the theory and characteristics of TCM. By clarifying the effective constituents, targets and pathways of medicines, it can promote the discovery of lead compounds and the research of innovative drugs, and continuously promote the modernization of TCM.

**Keywords:** Chinmedomics, Effective constituents of TCM, New drug discovery strategy, Serum pharmacology of TCM, Traditional Chinese medicine

**Graphical abstract:** <http://links.lww.com/AHM/A64>.

## Introduction

Traditional Chinese medicine (TCM) has played an important role in the medical and health system in China for thousands of years, and the discovery of effective TCM

constituents is a critical step in drug innovation<sup>[1]</sup>. For example, artemisinin, an antimalarial drug originating from *Artemisia Annua* herba, has saved 200 million people worldwide in recent years<sup>[2]</sup>; berberine, an isoquinoline alkaloid isolated from the Chinese herb *Coptidis rhizoma* and other *Berberis* plants, has been demonstrated to be a promising multifunctional drug that can be used to treat digestive, metabolic, cardiovascular, and neurological diseases<sup>[3-4]</sup>; paclitaxel (Taxol) is a natural antineoplastic drug that is widely used in treating breast, ovarian, and lung cancers<sup>[5]</sup>. Moreover, in the prevention and control of serious infectious diseases, such as COVID-19, TCM has exhibited an extensive advantage<sup>[6]</sup>. New formulas such as Qingfei Paidu granules, Huashi Baidu granules, and Xuanfei Baidu granules have emerged<sup>[7]</sup>, and a novel effective compound wogonoside in Qingfei Paidu granule has been demonstrated to exert anti-inflammatory effects through the downregulation of USP14 to promote the degradation of activating transcription factor 2<sup>[8]</sup>. In addition, TCM has unique advantages in treating cardiovascular, cerebrovascular<sup>[9]</sup>, and complex metabolic diseases<sup>[10]</sup>. Tanshinone IIA, the main fat-soluble component of *Salviae miltiorrhizae* Radix et Rhizoma, has shown remarkable cardioprotective effects through enhancing angiogenesis, which may serve as an effective treatment against cardiovascular diseases; it affects multiple signaling pathways including antiproliferative, antiapoptotic, anti-inflammatory, and antioxidative stress pathways<sup>[11]</sup>. The efficacy and treatment concept of TCM has attracted

<sup>1</sup> State Key Laboratory of Integration and Innovation of Classical Formula and Modern Chinese medicines, National Chinmedomics Research Center, National TCM Key Laboratory of Serum Pharmacology, Heilongjiang University of Chinese Medicine, Harbin 150040, China; <sup>2</sup> State Key Laboratory of Dampness Syndrome, The Second Affiliated Hospital Guangzhou University of Chinese Medicine, Guangzhou, 510120, China; <sup>3</sup> Haihe Laboratory of Modern Chinese Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin 301617, China

\* Corresponding author. Xijun Wang, State Key Laboratory of Integration and Innovation of Classical Formula and Modern Chinese medicines, National Chinmedomics Research Center, National TCM Key Laboratory of Serum Pharmacology, Heilongjiang University of Chinese Medicine, Harbin, 150040, China, E-mail: xijunw@sina.com; Hui Sun, Department of Pharmaceutical Analysis, Heilongjiang University of Chinese Medicine, Harbin 150040, China, E-mail: Sunhui7045@163.com.

Copyright © 2023 Tianjin University of Traditional Chinese Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Acupuncture and Herbal Medicine (2023) 3:3

Received 14 March 2023 / Accepted 26 June 2023

<http://dx.doi.org/10.1097/HM9.000000000000071>

increasing attention worldwide. The growing international demand for TCM has also provided a broad scope for the development of TCM and drug discovery.

However, the theoretical system, diagnosis, and treatment methods of TCM are notably different from those of Western medicine. From the perspective of TCM, the disease is mostly described as a syndrome (*Zheng*), and treatment is based on a combination of herbal medicines called formula; the correspondence between *Zheng* and formula is the premise of efficacy<sup>[12]</sup>. Different syndromes require different formulas to reverse imbalance and restore homeostasis<sup>[13]</sup>. Moreover, the complexity of the formula and the vagueness of the syndrome make the identification of the effective constituents and mechanisms related to clinical efficacy challenging. Discovering the active constituents of TCMs, evaluating their efficacy, and elucidating the targets and pathway are crucial steps for interpreting the principles of TCM, revealing the internal law of formula compatibility, and designing new formulas for treating current chronic diseases.

Over decades, researchers have developed strategies for identifying effective constituents. With the progress of modern analytical technology and systems biology, the methods for identifying the effective constituents of TCM are based on systematicity and integrity. From ephedrine, an active drug for relaxing tracheal smooth muscles<sup>[14]</sup>, to artemisinin, a new antimalarial drug that is safe and effective and can resolve drug resistance<sup>[15]</sup>, the method had advanced from the phytochemical approach to bioactivity-guided separation. More recently, the emergence of TCM serum pharmacology established a foundation for rapid screening of *in vivo* effective constituents of TCMs<sup>[16]</sup>. The development of metabolomics, the most phenotypic omics research, has enabled characterization of metabolic profiles and biomarkers of TCM syndromes and created the research model of correlation analysis between active formula constituents *in vivo* and endogenous biomarkers of TCM syndromes. Therefore, the research strategy of Chinmedomics was born<sup>[17]</sup>. The present review based on CNKI and PubMed, summarized different versions of effective constituent discovery methods of TCM by time and technological advancement, which helps clarify the effective substances of TCM, promotes the discovery of lead compounds and innovative drugs, and assists the modernization of TCM.

### Version updates of strategies for the effective constituent discovery of TCM

*Phytochemical method based on extraction, separation, purification, structure identification, and activity testing: version 1.0*

Initially, the discovery of effective constituents was mainly based on the phytochemical method. Extraction is the first step to identify effective constituents of herbal medicines by decoction, refluxion, and organic solvent ultrasound. The extracts had to be separated and purified using columns according to the properties (solubility, pH, adsorption, molecular weight) to obtain pure compounds, followed by identification of the chemical structure using infrared spectrum, ultraviolet-visible absorption spectrum, nuclear magnetic resonance, and mass spectrometry. Subsequently, biological methods were used to test the activity of the single compounds to ultimately determine the active

constituents<sup>[18]</sup>. Common separation technologies to purify compounds include low-medium-pressure column<sup>[19]</sup>, thin-layer<sup>[20]</sup>, and preparative chromatography<sup>[21]</sup>. The specific workflow is shown in Figure 1. This method is characterized by strong maneuverability, which can separate and prepare a large number of compounds and has been the common widely used method in the early stage of searching for effective constituents of TCM, achieving remarkable results.

The most representative and earliest example of the application of this strategy is the discovery of ephedrine in TCM *ephedra herba* (*Ephedra sinica* Stapf, *Ephedra intermedia* Schrenk et C.A. Mey., *Ephedra equisetina* Bge.). Nagai isolated an alkaloid from this *Ephedra sinica* in 1887, named it ephedrine, and discovered that it has a mydriatic effect; Amatsu and Kubota described a rise in blood pressure and relaxation of the intestines after ephedrine in 1918<sup>[22]</sup>. The Chinese pharmacologist Chen K.K. conducted a systematic study on the pharmacological effects of ephedrine in 1927, and found that ephedrine is clinically stable, so the age, exposure to light, or boiling does not alter its action; ephedrine has a persistent and uniform action, in contrast with that of adrenaline; ephedrine has low toxicity and an exceptionally wide margin of safety, which can be conveniently administered, either orally or through intramuscular injection without local irritation<sup>[23]</sup>.

Camptothecin, an alkaloid, was isolated from the tree *Camptotheca acuminata* Nyssaceae in 1966. The stem wood was extracted using the standard method of continuous, hot hexane–heptane extraction from dried plant material followed by similar extraction with 95% ethanol. The ethanol extract concentrate is partitioned between water and chloroform. Silica gel chromatography of the methanol-insoluble material from the chloroform extract followed by crystallization from methanol–acetonitrile showed camptothecin as pale yellow needles. Camptothecin (0.25–1.0 mg/kg/day) tested in L1210 leukemic mice prolonged survival by up to 100%; in Walker 256 (intramuscular) rat tumor, concentrations as low as 1.25 mg/kg significantly inhibited growth; moreover, it showed moderate cytotoxicity against KB cell culture ( $ED_{50} = 0.07 \mu\text{g/mL}$ )<sup>[24]</sup>.

Schizandrin provides another example; its plane structure was elucidated by Kochetkov et al. in 1961; subsequently, gomisins A, B, C, F, and G in *Schisandra chinensis* Fructus (*Schisandra chinensis* (Turcz.) Baill.) were gradually isolated<sup>[25]</sup>. Assessment of sleep time, activity, and muscle state of mice revealed that both schizandrin and gomisins A had sedating or tranquilizing effects. The efficacy of gomisins A was durable and intense and that of schizandrin was transient. However, the analgesic effect of gomisins A was inferior to that of schizandrin<sup>[26]</sup>. Other representative studies are shown in Table 1<sup>[27–46]</sup>.

In addition to the above-mentioned achievements, this method also has some disadvantages: cumbersome operation, long experimental cycle, lower clarity, and greater consumption. Moreover, the separated compounds usually have no or weak activity. In addition, some active ingredients with low content may be lost in the process of separation and purification. Owing to the yield limitation of the separated compounds and activity testing methods, several ingredients barely stay at the level of structural identification. In contrast, most screening

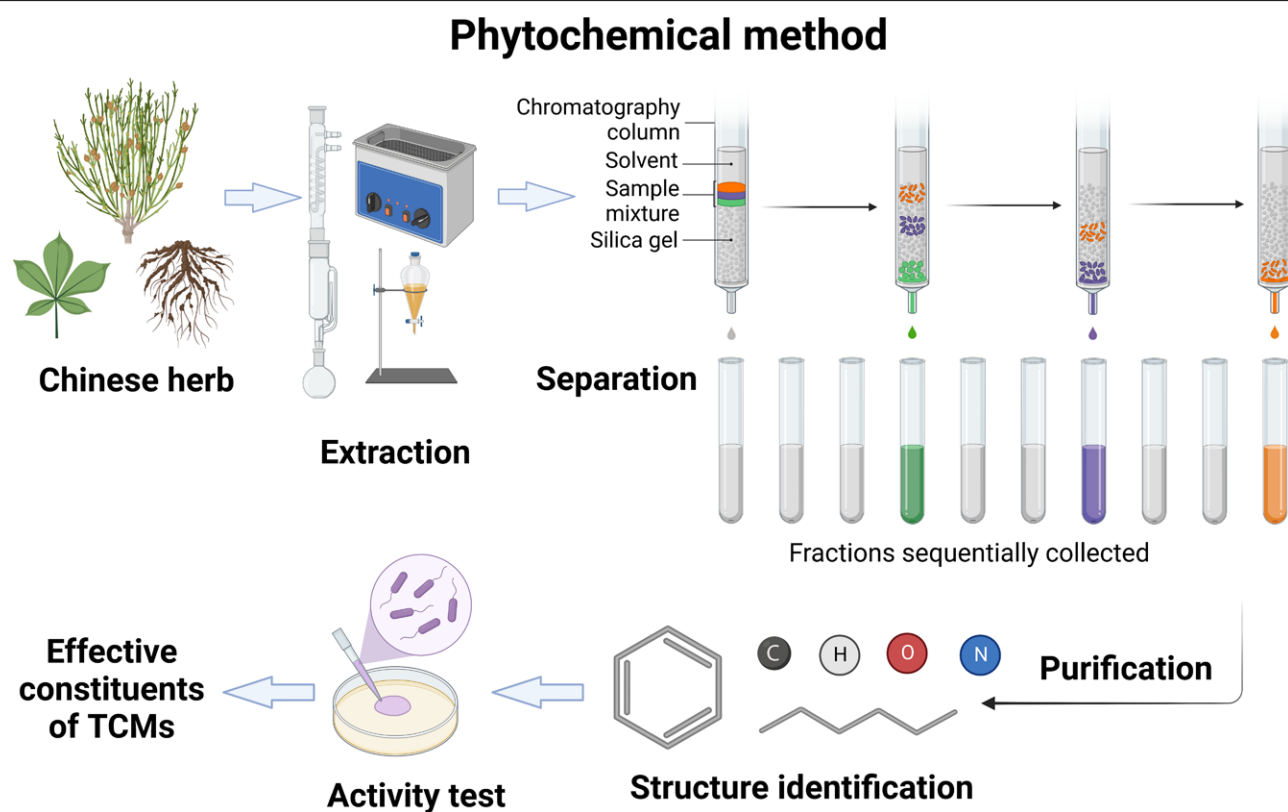
methods using the systematic separation focus on novel chemical structures and the discovery of new compounds. Furthermore, the activity evaluation model has a low correlation with traditional efficacy; therefore, the research results provide limited assistance with elucidating effective constituents of TCMs.

#### Bioactivity-guided separation: version 2.0

Bioactivity-guided separation is a pharmacological activity-oriented screening method for the effective constituent of TCMs. The main method started with total extraction under the guidance of sensitive and reliable activity evaluation and tested the activity of the fractions/constituents separated at each step, tracked and separated the active constituents, and finally, obtained the effective constituents<sup>[47]</sup>. The process can be simply summarized into two steps. The first step involves rough separation; TCMs are successively further extracted using different polar solvents to obtain corresponding extraction sites or macroporous adsorption resin, ion exchange resin, and glucan gel to separate the components. All obtained components are detected using an activity test, and the crudely separated components, which show biological activity, are called effective components. The second step involves fine separation; the effective components are loaded into a separation column and extracted with an eluent, and the monomer compound is finally obtained using repeated column chromatography separation. Subsequently, the constituents obtained from each separation are tested for activity. The structures of the single compounds obtained through tracking and

separation are determined based on their physical and chemical properties and spectral data. The workflow is summarized in Figure 2. Bioactivity-guided separation with simultaneous chemical separation and activity evaluation greatly reduced the blindness of the analysis and saved unnecessary separation and preparation of the inactive fraction. Based on this method, numerous active constituents have been discovered, with clear structures, significant pharmacological effects, and relatively clear mechanisms. Some of them have been developed into clinical medicine and a new drug development model using natural products was gradually created.

*Artemisia annua* L. (Qinghao) was first indicated to reduce fevers relevant to the symptoms of malaria in Handbook of Prescriptions for Emergency Treatments (Zhǒu Hòu Bèi Jí Fāng ,肘后备急方) written by Ge Hong in 340 AD. The antimalarial activity of the extract obtained *via* the conventional heating extraction method is not optimal. The investigators proposed that active constituents of *Artemisia annua* L. may be destroyed during the extraction process under heating. A low-temperature extraction method for the plant was then proposed, inspired by Ge Hong's handbook, instructing to "immerse a handful of Qinghao in 2L water, squeeze out the juice, and drink it all." Ether extracts produced encouraging results in mice infected with *Plasmodium berghei* and finally, a non-toxic neutral extract was obtained on October 4, 1971, by immersing or distilling Qinghao using ethyl ether<sup>[2]</sup>, with 100% efficacy in mice infected with *P. berghei* and in monkeys infected with *Plasmodium cynomolgi*. The extraction was further separated by antimalarial



**Figure 1.** Workflow of the classical phytochemical method for screening effective constituents of TCMs. TCM: Traditional Chinese medicine. Created by Biorender.com.

