

Comprehensive profiling of Lingzhihuang capsule by liquid chromatography coupled with mass spectrometry-based molecular networking and target prediction

Mengliang Huang¹, Sijia Yu², Qing Shao², Hao Liu², Yingchao Wang², Hongzhang Chen³, Yansheng Huang³, Yi Wang^{1,2,*}

Abstract

Objective: Lingzhihuang capsule (LZHC) is a natural product that consists of 10 commonly used medicinal plants, and it is used in traditional Chinese medicine to promote people's overall health. Previously, LZHC was successfully used as adjuvant therapy for treating patients with cancer. However, the chemical constituents of LZHC and their potential biological functions remain unclear. The aim of this study is to investigate the major bioactive compounds in LZHC and predict their pharmacological targets.

Methods: The LZHC constituents were putatively identified by ultra-high performance liquid chromatography coupled with time-of-flight mass spectrometry combined with mass spectrometry-based molecular networking. The targets were predicted using SwissTargetPrediction software, and the associated gene ontology and Kyoto encyclopedia of genes and genomes pathways were analyzed using the Database for Annotation, Visualization, and Integrated Discovery. The mass spectrometry-based molecular network and compound-target-pathway network were constructed using Cytoscape 3.8.0 software.

Results: We putatively identified 94 compounds of LZHC by mass spectrometry-based molecular networking, including triterpene (the main structural type) and other clusters (ie, flavonoids and organic acids). Our results suggested that multiple pivotal targets were regulated by LZHC, including tumor necrosis factor, nitric oxide synthase 2, glucocorticoid receptor, estrogen receptor, 3-oxo-5- α -steroid 4-dehydrogenase 2, prostaglandin e2 receptor ep4 subtype, estrogen receptor beta, phosphatidylinositol 4,5-bisphosphate 3-kinase catalytic subunit alpha isoform, mitogen-activated protein kinase 3, and rac-alpha serine, which are related to signal transduction, positive regulation of transcription from RNA polymerase II promoters, oxidation-reduction processes, inflammatory responses, and other biological processes. Functional annotation of those targets suggested that several signaling pathways may be regulated by LZHC, such as cancer-related proteoglycans, the PI3K-Akt-signaling pathway, and the cAMP-signaling pathway.

Conclusions: Our findings reveal the chemical constituents of LZHC and provided scientific support for the efficacy of LZHC in terms of immune regulation, anti-aging, and tumor suppression.

Keywords: Lingzhihuang capsule, Molecular networking, Network pharmacology, UPLC-Q-TOF/MS

Graphical abstract: <http://links.lww.com/AHM/A12>

Introduction

Lingzhihuang capsule (LZHC) is a natural product used to promote human health that consists of *Ganoderma lucidum*, *Atractylodes macrocephala*, *Polygonatum sibiricum*, *Lycium barbarum*, *Coix lacryma-jobi*, *Poria cocos*, *Ophiopogon japonicus*, *Cuscuta chinensis*, *Schisandra chinensis*, and *Glycyrrhiza uralensis* Fisch. Among these medicinal herbs, previous pharmacological data have suggested that *G. lucidum*, *G. uralensis* Fisch, *C. chinensis*, and *P. cocos* exert anti-inflammatory, antitumor, and anti-aging effects^[1-3]. For example, they have

demonstrated primary or auxiliary effects against gastric and throat cancer^[4], acute pancreatitis^[5], breast cancer^[6], liver disease^[7], diabetes^[8], dementia^[9], and other diseases. Previously, LZHC was recommended for middle-aged or elderly people with a weak physique, and for patients with cancer. Data from a previous report indicated that combined use of Yanggujian and LZHC was more beneficial when managing patients with *qi*-deficiency syndrome^[10]. However, the chemical composition and downstream targets of LZHC remain elusive, and little scientific evidence is available.

¹ State Key Laboratory of Component-Based Chinese Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin, China;

² Pharmaceutical Informatics Institute, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou, China; ³ Infinitus (China) Company, Ltd., Guangzhou, China

* Corresponding author. Yi Wang, State Key Laboratory of Component-based Chinese Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin 301617, China; Pharmaceutical Informatics Institute, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou 310058, China, E-mail: zjuwangyi@zju.edu.cn.

