






Review

# The Role of T Cells in Multiple Sclerosis: Molecular Mechanisms, Drivers, and Therapeutic Targets

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Academic Editor: Thomas Müller

Submitted: 27 October 2025 Revised: 10 January 2026 Accepted: 23 January 2026 Published: 16 March 2026

## Abstract

Multiple sclerosis is a chronic immune-mediated disease of the central nervous system, marked by demyelination, axonal damage, and progressive neurological decline. T lymphocytes—particularly CD4<sup>+</sup>, T helper (Th)1 and Th17 cells, as well as cytotoxic CD8<sup>+</sup> cells—play a pivotal role in initiating and sustaining central nervous system inflammation. Acute inflammation is driven by peripheral immune activation, while progressive disease reflects compartmentalized, smouldering inflammation within the central nervous system, dominated by CD8<sup>+</sup> T cells and microglia. A relative deficiency or dysfunction of regulatory T cells contributes to immune tolerance loss and ongoing neurodegeneration. Although T lymphocytes play a central role, the pathogenesis of multiple sclerosis involves a broader cellular network, including antigen-presenting cells, B lymphocytes, microglia, and astrocytes. While recent therapeutic strategies have increasingly focused on B lymphocytes, most disease-modifying therapies—and many emerging ones—exert at least partial effects by modulating T cell-mediated mechanisms. These insights underpin current T cell-targeted therapies and highlight unmet needs in multiple sclerosis.

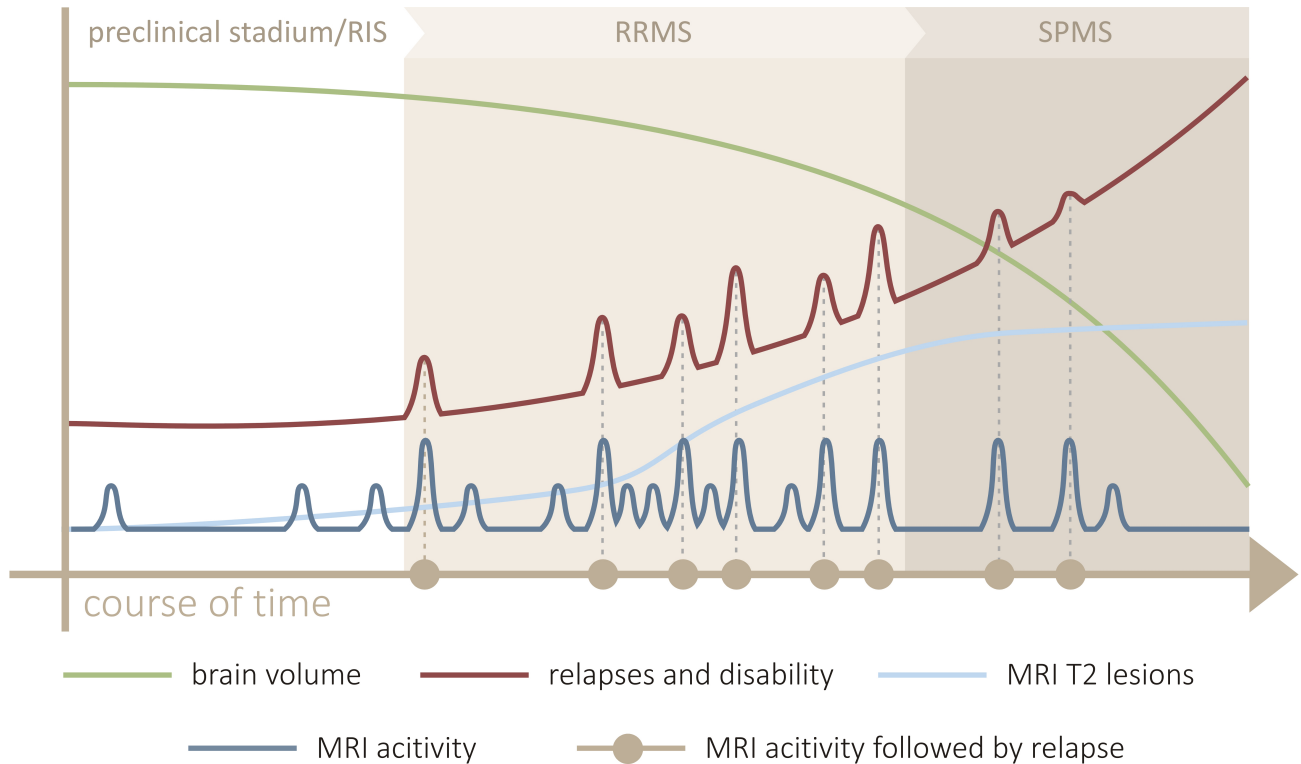
**Keywords:** multiple sclerosis; T cells; Th17; CD8 T cells; B cells; cytokines; molecular mechanisms; pathophysiology; smouldering inflammation; disease-modifying therapy

## 1. Introduction

Multiple sclerosis (MS) is a chronic demyelinating disease of the central nervous system (CNS) in whose pathogenesis, in addition to cell-mediated inflammatory activity, neurodegeneration also plays a role from the beginning. This disease most commonly affects young adults of productive age, with the average age at diagnosis around 30 years [1]. However, recent studies show a shift toward older age at diagnosis, with approximately 5% of patients being diagnosed in their sixth decade of life [2,3]. The prevalence in Europe and North America is estimated at around 111–300 cases per 100,000 inhabitants [4,5]. The aetiology of the disease remains unclear, but available evidence suggests a combination of genetic, epigenetic, and environmental factors (including Epstein-Barr virus (EBV) infection, vitamin D deficiency, smoking, and lifestyle factors) [6–10]. The influence of the gut microbiome and hormonal factors has also been described, with MS incidence being up to three times higher in women, while male sex is considered a negative prognostic factor [1,11].