**Table 1.****Representative studies on the effective constituents of TCM using classical systematic separation and screening methods**

Herb name	Monomer compound	Effective constituent	Activity evaluation	References
Ephedrae Herba (Mahuang)	Ephedrine	Ephedrine	Animal <i>in vivo</i> experiments (dogs, rabbits, rats, etc)	[23]
Camptotheca acuminata (Xishu)	Camptothecin	Camptothecin	Leukemia L1210 in mice, Walker 256 (intramuscular) tumor (rats)	[24]
Schisandra chinensis (Wuweizi)	Schizandrin, gomisin A, gomisin B, gomisin C, gomisin F, and gomisin G	Schizandrin, gomisin A	Animal <i>in vivo</i> experiments (mice, pigs, and rats)	[25–26]
Berberis vulgaris (Sankezhen)	Berberine	Berberine	Berberine hydrochloride (0.5%) inhibited the growth of <i>Bacterium coli</i> , inhibited the formation of indol and volatile fatty acids in protein decomposition, and acted as a strong poison to rabbits, frogs, hens, and mice, causing leucocytosis and nephritis and affecting the nervous system	[27]
Rehmannia glutinosa (Dihuang)	Catalpol, ajugol, aucubin, melittoside, rehmannioside A, rehmannioside B, rehmannioside C, and rehmannioside D	Rehmannioside D	Rehmannioside D showed a weak hypoglycemic action in spontaneously diabetic mice	[28]
Chelidonium majus (Baiqucai)	Chelidocystatin	Chelidocystatin	Chelidocystatin exhibited a strong inhibition of cathepsin L, papain, and cathepsin H	[29]
Poria cocos (Fuling)	3 $\beta$ -p-hydroxybenzoyldehydrotumulosic acid	3 $\beta$ -p-hydroxybenzoyldehydrotumulosic acid	TPA- and AA-induced ear inflammation in mice	[30]
Gentiana tibetica (Xizangqinjiao)	8-Hydroxy-10-hydrosveroside, isomacrophylloside and ethyl N-docosanoylanthranilate, $\beta$ -sitosterol, daucosterol, oleanolic acid, loganic acid, gentiopicroside, sweroside, 2'-(2,3-dihydroxybenzoyl) sweroside, trifloroside, rindoside, and macrophyllouside A	Ethyl N-docosanoylanthranilate	Ethyl N-docosanoylanthranilate inhibited the growth of the human pathogenic fungi <i>Candida albicans</i> and <i>Aspergillus flavus</i>	[31]
Ganoderma lucidum (Lingzhi)	Ganoderic acid alpha, ganoderic acids A, ganoderic acids B, ganoderic acids C1, ganoderic acids H, ganoderiols A, ganoderiols B, ganoderiols F, ganodermanontriol, ergosterol, ergosterol peroxide, cerevisterol, 3 $\beta$ -5 alpha-dihydroxy-6 $\beta$ -methoxyergosta-7,22-diene	Ganoderiol F, ganodermanontriol, ganoderic acid B, ganoderiol B, ganoderic acid C1, 3 $\beta$ -5 alpha-dihydroxy-6 $\beta$ -methoxyergosta-7,22-diene, ganoderic acid alpha, ganoderic acid H, and ganoderiol A	Active as anti-HIV-1 agents, moderately active inhibitors against HIV-1 PR	[32]
Plumbago zeylanica (Baihuadan)	3'-O- $\beta$ -glucopyranosyl plumbagic acid, 3'-O- $\beta$ -glucopyranosyl plumbagic acid methylester, plumbagin, chitranone, maritinone, elliptinone and isoshinanolone, seselin, 5-methoxyseselin, suberosin, xanthyletin, and xanthoxyletin	Plumbagin	Cytotoxicity of all compounds against various tumor cells lines was evaluated, and plumbagin was found to significantly suppress growth of Raji, Calu-1, HeLa, and Wish tumor cell lines	[33]
Scutellaria barbata (Banzhilian)	Barbatin A, barbatin B, barbatin C, scutebarbatine B	Barbatins A, Barbatins B, Barbatins C, scutebarbatine B	Human cancer lines, namely, HONE-1 nasopharyngeal, KB oral epidermoid carcinoma, and HT29 colorectal carcinoma cells	[34]
Salvia miltiorrhiza (Danshen)	Salvianonol, salviamone, 2a-acetoxysugiol, palmitoyl arucadiol, (E)-4-[5-(hydroxymethyl) furan-2-yl] but-3-en-2-one, along with 13 known compounds	Palmitoyl arucadiol	Cancer cell lines (HeLa and OVCAR-3 cells)	[35]
Gentiana rigescens (Dianlongdancao)	Gentirigenic acid, five glycosides, gentirigeosides A–E	Gentirigeosides A, Gentirigeosides C, Gentirigeosides E	Antifungal activity against the plant pathogen <i>Glomerella cingulata</i>	[36]
Datura metel (Yangjinhua)	Withametelins I-P, 1,10-seco-withametelin B, and 12 $\beta$ -hydroxy-1,10-seco-withametelin B, together with seven known withanolides	Withametelin I, withametelin K, withametelin L, withametelin N	Exhibited cytotoxic activities against A549 (lung), BGC-823 (gastric), and K562 (leukemia) cancer cell lines	[37]

(Continued)

**Table 1.**  
(Continued)

Herb name	Monomer compound	Effective constituent	Activity evaluation	References
Sinomenium acutum (Qingfengteng)	2,2'-disinomenine, 7',8'-dihydro-1,1'-disinomenine, 1,1'-disinomenine	2,2'-disinomenine, 1,1'-disinomenine	Weak inhibition against A549 and HeLa cells	[38]
Uncaria rhynchophylla (Gouteng)	18,19-dehydrocorynoxinic acid, B, 18,19-dehydrocorynoxinic acid, corynoxine, isocorynoxine, rhynchophylline, isorhynchophylline, vincoside lactam	Corynoxine, isocorynoxine, rhynchophylline, isorhynchophylline, vincoside lactam	Exhibited inhibitory activity on LPS-induced NO release in primary cultured rat cortical microglia (IC <sub>50</sub> : 13.7–19.0 μM)	[39]
Dioscorea septemloba (Mianbixie)	Cholestane glycosides, dioseptemlosides A and B, together with six spirostane glycosides, dioseptemlosides C–H	None	These compounds did not show considerable inhibitory antitumor activities at a concentration of 10 mM	[40]
Euphorbia helioscopia (Zeqi)	Four new jatrophone-type diterpenoids, together with 16 known related compounds: euphornin, euphornin A, euphornin B, euphornin G, euphoheliosnoid A, euphoheliosnoid B, euphoheliosnoid C, euphoscopin A, euphoscopin B, euphoscopin C, euphoscopin E, euphoscopin J, helioscopinolide A, dehydrovomifoliol, fusic acid, loliolide	Helioscopinolide A and euphornin	Cytotoxicity against HeLa and MDA-MB-231 cells was evaluated, with only helioscopinolide A and euphornin showing activity	[41]
Schisandra propinqua (Wuweizi)	14 new lignans, tiegusanins A–N, together with 13 known compounds	Tiegusanin G	Anti-HIV-1 activity with an EC <sub>50</sub> value of 7.9 mM and a TI of more than 25	[42]
Gentiana loureirii (Huananlongdan)	Iridoid glycosides, 2',3',6'-tri-O-acetyl-4'-O-trans-p-(O-β-d-glucopyranosyl) coumaroyl-7-ketologanin, 2'-O-caffeoylloganic acid, 2'-O-p-hydroxybenzoylloganic acid, 2'-O-trans-p-coumaroylloganic acid, and 2'-O-cis-p-coumaroylloganic acid, along with six known iridoids, 7-ketologanin, loganin, loganic acid, sweroside, boonein, isoboonein, and three other known compounds	None	The isolated iridoids were evaluated for antibacterial and antioxidant activities but were either inactive or very weakly active	[43]
Capparis spinosa (Cishangan)	Capparisine A, capparisine B, capparisine C, 2-(5-hydroxymethyl-2-formylpyrrol-1-yl) propionic acid lactone, and N-(3'-maleimidyl)-5-hydroxymethyl-2-pyrrole formaldehyde	None	The five compounds had no inhibitory effect on the human hepatocyte cell line HL-7702 apoptosis induced by Act D and TNF-α	[44]
Valeriana jatamansi (Zhizhuxiang)	Jatamanins A–M, (+)-9'-isovaleroxylariciresinol, together with seven known iridoids and 13 lignans	(+)-9'-isovaleroxylariciresinol	Showed significant <i>in vitro</i> cytotoxicity against PC-3M and HCT-8 cell lines	[45]
Asparagus filicinus (Yangchitianmendong)	Filiasparoside E, filiasparoside F, filiasparoside G, stachysterone A-20, 22-acetonide, asparagasin, filiasparoside A, filiasparoside B, aspaflioside A, aspaflioside B, and filiasparoside C	Aspaflioside A, aspaflioside B, filiasparoside C	Human breast adenocarcinoma MDA-MB-231 cell line	[46]

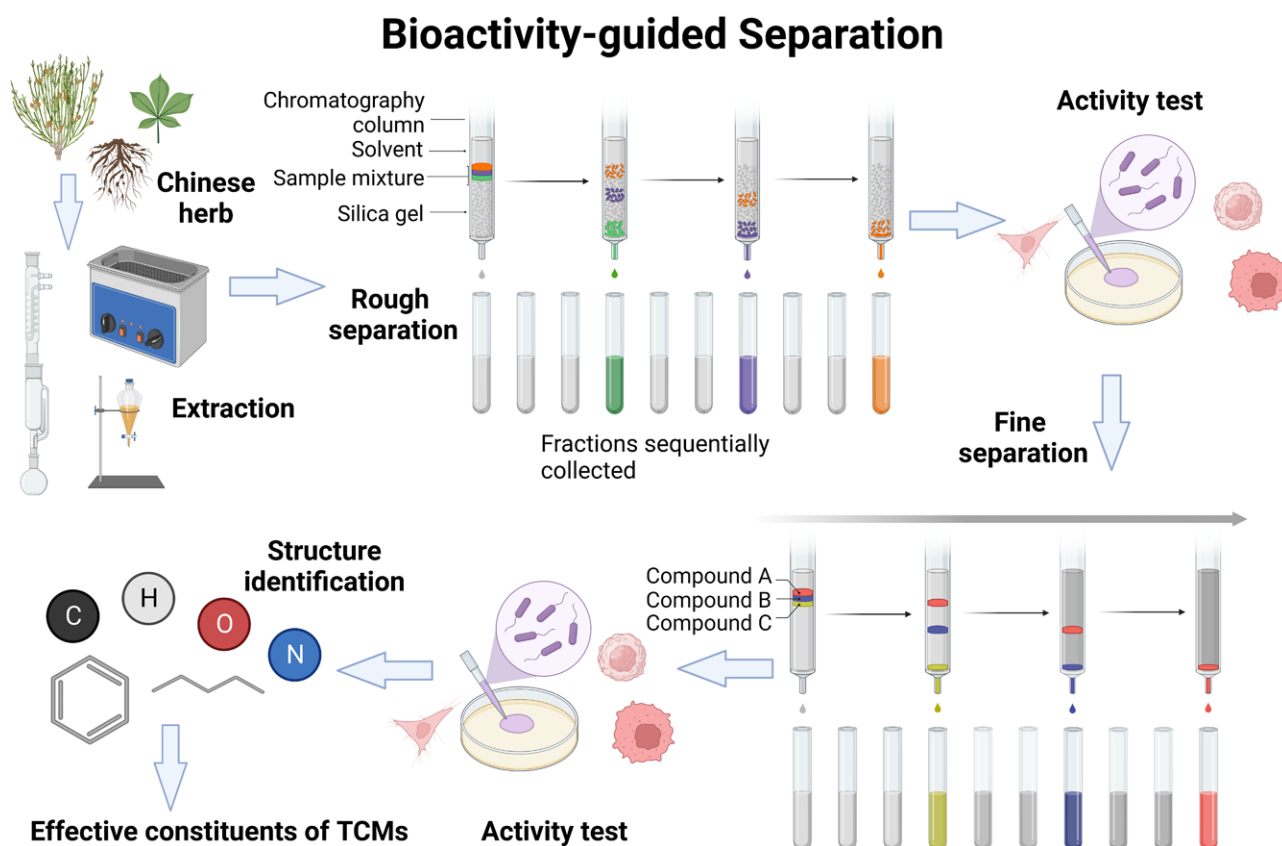
AA: Arachidonic acid; LPS: Lipopolysaccharide; PR: Protease receptor; TCM: Traditional Chinese medicine; TI: Therapeutic index; TNF: Tumor necrosis factor; TPA: 12-O-tetradecanoylphorbol-13-acetate.

activity; a sesquiterpene active constituent was identified as artemisinin and reported in 1979<sup>[48]</sup>.

Taxol, isolated from the stem bark of the western yew, *Taxus breuifolia*, has potent antileukemic and antitumor properties; it is the first compound possessing a taxane ring that has been demonstrated to have such activities. The alcohol extract of the stem bark was concentrated and partitioned between water and chloroform. Guided by assay in 9 KB and various leukemia systems, three successive chromatographies of the residue from the chloroform extract on Sephadex LH-20, and silica

gel followed by crystallization from aqueous methanol showed taxol as needles<sup>[15]</sup>. Further representative studies are summarized in Table 2<sup>[49–84]</sup>.

In general, the majority of single compounds isolated using bioactivity-guided separation have adequate activities, and the hit rate of the effective constituents found is considerably higher than that of version 1.0 (phytochemical method). However, this method has certain limitations; extensive separation and preparation is still required, which is time-consuming and labor-intensive. In addition, TCM have multi-component and multi-target



**Figure 2.** Workflow of the bioactivity-guided separation method for screening effective constituent of TCMs. TCM: Traditional Chinese medicine. Created by Biorender.com.

characteristics; therefore, the active ingredients screened using a single or a few indicators cannot fully reflect all effective constituents of TCM. Importantly, this method is still in the *in vitro* stage, which cannot reflect the bio-transformation of various constituents in the body, nor can it demonstrate the interaction among drugs during the process of absorption, distribution, metabolism, and elimination *in vivo*. Therefore, bioactivity-guided separation is a basic method for initial screening of active TCM constituents.

### Serum pharmacology of TCM: version 3.0

In the 1980s, Japanese scholar Masakazu Tashiro first proposed the concepts of “serum pharmacology” and “serum pharmacology.” Some constituents are selectively absorbed after oral administration of crude drugs, and those compounds reach their target by circulation and play an effective role<sup>[85]</sup>. Based on serum pharmacology, Professor Xijun Wang performed a large number of studies on TCM formulas, such as Yin Chen Hao Tang (YCHD)<sup>[86]</sup>, Yinchen Wuling powder<sup>[87]</sup>, Liuwei Dihuang pill<sup>[88–89]</sup>, among others. By integrating serum pharmacology with TCM theory and clinical practice, he proposed and established the theory and method of TCM serum pharmacology<sup>[16]</sup>. This method involves TCM sample preparation and quality control, sample administration, blood collection and serum preparation, analysis, and detection. In general, the method uses modern separation technology to analyze and identify the constituents in human/animal blood after oral administration of TCM formulas to identify

the compounds in blood that originate from the formula. This approach reflects the synergistic effect of the combination of the formula drug and its ingredients and the interaction between the herbal drugs, intestinal flora, and human body, ultimately identifying the directly effective constituents *in vivo*<sup>[90]</sup>. The specific steps are shown in Figure 3. Serum pharmacology of TCMs can effectively avoid the false positives or false negatives caused by the blind separation of chemical constituents *in vitro*. Based on this method, numerous herbal medicines and TCM formulas have been analyzed. Studies on YCHD and Ciwujia (*Acanthopanax senticosus* (Rupr. et Maxim.) Harms, AS) are used as typical examples to provide details of the method.