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Liquid chromatography-coupled with mass spectrometry (LC-MS) has become a powerful tool for revealing the complex chemical compositions of herbal medicines. For example, Fu et al.^[11] analyzed the chemical constituents of Xuefu Zhuyu decoction by ultra-performance liquid chromatography (UPLC) coupled to ion-trap time-of-flight (TOF)-MS and inferred 34 compounds. Likewise, Yang et al.^[12] studied the chemical constituents of *Glycyrrhizae Radix et Rhizoma* in *Pinelliae Rhizoma Praeparatum* (PRP) by UPLC coupled to quadrupole (Q)-TOF-MS/MS, and identified 15 chemical compounds present in PRP licorice. Yang et al.^[13] also used UPLC-Q-TOF/MS to study Suanzaoren decoction, from which 22 compounds were identified or inferred. Moreover, MS-based molecular networking (MN) is an emerging tool used to accelerate the pace of analyzing natural plants. Wang et al.^[14] combined MS-MN with a virtual screening-affinity MS-screening technique to examine the active components from 12 anti-cancer Chinese medicinal herbs, and two compounds were identified that regulate the activity of the Ras protein. This method significantly improved the throughput of ligand screening and the accuracy of the screening results. Yang et al.^[15] used LC-high resolution MS combined with MS-MN to study the chemical constituents and pharmacological mechanism of Qingfei Paidu decoction (a traditional Chinese medicine prescription that has been used for clinical treatment of coronavirus disease 2019) and 129 compounds were identified and classified.

Those applications suggest that integrated LC-MS analysis and MN can be used to efficiently determine the complex chemical composition of LZHC. Network pharmacology^[16] is a new strategy that combines pharmacology with network analysis, which was originally proposed by Hopkins in 2007. The strategy involves searching for the potentially active ingredients of the mixture through network databases, such as the Traditional Chinese Medicine Systems Pharmacology (TCMSP) database, followed by predicting their targets and relevant diseases, as well as signal-pathway enrichment analysis to establish network relationships or drug-gene-target-disease interactions. The possible effective components of drugs and their molecular mechanisms can be predicted by the network relationship, which provides a basis for further research and applications. This method is especially suitable for studying compound prescriptions used in traditional Chinese medicine for which the pharmacological mechanism involves multiple components, targets, and pathways. Therefore, network pharmacology has become a popular approach for studying the molecular mechanisms of traditional Chinese medicines in recent years^[17–20].

In this study, the chemical components and potential biological mechanisms of LZHC were comprehensively analyzed by UPLC-Q-TOF/MS combined with network pharmacology. MN was employed to process the LC-MS data and classify the identified compounds into different categories, based on MS/MS similarity. Furthermore, the potential targets of the major constituents were predicted by network pharmacology. Functional-annotation results indicated that the pivotal targets regulated by LZHC include tumor necrosis factor (TNF), nitric oxide synthase 2 (NOS2), glucocorticoid receptor (NR3C1), and rac-alpha serine (AKT1), which suggested that multiple pathways associated with cancer, aging, and inflammation may be regulated by LZHC. Our findings provided scientific support for the

efficacy of LZHC in terms of immune regulation, anti-aging, and combinatory therapy against tumors.

Materials and methods

Materials and reagents

The reference compounds quercetin, hyperoside, liquiritin apioside, licochalcone A, glycyrrhizic acid, and liquiritigenin were purchased from Shanghai Yuanye Biotechnology Co., Ltd. (Shanghai, China). Chlorogenic acid, isoliquiritin apioside, 5-hydroxymethylfurfural, 3,4-dihydroxybenzaldehyde, liquiritin, isoliquiritin, kaempferol, ophiopogonin D, methyllophiopogonanone A, and methyllophiopogonanone B were purchased from Shanghai Ronghe Medical Technology Development Co., Ltd. (Shanghai, China). 20-Hydroxyganoderic acid G; gano-lactone B; ganoderic acid I; ganoderlactone D; ganoderenic acid C; ganoderic acid C2, (3 β ,7 β ,12 β ,20Z)-3,7,12-trihydroxy-11,15,23-trioxo-lanost-8,20-dien-26-oic acid; ganoderic acid C6; ganoderic acid G; ganoderenic acid B; ganoderic acid N; ganoderic acid B; ganoderic acid LM2; ganoderenic acid A; chol-8-en-24-oic acid,7,15-dihydroxy-4,4,14-trimethyl-3,11-dioxo-, (5 α); ganoderenic acid E; ganoderic acid A; ganoderic acid H, 20(21)-dehydro-lucidinic acid A; deacetyl ganoderic acid F; ganoderic acid D2; 12-hydroxyganoderic acid D; lucidinic acid A; ganoderenic acid D; ganoderic acid D; and ganoderic acid C1 were purchased from Chengdu Pusi Biotechnology Co., Ltd. (Shanghai, China). LZHCs were obtained from Wuxianji Co., Ltd. (Guangzhou, China). High-performance liquid chromatography (HPLC)-grade formic acid was obtained from Roe Scientific (Newark, United States). HPLC-grade acetonitrile and methanol were obtained from Merck (Darmstadt, Germany). Deionized water was prepared with an ELGA PURELAB flex system (ELGA LabWater, United Kingdom). Analytical-grade n-butanol was from Hangzhou Gaojing Jingxi Chemical Co., Ltd. (Zhejiang, China).