Although the classification of MS based on clinical phenotypes is increasingly viewed as outdated from a pathogenetic perspective, it is still used in clinical practice, primarily for practical reasons. The most common disease course is relapsing-remitting MS (RRMS), characterized by alternating periods of flare-ups and relative stability. During a relapse, (multi)focal neurological dysfunction develops, which may even resolve completely. As the disease progresses, relapse frequency decreases, while neurological impairment, dependent on and independent of relapses (progression independent of relapse activity, PIRA), increases (Fig. 1). This is associated with advanced demyelination, accumulation of axonal and neuronal damage, and reduced neuronal connectivity [12]. Over time, the disease often transitions into a secondary progressive form. “Smouldering” inflammation behind an already closed blood-brain barrier (BBB) typically localizes at the edges of existing lesions and is mainly mediated by activated microglia. Magnetic resonance imaging (MRI) reveals slowly expanding lesions (SEs) and progressive brain atrophy. While the primary target of inflam-





**Fig. 1. Course of relapsing-remitting multiple sclerosis.** Although we divide MS into three stages - RIS, RRMS and SPMS, the biological course of the disease is continual. Inflammatory activity in the CNS leads to accumulation of T2-weighted lesions and later to axonal loss manifested by progressive brain atrophy. Only some inflammatory flare-ups are accompanied by a clinical relapse, while others are subclinical, but gradually contribute to slowly progressing disability. RIS - A subclinical stage of MS until the first relapse, but already with evident biological and radiological activity. RRMS - A clinical stage of MS associated with relapses, accumulation of T2-weighted lesions, brain atrophy and clinical disability. SPMS - A clinical stage of MS with minimum relapses, but dominant progressive axonal loss, brain atrophy and clinical disability independent of relapses. The line between RR and SP grade is not sharp. MS, multiple sclerosis; RIS, radiologically isolated syndrome; RRMS, relapsing-remitting multiple sclerosis; SPMS, secondary progressive multiple sclerosis; MRI, magnetic resonance imaging; CNS, central nervous system.

mation is the myelin sheath, the ultimate cause of permanent damage and PIRA is axonal loss. This is attributed to inflammation, accumulation of lesions with subsequent retrograde and anterograde degeneration, mitochondrial dysfunction and oxidative stress, iron accumulation in myelin and oligodendrocytes, ectopic meningeal lymphoid follicles, age-related neurodegeneration, and loss of functional reserves [13]. The degree of disability is also influenced by the development of neural network dysfunction and inadequate repair mechanisms [14]. A rarer form of MS (10–15% of cases) is primary progressive MS (PPMS), which is characterized by the gradual progression of symptoms from the onset, without relapses [4].

According to the currently most widely accepted model (“outside-in”), MS is considered a disease that begins with peripheral activation of an autoimmune process, which subsequently extends into the CNS. The interaction between infiltrating immune cells (T and B lympho-

cytes) and resident CNS cells (particularly microglia and astrocytes) plays a key role [15]. Primary activation of T lymphocytes occurs in the periphery via antigen-presenting cells (APCs). Activated CD4<sup>+</sup> T lymphocytes (especially T helper (Th)1 and Th17 subsets) express adhesion molecules and chemokine receptors (e.g., very late antigen-4 [VLA-4], lymphocyte function-associated antigen 1 [LFA-1], CC chemokine receptor 6 [CCR6]) that allow them to interact with BBB endothelial cells [16,17]. This interaction leads to endothelial activation, resulting in increased BBB permeability. Cytokines modulate blood–brain barrier properties in a context-dependent manner. Interleukin (IL)-17 and IL-22 can promote leukocyte trafficking by upregulating endothelial adhesion and altering the function of tight junctions. In contrast, cytokine interferon-gamma (IFN- $\gamma$ ) exhibits bidirectional effects that, depending on the cellular state and timing, may either amplify inflammation or contribute to barrier stabilisation. The role of IL-26 in BBB

regulation has emerging support but remains incompletely defined in MS. According to Broux *et al.* [18], IL-26 has a protective effect on the integrity of the BBB, induces a downregulation of pro-inflammatory and oxidative pathways in endothelial cells, concomitantly stimulating the upregulation of tight junction protein transcripts (Fig. 2). Once T lymphocytes infiltrate the CNS, they release cytokines (e.g., IFN- $\gamma$ , tumour necrosis factor-alpha [TNF- $\alpha$ ]) that amplify the expression of adhesion molecules on the endothelium, creating an inflammatory environment that attracts monocytes, B lymphocytes, additional T lymphocytes, and other effector cells. These cytokines are also trafficked via exosomes, which serve as critical mediators of intercellular communication and can modulate target cell gene expression in MS pathogenesis [19]. These additional immune cells cross the BBB and attack the myelin in the brain, forming multiple, irregularly distributed foci densely populated by phagocytic cells in perivascular areas called plaques [20]. Chronic inflammation and destruction of the myelin sheath in the brain and spinal cord white matter occur, accompanied by varying degrees of axonal loss. Even in white matter that appears normal morphologically and on MRI, many axons are already damaged in MS patients [21]. The potential for repair depends on the preservation of oligodendrocytes and, critically, on axonal integrity. Remyelination can restore conduction efficiency where axons remain intact, but once axons are lost, functional recovery is no longer achievable. Chronic inactive plaques are sharply demarcated without active myelin degradation.