YCHD, recorded in “Treatise on Cold Damage Diseases (Shāng Hǎn Lùn, 伤寒论)” written by Zhang Zhongjing during the Eastern Han Dynasty (25–220 AD), is the representative classical formula for treating the jaundice syndrome (Yang Huang syndrome, YHS). The formula comprises three Chinese herbs: *Artemisia scopariae* Herba (*Artemisia scoparia* Waldst. et Kit. / *Artemisia capillaris* Thunb.), *Gardeniae Fructus* (*Gardenia jasminoides* Ellis), and *Rhei Radix et Rhizoma* (*Rheum palmatum* L. / *Rheum tanguticum* Maxim. ex Bal. / *Rheum officinale* Bail.). Under strict quality control, a total of 45 compounds were detected and 21 compounds were identified in blood, including picrocrocin acid, chlorogenic acid, 6-methoxy coumarin-7-hydroxyl sulfate, 7-methoxy coumarin-6-hydroxyl sulfate, geniposide, geniposide, saffloryellow A, capillaridin A, isofraxidin, quercetin-3-O-glycoside, 6,7-dimethoxy coumarin, 2,5-dimethyl-7-hydroxy chromone, chimaphyllin,

**Table 2.****Representative studies on the effective constituents of TCM using bioactivity-guided separation**

Herb name	Monomer compound	Effective constituent	Activity evaluation model	References
Artemisia annua (Qinghao)	Artemisinin	Artemisinin	Antimalarial activity in a mouse model of infection	[48]
Taxus breiofolia (Duanyehongdoushan)	Taxol	Taxol	Various leukemia systems	[15]
Gentiana olivieri (Xiewanqueqinjiao)	Isorientin	Isorientin	Glucose-hyperglycemic and streptozotocin-induced diabetic rats	[49]
Kaempferia galanga (Shannai)	Ethyl cinnamate, ethyl p-methoxycinnamic acid	Ethyl cinnamate	Anesthetized rats (determination of mean arterial pressure)	[50]
Spatholobus suberectus (Jixueteng)	3',4',7-trihydroxyflavone, eriodictyol, plathymenin, dihydroquercetin, butin, neoisoliquiritigenin, dihydrokaempferol, liquiritigenin, 6-methoxyeriodictyol	Butin	HEMn	[51]
Senecio scandens (Qianliguang)	Jacaranone glycoside 1, jacaranone glycoside 2, jacaranone glycoside 3, jacaranone glycoside 4	Jacaranone glycoside 1, jacaranone glycoside 2, jacaranone glycoside 3, jacaranone glycoside 4	Moderate cytotoxicities against five tumor cell lines	[52]
Dipsacus asper (Chuanxuduan)	Caffeic acid, 2,6-dihydroxycinnamic acid, vanillic acid, 2'-O-caffeoyl-D-glucopyranoside ester, caffeoylquinic acid, loganin, cantleyoside, triptostoid A, lisianthioside, 6'-O-β-D-apiofuranosyl sweroside, as well as triterpenoids oleanic acid and akebiasaponin D, dipsanosides C-G, 3'-O-β-D-glucopyranosyl sweroside	Caffeic acid, 2,6-dihydroxycinnamic acid, vanillic acid, 2'-O-caffeoyl-D-glucopyranoside ester, and caffeoylquinic acid	Five human tumor cell lines including lung carcinoma A549, hepatoma Bel7402, gastric carcinoma BGC-823, colon cancer HCT-8, and ovary cancer A278052	[53]
Tripterygium wilfordii (Leigongteng)	Tripterygiol, (-)-syringaresinol, triptolide, triptidiolide, triptriolide, euonine, wilforine, cangorinine E-I, and 3,4,5-trimethoxyphenol	Tripterygiol, (-)-syringaresinol, triptolide, triptidiolide, triptriolide, euonine, wilforine, cangorinine E-I, and 3,4,5-trimethoxyphenol	Mouse monocyte/macrophage cell line RAW 264.7	[54]
Valeriana sorbifolia (Xiecao)	Sorbifolivaltrates A-D, isovaltrate, valtrate, seneciovaltrate, valtrate hydrine B3, and valtrate hydrine B7	Sorbifolivaltrates A-D, isovaltrate, valtrate, seneciovaltrate, valtrate hydrine B3, and valtrate hydrine B7	All compounds exhibited weak to moderate cytotoxicity against the human metastatic prostate cancer cell line, PC-3M	[55]
Rabdosia coetsa (Xiangchacai)	Yielded ethyl caffeate, rosmarinic acid, and methyl rosmarinic acid	Yielded ethyl caffeate, rosmarinic acid, and methyl rosmarinic acid	Inhibited ACE activity	[56]
Scrophularia ningpoensis (Xuanshen)	6''-O-caffeoylharpagide, 6''-O-feruloylharpagide, 6''-O-β-glucopyranosylharpagide, 2-(3-hydroxy-4-methoxyphenyl) ethyl O-alpha-arabinopyranosyl-(1→6)-O-alpha-rhamnopyranosyl-(1→3)-O-β-glucopyranoside, phenyl O-β-xylopyranosyl-(1→6)-O-β-glucopyranoside, 3-methylphenyl O-β-xylopyranosyl-(1→6)-O-β-glucopyranoside, 6-O-cinnamoyl β-fructofuranosyl-(2→1)-O-alpha-glucopyranosyl-(6→1)-O-alpha-glucopyranoside, 6-O-feruloyl β-fructofuranosyl-(2→1)-O-alpha-glucopyranosyl-(6→1)-O-alpha-glucopyranoside, 6''-O-alpha-D-galactopyranosyl harpagoside, 6''-O-(p-coumaroyl) harpagide, harpagoside, angoroside C	6''-O-caffeoylharpagide, phenyl O-β-xylopyranosyl-(1→6)-O-β-glucopyranoside, 3-methylphenyl O-β-xylopyranosyl-(1→6)-O-β-glucopyranoside, 6''-O-(p-coumaroyl) harpagide, and harpagoside	Rat cardiac myocytes	[57]

(Continued)

**Table 2.**  
(Continued)

Herb name	Monomer compound	Effective constituent	Activity evaluation model	References
Saussurea lappa (Yunmuxiang)	Dehydrocostus lactone, 11 $\beta$ ,13-dihydrodehydrocostus lactone, costunolide, 11 $\beta$ ,13-dihydrocostunolide, 11 $\beta$ ,13-dihydrosantamarin, 11 $\beta$ ,13-dihydro-reynosin, a-cyclocostunolide, 11 $\beta$ ,13-dihydro-acyclocostunolide, 1 $\beta$ -hydroxycolartin, ilicol, 11 $\beta$ ,13-dihydro-b-cyclocostunolide, alantolactone, isoalantolactone	Dehydrocostus lactone, costunolide, $\alpha$ -cyclocostunolide	Mouse macrophage RAW 264.7 cells	[58]
Angelica sinensis (Danggui)	Z-ligustilide, 11-angeloylsenkyunolide F, coniferyl ferulate, and ferulic acid	Z-ligustilide, 11-angeloylsenkyunolide F, coniferyl ferulate, and ferulic acid	Aggregated amyloid $\beta$ -peptide (agg A $\beta$ 1-40) induced damage of differentiated PC-12 cells	[59]
Mori Cortex Radicis (Sangbaipi) / Morus Bombycis (Sangye)	Moracin O, moracin P, moracin Q, moracin M, mulberrofuran H, mulberrofuran G, sanggenon O, sanggenon C, albafuran A, mulberrofuran D, mulberrofuran W, kuwanon J, kuwanon Q, kuwanon R, kuwanon V	Moracin O, moracin P, moracin Q, mulberrofuran H, sanggenons O, albafuran A, mulberrofurans D, kuwanons J	Human hepatocellular carcinoma cell-line Hep3B cells	[60]
Polygonum cuspidatum (Huzhang)	Citreorosein, together with emodin and its glucoside	Citreorosein	Recombinant yeast was used to assess the estrogenic activity	[61]
Garcinia oblongifolia (Lingnanshanzhuzi)	Oblongixanthenes A-C, oblongifolins E-G, and 12 known compounds	Oblongifolin C	All isolates were assayed for their apoptosis-inducing effects against HeLa-C3 cells	[62]
Paeonia lactiflora (Chishao)	1,2,3,4,6-penta-O-galloyl-D-glucopyranose	1,2,3,4,6-penta-O-galloyl-D-glucopyranose	<i>In vitro</i> enzyme assay with human recombinant PTP1B	[63]
Actaea racemosa (Shengma)	Actein, 26-deoxyactein, 23-epi-26-deoxyactein, 23-O-acetylshengmanol 3-O- $\beta$ -D-xylopyranoside, cimircemoside F, cimigenol 3-O- $\beta$ -D-xylopyranoside, cimircemoside D, 25-O-acetylcimigenol 3-O- $\alpha$ -L-arabinopyranoside, 25-O-anhydrocimigenol 3-O- $\alpha$ -L-arabinopyranoside, 25-O-anhydrocimigenol 3-O- $\beta$ -D-xylopyranoside, cimircemoside K	23-O-acetylshengmanol 3-O- $\beta$ -D-xylopyranoside, actein, cimigenol 3-O- $\beta$ -D-xylopyranoside, and 25-O-acetylcimigenol 3-O- $\alpha$ -L-arabinopyranoside	GABA(A) receptor-modulating constituents in <i>Xenopus laevis</i> oocytes	[64]
Euphorbia kansui (Gansui)	Kansuinine B, kansuinine A, kansuiphorin C, 3-O-benzoyl-20-deoxyingenol, and 3-O-(2'E,4'Z-decadienyl)-20-O-acetylingenol	Kansuinine B, kansuinine A, 3-O-(2'E,4'Z-decadienyl)-20-O-acetylingenol	Models of inflammation using exoteric mice SPL and rat peritoneal macrophages (PM $\phi$ )	[65]
Glycyrrhiza uralensis (Gancao)	Glycyrrhisoflavone	Glycyrrhisoflavone	Alpha-glucosidase inhibitory activity	[66]
Tinospora sagittata (Qingniudan)	Palmatine, columbamine, and columbinyl glucoside	Palmatine, columbamine, and columbinyl glucoside	RAW264.7 macrophage cells	[67]
Salvia miltiorrhiza (Danshen)	Royleanone, horminone, 7-O-methyl-horminone and 7-acetyl-horminone, erythrodiol-3-acetate, $\beta$ -sitosterol	Horminone, 7-acetyl-horminone and erythrodiol-3-acetate, 7-O-methyl-horminone	Human cervix adenocarcinoma (HeLa), skin carcinoma (A431), and breast adenocarcinoma (MCF7) cells using the MTT assay	[68]
Acanthopanax senticosus (Ciwujia)	Eleutheroside E, eleutheroside E (2)	Eleutheroside E, eleutheroside E (2)	Sleep-deprived mice (mental fatigue model)	[69]

(Continued)

**Table 2.**  
(Continued)

Herb name	Monomer compound	Effective constituent	Activity evaluation model	References
Gardenia jasminoides (Zhizi)	Shanzhiside, gardenoside, geniposide, geniposidic acid, chlorogenic acid, 10-O-acetyl geniposide, shanzhiside methyl ester, scandoside methyl ester or deacetyl asperulosidic methyl ester, 6"-O-E-caffeoyl deacetyl asperulosidic methyl ester, 6"-O-sinapoyl gardoside, 3-O-caffeoyl-5-O-sinapoyl quinic acid or 3-O-caffeoyl-4-O-sinapoyl quinic acid, 3,4-di-O-caffeoyl quinic acid or 3,5-diO-caffeoyl quinic acid, 6"-O-sinapoyl geniposide	Shanzhiside, gardenoside, geniposide, geniposidic acid, chlorogenic acid, 10-O-acetyl geniposide, shanzhiside methyl ester, scandoside methyl ester or deacetyl asperulosidic methyl ester, 6"-O-E-caffeoyl deacetyl asperulosidic methyl ester, 6"-O-sinapoyl gardoside, 3-O-caffeoyl-5-O-sinapoyl quinic acid or 3-O-caffeoyl-4-O-sinapoyl quinic acid, 3,4-di-O-caffeoyl quinic acid or 3,5-diO-caffeoyl quinic acid, 6"-O-sinapoyl geniposide	Antiviral activity: influenza virus strain A/FM/1/47-MA in rats	[70]
Salvia miltiorrhiza (Danshen)	15,16-dihydrotanshinone I, cryptotanshinone, tanshinone I, tanshinone IIA, and dansenspiroketallactone	15,16-dihydrotanshinone I, cryptotanshinone, tanshinone I, tanshinone IIA	<i>In vitro</i> FAS inhibitory activity	[71]
Ligusticum chuanxiong (Chuanxiong)	3,8-Dihydro-diligustilide, 4,5-dehydro-diligustilide, levistolide A, tokinolide B, riligustilide	3,8-Dihydro-diligustilide, riligustilide	Estrogenic activity was measured <i>via</i> ERa and ERb expression in the presence of the estrogen-responsive MMTV-ERE-Luc reporter gene (HeLa cells)	[72]
Uncaria rhynchophylla (Gouteng)	Corynoxine, corynoxine B, corynoxene, isorhynchophylline, isocorynoxene, rhynchophylline	Rhynchophylline, isorhynchophylline	Cellular model of Alzheimer disease, $\beta$ -amyloid-(A $\beta$ -) induced neurotoxicity in PC12 cells	[73]
Garcinia nuijiangensis (Nuijiangtenghuang)	Nuijiangexanthones A and B, nuijiangefolins A–C, and 10 known related analogs	Isojacareubin	11 cancer cell lines and immortalized MIHA normal liver cells	[74]
Polygonum cuspidatum (Huzhang)	2-methoxystyandrone, as well as three anthraquinones	2-Methoxystyandrone	A potent inhibitory effect on STAT3 activation and significant inhibition of cell proliferation in human breast cancer cells, especially those with constitutively activated STAT3	[75]
Astragalus membranaceus (Huangqi)	Formononetin, calycosin, and astragaloside IV	Formononetin	Mouse macrophage RAW 264.7 cells (anti-inflammatory effect)	[76]
Picriafel terrae (Kuxuanshen)	Picfeltarraenin IA, picfeltarraenin IB, picfeltarraenin IV, picfeltarraenin X, picfeltarraenin XI, and one unknown compound	Picfeltarraenin IA, picfeltarraenin IB, picfeltarraenin IV, picfeltarraenin X, picfeltarraenin XI, and one unknown compound	One 96-well plate was used for a bioassay (AChE-inhibitory assay)	[77]
Scutellaria barbata (Banzhilian)	Scutebata P–R, scutebata E, and scutebarbatine B	All compounds except scutebata R showed weak cytotoxicity	K562 cell lines, HL60 cell lines	[78]
Ligustrum lucidum (Nvzhenzi)	Tyrosol, tyrosyl acetate, hydroxytyrosol, salidoside, oleoside dimethyl ester, oleoside-7-ethyl-11-methyl ester, nuzhenide, and G13	Tyrosol, tyrosyl acetate, hydroxytyrosol, salidoside, oleoside dimethyl ester, oleoside-7-ethyl-11-methyl ester, nuzhenide, and G13	Osteoblast-like UMR-106 cells	[79]
Cinnamomum cassia (Rougui)	Cinnamoids A–D, cinnamoid E, and 18 known compounds	Cinnamoid E, (–)-15-hydroxy-T-muurolol, 10 $\alpha$ -dihydroxy aromadendrane, and cinnacasside A	High-glucose-stimulated mesangial cells (diabetic nephropathy)	[80]
Limonium bicolor (Ersebuxuecao)	Luteolin, acacetin, quercetin, isorhamnetin, kaempferol, eriodictyol, kaempferol-3-O- $\alpha$ -L-rhamnoside, kaempferol-3-O- $\beta$ -D-glucoside, quercetin-3-O- $\alpha$ -L-rhamnoside, quercetin-3-O- $\beta$ -D-glucoside, quercetin-3-O- $\beta$ -D-galactoside, myricetin-3-O- $\alpha$ -L-rhamnoside, kaempferol-3-O-(6"-O-galloyl)- $\beta$ -D-glucoside, hesperidin, rutin	Luteolin, quercetin, kaempferol	Human colon cancer cells, human breast cancer cells, and osteosarcoma cell lines	[81]