Sample preparation

The contents of LZHC (10 g) were refluxed for 2 h in a water bath with 150 mL water and 40 mL water-saturated n-butanol, and then cooled to room temperature (10°C–30°C) for stratification. The upper layer was collected, subjected to rotary evaporation, and re-dissolved in 70% methanol. The sample was centrifuged at 10,000 rpm for 10 min, and the supernatant was taken for LC-MS analysis.

UPLC-Q-TOF/MS analysis

An Acquity UPLC system (Waters, Milford, United States) coupled with a Triple TOF SYNAPT XS system (Waters, Milford, United States) was employed for chemical identification.

Chromatographic separation was performed on a Waters ACQUITYUPLC HSS T₃ column (100 mm \times 2.1 mm, 1.8 μ m) at a column temperature of 30°C with mobile phase A (water containing 0.05% formic acid; v/v) and mobile phase B (100% acetonitrile). The flow rate was 0.25 mL/min and linear gradient elution was performed under the following optimized conditions: 0 to 8 min, 2% to 2% B; 8 to 15 min, 2% to 15% B; 15 to 19 min, 15% to 15% B; 19 to 30 min, 15% to 25% B; 30 to

40 min, 25% to 27% B; 40 to 52 min, 27% to 29% B; 52 to 75 min, 29% to 35% B; and 75 to 95 min, 35% to 100% B. The detection wavelength range was set to 190 nm to 400 nm, and the injection volume was 3 μ L.

The analysis was carried out in negative-ion mode using the following parameters: scan range, mass/charge (m/z) ratio of 50 to 1,200 Da; capillary voltage, 2.5 kV; cone gas flow, 50 L/h; desolvation gas flow, 700 L/h; source temperature, 140°C; desolvation temperature, 20°C; collision energy, low-mass (50 Da) CE ramp (8V–12 V), and high-mass (1,200 Da) CE ramp (60 V–90 V). The MS data was collected in data-dependent analysis mode.

Raw MS data preprocessing

Raw MS data was first preprocessed with Progenesis QI 4.1, Nonlinear Dynamics)^[21]. The picking limits were set using a minimum intensity of 15,000 and a minimum width of 0.5 min. Then, the MSP data file and

corresponding CSV metadata files (containing compound intensities and annotations) were exported.

MN of MS data

MS-MN data for LZHC was generated using the on-line Feature-Based Molecular Networking (FBMN) workflow on the Global Natural Product Social Molecular Networking (GNPS) platform^[22–24] (<http://gnps.ucsd.edu>). Data processing was performed using Progenesis QI software. The parameter settings used to construct the molecular network are shown in Table 1, and the other parameters used were the default parameters recommended on the website. The constructed MN was visualized using Cytoscape software 3.8.0.

Compound target prediction for LZHC

Target information for the tentatively identified compounds was collected from the Swiss Target Prediction database^[25] (<http://www.swisstargetprediction.ch/>). The selection criteria for the compound targets were the top 10 targets with probability >0. The protein-target information obtained was standardized using the UniProt database (<https://www.uniprot.org/>), and the gene symbols were converted into official HUGO Gene Nomenclature Committee symbols (<https://www.genenames.org/>).

Duplicate genes were deleted and the target list was uploaded to the Database for Annotation, Visualization, and Integrated Discovery (DAVID; <https://david.ncifcrf.gov/>). Official gene symbols were selected as identifiers, annotated using *Homo sapiens* as the species, and outputted using “gene list” as the list type. Next, the main LZHC biological processes and metabolic pathways were analyzed by carrying out an enrichment analysis.

LZHC compound-target-pathway networks were generated using Cytoscape software 3.8.0. The built-in software tools were used to analyze different network topology parameters of the targets, including the degree,

Table 1
Parameters used to construct the molecular network.