T lymphocytes are pivotal drivers of relapse activity—particularly in RRMS. In contrast, disability accrual and progression reflect an integrated network involving B cells, innate immunity, and CNS-resident glia within compartmentalized inflammation [22]. The predominant lymphocyte type at all stages of MS is the cytotoxic CD8<sup>+</sup> T cell, which closely correlates with axonal damage [23]. Regulatory T lymphocytes (Tregs), which protect the brain from further immune-mediated injury, are less abundant [24]. This article aims to provide an up-to-date overview of the influence of T lymphocytes on MS pathophysiology and to highlight the mechanisms of treatment targeting T-cell modulation.

## 2. Genetic and Epigenetic Factors in the Etiopathogenesis of MS

Genome-wide association studies (GWAS) have brought about a fundamental shift in our understanding of genetic and epigenetic factors in the etiopathogenesis of MS. These studies have identified more than 200 single-nucleotide polymorphisms associated with genetic predisposition to MS, most of which are located on chromosome 6 in a region containing genes that regulate the immune response. However, the odds ratio (OR) associated with most of these variants is low (1.1–1.2) [25]. The largest group of genes identified in GWAS is involved in either the sig-

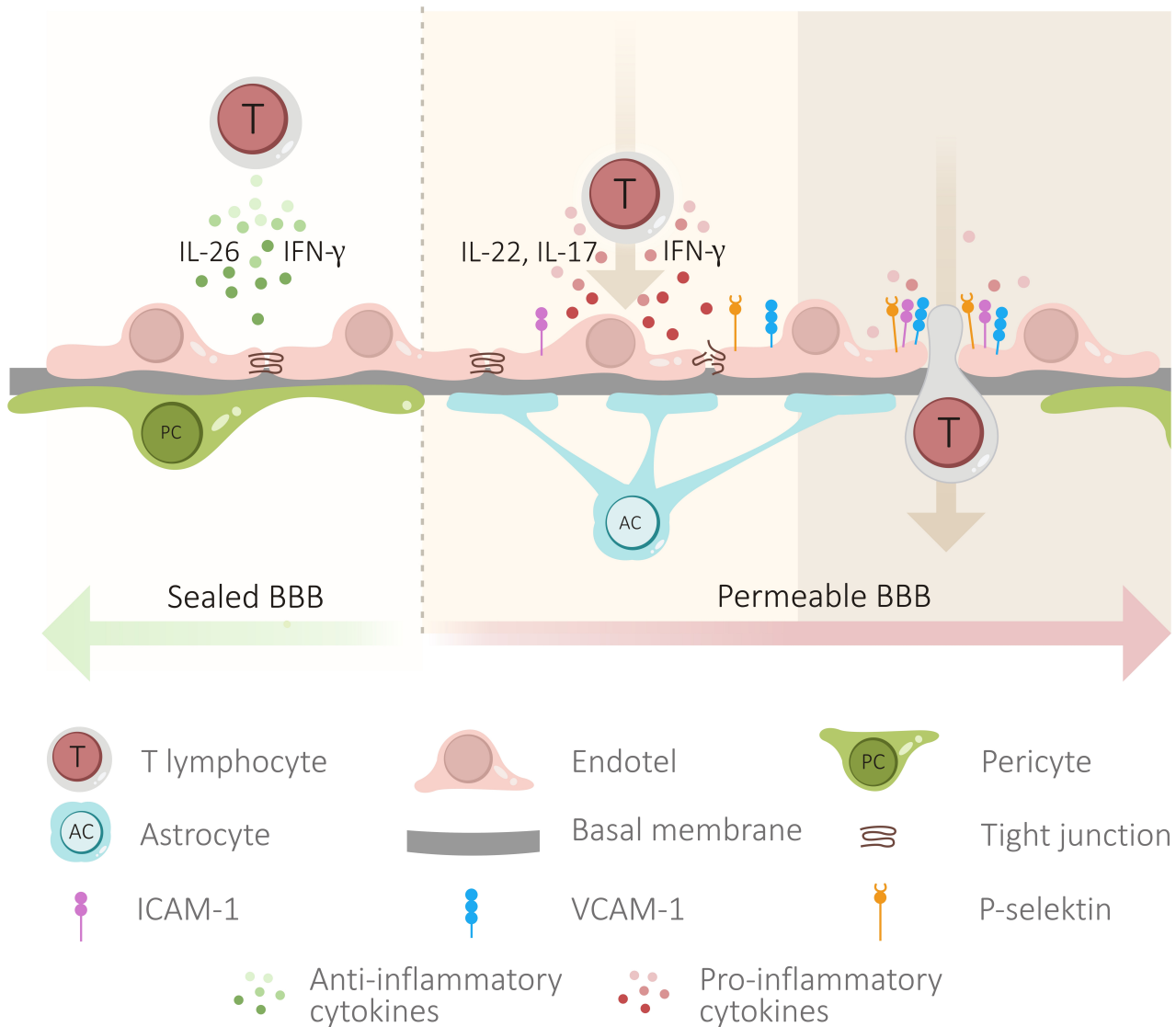
nalling, development, and differentiation of T lymphocytes or in the presentation of antigens to these cells [26]. The human leukocyte antigen DR15 (*HLA-DR15*) haplotype plays a crucial role in promoting the development of an autoreactive CD4<sup>+</sup> T cell repertoire in conjunction with environmental factors [27]. Self-reactivity, defined as the “auto-proliferation” of peripheral Th1 cells, is increased in individuals carrying the *HLA-DRB1\*15:01* allele [28]. This auto-proliferation is mediated by memory B cells in an HLA-DR-dependent manner, with its intensity decreasing both during *in vitro* depletion of B cells and during *in vivo* anti-CD20 therapy [29].