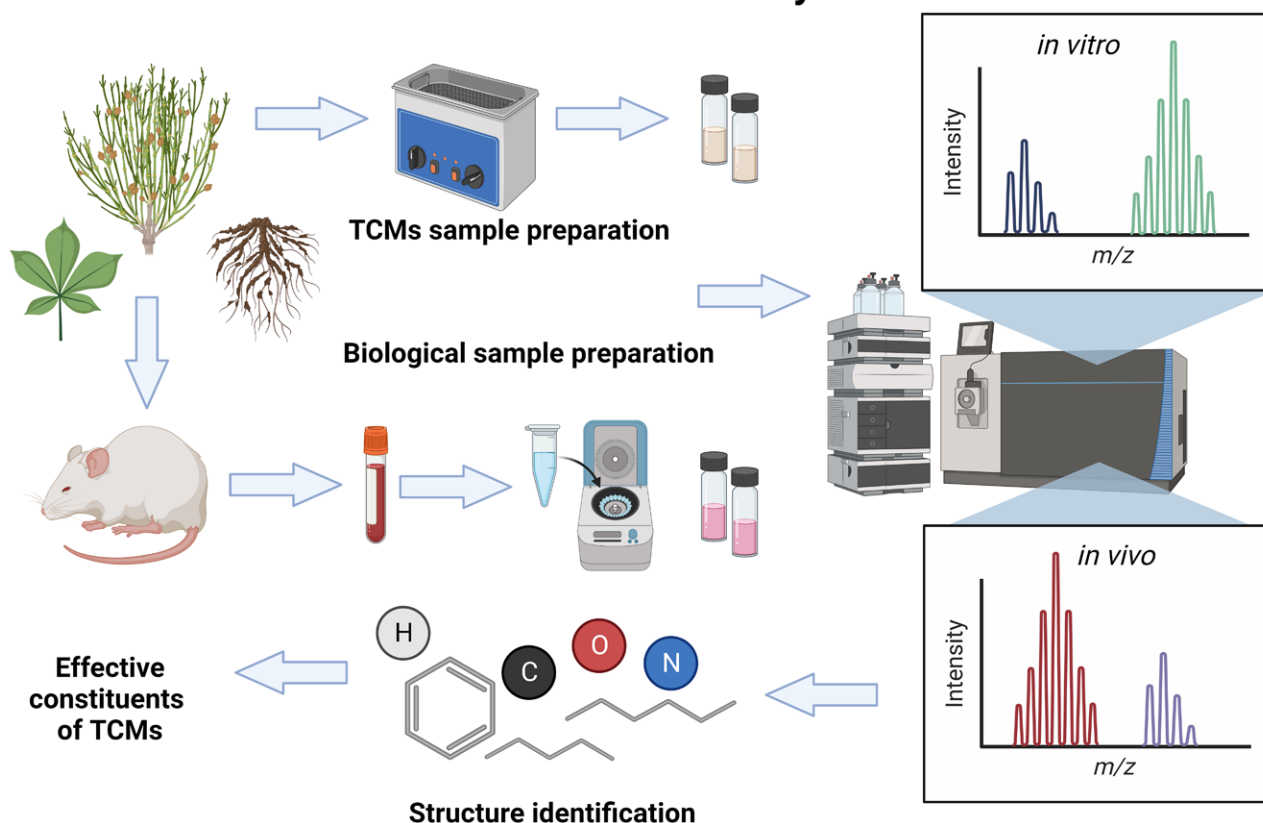
(Continued)

**Table 2.**  
(Continued)

Herb name	Monomer compound	Effective constituent	Activity evaluation model	References
Cimicifuga yunnanensis (Yunnanshengma)	Actein, cimigenol-3-O- $\beta$ -d-xylopyranoside (23R,24S), 26-deoxyacetylacteol-3-O- $\alpha$ -l-arabinopyranoside, 23-epi-26-deoxyactein, cimifugin, cimifugin glucoside, norcimifugin glucoside, norkhellol, 25-O-acetylcimigenol, 26-deoxyacetylacteol, acetylacteol, 25-O-methylcimigenol, cimigenol, 25-O-acetylcimigenol-3-O- $\beta$ -d-xylopyranoside, 24-O-acetylhydroshengmanol-3-O- $\beta$ -d-xylopyranoside, 26-deoxyactein	23-epi-26-deoxyactein and cimigenol	Triple-negative breast cancer cells	[82]
Uncaria gambier (Erchagouteng)	Uncariitannin	Uncariitannin	Mouse models of intraperitoneal infection ( <i>Staphylococcus aureus</i> )	[83]
Morinda officinalis (Bajitian)	Two inulin-type fructans with a backbone consisted of $\alpha$ -D-Glcp-(1 $\rightarrow$ , 4 $\rightarrow$ 1)- $\beta$ -D-Fruf-(2 $\rightarrow$ and $\beta$ -D-Fruf-(2 $\rightarrow$ residues (DP = 7 and 13, respectively)	Two inulin-type fructans	Classic ovariectomized rat model and MC3T3-E1 cells	[84]

ACE: Angiotensin-converting enzyme; FAS: Fatty acid synthase; HEMn: Human epidermal melanocytes; MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; MIHA: cell code, without extension; PYP1B: Protein tyrosine phosphatase 1B; SPL: Splenic lymphocytes; TCM: Traditional Chinese medicine.

## Serum Pharmacochemistry of TCM



**Figure 3.** Workflow of serum pharmacochemistry of TCM method for screening effective constituent of TCMs. TCM: Traditional Chinese medicine. Created by Biorender.com.

6-dementhocycapillarasin, capillarasin, rhein, and emodin<sup>[91]</sup>. Serum sample analysis after the administration of YCHD with different compositions (complete formula, removal of one combination herbal drug or single herbal drug) showed that eight of the 21 compounds were

absorbed from the complete formula alone; interestingly, these eight compounds have stronger hepatoprotective and choleric activities than those of the others. These findings illustrated the scientific significance of TCM formula compatibility<sup>[92]</sup>. Pharmacokinetic analysis of the

main compounds in blood, namely 6,7-dimethoxyesculetin, geniposide, and rhein, demonstrated considerably better pharmacokinetic properties of these compounds from the complete formula; the three herbal drugs work synergistically *in vivo*<sup>[93]</sup>. Bioanalysis Zone described this phenomenon as “Three’s a crowd: but could three be better than one”<sup>[94]</sup>.

AS has anti-stress, antiulcer, anti-irradiation, anti-cancer, anti-inflammatory, and hepatoprotective activities<sup>[95]</sup>. After oral administration of AS extracts, 19 compounds were detected and identified in the blood; of which, 11 were prototype constituents, for example, protocatechuic acid, eleutheroside B, chlorogenic acid, caffeic acid, eleutheroside E, isofraxidin; eight were metabolites, mostly metabolites of eleutheroside B. In addition, the results showed that the content of isofraxidin was small *in vitro* but was highly increased *in vivo*. The time course of isofraxidin in serum after oral administration of AS extract exhibited a bimodal absorption, which is completely different from single compound administration of isofraxidin; this finding may be the combined behavior of free-type isofraxidin and the biotransformation of related precursor compounds (eleutheroside B1). The results demonstrated that the AS extract is a natural slow-release system of isofraxidin, which shows a higher bioavailability and longer-lasting effect compared to-with that of the single compound. These results explain the difference between administration of the AS extract and single isofraxidin and reveals the inherent lasting effect of TCM<sup>[96]</sup> and shows the advantages of serum pharmacochimistry of TCM. Furthermore, with the continuous progress of detection technology, serum pharmacochimistry of TCM has been widely used; the relevant studies are shown in Table 3<sup>[97-120]</sup>.

The emergence of serum pharmacochimistry of TCM was a revolutionary breakthrough in the research on active constituents of TCM and a leap from separation *in vitro* to analysis *in vivo*. The results reflect not only the interaction between formula and herbal drugs but also the effect of the human body on the drugs. The method has been widely used in the field of pharmacodynamics and pharmacokinetics of TCM. Nevertheless, some limitations persist. The most crucial weakness is that the identified components found in the blood are still not associated with disease targets and pathways; in particular, these components are not associated with the biomarkers of TCM syndromes or reflect the clinical efficacy of the TCM formula. This weakness will be overcome by integrating the serum pharmacochimistry of TCM with syndrome metabolomics, establishing associations between *in vivo* (not exclusively in blood) chemical constituents from herbal medicines and syndrome markers to find the constituents highly associated with clinical efficacy. Moreover, other problems remain to be solved. For example, some constituents of herbal medicines with strong activity may not be absorbed; instead, they affect the intestinal flora, and those constituents are ignored using this approach. The enrichment and detection of low-content constituents in serum is also a challenge for active constituent screening, and the time for each TCM constituent to reach peak concentration in serum is not uniform; therefore, more comprehensive

and accurate methods for detecting constituents in blood need to be established to improve the theory and method of serum pharmacochimistry of TCM.

#### *Chinmedomics method: version 3.0 plus*

The effective constituents of TCM are compounds that show clinical efficacy *in vivo* and are related to the targets of disease. According to TCM theory and clinical practice, the disease is described as a syndrome (*Zheng*), and the combination of herbal drugs called formula used as a clinical drug must correspond to the syndrome<sup>[121]</sup>. One herbal drug may contain thousands of chemical constituents with individual bioactivities; nevertheless, owing to the different combinations in formulas for different syndromes, the absorbed constituents for the same herbal drug are different, resulting from formula compatibility and the corresponding syndrome<sup>[122-123]</sup>. Therefore, analyzing the *in vivo* constituents when the formula shows efficacy for the syndrome is a better choice for identifying the compounds related to the treatment of the disease. However, for a long time, TCM syndromes did not have objective diagnostic criteria, and the diagnosis depended on the subjective cognitive analysis and judgment by the practitioner<sup>[124]</sup>. The absence of diagnostic parameters for TCM syndromes resulted in the inability to accurately evaluate the effectiveness of the formula; therefore, identification of the effective constituents of the formula *in vivo* related to clinical efficacy was not possible. Hence, interpretation of the biological characteristics of a syndrome and syndrome biomarkers are the premise for evaluating the efficacy and discovering effective constituents of TCMs<sup>[125]</sup>.

Traditional Chinese medicine syndromes are functional disorder states, which are the body’s response to various environmental changes and internal and external pathogenic factors. The essence of the syndrome is based on the metabolic network changes caused by body imbalance<sup>[121]</sup>. Metabolites represent both the downstream output of the genome and the upstream input from the environment, and they can provide unique insights into the fundamental causes of disease<sup>[126]</sup>. Therefore, metabolomics technology can facilitate the discovery of biomarkers of syndromes/diseases and the exploration of targets and metabolic pathways of a syndrome through non-discriminatory analysis of endogenous small-molecule metabolites. Professor Xijun Wang integrated serum pharmacochimistry of TCM and metabolomics, thereby proposing and establishing the research strategy of “Chinmedomics”<sup>[127]</sup>. On one hand, metabolomics technology is used to clarify the biomarkers of TCM syndromes and evaluate the efficacy of the formula based on metabolomic profiles and biomarkers of the syndromes. On the other hand, serum pharmacochimistry of TCM is used to identify the active constituents *in vivo* when the formula shows efficacy, further mining the correlation between formula constituents *in vivo* and endogenous biomarkers of syndromes using big data analytics and bioinformatics to identify constituents highly related to the efficacy of the formula as the effective constituents of TCM<sup>[128]</sup>. The flow chart of the strategy is shown in Figure 4. This method consists of four key techniques: identification



**Table 3.**  
(Continued)

Single herb/ formula	Chemical component ( <i>in vitro</i> )	Constituent ( <i>in vivo</i> )	References
Shuang Huang Lian formula	46 peaks were obtained and tentatively characterized	Quinic acid, 3,4-dihydroxyphenylglycol sulfate, dihydrosecologanic acid, N-acetyl cysteine S conjugate, isomer of secoxyloganin2-(3,4-dihydroxyphenyl) ethanol sulfate, 3,4-dihydroxyphenylethanol, isomer of secologanic acid, dihydrosecologanic acid, secologanoside-7-methyl ester, demethyldihydrophillygeninglucuronide, secologanic acid, isomer of sweroside, caffeic acid, isomer of secoxyloganin, demethyldihydrophillygenin glucuronide, sweroside, secoxyloganin, demethyldihydrophillygenin glucuronide, p-hydroxyphenyl-propionic acid, chrysin-6-C-arabinosyl-8-C-glucoside, 5,7,3,2',6'-Ppntahydroxyflavone, (+)-pinoresinol glucuronidehydroxylphillygenin glucuronide, (+)-pinoresinol glucuronide, (+)-dehidropinoresinol, (+)-hydroxylpinoresinol, dihydroxy-trimethoxy-flavanone, baicalin, 3,4'-dihydroxy-5,6-dimethoxy-7-O-glucuronide, phillygenin glucuronide, isomer of baicalin, phillygenin glucuronide, 5,7,8-trihydroxy-8-methoxyflavone-7-O-glucuronide, norwogonin-7-O-glucuronide, chrysin-7-O-glucuronide, oroxylin A-7-O-glucuronide, 5,6,7-trihydroxy-8-methoxyflavone-7-O-glucuronide, dedydrophillygenin, isomer of baicalin, wogonoside, dihydroxwogonin, 5,7-dihydroxy-2'-methoxyflavanone-7-O-glucuronide, dihydroxy-dimethoxy flavoneglucuronide, dihydroxy-dimethoxy flavoneglucuronide, matairesinol, hesperetin, skullcapflavon II glucuronide, 5,7,3-trihydroxy-4'-methoxyflavone, norwogonin, 5,7,4'-trihydroxy-8-methoxyflavone, trihydroxy-dimethoxy flavone, baicalein, trihydroxy-dimethoxy flavone, trimethoxy flavone, (+)-epipinoresinol5,2'-dihydroxy-7,8,6'-trimethoxy flavone, wogonin, chrysin, 5,7-dihydroxy-6,8-dimethoxyflavone, skullcapflavone II, oroxylin A	[103]
Zhibai Dihuang pill	93 constituents were identified or tentatively characterized, 23 of which were from Phellodendri amurensis Cortex	Hydroxymethyl armepavine-O-glucuronide, hydroxymethyl armepavine-O-glucuronide, phellodendrine-O-glucuronide, phellodendrine, armepavine metabolites, 3-O-feruloylquinic acid-O-glucuronide, magnoflorine, tetrahydrojatrorrhizine-O-glucuronide, 3-O-feruloylquinic acid, demethyleneberberine-O-glucuronide, jatrorrhizine-O-glucuronide, menisperine, N-methyltetrahydrocolumbamine, berberine-O-sulfate-O-glucuronide, hydroxyberberine-O-glucuronide, hydroxyberberine, hydroxyjatrorrhizine-O-glucuronide, palmatine, berberine, $\gamma$ -hydroxybutenolide derivatives II, sanleng acid-O-glucuronide, sanleng acid, rutaevin, obaculactone, obacunone, 3-hydroxyl feruloylquinic acid, tri-hydroxyjatrorrhizine, rutaevin-O-sulfate 3-O-feruloylquinic acid-O-glucuronide, hydroxyjatrorrhizine, sanleng acid-O-sulfate, rutaevin-O-glucuronide	[104]
Phellodendri Amurensis Cortex	A total of 46 peaks were obtained, 41 of which were tentatively characterized from GHB	24 ions of interest were extracted, 12 prototype constituents of GHB, phellodendrine, magnoflorine, 3-O-feruloylquinic acid, menisperine, tetrahydropalmatine, palmatine, berberine prototype, sanleng acid, rutaevin, obaculactone, obacunone, and 12 metabolites	[105]
Shaoyao Gancao decoction	-	Gallic acid, albiflorin, liquiritigenin-4-O-glucuronide, liquiritin, formononetin glucuronide, pallidiflorin, liquiritigenin, isoliquiritigenin, formononetin, isolicoflavonol, licoricone, one unknown (12 constituents including nine prototype constituents and two metabolites)	[106]
Xianling Gubao capsule	-	Diphyllaside A, epimedeside E, diphyllaside B, epimedeside A, epimedin A, epimedin B, epimedin C, icariin, wanepimedeside A, ikarisoside B, ikarisoside F, 2''-O-rhamnosylkariside A, ikarisoside A, sagittatoside A, icariside I, sagittatoside B, 2''-O-rhamnosylkariside I, icariside II, icaritin, psoralenollsobavachin, neobavaisoflavone, bavachin, corylin, bavachalcone, isobavachalcone, bavachinin, corylifol A, bavachromene, 4-methylbavachalcone, neocorylin, psoralidin, psoralenoside, isopsoralenoside, psoralen, isopsoralen, asperosaponin VI, hederagenin B, sophorosyl ester, akebia saponin PA, timosaponin B-II, timosaponin B-III, timosaponinA- III, timosaponin A-I, protocatec huicaldehyde, caffeic acid, danshensu, ferulic acid, isoerulic acid, 15,16-dihydrotanshinone I, tanshinol B, 1,2-dihydrotanshinone I, tanshinone v, cyptotanshinone, dehydrotanshinone II, A, tanshinone IIA, magnoflorine, cycloolivil, 3-hydroxybakuchiol, bakuchiol, loganin, sweroside, cis-hinoliresinol, etc (57 prototype components and 77 metabolites)	[107]
Sheng Mai powder	-	7,8-dihydroxy-3-demethyl schizandrin, 7,8-dihydroxy-2-demethyl schizandrin, 7,8-dihydroxy-2-demethyl schizandrinisomer, 7,8-dihydroxy-schizandrin, schisandrin, gomisin D, schisandrol B, angeloygomisin H, schisantherin A, schisantherin B, deoxyschisandrin, $\gamma$ -schisandrin, schisandrin B, 20(S)-ginsenoside Rh1, 20(R)-ginsenoside Rh1, ginsenoside Rk3, ginsenoside Rh4, gomisin P isomer, gomisin H isomer, isoschisandrin, schisandrin isomer, tigloylgomisin H, epi-gomisin O, benzoylgomisin Q, angeloylgomisin Q, gomisin F, schisandrin C	[108]