Workflow selection	Parameter	Value
Basic options	Quantification table source	Progenesis QI
	Precursor ion mass tolerance	0.01
	Fragment ion mass tolerance	0.01
Advanced network options	Min Pairs Cos	0.8
	Network TopK	10
	Minimum matched fragment ions	4
	Maximum connected component size	100
	Maximum shift between precursors	500
Advanced filtering options	Advanced filtering options	Do not filter
	Filter peaks in 50 Da Window	Do not filter
Advanced library search options	Library search min matched peaks	6
	Score threshold	0.7

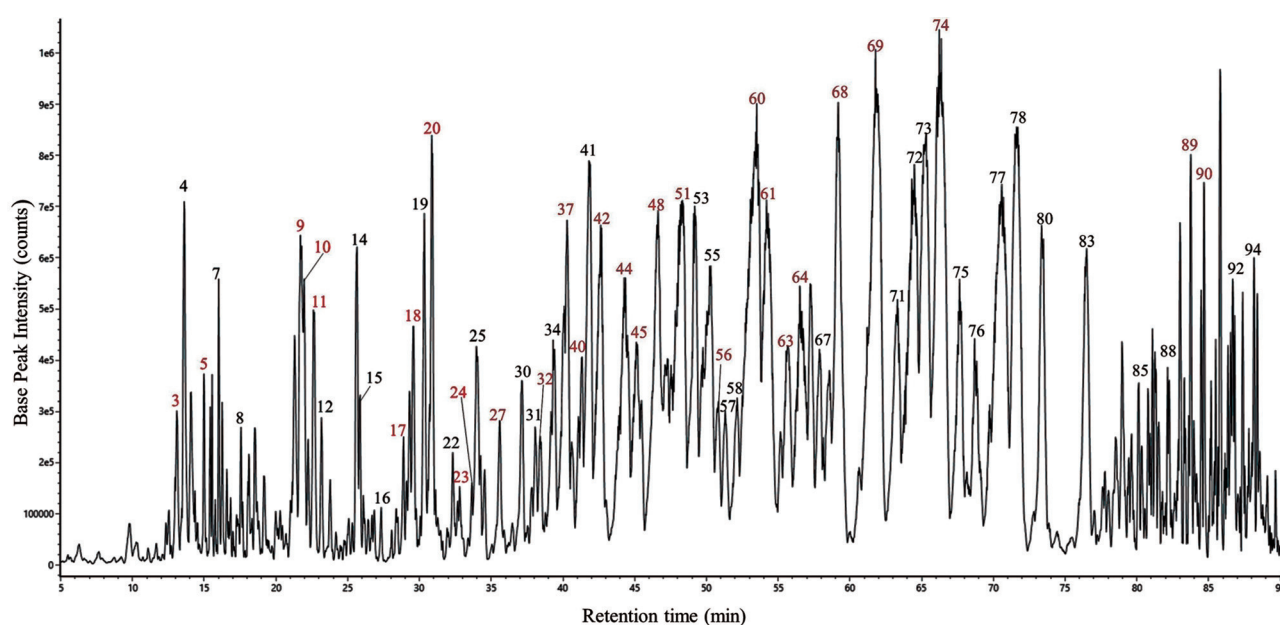


Figure 1. Mass spectrometry chromatogram of an Lingzhihuang capsule obtained in negative-ion mode. Number 1–94 represent compounds identified. The red numbers represent compounds identified based on reference compound matching.