In addition to genetic predisposition, epigenetic mechanisms that influence the expression of genes associated with the inflammatory response also contribute to the pathogenesis of MS. Active and inactive MS lesions exhibit a different spectrum of expressed small non-coding RNAs (microRNAs, miR) miR-155, miR-34A, and miR-326 predominate in active lesions, influencing the expression of pathological cell genes, including by regulating the transcription of integrin-associated protein CD47, which is involved in macrophage activity [30]. Furthermore, miR-155 expression contributes to the differentiation of Th1 and Th17 lymphocytes and worsens the course of experimental autoimmune encephalomyelitis (EAE) and MS [31]. The important proinflammatory cytokine IL-17, produced by Th17 lymphocytes, is elevated in the cerebrospinal fluid of patients with RRMS, and its blockade reduces inflammatory activity in the experimental autoimmune encephalomyelitis model [32]. In addition, hypomethylation of the promoter region of the *IL-17A* gene has been demonstrated in patients with MS, which is associated with increased expression of this cytokine.

## 3. Environmental Factors and T Lymphocytes

In addition to genetic predisposition and epigenetic factors, external environmental factors also play a significant role in the aetiology and pathogenesis of MS. The importance of environmental factors is underscored by numerous migration studies. Adult migrants from countries with a lower risk of developing MS do not experience an increase in risk after moving to high-risk regions, but an increase is already apparent in migrant children born in high-risk countries [33]. Likely, we are already exposed to risk factors for the development of MS in utero. Early-life influences are suggested by the modest and geographically variable month-of-birth effect, as well as familial concordance patterns. Concordance is highest in monozygotic twins, lower in dizygotic twins, and slightly lower still in non-twin siblings, indicating contributions from both genetics and shared early environmental factors [34].

Among modifiable environmental risk factors, smoking is one of the most significant. Some studies report that it increases the risk of developing MS by up to 50%



**Fig. 2. Transfer of inflammation across the BBB.** An autoimmune inflammation activated in the periphery leads to increased BBB permeability. Activated  $CD4^+$  T lymphocytes produce pro-inflammatory cytokines (IL-22, IL-17 and  $IFN-\gamma$ ), which promote the expression of adhesive molecules on endothelial cells (ICAM-1, VCAM-1 and P-selectin) and disruption of tight junction sealing the intercellular spaces of the BBB. This process ensures the passage of  $CD4^+$  T lymphocytes across the BBB and settling of the inflammation in the CNS. Resting  $CD4^+$  T lymphocytes produce anti-inflammatory cytokines (IL-26 and  $IFN-\gamma$ ) causing downregulation of adhesive molecules on endothelial cells and strengthen tight junctions.  $IFN-\gamma$  can act as a pro- and anti-inflammatory agent depending on the overall cytokine and immune environment. IL, interleukin;  $IFN-\gamma$ , interferon-gamma; BBB, blood-brain-barrier; ICAM-1, Intercellular Adhesion Molecule 1/CD54; VCAM-1, Vascular cell adhesion molecule 1/CD106.

[35]. A major Swedish study involving more than 9000 patients with MS and an equal number of age-matched controls determined the population attributable fraction to be 13%, meaning that eliminating smoking would potentially prevent the development of MS in this proportion of patients [9]. Smoking is thus one of the few modifiable risk factors. Its influence is well documented not only epidemiologically but also at the level of molecular mechanisms. Cigarette smoke induces oxidative stress and activates various intracellular signalling pathways, including Toll-like

receptors (TLRs), via pathogen-associated molecular patterns (PAMPs), which can lead to epigenetic changes in the regulation of immune genes. In addition, changes in T-cell differentiation occur in the lung microenvironment; in an experimental model of EAE, it has been shown that autoreactive T cells acquire the ability to penetrate the CNS after transient exposure in lung tissue. Smoke-induced signals can transform potentially autoreactive T cells into a pathogenic phenotype. A key molecular mechanism involves the aryl hydrocarbon receptor (AHR), a ligand-

activated transcription factor expressed in T cells. Polycyclic aromatic hydrocarbons present in cigarette smoke activate AHR, which promotes Th17 cell differentiation and IL-17A production while modulating the Th17/Treg balance [36–38]. This AHR-mediated pathway provides a mechanistic link between smoking and the Th17-driven neuroinflammatory environment characteristic of MS.

Another important environmental factor is vitamin D deficiency. Its effect on the immune system is mediated, among other things, by epigenetic mechanisms. Vitamin D plays a key anti-inflammatory role in modulating the immune response, and its deficiency has been repeatedly associated with an increased risk of developing MS [8]. The active form of vitamin D acts through the vitamin D receptor, which binds to specific sequences in the genome and regulates the transcription of hundreds of target genes, including those with immunological functions. This mechanism interferes with both innate immunity (e.g., monocytes, dendritic cells) and adaptive immunity, particularly by modulating T and B lymphocytes. Vitamin D inhibits the polarization of pathogenic Th1 and Th17 cells, whose activity plays a key role in the pathogenesis of MS. At the same time, it promotes the formation of regulatory T cells, preferentially stimulating the Tr1 (Type 1 regulatory T cells) subpopulation, which produces the anti-inflammatory cytokine IL-10 [39,40]. The result is a shift in the immune response towards a tolerogenic profile and a reduction in autoreactivity. The epigenetic effects of vitamin D include histone acetylation and the modulation of microRNAs that regulate gene expression, which contributes to maintaining the balance between effector and regulatory immune responses [41–43].