(Continued)

**Table 3.**

(Continued)

Single herb/ formula	Chemical component ( <i>in vitro</i> )	Constituent ( <i>in vivo</i> )	References
Kai Xin powder	-	Bergapten, mannose, valine, dimethyl(R)-(+)-malate, sucrose, nicotinic acid, benzoic acid, adenine, adenosine, sibiricose Al, ethyl benzoylacetate, salicylic acid, polygalaxanthone III, 1-(3,4-dimethoxyphenyl) ethan-1-one, 2-O-methyl-a-D-glucopyranose, 2-hydroxybenzoic acid[(2S)-5-oxotetrahydro-2-furanyl] methyl, benzoate, dibutyl oxalate, 3,4,5-trimethoxy cinnamic acid, acoronene, poricoic acid H, poricoic acid G, gomisin A, 3β,16a-dihydroxylanosta-7,9(11),24-trien-21-oic acid, poricoic acid B, 6,7-dehydroporicoic acid H, 16-deoxyporicoic acid B, tumulosic acid, cis/trans-methylisoeugenol, 3β-hydroxy-16a-acetoxy-lanosta-7,9(11),24-trien-21-oic acid, daedaleanic acid A, (5E,20S)-24-methylene-3-oxolanosta-7,9(11)-dien-21-oic acid, dehydroeburiconic acid, dehydrotumulosic acid, 2,6-di-sec-buty1-4-methylphenol, pachymic acid, 3-(2-furyl)-3-oxopropanenitrile, N-(2,5-dimethoxyphenyl)-2-methoxyacetamide	[109]
Zi Shen pill	92 peaks were characterized	7-O-glucopyranosyl mangiferin, amurenlactone A/B, mangiferin/isomangiferin, phellodendrine, mangiferin/isomangiferin, magnoflorine, lotusine, menisperine, tetrahydropalmatine, jatrorrhizine, palmatine, berberine, γ-hydroxybutenolide derivatives II, anemarsaponin I/anemarsaponin II, sanleng acid, rutaevin, obaculactone, obacunone	[110]
Acanthopanax senticosi Radix et Rhizoma Seu Caulis (Leaf)	131 constituents were identified or tentatively characterized including triterpenoid saponins, phenols, flavonoids, lignans, coumarins, polysaccharides, and other compounds	Protocatechuic acid, 1-O-butyl-xylopyranose, quinic acid, caffeic acid, feruloyl quinic acid, rutin, hyperoside, quercitrin, 2,3,5,4-tetrahydroxystilbene 2-O-glucopyranoside, 4,15-dioxo-5,8,11,14-tetraoxaocadecane-1,18-dioic acid, malyngic acid, 3-[[2-O-(6-deoxy-mannopyranosyl)- arabinopyranosyl] oxy] olean-12-ene-28,29-dioic acid	[111]
Da Bu Yin pill	70 constituents including alkaloids, flavonoids, polysaccharide, limonoids, flavonoids	Z-6,7-epoxyiligustilide, ginsenoside Re, butylidenephthalide, 3-butylphthalide, ginsenoside Rc, glycyrrhizin, ginsenoside Rd, gypenoside XVI, zingibroside R1, paconol + O + SO <sub>3</sub> , liquiritigenin + SO <sub>3</sub> + GluA, liquiritigenin + GluA, liquiritigenin + GluA, davidigenin + GluA + SO <sub>3</sub> , liquiritigenin + SO <sub>3</sub> , liquiritigenin + SO <sub>3</sub> + GluA, davidigenin + GluA + SO <sub>3</sub> , davidigenin + O + GluA, davidigenin + GluA, davidigenin + GluA, isoliquiritigenin + GluA, isoliquiritigenin + SO <sub>3</sub> , hydroxyglycyrrhetinic acid, hydroxyglycyrrhetinic acid, glyasperinD + GluA, hydroxyglycyrrhetinic acid, hydroxyglycyrrhetinic acid	[112]
Xianling Gubao capsule	-	Sweroside, magnoflorine, psoralenoside, isopsoralenoside, psoralen, timosaponin B II, isopsoralen, epimedin C, icariin, akebia saponin D, icariside I, 2''-O-rha-icariside II, neobavaisoflavone, icariside II, corylin, akebia saponin PA, psoralidin, bavachinin b-glucogallin, gallic acid, mudanoside B, (+)-catechin, hyemaloside B, oxypaeoniflorin, dihydroxy acetophenone, ethyl gallate, dihydroxy methoxy acetophenone, paeonol, paeoniflorin, ethyl gallate, suffruticoside B or D, galloyl paeoniflorin, paeonol, benzoyloxypaeoniflorin, mudanpioside C, dihydroxy methoxy acetophenone	[113]
Moutan Cortex	46 constituents	Syringic acid-4-B-D-glucopyranoside, protocatechuic acid-3-glucoside, oxypaeoniflorin, magnoflorine, albiflorin, paeoniflorin, linalool 3,6-oxide 6-O-β-D-xylopyranosyl-β-D-glucopyranoside, berberrubine, galloylpaeoniflorin isomer, coptisine, epiberberine, jatrorrhizine, berberine, palmatine, 3-methyl ellagic acid, 3,3-di-O-methyl ellagic acid, ginsenoside Rg1, 9(S),12(S),13(S)-trihydroxy-10(E),15(Z)-octadecadienoic acid, ginsenoside Rb1, 5-O-feruloylquinic acid/3-O-feruloylquinic acid, berberrubine-9-O-3-D-glucuronide, jatrorrhizine-3-O-B-D-glucuronide, demethyleneberberine-2-O-3-D-glucuronide, demethylberberubine-2-O-3-D-glucuronide, demethyleneberberine, 9(S),10(S),11(R)-trihydroxy-12(Z),15(Z)-octadecadienoic acid, ginsenoside Rf, 9(S),10(S),13(S)-trihydroxy-11(E),15(Z)-octadecadienoic acid	[114]
Wen Xin formula	97 constituents	Vanillic acid glucoside, protocatechuic acid, vanillic acid, neochlorogenic acid, coniferol, sinapyl alcohol, ferulic acid, sinapyl alcohol, caffeic acid, isofraxidin, vanillic acid, isofraxidin, caffeic acid, isofraxidin, guaiacylglycerolconiferol glucoside, guaiacylglycerol hydroconiferolglucoside, 2-[2-hydroxy-5-(1E)-3-hydroxy-1-propen-1-yl]-3-methoxyphenyl]-3-(4-hydroxy-3-methoxyphenyl) propylhexopyranoside, isofraxidin, guaiacylglycerol coniferol glucoside, salvadoraside, guaiacylglycerolhydroconiferolglucoside, 2-[2-hydroxy-5-[(1E)-3-hydroxy-1-propen-1-yl]-3-methoxyphenyl]-3-(4-hydroxy-3-methoxyphenyl) propylhexopyranoside, guaiacylglycerolconiferol glucoside, guaiacylglycerol hydro-coniferol glucoside, feruloylquinic acid, (2R,3R)-2,3-bis (4-hydroxy-3,5-dimethoxybenzyl)-1,4-butanediol, ciwujiatone, secoisolariciresinol, dimethoxyl lariciresinol, ciwujiatone, lariciresinol 3-α-O-β-D-glucopyranoside, dimethoxyl lariciresinol, methoxyl lariciresinol, isofraxidin, pinosresinol glucoside, syringaresinol, 4-O-β-D-glucopyranoside, matairesinol	[115]
Acanthopanax senticosi Radix et Rhizoma Seu Caulis (Stem)	115 constituents	Vanillic acid glucoside, protocatechuic acid, vanillic acid, neochlorogenic acid, coniferol, sinapyl alcohol, ferulic acid, sinapyl alcohol, caffeic acid, isofraxidin, vanillic acid, isofraxidin, caffeic acid, isofraxidin, guaiacylglycerolconiferol glucoside, guaiacylglycerol hydroconiferolglucoside, 2-[2-hydroxy-5-(1E)-3-hydroxy-1-propen-1-yl]-3-methoxyphenyl]-3-(4-hydroxy-3-methoxyphenyl) propylhexopyranoside, isofraxidin, guaiacylglycerol coniferol glucoside, salvadoraside, guaiacylglycerolhydroconiferolglucoside, 2-[2-hydroxy-5-[(1E)-3-hydroxy-1-propen-1-yl]-3-methoxyphenyl]-3-(4-hydroxy-3-methoxyphenyl) propylhexopyranoside, guaiacylglycerolconiferol glucoside, guaiacylglycerol hydro-coniferol glucoside, feruloylquinic acid, (2R,3R)-2,3-bis (4-hydroxy-3,5-dimethoxybenzyl)-1,4-butanediol, ciwujiatone, secoisolariciresinol, dimethoxyl lariciresinol, ciwujiatone, lariciresinol 3-α-O-β-D-glucopyranoside, dimethoxyl lariciresinol, methoxyl lariciresinol, isofraxidin, pinosresinol glucoside, syringaresinol, 4-O-β-D-glucopyranoside, matairesinol	[116]

(Continued)

**Table 3.**  
(Continued)

Single herb/ formula	Chemical component ( <i>in vitro</i> )	Constituent ( <i>in vivo</i> )	References
Schisandrae chinensis Fructus	-	Schisandrin, gomisin D, schisandrol B, benzoylgomisin H, angeloylgomisin Q, gomisin G, gomisin K, gomisin E, deoxyschizandrin, schisandrin B, and 39 metabolites	[117]
Ding Kun pellet	A total of 234 constituents were identified, including 47 triterpenoid saponins, 55 flavonoids, and 38 alkaloids	1,2,3-tri-O-methyl-D-arabino-L-erythro-4-octulose, safflomin A, isovanillic acid, D-phenylalanine, (+)-laurotetanine, albiflorin, picrocrocin, mudanpioside E, liquiritin, ferulic acid, rotundin, coryphenanthrine, homochelidonine, tetrahydropalmatine, tetrahydrocoptisine, notoginsenoside R <sub>1</sub> , baicalin, ginsenoside Rg <sub>1</sub> , corydaline, liquiritigenin, oroxylin-A-7-O-glucuronide, hispidulin-7-O-β-D-glucuronopyranoside, wogonoside, ginsenoside Rb1, glyasperins M, isoliquiritigenin, ginsenoside Rd, 3-(2,4-dihydroxyphenyl)-8-(1,1 dimethylprop-2-enyl)-7-hydroxy-5-methoxy-coumarin, glycycomarin or isomer, palmitic acid, senkyunolide C, phytosphingosine, yunnankadsurin A, genkwanin, acetoxyl oxokadsurane, heteroclitin G, ligustilide, eudesm-4 (15) en-6-one, glycyrin, angeloylgomisin R, interiotherin A, senkyunolide O, levistolide A	[118]
Zuo Jin pill	90 constituents were identified in ZJP	3,4-dihydroxybenzoic acid, magnoflorine, berberrubine, 5-O-feruloylquinic acid, coptisine, dehydroevodiamine, epiberberine, columbamine, jatrorrhizine, berberine, palmatine, dehydrocorydaline, diosmin, skimmianine, formylidihydrorutaecarpine, 8-oxo-epiberberine, 8-oxocoptisine, evodiamine, rutaecarpine, 1-Methyl-2- undecyl – 4(1H)-quinolone, evocarpine, dihydroevocarpine	[119]
Da Cheng Qi decoction	-	5,7-dihydroxychromone, rhein, diglucoside, cinnamic acid, 2-cinnamoyl-glucose, benzylideneacetone, safrrole, (–)-epicatechin gallate, rhein-8-O-3-D-glucopyranoside, physcion 8-glucoside, toralactone, narirutin, naringin, aloe-emodin-glucoside, hesperidin, emodinanthrone, 5,7,4'-trimethylapigenin, tetramethoxyluteolin, 2-methoxy-4-vinylphenol, hesperetin, quercetin, chrysophanol, glucoside, neohesperidin, 4',5,7,8-tetramethoxyflavone, anthraglycoside B, eupatin, apigenin, naringenin, poncirin, magnolignan C, bergapten, rheochrysin, nobiletin, randaiol, luteolin, ammidin, magnolignan E, isosinensetin, tangeretin, aloemodin, rhein, 5-O-demethylnobiletin, emodin, obovatol, chrysophanol, physcion	[120]

GHB: Guanhuangbo (Phellodendri amurensis Cortex); MS: Mass spectrum; TCM: Traditional Chinese medicine.

of syndrome biomarkers, precise efficacy evaluation of formula, characterization of constituents of formula *in vivo*, and mining the correlation between constituents and syndrome biomarkers<sup>[17]</sup>. This method has been widely accepted and cited; *Nature* recommended it as a powerful approach for the efficacy evaluation of TCM, a biological language between TCM and Western medicine<sup>[129]</sup>. Professor Rahman described it as an emerging technology that could be used for disease fingerprinting or predicting drug effects<sup>[130]</sup>. In 2015, Professor Xijun Wang published a monograph on Chinmedomics by Elsevier<sup>[131]</sup>, and he published six series of monographs in yearly volumes of Research Progress in Chinmedomics by Science Press (2016–2021)<sup>[132–137]</sup>.