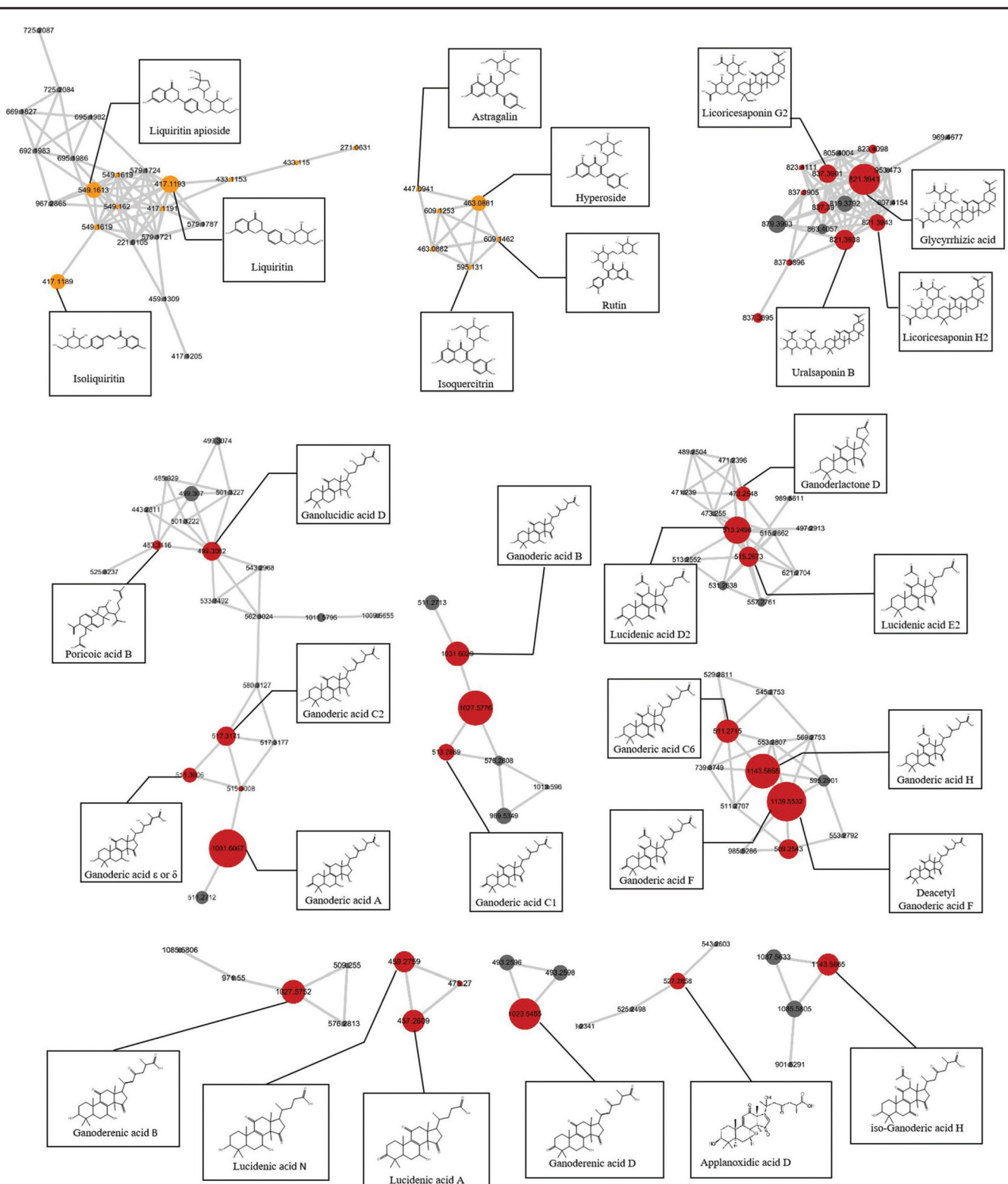


Figure 3. Representative molecular networking clusters of a Lingzhihuang capsule. In the network, the red nodes represent triterpenoids and the orange nodes represent flavonoids, each node represents the mass spectrometry information of a compound, and hub nodes are represented with a larger node size. The line between nodes represents the correlation between the mass spectrometry spectrum of two compounds. The correlation can be expressed by the thickness of the line.

herbs of these chemical constituents were further assigned using the TCMSP platform and information in the literature. The compound-identification results are shown in Table S1, <http://links.lww.com/AHM/A6>.

The MN was visualized using Cytoscape 3.8.0 and was found to contain 954 nodes and 1,174 edges (Figure 2).

The node size corresponds to the ion intensity, and the node color represents the structural category of the compound. The represent compounds were annotated on the nodes, and triterpene was identified as the main structural type, and the other clusters were mainly flavonoids and organic acids (Figure 3).

with reference compounds. Ninety-four chemical constituents were putatively identified, including triterpenes, flavonoids, and organic acids. The results of compound target prediction and pathway enrichment showed that LZHC may play functional roles in immune regulation, anti-aging, and combination therapy against tumors by regulating potential targets such as TNF, NOS2, NR3C1, ESR1, and AKT1 and related signal pathways, such as the PI3K-Akt signaling pathway. These possible mechanisms have been rarely validated in experimental studies, although this topic merits further investigation.

Conflict of interest statement

The authors declare no conflict of interest.

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Author contributions

Mengliang Huang, Qing Shao, and Yi Wang designed the research. Mengliang Huang performed the experimental work. Mengliang Huang, Sijia Yu, and Hao Liu analyzed and visualized the data. Mengliang Huang, Sijia Yu, Hao Liu, Yingchao Wang, and Yi Wang participated in the preparation of the manuscript. Hongzhang Chen and Yansheng Huang provided the research materials. All authors proofread the paper and approved the final version of the manuscript.

Ethical approval of studies and informed consent

Not applicable.

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