Among environmental risks, EBV exposure stands out as one of the most significant. A history of infectious mononucleosis approximately doubles the risk of MS [44,45]. A large cohort study using data from more than 10 million US military recruits found up to a 32-fold higher risk of developing MS in individuals with serologically proven contact with EBV, opening up potential opportunities for MS prevention [46]. In patients with MS, an increased and expanded repertoire of antibody and T-cell immune responses to EBV-encoded antigens, particularly against the dominant CD4<sup>+</sup> T-cell EBV nuclear antigen 1 (EBNA1), has been observed [47]. Two main hypotheses have been proposed: (1) molecular mimicry, whereby EBV antigens cross-react with CNS components, and (2) repeated EBV reactivation during the alternation between latent and lytic phases of infection, leading to chronic stimulation of the immune system. Consistent with the first, cross-reactivity between EBV and CNS antigens has been documented in CD4<sup>+</sup> T cells, while CD8<sup>+</sup> T cells display increased frequency but reduced functional capacity [48]. Reactivity to antigens of both phases of infection—latent and lytic—further suggests that cyclic reactivation of EBV promotes diversity and long-term survival of mem-

ory T cells in patients with MS [7]. Nevertheless, the near-universal prevalence of EBV (up to 99%) suggests that contact with the virus alone is not sufficient for the development of MS. The development of the disease probably requires a combination of other genetic predispositions and environmental factors [25,33]. Recent exosome profiling has revealed that EBV-encoded miRNAs (BART9-3p and BART15) are significantly elevated in the cerebrospinal fluid of RRMS patients, accompanied by upregulation of proinflammatory cytokines including IFN- $\gamma$ , IL-17, and TNF- $\alpha$ , further supporting a mechanistic link between EBV activity and neuroinflammation [49]. Moreover, a recent study has shown that in early MS patients, 13% of T lymphocytes in the cerebrospinal fluid specifically target and attack autologous B lymphocytes infected with EBV, not just the virus itself. In the most expanded T-cell clones, this proportion rises to 47%. This suggests a novel mechanism: T cells attempting to control EBV-infected B cells in the CNS may inadvertently trigger or perpetuate the inflammatory cascade characteristic of MS [50].

Obesity, particularly during adolescence, is another important modifiable risk factor. Elevated body mass index has been repeatedly linked to increased MS risk, with the pathophysiological basis of this relationship lying in impaired immunological and metabolic regulation. Adipose tissue in obesity exhibits an overproduction of proinflammatory adipokines (e.g., leptin) and reduced levels of anti-inflammatory adipokines (e.g., adiponectin), leading to chronic subclinical inflammation that may facilitate the development of the autoimmune process. Obese individuals also show elevated circulating proinflammatory cytokines, including IL-6 and TNF- $\alpha$ . Furthermore, during the shift from lean to obese states, adipose tissue undergoes changes in T-cell composition, with reduced regulatory and anti-inflammatory CD4<sup>+</sup> T cells and expansion of effector Th1 and Th17 subsets, which play a key role in the pathogenesis of MS, predominate [51].

#### 4. The Microbiome as a Modulator of T-Lymphocyte Response

Various organs, particularly the intestine and possibly the lungs, perceive environmental signals and shape immune responses via lymphoid tissue. Physiological colonization of the gut is essential for maintaining the balance of mucosal and systemic immunity. Disruption of the normal intestinal microbiome (dysbiosis) predisposes individuals to immunopathological conditions and has been repeatedly linked to the onset and progression of MS [11,52,53]. The microbiome produces a number of biologically active substances, including short-chain fatty acids (SCFAs) such as propionate and butyrate. These metabolites induce histone deacetylation, thereby epigenetically regulating gene expression and also promoting the maturation of protective Tregs [54]. Their action leads to a reduction in the activity of Th1 and Th17 effector cells. Functional microbiome

analysis has shown that oral administration of propionate leads to increased expression of Treg-inducing genes in the intestine and normalization of mitochondrial function and morphology of these cells in patients with MS [55]. SCFAs also promote the production of the anti-inflammatory cytokine IL-10, thereby further contributing to immunological tolerance [56].

MS patients consistently exhibit reduced gut microbiome diversity compared to healthy controls, with notable depletion of SCFA-producing species, especially certain Clostridia, which support not only remyelination but also the development of Tregs. The result is an imbalance in the T-cell response—a reduced occurrence of Tregs in the lamina propria mucosae, Peyer's patches, and mesenteric lymph nodes, and at the same time, an increased prevalence of proinflammatory Th1 and Th17 cells [57]. This immunological imbalance can lead to a lower threshold for the development or exacerbation of MS. Other mechanisms are also likely to be involved in the link between the gut microbiome and MS. These include molecular mimicry (so far mainly documented in mouse models), increased binding of intestinal immunoglobulin A (IgA) to specific types of bacteria, and the effects of oxidative stress [58].

Increasing attention is also being paid to the link between the microbiome and the progression of MS. In 2020, a study using metagenomic sequencing of the gut microbiome was published, revealing differences between patients with RRMS and secondary progressive MS (SPMS). These observations support the hypothesis that changes in the microbiome may contribute to the transition to the progressive phase of the disease, among other things by influencing T-lymphocyte balance [59,60]. Changes in the gut microbiome thus significantly modulate the balance between regulatory and effector T lymphocytes and influence the immunological microenvironment in the CNS. However, microbiome findings show considerable variability between studies, with conflicting results for the same bacterial species across different cohorts. This heterogeneity, likely reflecting geographic variation, dietary differences, and disease modifying therapies (DMT) effects, complicates the translation of these findings into therapeutic interventions [11,52,53,59,60]. Despite these challenges, the microbiome remains a promising modulator of the autoimmune response and a potential therapeutic target in MS.