Chinmedomics has been used to evaluate the efficacy of approximately 20 classical TCM formulas and elucidate their mechanisms, a summary of these is provided in Table 4<sup>[138–154]</sup>. We will use YHS and YCHD as examples to introduce the detail of Chinmedomics. YHS is a damp-heat phenotype of jaundice syndrome, and its clinical symptoms are fever; polydipsia; bright yellowish discoloration of the whites of the eyes, mucous membranes, and the skin; strong-tea-colored dark urine; accompanied by loss of appetite or nausea and vomiting<sup>[155]</sup>. YCHD is a classical prescription commonly used in the treatment of YHS; thousands of years of experiments on human subjects indicate that the clinical effect is remarkable<sup>[156]</sup>. However, it remains unclear which specific active ingredients act

on which targets to produce therapeutic effects, and Chinmedomics provides a new research paradigm for solving this problem. For the first time, researchers introduced the metabonomics technology for small-molecule metabolite-level characterization of the TCM syndrome. Comparing patients accompanied YHS with healthy subjects, 40 urinary metabolic differential markers were successfully identified; these mainly included kinetin, porphobilinogen, and indoleacrylic acid<sup>[157]</sup>. The approach achieved accurate quantitative characterization of the TCM syndrome, which laid a foundation for an accurate evaluation of the TCM curative effect. In a recent study of blood metabolomics in jaundice patients, 14 different metabolites were successfully identified, including taurocholic acid, bilirubin glucuronide, bilirubin, and biliverdin, which suggests that they can be used as an effective supplement to clinically diagnose jaundice. In addition, after treatment with YCHD, the clinical symptoms of patients were markedly improved, and the contents of 13 blood metabolic biomarkers were notably altered, returning to normal levels. Moreover, 26 prototype components and three metabolites were successfully identified *in vivo* using serum pharmacology of YCHD in clinical subjects with significant efficacy, and the correlation analysis with biomarkers revealed that geniposide, scoparone, isorhamnetin, quercetin, naringenin, rhein, chlorogenic acid, and kaempferol were related to clinical efficacy<sup>[158]</sup>.



**Table 4.****Representative studies on the effective constituents of TCMs using Chinmedomics**

Prescription	Syndrome/ diseases	Effective constituent	References
Wen Xin formula	Myocardial ischemia syndrome	Ginsenosides, paeoniflorin, galloyl glucose, berberis alkaloids, phenolic, phenolic glycosides and unsaturated fatty acids, glucuronide products of original berberis alkaloids	[138]
Shen Qi pill	Kidney- <i>yang</i> deficiency syndrome	Azelaic acid-O-glucuronide, jionoside D, azelaic acid, poricoic acid B-O-sulfate, tumulosic acid-O-glucuronide, poricoic acid A, eugenol methyl ether, and dehydroeburicoic acid	[139]
Nan Shi Oral liquid	Kidney- <i>yang</i> deficiency syndrome	Scandoside, monotropeine, gallic acid, stevioside C, steviolbioside, and morroniside glucuronide conjugation	[140]
AS1350	Kidney- <i>yang</i> deficiency syndrome	Schizandrin, gomisin S, $\beta$ ine, scoparone, clovene, stepharine, longipedunin C, gomisins, auxin A, and 1,11-undecanedicarboxylic acid	[141]
Nan Shi capsule	Kidney- <i>yang</i> deficiency syndrome	Caffeic acid, 2-hydroxy-1-methoxy-antraquinone, 1-hydroxy-2-methoxyantraquinone, ferulic acid glucuronide conjugation, deacetylasperulosidic acid, cynaroside, $\beta$ ine, and umbelliferone	[142]
Phellodendri amurense Cortex (GHB)	Prostate cancer	Magnoflorine-O-glucuronide, (p-hydroxybenzyl)-6,7-dihydroxy-N-methyltetrahydro isoquinoline-7-O-p-D-glucopyranosid, magnoflorine, menisperine-O-glucuronide, menisperine, berberine, jatrorrhizine, obaculactone, and obacunone	[143]
Sheng Mai powder	Alzheimer disease	Schisandrin, isoschisandrin, angeloylgomisin Q, gomisin D, angeloylgomisin H, gomisin M2, ginsenoside F1, and 20(R)-ginsenoside Rg3	[144]
Kai Xin powder	Alzheimer disease	Ginsenoside Rf, ginsenoside F1, 20-O-glucopyranosyl ginsenoside Rf, dehydropachymic acid, and E-3, 4,5-trimethoxycinnamic acid	[145]
Yin Chen Hao decoction	Dampness-heat jaundice syndrome	2-ethyl-2-hexenal, isofraxidin, 2,5-dimethyl-7-hydroxy chromone, 6,7-dimethoxy coumarin, geniposide, capillarasin, neochlorogenic acid, chimaphyllin, and isorhamnetin-3-glucoside	[146]
Yun Nan Bai Yao	Blood stasis syndrome	Karacolidine, senbusine B, isotalatizidine, karakoline, denudatine, talatisamine, and chasmanine	[147]
Gelan Xinning capsule	Coronary heart disease	11 compounds as quality markers responsible for the efficacy of Ge Lan Xin Ning capsule	[148]
Fangji Huangqi decoction	Nephrotic syndrome	(+)-Tetrandrine demethylation, fengfangjine G hydrogenation, tetrandrine, N-methylfangchinoline, tetrandrine demethylation, fangchinoline, glycyrrhetic acid, astragaloside II alcohol dehydration, atractylenolide III demethylation + hydrogenation, atractylenolide III demethylation + hydrogenation, and licoricone-N-acetylcysteine conjugation	[149]
Baizhu Shaoyao powder	Ulcerative colitis	Atractylenolide I-M5, hesperidin-M2, cimifugin, albiflorin-M2, cimifugin-M3, 5-O-methylvisaminol-M2, sec-O-glucosyl-hamaudol, atractylenolide II, atractylenolide III-M5, and atractylenolide III-M4	[150]
Compound Zao fan pill (consist of Panacis quinquefolii Radix)	<i>Qi</i> -blood deficiency syndromes	24-methyl-7-cholesten-3 $\beta$ -ol, zizyboside II, betulin, ginsenoside Rd, cinnamyl alcohol, and pseudoginsenoside F11 considered a potential Q-marker for the compound Zao fan pill; pseudoginsenoside F11 and ginsenoside Rd, considered a potential Q-marker for Panax quinquefolium L	[151]
Si Jun Zi decoction	Spleen <i>qi</i> deficiency syndrome	Malonyl-ginsenoside Rb2 and ginsenoside Ro as the q-markers of ginseng; dehydrotumulosic acid and dihydroxy lanostene-triene-21-acid as the q-markers of poria; glycyrrhizic acid, isoglabrolide, and glycyrrhetic acid as the q-markers of licorice; and 2-atractylenolide as the q-marker of macrocephala	[152]
GHB in Zhi Bai Di Huang pill	Kidney- <i>yin</i> deficiency	Magnoflorine and jatrorrhizine	[122]
Zhizi Baiqi decoction	Damp-heat jaundice syndrome	Isoformononetin, 3-O-feruloylquinic acid, glycyrrhizic acid, oxyberberine, obaculactone, and five metabolites	[123]
Ke Luo Xin capsule	Diabetic retinopathy	Emodin, rhein, astragaloside IV, chrysophanol, chrysophanol-8-O- $\beta$ -d-glucopyranoside, aloe-emodin, aloin, biochanin A, 10-hydroxyoleuropein, 3,4',7-trihydroxyflavone, and 3'-O-methyl(-)-epicatechin 7-O-glucuronide	[153]
Danggui Jianzhong decoction	Primary dysmenorrhea	Ferulic acid, zizyphusin, cinnamic acid, protocatechuic acid-3-glucoside, and azelaic acid	[154]

GHB: Guanhuangbo (Phellodendri amurense Cortex); TCM: Traditional Chinese medicine.

**Conclusion**

Analysis and identification of effective constituents from the complex mega system of TCMs are crucial for interpreting the principles of TCMs. They are also key steps

to the discovery of new drugs based on clinical experience. With the continuous innovation of technology, researchers have innovatively explored and established a series of strategies for TCM. The discovery methods



Ye Sun, Ying Han, Qiqi Zhao, and Shuyu Kang participated in the performance of some research and reference material collection.

### Ethical approval of studies and informed consent

Not applicable.

### Acknowledgments

None.

### Data Availability

All data generated or analyzed during this study are included in this published article.

### References

- [1] Cheung F. TCM: made in China. *Nature* 2011;480(7378):S82–S83.
- [2] Tu Y. Artemisinin—a gift from Traditional Chinese Medicine to the World (Nobel Lecture). *Angew Chem Int Ed Engl* 2016;55(35):10210–10226.
- [3] Hou Q, He WJ, Wu YS, et al. Berberine: a traditional natural product with novel biological activities. *Altern Ther Health Med* 2020;26(S2):20–27.
- [4] Song D, Hao J, Fan D. Biological properties and clinical applications of berberine. *Front Med* 2020;14(5):564–582.
- [5] Shao F, Wilson IW, Qiu D. The research progress of taxol in Taxus. *Curr Pharm Biotechnol* 2021;22(3):360–366.
- [6] Lyu M, Fan G, Xiao G, et al. Traditional Chinese medicine in COVID-19. *Acta Pharm Sin B* 2021;11(11):3337–3363.
- [7] National Medical Products Administration. National Medical Products Administration approved the marketing of Qingfei Didu Granules, Huashi Baidu Granules and Xuanfei Baidu Granules. Available from: <https://www.nmpa.gov.cn/zhuanti/yqyjxd/yqyjxd/20210302190503177.html>. Accessed March 2, 2021.
- [8] Xu X, Xia J, Zhao S, et al. Qing-Fei-Pai-Du decoction and wogonoside exert anti-inflammatory action through down-regulating USP14 to promote the degradation of activating transcription factor 2. *FASEB J* 2021;35(9):e21870.
- [9] Hao P, Jiang F, Cheng J, et al. Traditional Chinese Medicine for cardiovascular disease: evidence and potential mechanisms. *J Am Coll Cardiol* 2017;69(24):2952–2966.
- [10] Zhang HY, Tian JX, Lian FM, et al. Therapeutic mechanisms of traditional Chinese medicine to improve metabolic diseases via the gut microbiota. *Biomed Pharmacother* 2021;133:110857.
- [11] Guo R, Li L, Su J, et al. Pharmacological activity and mechanism of Tanshinone IIA in related diseases. *Drug Des Devel Ther* 2020;14:4735–4748.
- [12] Wang P, Chen Z. Traditional Chinese medicine ZHENG and Omics convergence: a systems approach to post-genomics medicine in a global world. *Omics* 2013;17(9):451–459.
- [13] Tang JL, Liu BY, Ma KW. Traditional Chinese medicine. *Lancet* 2008;372(9654):1938–1940.
- [14] Chen KK, Schmidt CF. The action and clinical use of ephedrine, an alkaloid isolated from the Chinese drug ma Huang; historical document. *Ann Allergy* 1959;17:605–618.
- [15] Wani MC, Taylor HL, Wall ME, et al. Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemic and antitumor agent from *Taxus brevifolia*. *J Am Chem Soc* 1971;93(9):2325–2327.
- [16] Wang XJ. Study on serum pharmacology of Traditional Chinese Medicine. *Modernization of Traditional Chinese Medicine and Materia Medica-World Science and Technology* 2002;4(2):1–4 + 78.
- [17] Han Y, Sun H, Zhang A, et al. Chinmedomics, a new strategy for evaluating the therapeutic efficacy of herbal medicines. *Pharmacol Ther* 2020;216:107680.
- [18] Phillipson JD. Phytochemistry and medicinal plants. *Phytochem* 2001;56(3):237–243.
- [19] Ji D, Wang Q, Wang H, et al. Preparative separation of gallic acid from *Fallopia aubertii* using middle-pressure chromatogram isolated gel coupled with reversed-phase chromatography with hydrophilic groups. *RSC Adv* 2021;11(44):27276–27282.
- [20] Santiago M, Strobel S. Thin layer chromatography. *Methods Enzymol* 2013;533:303–324.
- [21] McChesney JD, Rodenburg DL. Preparative chromatography and natural products discovery. *Curr Opin Biotechnol* 2014;25:111–113.
- [22] Chen KK, Schmidt CF. The action of ephedrine, an alkaloid from Ma Huang. *Proc Soc Exp Biol Med* 1924;21(6):351–354.
- [23] Chen KK. A study of ephedrine. *Br Med J* 1927;2(3482):593.
- [24] Wall ME, Wani MC, Cook CE, et al. Plant antitumor agents. I. The isolation and structure of camptothecin, a novel alkaloidal leukemia and tumor inhibitor from *Camptotheca acuminata* 1,2. *J Am Chem Soc* 1966;88(16):3888–3890.
- [25] Ikeya Y, Taguchi H, Yoshioka I, et al. The constituents of *Schizandra chinensis* Baill. I. Isolation and structure determination of five new lignans, gomisin A, B, C, F and G, and the absolute structure of schizandrin. *Chem Pharm Bull (Tokyo)* 1979;27(6):1383–1394.
- [26] Maeda S, Sudo K, Aburada M, et al. Pharmacological studies on *Schizandra* fruit. I. General pharmacological effects of gomisin A and schizandrin (author's transl). *Yakugaku Zasshi* 1981;101(11):1030–1041.
- [27] Schulz ER. Berberine in the common barberry (*Berberis vulgaris* L.). *J Am Pharm Assoc (Wash)* 1926;15(1):33–39.
- [28] Oshio H, Inouye H. Iridoid glycosides of *Rehmannia glutinosa*. *Phytochem* 1982;2(1):133–138.
- [29] Rogelj B, Popovic T, Ritonja A, et al. Chelidocystatin, a novel phytoconstituent from *Chelidonium majus*. *Phytochem* 1998;49(6):1645–1649.
- [30] Yasukawa K, Kaminaga T, Kitanaka S, et al. 3 beta-p-hydroxybenzoyldehydrotramulosic acid from *Poria cocos*, and its anti-inflammatory effect. *Phytochem* 1998;48(8):1357–1360.
- [31] Tan RX, Kong LD, Wei HX. Secoiridoid glycosides and an anti-fungal anthranilate derivative from *Gentiana tibetica*. *Phytochem* 1998;47(7):1223–1226.
- [32] el-Mekkawy S, Meselhy MR, Nakamura N, et al. Anti-HIV-1 and anti-HIV-1-protease substances from *Ganoderma lucidum*. *Phytochem* 1998;49(6):1651–1657.
- [33] Lin LC, Yang LL, Chou CJ. Cytotoxic naphthoquinones and plumbagic acid glucosides from *Plumbago zeylanica*. *Phytochem* 2003;62(4):619–622.
- [34] Dai SJ, Tao JY, Liu K, et al. neo-Clerodane diterpenoids from *Scutellaria barbata* with cytotoxic activities. *Phytochem* 2006;67(13):1326–1330.
- [35] Don MJ, Shen CC, Syu WJ, et al. Cytotoxic and aromatic constituents from *Salvia miltiorrhiza*. *Phytochem* 2006;67(5):497–503.
- [36] Xu M, Wang D, Zhang YJ, et al. Dammarane triterpenoids from the roots of *Gentiana rigescens*. *J Nat Prod* 2007;70(5):880–883.
- [37] Pan Y, Wang X, Hu X. Cytotoxic withanolides from the flowers of *Datura metel*. *J Nat Prod* 2007;70(7):1127–1132.
- [38] Jin HZ, Wang XL, Wang HB, et al. Morphinane alkaloid dimers from *Sinomenium acutum*. *J Nat Prod* 2008;71(1):127–129.
- [39] Yuan D, Ma B, Wu C, et al. Alkaloids from the leaves of *Uncaria rhynchophylla* and their inhibitory activity on NO production in lipopolysaccharide-activated microglia. *J Nat Prod* 2008;71(7):1271–1274.
- [40] Liu XT, Wang ZZ, Xiao W, et al. Cholestane and spirostane glycosides from the rhizomes of *Dioscorea septemloba*. *Phytochem* 2008;69(6):1411–1418.
- [41] Lu ZQ, Guan SH, Li XN, et al. Cytotoxic diterpenoids from *Euphorbia helioscopia*. *J Nat Prod* 2008;71(5):873–876.
- [42] Li XN, Pu JX, Du X, et al. Lignans with anti-HIV activity from *Schisandra propinqua* var. *sinensis*. *J Nat Prod* 2009;72(6):1133–1141.
- [43] Wu M, Wu P, Liu M, et al. Iridoids from *Gentiana loureirii*. *Phytochem* 2009;70(6):746–750.
- [44] Tao Y, Wang CH, Chou GX, et al. New alkaloids from *Capparis spinosa*: Structure and X-ray crystallographic analysis. *Food Chem* 2010;123(3):705–710.
- [45] Lin S, Chen T, Liu XH, et al. Iridoids and lignans from *Valeriana jatamansi*. *J Nat Prod* 2010;73(4):632–638.
- [46] Wu JJ, Cheng KW, Zuo XF, et al. Steroidal saponins and ecdysterone from *Asparagus filicinus* and their cytotoxic activities. *Steroids* 2010;75(10):734–739.
- [47] Dettweiler M, Marquez L, Bao M, et al. Quantifying synergy in the bioassay-guided fractionation of natural product extracts. *PLoS One* 2020;15(8):e0235723.
- [48] Ma N, Zhang Z, Liao F, et al. The birth of artemisinin. *Pharmacol Ther* 2020;216:107658.
- [49] Sezik E, Aslan M, Yesilada E, et al. Hypoglycaemic activity of *Gentiana olivieri* and isolation of the active constituent through bioassay-directed fractionation techniques. *Life Sci* 2005;76(11):1223–1238.

- [50] Othman R, Ibrahim H, Mohd MA, et al. Bioassay-guided isolation of a vasorelaxant active compound from *Kaempferia galanga* L. *Phytomedicine* 2006;13(1-2):61–66.
- [51] Lee MH, Lin YP, Hsu FL, et al. Bioactive constituents of *Spatholobus suberectus* in regulating tyrosinase-related proteins and mRNA in HEMn cells. *Phytochem* 2006;67(12):1262–1270.
- [52] Tian XY, Wang YH, Yang QY, et al. Jacaranone glycosides from *Senecio scandens*. *J Asian Nat Prod Res* 2006;8(1-2):125–132.
- [53] Tian XY, Wang YH, Liu HY, et al. On the chemical constituents of *Dipsacus asper*. *Chem Pharm Bull (Tokyo)* 2007;55(12):1677–1681.
- [54] Ma J, Dey M, Yang H, et al. Anti-inflammatory and immunosuppressive compounds from *Tripterygium wilfordii*. *Phytochem* 2007;68(8):1172–1178.
- [55] Xu YM, McLaughlin SP, Gunatilaka AA. Sorbifolivaltrates A-D, diene valepotriates from *Valeriana sorbifolia*(1). *J Nat Prod* 2007;70(12):2045–2048.
- [56] Li QL, Li BG, Zhang Y, et al. Three angiotensin-converting enzyme inhibitors from *Rabdosia coetsa*. *Phytomedicine* 2008;15(5):386–388.
- [57] Chen B, Liu Y, Liu HW, et al. Iridoid and aromatic glycosides from *Scrophularia ningpoensis* Hemsl. and their inhibition of  $[Ca^{2+}]_i$  increase induced by KCl. *Chem Biodivers* 2008;5(9):1723–1735.
- [58] Zhao F, Xu H, He EQ, et al. Inhibitory effects of sesquiterpenes from *Saussurea lappa* on the overproduction of nitric oxide and TNF- $\alpha$  release in LPS-activated macrophages. *J Asian Nat Prod Res* 2008;10(11–12):1045–1053.
- [59] Ho CC, Kumaran A, Hwang LS. Bio-assay guided isolation and identification of anti-Alzheimer active compounds from the root of *Angelica sinensis*. *Food Chem* 2009;114(1):246–252.
- [60] Dat NT, Jin X, Lee K, et al. Hypoxia-inducible factor-1 inhibitory benzofurans and chalcone-derived diels-alder adducts from *Morus* species. *J Nat Prod* 2009;72(1):39–43.
- [61] Zhang C, Wang X, Zhang X, et al. Bioassay-guided separation of citreosein and other oestrogenic compounds from *Polygonum cuspidatum*. *Phytother Res* 2009;23(5):740–741.
- [62] Huang SX, Feng C, Zhou Y, et al. Bioassay-guided isolation of xanthenes and polycyclic prenylated acylphloroglucinols from *Garcinia oblongifolia*. *J Nat Prod* 2009;72(1):130–135.
- [63] Baumgartner RR, Steinmann D, Heiss EH, et al. Bioactivity-guided isolation of 1,2,3,4,6-Penta-O-galloyl-D-glucopyranose from *Paeonia lactiflora* roots as a PTP1B inhibitor. *J Nat Prod* 2010;73(9):1578–1581.
- [64] Cicek SS, Khom S, Taferner B, et al. Bioactivity-guided isolation of GABA(A) receptor modulating constituents from the rhizomes of *Actaea racemosa*. *J Nat Prod* 2010;73(12):2024–2028.
- [65] Shu X, Yu L, Tang Y, et al. Bioassay-guided separation of the proinflammatory constituents from the roots of *Euphorbia kansui*. *J Nat Med* 2010;64(1):98–103.
- [66] Li W, Li S, Lin L, et al. Bioassay-guided isolation and quantification of the alpha-glucosidase inhibitory compound, glycyrrhisoflavone, from *Glycyrrhiza uralensis*. *Nat Prod Commun* 2010;5(7):1049–1053.
- [67] Liu X, Hu Z, Shi Q, et al. Anti-inflammatory and anti-nociceptive activities of compounds from *Tinospora sagittata* (Oliv.) Gagnep. *Arch Pharm Res* 2010;33(7):981–987.
- [68] Janicsak G, Zupko I, Nikolovac MT, et al. Bioactivity-guided study of antiproliferative activities of *Salvia* extracts. *Nat Prod Commun* 2011;6(5):575–579.
- [69] Huang LZ, Huang BK, Ye Q, et al. Bioactivity-guided fractionation for anti-fatigue property of *Acanthopanax senticosus*. *J Ethnopharmacol* 2011;133(1):213–219.
- [70] Yang Q, Wu B, Shi Y, et al. Bioactivity-guided fractionation and analysis of compounds with anti-influenza virus activity from *Gardenia jasminoides* Ellis. *Arch Pharm Res* 2012;35(1):9–17.
- [71] Jang TS, Zhang H, Kim G, et al. Bioassay-guided isolation of fatty acid synthase inhibitory diterpenoids from the roots of *Salvia miltiorrhiza* Bunge. *Arch Pharm Res* 2012;35(3):481–486.
- [72] Lim LS, Shen P, Gong YH, et al. Dimeric progestins from rhizomes of *Ligusticum chuanxiong*. *Phytochem* 2006;67(7):728–734.
- [73] Xian YF, Lin ZX, Mao QQ, et al. Bioassay-guided isolation of neuroprotective compounds from *Uncaria rhynchophylla* against beta-amyloid-induced neurotoxicity. *Evid Based Complement Alternat Med* 2012;2012:802625.
- [74] Xia ZX, Zhang DD, Liang S, et al. Bioassay-guided isolation of prenylated xanthenes and polycyclic acylphloroglucinols from the leaves of *Garcinia nuijiangensis*. *J Nat Prod* 2012;75(8):1459–1464.
- [75] Liu J, Zhang Q, Chen K, et al. Small-molecule STAT3 signaling pathway modulators from *Polygonum cuspidatum*. *Planta Med* 2012;78(14):1568–1570.
- [76] Lai PK, Chan JY, Cheng L, et al. Isolation of anti-inflammatory fractions and compounds from the root of *Astragalus membranaceus*. *Phytother Res* 2013;27(4):581–587.
- [77] Wen L, Wei Q, Chen G, et al. Bioassay- and liquid chromatography/mass spectrometry-guided acetylcholinesterase inhibitors from *Picriafel-terrae*. *Pharmacogn Mag* 2013;9(Suppl 1):S25–S31.
- [78] Li YY, Tang XL, Jiang T, et al. Bioassay-guided isolation of neoclerodane diterpenoids from *Scutellaria barbata*. *J Asian Nat Prod Res* 2013;15(9):941–949.
- [79] Chen Q, Yang L, Zhang G, et al. Bioactivity-guided Isolation of antiosteoporotic compounds from *Ligustrum lucidum*. *Phytother Res* 2013;27(7):973–979.
- [80] Yan YM, Fang P, Yang MT, et al. Anti-diabetic nephropathy compounds from *Cinnamomum cassia*. *J Ethnopharmacol* 2015;165:141–147.
- [81] Chen J, Teng J, Ma L, et al. Flavonoids isolated from the flowers of *Limonium bicolor* and their in vitro antitumor evaluation. *Pharmacogn Mag* 2017;13(50):222–225.
- [82] Li X, Wang W, Fan Y, et al. Anticancer efficiency of cycloartane triterpenoid derivatives isolated from *Cimicifuga yunnanensis* Hsiao on triple-negative breast cancer cells. *Cancer Manag Res* 2018;10:6715–6729.
- [83] Zhang H, Jiang JM, Han L, et al. Uncariitannin, a polyphenolic polymer from *Uncaria gambier*, attenuates *Staphylococcus aureus* virulence through an MgrA-mediated regulation of alpha-hemolysin. *Pharmacol Res* 2019;147:104328.
- [84] Zhang D, Zhang S, Jiang K, et al. Bioassay-guided isolation and evaluation of anti-osteoporotic polysaccharides from *Morinda officinalis*. *J Ethnopharmacol* 2020;261:113113.
- [85] Shinichi T. “serumpharmacology” and “serum pharmacology” - the new world of the determination of blood concentration in Kambo. *TDM Res* 1988;5:54.
- [86] Kano Y, Wang XJ, Shirakawa J, et al. Pharmacological properties of galenic preparations (IX,X) pharmacokinetics study of 6,7-dimethylesculetin in rats. *J Tradit Med* 1994;11(3):176–180.
- [87] Wang XJ, Sun H, Zhu DM, et al. Chemical analysis of Yinchen Wuling powder. *National Sympos Chin Patent Med* 1994;1(1):221.
- [88] Wang XJ, Sun WJ, Zhang N, et al. Isolation and identification of constituents absorbed into blood after oral administration of Liuwei Dihuang Pill. *Chin J Nat Med* 2007;5(4):277–280.
- [89] Xijun W, Ning Z, Hui S, et al. Study on serum medicinal chemistry of liuwe dihuang pills. *Chin Nat Med* 2004;2(4):219–222.
- [90] Yan GL, Sun H, Zhang AH, et al. Progress of serum pharmacology of traditional Chinese medicine and further development of its theory and method. *China J Chin Mat Med* 2015;40(17):3406–3412.
- [91] Wang X, Sun W, Sun H, et al. Analysis of the constituents in the rat plasma after oral administration of Yin Chen Hao Tang by UPLC/Q-TOF-MS/MS. *J Pharm Biomed Anal* 2008;46(3):477–490.
- [92] Wang XJ. Serum Pharmacology of Yin Chen Hao Decoction. *Serum Pharmacology of Traditional Chinese Medicine*. Beijing: Science Press; 2010:72–160.
- [93] Zhang A, Sun H, Wang X, et al. Simultaneous in vivo RP-HPLC-DAD quantification of multiple-component and drug-drug interaction by pharmacokinetics, using 6,7-dimethylesculetin, geniposide and rhein as examples. *Biomed Chromatogr* 2012;26(7):844–850.
- [94] Bioanalysis Zone. Sutton CE. Three’s a crowd: but could three be better than one. Available from: <https://www.bioanalysis-zone.com/interview-with-maureenhighkin-on-mass-spectrometry-analysis-to-assist-discovery-of-sphingosine-kinase-inhibitors/>. Accessed March 2, 2021.
- [95] Huang L, Zhao H, Huang B, et al. *Acanthopanax senticosus*: review of botany, chemistry and pharmacology. *Pharmazie* 2011;66(2):83–97.
- [96] Wang XJ. Serum pharmacology of Ci Wu Jia. *Serum Pharmacology of Traditional Chinese Medicine*. Beijing: Science Press; 2010:395–416.
- [97] Jiang P, Liu R, Dou S, et al. Analysis of the constituents in rat plasma after oral administration of Shexiang Baoxin pill by HPLC-ESI-MS/MS. *Biomed Chromatogr* 2009;23(12):1333–1343.
- [98] Su S, Guo J, Duan JA, et al. Ultra-performance liquid chromatography-tandem mass spectrometry analysis of the bioactive components and their metabolites of Shaofu Zhuyu decoction active extract in rat plasma. *J Chromatogr B Analyt Technol Biomed Life Sci* 2010;878(3–4):355–362.
- [99] Wang XJ, Zhang AH, Sun H, et al. Serum pharmacology of Yin Chen Si Ni Decoction. *Serum Pharmacology of Traditional Chinese Medicines*. Beijing: Science Press; 2017:231–301.