## 5. The Pathophysiology of Damaging Inflammation

When nervous tissue is damaged, for example, by a viral agent or toxic substance, antigenic peptides are presented by APCs via major histocompatibility complex (MHC) molecules to T cells. Among APCs, dendritic cells—a functionally heterogeneous population—are the most effective, as they are capable of processing complex antigen structures. These cells possess pattern-recognition receptors (PRRs), enabling them to identify external threats

in the form of PAMPs and internal danger signals known as damage-associated molecular patterns (DAMPs). Recognition of DAMPs or PAMPs via PRRs leads to dendritic cell activation and their targeted migration into secondary lymphoid tissues. There, dendritic cells functionally differentiate based on the specific PRR stimulus and begin to present processed antigenic peptides in the context of MHC molecules to T cells. This interaction triggers the activation of specific T cells, followed by their clonal expansion and functional polarization. Polarization is mediated by co-stimulatory signals from dendritic cells and by the cytokine milieu, largely shaped by these same dendritic cells. The initiation of damaging inflammation takes place in secondary lymphoid organs [61].

T cell development primarily occurs in the thymus, where two major populations emerge: precursors of cytotoxic T cells (CD8<sup>+</sup>) and helper T cells (CD4<sup>+</sup>, Th). Upon encountering an antigen, these cells differentiate into effector T cells [62]. For T cells to participate in damaging inflammation, they must exit the secondary lymphoid organs and migrate to target sites. Chemokines regulate this migration but also depend on the sphingosine-1-phosphate (S1P) gradient formed through the metabolism of ceramides. Lymphocytes sense this gradient via S1P receptors, primarily S1P1, which signal through intracellular GTPases. The binding of S1P to the S1P1 receptor induces controlled egress of lymphocytes from secondary lymphoid tissues and supports their subsequent chemokine-guided migration [63]. Specifically, this involves interaction with the chemokine CCL21 bound to intercellular matrix components, mediated through CCR7 receptors on the surface of central memory T cells [64].

Under physiological conditions, the entry and exit of immune cells and macromolecules from the brain are tightly regulated by the BBB, which protects the CNS from circulating immune cells and potentially harmful molecules [65]. In MS, however, the BBB is compromised, leading to increased permeability. T cells adhere to the endothelium during CNS infiltration via adhesion molecules such as LFA-1/Intercellular Adhesion Molecule 1/CD54 (ICAM-1) and VLA-4/Vascular cell adhesion molecule 1/CD106 (VCAM-1) [17]. B-cell trafficking involves classical adhesion pathways (e.g., VLA-4/VCAM-1 and LFA-1/ICAM-1), with contributions varying by activation state and tissue context [66]. Within the CNS, T cells are reactivated through renewed antigen presentation, which initiates a cascade of inflammation and recruits additional immune cells. Autoreactive T cells recognize immunodominant epitopes of neural membrane components, particularly myelin basic protein (MBP), myelin oligodendrocyte glycoprotein (MOG), myelin-associated glycoprotein (MAG), proteolipid protein (PLP), and cyclic nucleotide phosphodiesterase. This cascade results in focal oedema, demyelination, oligodendrocyte death, and damage to axons and neu-

rons [4,67]. Regarding cellular immunity, macrophages, microglial cells, cytotoxic CD8<sup>+</sup> T cells, and natural killer (NK) cells are activated.

The role of perivascular B cells associated with active white matter lesions is likely to be to reactivate proinflammatory CD4<sup>+</sup> and CD8<sup>+</sup> T cells. In later stages of the disease, B cells are, at least in part, gradually transformed into plasma cells [68]. In addition to producing antibodies, B cells contribute to inflammation through cytokine secretion and their role as APCs activating T cells [69,70]. Humoral immunity further participates via antibody-mediated recognition of myelin surface structures, leading to complement activation. Conversely, regulatory B cells (Bregs) have immunosuppressive properties—they modulate macrophage and dendritic cell function, inhibit CD4<sup>+</sup> T-cell proliferation, and enhance the expansion of Tregs [71,72].

The pathological landscape of MS is highly complex. Mounting evidence suggests the coexistence of two forms of inflammation—one driving multifocal active lesions and another associated with smouldering, slowly evolving lesions (Fig. 3) [73]. Data from MRI and histopathological studies show that progressive neuroaxonal loss, which drives long-term disability, begins early in the disease, indicating a continuum between the relapsing and progressive phases of MS [13].

In acute inflammation, peripheral immune responses dominate. Autoreactive T cells generated in secondary lymphoid tissues produce proinflammatory cytokines that disrupt the BBB and enable immune cell entry into the CNS. The predominant inflammatory cells in early active lesions are CD8<sup>+</sup> T cells, which proliferate and contribute to lesion formation [68]. These lesions also include significant numbers of CD4<sup>+</sup> T cells, activated macrophages, microglial cells, B cells, antibodies, and complement components [74]. Lymphocyte distribution within lesions appears stratified: CD8<sup>+</sup> T cells are often located at lesion borders, while CD4<sup>+</sup> T cells are found deeper within [75]. These classic active white matter lesions are common in RRMS but become infrequent as patients transition to the progressive stage [76].