- [100] Lv YH, Zhang X, Liang X, et al. Characterization of the constituents in rat biological fluids after oral administration of Fufang Danshen tablets by ultra-performance liquid chromatography/quadrupole time-of-flight mass spectrometry. *J Pharm Biomed Anal* 2010;52(1):155–159.
- [101] Hu Y, Jiang P, Wang S, et al. Plasma pharmacology based approach to screening potential bioactive components in Huang-Lian-Jie-Du-Tang using high performance liquid chromatography coupled with mass spectrometric detection. *J Ethnopharmacol* 2012;141(2):728–735.
- [102] Miao WJ, Wang Q, Bo T, et al. Rapid characterization of chemical constituents and rats metabolites of the traditional Chinese patent medicine Gegen-Qinlian-Wan by UHPLC/DAD/qTOF-MS. *J Pharm Biomed Anal* 2013;72:99–108.
- [103] Yan GL, Zhang AH, Sun H, et al. An effective method for determining the ingredients of Shuanghuanglian formula in blood samples using high-resolution LC-MS coupled with background subtraction and a multiple data processing approach. *J Sep Sci* 2013;36(19):3191–3199.
- [104] Wang H, Sun H, Zhang A, et al. Rapid identification and comparative analysis of the chemical constituents and metabolites of Phellodendri amurensis cortex and Zhibai dihuang pill by ultra-performance liquid chromatography with quadrupole TOF-MS. *J Sep Sci* 2013;36(24):3874–3882.
- [105] Wang H, Yan G, Zhang A, et al. Rapid discovery and global characterization of chemical constituents and rats metabolites of Phellodendri amurensis cortex by ultra-performance liquid chromatography-electrospray ionization/quadrupole-time-of-flight mass spectrometry coupled with pattern recognition approach. *Analyst* 2013;138(11):3303–3312.
- [106] Wang P, Yin QW, Zhang AH, et al. Preliminary identification of the absorbed bioactive components and metabolites in rat plasma after oral administration of Shaoyao-Gancao decoction by ultra-performance liquid chromatography with electrospray ionization tandem mass spectrometry. *Pharmacogn Mag* 2014;10(40):497–502.
- [107] Geng JL, Dai Y, Yao ZH, et al. Metabolites profile of Xian-Ling-Gu-Bao capsule, a traditional Chinese medicine prescription, in rats by ultra performance liquid chromatography coupled with quadrupole time-of-flight tandem mass spectrometry analysis. *J Pharm Biomed Anal* 2014;96:90–103.
- [108] Han Y, Wu FF, Zhang AH, et al. Characterization of multiple constituents in rat plasma after oral administration of Shengmai San using ultra-performance liquid chromatography coupled with electrospray ionization/quadrupole-time-of-flight high-definition mass spectrometry. *Anal Methods* 2015;7(3):830–837.
- [109] Liu C, Zhang AH, Han Y, et al. Ultra-high performance liquid chromatography coupled with time-of-flight mass spectrometry screening and analysis of potential bioactive compounds from traditional Chinese medicine Kai-Xin-San, using a multivariate data processing approach and the MetaboLynx tool. *RSC Adv* 2015;5(1):85–92.
- [110] Li XN, Zhang A, Sun H, et al. Rapid discovery of absorbed constituents and metabolites in rat plasma after the oral administration of Zi Shen Wan using high-throughput UHPLC-MS with a multivariate analysis approach. *J Sep Sci* 2016;39(24):4700–4711.
- [111] Zhang Y, Zhang A, Zhang Y, et al. Application of ultra-performance liquid chromatography with time-of-flight mass spectrometry for the rapid analysis of constituents and metabolites from the extracts of *Acanthopanax senticosus* Harms Leaf. *Pharmacogn Mag* 2016;12(46):145–152.
- [112] Li X, Sun H, Zhang A, et al. High-throughput LC-MS method for the rapid characterization of multiple chemical constituents and metabolites of Da-Bu-Yin-Wan. *J Sep Sci* 2017;40(21):4102–4112.
- [113] Yao ZH, Qin ZF, He LL, et al. Identification, bioactivity evaluation and pharmacokinetics of multiple components in rat serum after oral administration of Xian-Ling-Gu-Bao capsule by ultra performance liquid chromatography coupled with quadrupole time-of-flight tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 2017;1041–1042:104–112.
- [114] Wang XJ, Liu JH, Zhang AH, et al. Serum pharmacology of TCM screening the bioactive components from Moutan Cortex. *Serum Pharmacology of Traditional Chinese Medicine*. London: Elsevier; 2017:287–302.
- [115] Wang XJ, Wu FF, Sun H, et al. Serum pharmacology of TCM for screening the active ingredients from Wen-Xin Formulae. *Serum Pharmacology of Traditional Chinese Medicine*. London: Elsevier; 2017:73–101.
- [116] Wang XJ, Liu JH, Zhang AH, et al. Systematic characterization of the absorbed components of *Acanthopanax senticosus* Stem. *Serum Pharmacology of Traditional Chinese Medicine*. London: Elsevier; 2017:313–336.
- [117] Wei M, Liu Y, Pi Z, et al. Systematically characterize the anti-Alzheimer's disease mechanism of Lignans from *S. chinensis* based on in vivo ingredient analysis and target-network pharmacology strategy by UHPLC-Q-TOF-MS. *Molecules* 2019;24(7):1203.
- [118] Dou XX, Lin S, Tian XH, et al. Systematic characterization of the chemical constituents in vitro and prototypes in vivo of Dingkun Dan using ultra-high-performance liquid chromatography quadrupole time-of-flight mass spectrometry combined with the UNIFI software. *Biomed Chromatogr* 2020;34(10):e4914.
- [119] Zhang J, Yin Y, Xu Q, et al. Integrated serum pharmacology and investigation of the anti-gastric ulcer effect of Zuojin pill in rats induced by ethanol. *Pharm Biol* 2022;60(1):1417–1435.
- [120] Yin FT, Zhou XH, Kang SY, et al. Prediction of the mechanism of Dachengqi Decoction treating colorectal cancer based on the analysis method of “into serum components-action target-key pathway”. *J Ethnopharmacol* 2022;293:115286.
- [121] Zhang AH, Sun H, Yan GL, et al. Chinmedomics: a powerful approach integrating metabolomics with serum pharmacology to evaluate the efficacy of Traditional Chinese Medicine. *Engineering (Beijing)* 2019;5(1):132–149.
- [122] Liu SB, Lu SW, Sun H, et al. Deciphering the Q-markers of nourishing kidney-yin of Cortex Phellodendri amurensis from ZhibaiDihuang pill based on Chinmedomics strategy. *Phytomedicine* 2021;91:153690.
- [123] Wei WF, Sun H, Liu SB, et al. Targets and effective constituents of ZhiziBaipi Decoction for treating damp-heat jaundice syndrome based on Chinmedomics coupled with UPLC-MS/MS. *Front Pharmacol* 2022;13:857361.
- [124] Bian L, Tang J, Zi M, et al. Discussions on Symptomatic Assessment in Clinical Evaluation of FGIDs. *Modernization of Traditional Chinese Medicine and Materia Medica-World Science and Technology* 2020;22(10):3636–3639.
- [125] Wang XJ. Methodology for systematic analysis of in vivo efficacy material base of traditional Chinese medicine--Chinmedomics. *China J Chin Mat Med* 2015;40(1):13–17.
- [126] Wishart DS. Emerging applications of metabolomics in drug discovery and precision medicine. *Nat Rev Drug Discov* 2016;15(7):473–484.
- [127] Wang X, Zhang A, Sun H. Future perspectives of Chinese medical formulae: chinmedomics as an effector. *OMICs* 2012;16(7-8):414–421.
- [128] Ren JL, Yang L, Qiu S, et al. Efficacy evaluation, active ingredients, and multitarget exploration of herbal medicine. *Trends Endocrinol Metab* 2023;34(3):146–157.
- [129] Wang XJ. Inside view. *Nature* 2015;5128(7582).
- [130] Rahman J, Rahman S. Mitochondrial medicine in the omics era. *Lancet* 2018;391(10139):2560–2574.
- [131] Wang X. Chinmedomics-preface. *Chinmedomics*. Boston: Academic Press; 2015:xxi–xxii.
- [132] Wang XJ. *Research Progress of Chinmedomics (Volume 2016)*. Beijing: Science Press; 2016.
- [133] Wang XJ. *Research Progress of Chinmedomics (Volume 2017)*. Beijing: Science Press; 2017.
- [134] Wang XJ. *Research Progress of Chinmedomics (Volume 2018)*. Beijing: Science Press; 2018.
- [135] Wang XJ. *Research Progress of Chinmedomics (Volume 2019)*. Beijing: Science Press; 2019.
- [136] Wang XJ. *Research Progress of Chinmedomics (Volume 2020)*. Beijing: Science Press; 2020.
- [137] Wang XJ. *Research Progress of Chinmedomics (Volume 2021)*. Beijing: Science Press; 2021.
- [138] Cao H, Zhang A, Zhang FM, et al. Ultra-performance liquid chromatography tandem mass spectrometry combined with automated MetaboLynx analysis approach to screen the bioactive components and their metabolites in Wen-Xin-Formula. *Biomed Chromatogr* 2014;28(12):1774–1781.
- [139] Wang X, Zhang A, Zhou X, et al. An integrated chinmedomics strategy for discovery of effective constituents from traditional herbal medicine. *Sci Rep* 2016;6:18997.
- [140] Zhang AH, Liu Q, Zhao HW, et al. A research of chinmedomics on the pharmacodynamic basis of nanshi oral liquid and its mechanism behind the efficacy on kidney-yang deficiency syndrome. *Modernization of Traditional Chinese Medicine and Materia Medica-World Science and Technology* 2016;18(10):1670–1683.
- [141] Liu Q, Zhang A, Wang L, et al. High-throughput chinmedomics-based prediction of effective components and targets from herbal medicine AS1350. *Sci Rep* 2016;6:38437.

- [142] Liu Q, Zhao HW, Zhang AH, et al. Chinmedomics strategy to discover effective constituents and elucidate action mechanism of Nanshi capsule against kidney-yang deficiency syndrome. *China J Chin Mat Med* 2016;41(15):2901–2914.
- [143] Li XN, Zhang A, Wang M, et al. Screening the active compounds of Phellodendri Amurensis cortex for treating prostate cancer by high-throughput chinmedomics. *Sci Rep* 2017;7:46234.
- [144] Zhang AH, Yu JB, Sun H, et al. Identifying quality-markers from Shengmai San protects against transgenic mouse model of Alzheimer's disease using chinmedomics approach. *Phytomedicine* 2018;45:84–92.
- [145] Wang XJ, Zhang AH, Kong L, et al. Rapid discovery of quality-markers from Kaixin San using chinmedomics analysis approach. *Phytomedicine* 2019;54:371–381.
- [146] Sun H, Zhang AH, Yang L, et al. High-throughput chinmedomics strategy for discovering the quality-markers and potential targets for Yinchenhao decoction. *Phytomedicine* 2019;54:328–338.
- [147] Yang B, Han Y, Zhang QY, et al. Study on absorbed components of Aconitum kusnezoffi under Yunnan Baiyao compatibility in effect of activating blood circulation and removing blood stasis. *China J Chin Mat Med* 2019;44(15):3349–3357.
- [148] Gao X, Hu X, Zhang Q, et al. Characterization of chemical constituents and absorbed components, screening the active components of gelanxin capsule and an evaluation of therapeutic effects by ultra-high performance liquid chromatography with quadrupole time of flight mass spectrometry. *J Sep Sci* 2019;42(22):3439–3450.
- [149] Liu X, Zhou QG, Zhu XC, et al. Screening for potential active components of Fangji Huangqi Tang on the treatment of nephrotic syndrome by using integrated metabolomics based on “Correlations Between Chemical and Metabolic Profiles”. *Front Pharmacol* 2019;10:1261.
- [150] Cai H, Xu Y, Xie L, et al. Investigation on spectrum-effect correlation between constituents absorbed into blood and bioactivities of Baizhu Shaoyao San before and after processing on ulcerative colitis rats by UHPLC/Q-TOF-MS/MS coupled with gray correlation analysis. *Molecules* 2019;24(5):940.
- [151] Xiong H, Zhang AH, Zhao QQ, et al. Discovery of quality-marker ingredients of Panax quinquefolius driven by high-throughput chinmedomics approach. *Phytomedicine* 2020;74:152928.
- [152] Zhao Q, Gao X, Yan G, et al. Chinmedomics facilitated quality-marker discovery of Sijunzi decoction to treat spleen qi deficiency syndrome. *Front Med* 2020;14(3):335–356.
- [153] Kong L, Sun Y, Sun H, et al. Chinmedomics strategy for elucidating the pharmacological effects and discovering bioactive compounds from Keloixin against diabetic retinopathy. *Front Pharmacol* 2022;13:728256.
- [154] Wang Y, Yang L, Zhang X, et al. Quality marker discovery of Danggui Jianzhong decoction for treating primary dysmenorrhoea based on chinmedomics strategy. *Phytomedicine* 2023;115:154724.
- [155] Cao HX, Sun H, Jiang XG, et al. Comparative study on the protective effects of Yinchenhao Decoction against liver injury induced by alpha-naphthylisothiocyanate and carbon tetrachloride. *Chin J Integr Med* 2009;15(3):204–209.
- [156] Zhang A, Sun H, Qiu S, et al. Advancing drug discovery and development from active constituents of yinchenhao tang, a famous traditional Chinese medicine formula. *Evid Based Complement Alternat Med* 2013;2013:257909.
- [157] Wang X, Zhang A, Han Y, et al. Urine metabolomics analysis for biomarker discovery and detection of jaundice syndrome in patients with liver disease. *Mol Cell Proteomics* 2012;11(8):370–380.
- [158] Xiong H, Zhang A-H, Guo Y-J, et al. A clinical and animal experiment integrated platform for small-molecule screening reveals potential targets of bioactive compounds from a herbal prescription based on the therapeutic efficacy of Yinchenhao Tang for jaundice syndrome. *Engineering* 2021;7(9):1293–1305.
- [159] Fang H, Zhang A, Yu J, et al. Insight into the metabolic mechanism of scoparone on biomarkers for inhibiting Yanghuang syndrome. *Sci Rep* 2016;6:37519.
- [160] Liu XY, Zhang AH, Fang H, et al. Serum metabolomics strategy for understanding the therapeutic effects of Yin-Chen-Hao-Tang against Yanghuang syndrome. *RSC Adv* 2018;8(14):7403–7413.
- [161] Sun H, Zhang AH, Song Q, et al. Functional metabolomics discover pentose and glucuronate interconversion pathways as promising targets for Yang Huang syndrome treatment with Yinchenhao Tang. *RSC Adv* 2018;8(64):36831–36839.
- [162] Sun H, Yang L, Li MX, et al. UPLC-G2Si-HDMS untargeted metabolomics for identification of metabolic targets of Yin-Chen-Hao-Tang used as a therapeutic agent of dampness-heat jaundice syndrome. *J Chromatogr B Analyt Technol Biomed Life Sci* 2018;1081–1082:41–50.
- [163] Fang H, Zhang A, Zhou X, et al. Study on the target of intervention of Genipine side in Yanghuang Syndrome based on metabolic regulation pathway. *Modernization of Traditional Chinese Medicine and Materia Medica-World Science and Technology* 2016;18(10):1697–1708.
- [164] Nie Q, Chen H, Hu J, et al. Dietary compounds and traditional Chinese medicine ameliorate type 2 diabetes by modulating gut microbiota. *Crit Rev Food Sci Nutr* 2019;59(6):848–863.
- [165] Jia Q, Wang L, Zhang X, et al. Prevention and treatment of chronic heart failure through traditional Chinese medicine: Role of the gut microbiota. *Pharmacol Res* 2020;151:104552.
- [166] Zhao H, He M, Zhang M, et al. Colorectal cancer, gut microbiota and Traditional Chinese Medicine: a systematic review. *Am J Chin Med* 2021;49(4):805–828.
- [167] Harvey AL, Edrada-Ebel R, Quinn RJ. The re-emergence of natural products for drug discovery in the genomics era. *Nat Rev Drug Discov* 2015;14(2):111–129.
- [168] Yu Z, Liao J, Chen Y, et al. Single-cell transcriptomic map of the human and mouse bladders. *J Am Soc Nephrol* 2019;30(11):2159–2176.
- [169] Giudice G, Petsalaki E. Proteomics and phosphoproteomics in precision medicine: applications and challenges. *Brief Bioinform* 2019;20(3):767–777.

**How to cite this article:** Ge N, Yan G, Sun H, Yang Le, Kong L, Sun Y, Han Y, Zhao Q, Kang S, Wang X. Version updating of strategy for drug discovery based on effective constituents of traditional Chinese medicines. *Acupunct Herb Med* 2023;3(3):158–179. doi: 10.1097/HM9.000000000000071