During the acute phase, CNS inflammatory infiltrates may resolve without causing axonal damage or gliosis, enabling recovery and remyelination. However, some infiltrates persist as chronic aggregates resembling tertiary lymphoid structures composed of CD4<sup>+</sup> and CD8<sup>+</sup> T cells, B cells, and plasma cells, leading to compartmentalized inflammation [77]. In the progressive phase—characterized by chronic or smouldering inflammation—the inflammatory process persists behind a re-sealed BBB. This compartmentalized inflammation drives neurodegeneration and disease progression. It is diffusely present in the meninges and perivascular Virchow–Robin spaces. It is associated with slow expansion of prior white matter lesions, widespread damage to normal-appearing white and grey matter, and subpial cortical demyelination in the brain and spinal cord.

These features are hallmarks of progressive MS and are often accompanied by active demyelination and neurodegeneration [73]. Significant infiltration by CD8<sup>+</sup> T cells and activated microglia in these regions likely contribute to smouldering inflammation, ongoing cognitive decline, and disability progression [78,79].

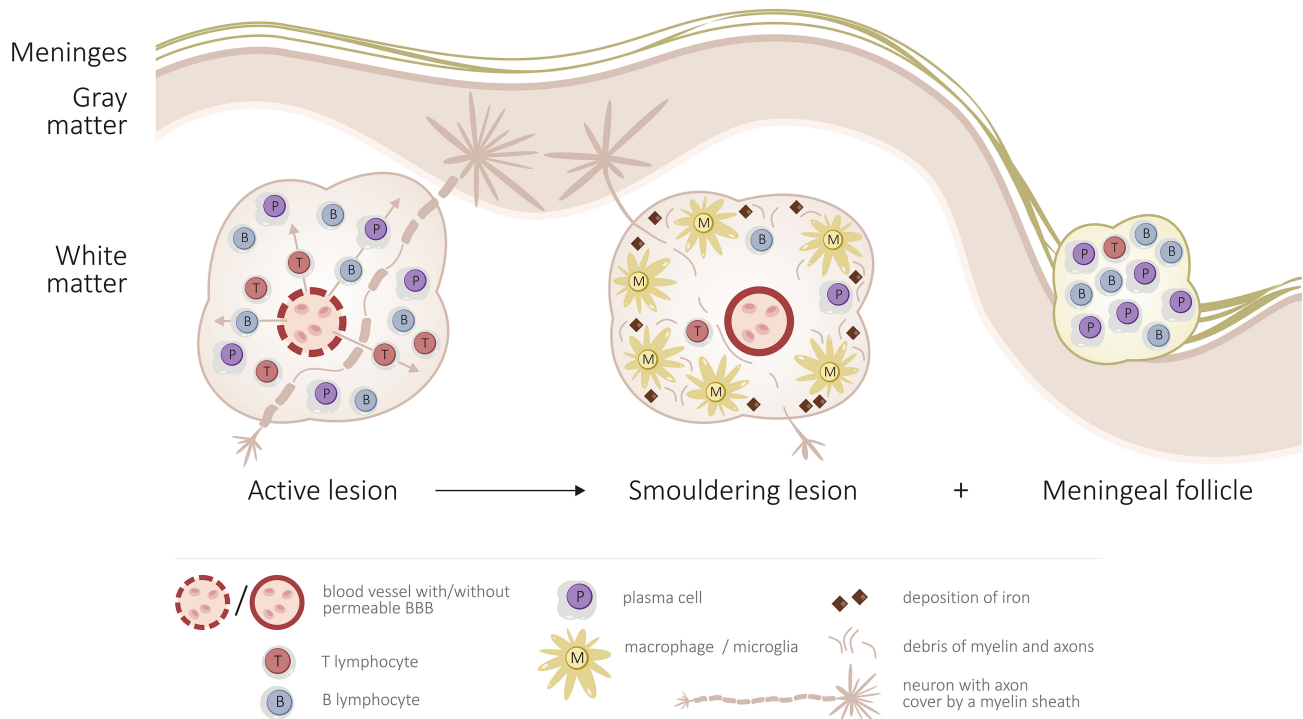
## 6. T Cell-Glia Interactions

Beyond their co-localization, these cell populations engage in bidirectional signalling that amplifies neuroinflammation. Microglia express MHC class I and II molecules, enabling them to present myelin antigens to CD8<sup>+</sup> and CD4<sup>+</sup> T cells, respectively, thereby reactivating infiltrated lymphocytes within the CNS parenchyma [80]. In turn, T cell-derived cytokines, particularly IFN- $\gamma$  and the granulocyte-macrophage colony-stimulating factor (GM-CSF), induce microglial activation toward a proinflammatory phenotype characterized by enhanced phagocytosis, reactive oxygen species production, and secretion of TNF- $\alpha$  and IL-1 $\beta$  [81]. Recent spatial transcriptomic analyses have revealed a lymphocyte-microglia-astrocyte axis at the rim of chronic active lesions, where activated microglia cluster with CD8<sup>+</sup> T cells and reactive astrocytes, perpetuating tissue damage in progressive MS [82]. However, T cell-glia interactions represent only one component of the complex cellular network driving MS pathology.

## 7. B Cell and T Cell Crosstalk in MS Pathogenesis

While this review focuses on T cells, emerging evidence challenges the traditional T-cell-centric view of MS pathogenesis. Despite CD8<sup>+</sup> T cells being the predominant infiltrating population in MS lesions, B-cell depletion therapies have demonstrated superior clinical efficacy compared to many T-cell-targeted approaches. Recent findings reveal that B cells can control responses of myeloid cells through oxidative phosphorylation-regulated cytokine release, revealing their previously underappreciated regulatory role rather than secondary players [83]. This paradox, coupled with the failure of atacept (which blocks the B-cell activating factor and a proliferation-inducing ligand), which actually worsens disease activity, demonstrates that B cells have complex pathogenic and protective functions that may be equally central to MS pathogenesis [84].

The understanding of the role of B cells in MS has evolved substantially in recent years. Intrathecal antibody production is a hallmark of multiple sclerosis, and humoral immunity plays an important role in the inflammatory response and development of demyelinated lesions. Interplay between B and T lymphocytes is a central feature of disease pathogenesis [85]. B cells are well-known efficient APCs, characterized by the expression of MHC class II, and specialized in capturing soluble and membrane-tethered antigens, with a high efficiency in presenting antigens and activating T cells. B cells produce pro-inflammatory cytokines



**Fig. 3. Development of inflammation and neurodegeneration in multiple sclerosis.** Active lesion – The pathogenesis of multiple sclerosis begins with active lesions, where inflammation originates in the peripheral immune system. A proinflammatory cytokine milieu leads to BBB disruption, enabling the infiltration of autoreactive T and B lymphocytes into the CNS. Cellular immunity ( $CD4^+$  and  $CD8^+$  T cells) and humoral immunity (B cells and plasma cells) contribute to demyelination through oligodendrocyte damage. Although axonal loss is less prominent in this phase, it is already underway. After crossing the BBB, immune cells localize in perivascular areas, which become the initial sites of inflammation. Smouldering lesion – In smouldering lesions, the BBB reseals, leading to compartmentalization of inflammation within the CNS. Innate immune cells (macrophages, activated microglia) and cytotoxic  $CD8^+$  T cells dominate the chronic inflammatory response. These cells accumulate at the lesion rim, where they interact with myelin and neuronal degradation products and contribute to iron deposition. This phase marks the transition toward a primarily neurodegenerative process characterized by progressive axonal loss. Tertiary meningeal follicle – Tertiary meningeal follicles represent another manifestation of compartmentalized inflammation beyond the BBB. These ectopic lymphoid structures form in the meninges and are rich in B cells, which undergo maturation into plasma cells, contributing to chronic immune activation in progressive MS.

IL-6, GM-CSF, TNF- $\alpha$ , and lymphotoxin alpha. IL-6 is a major mechanism of B cell-driven pathogenesis in T cell-mediated autoimmune disease, such as EAE and MS [86].

In early and active lesions,  $CD20^+$  B cells dominate and may have pro-inflammatory functions, while at later stages, plasma cells with possible anti-inflammatory functions increase in number [68]. B cells are mainly located in the meninges and in the large perivascular spaces around the cerebral ventricles. Especially within deep cortical sulci, prominent B cell-rich inflammatory aggregates of ectopic meningeal lymphoid follicles are found. These tertiary lymphoid follicles rich in B and T cells likely mediate the progressive loss of neurological function in MS [77].

MS brain-infiltrating lymphocytes also express and respond to IL-21, the cytokine that drives follicular T- and B-cell responses [87]. Additionally, IFN- $\gamma$ -triggered B cell activation promotes ectopic follicle formation in autoimmune mice, suggesting that the structures observed in the

CNS in MS are induced by B cells interacting with IFN- $\gamma$ -producing T cells [88]. IFN- $\gamma$ , and possibly also GM-CSF, can activate microglia or infiltrated macrophages to cause damage to oligodendrocytes, and IFN- $\gamma$  also drives IgG-producing plasmablasts in MS [70,89,90]. These findings underscore a fundamental shift in our understanding: rather than a T-cell-driven disease with B cell support, MS may represent a complex interplay where both cell types can serve as primary drivers depending on disease stage and patient subtype. Despite these advances in understanding B cell pathogenicity, critical therapeutic limitations persist. B cell depletion therapy fails to resolve chronic active lesions with paramagnetic rim lesions even after a 2-year follow-up, attributed to limited B-cell turnover in CNS tissue, inefficient anti-CD20 antibody passage across the blood-brain barrier, and paucity of B cells within established chronic active lesions [91]. Furthermore, anti-CD20 agents do not prevent progression independent of relapse activity, high-

lighting that compartmentalized inflammation remains unaddressed [92].

Collectively, these findings reveal that the B-T cell axis in MS is more complex than previously appreciated. B cells function not merely as antibody producers but as critical orchestrators of T-cell responses through antigen presentation, cytokine production, and formation of ectopic lymphoid structures. The bidirectional nature of B-T cell interactions—where each population can activate and sustain the other—creates self-perpetuating inflammatory loops that current therapies only partially address. Understanding these intricate cellular interactions will be essential for developing next-generation therapies that can effectively target compartmentalized inflammation and halt disease progression.

## 8. Subtypes of T Lymphocytes and Their Role in MS Pathogenesis

Although multiple immune cell types contribute to MS pathophysiology, T lymphocytes have emerged as central regulators of both peripheral and CNS immune responses. Their functional diversity—ranging from highly proinflammatory to regulatory phenotypes—helps explain the immunological heterogeneity observed in MS lesions. A closer look at individual T-cell subsets reveals how specific phenotypes contribute to demyelination, neurodegeneration, or, conversely, immune regulation and provides insight into the cellular targets of current and emerging therapies.

### 8.1 Cytotoxic CD8<sup>+</sup> T Cells

CD8<sup>+</sup> cytotoxic T cells protect the CNS from infectious agents and eliminate damaged or malignant cells. These cells express CD8 on their surface, which enables them to recognize antigens presented on MHC class I molecules. Upon recognition, CD8<sup>+</sup> cells induce apoptosis in the target cell. They also play a central role in the pathogenesis of MS. During acute demyelination, axons become vulnerable and temporarily express MHC class I molecules, making them targets for CD8<sup>+</sup> T cells and macrophages [74]. CD8<sup>+</sup> cells are the predominant lymphocyte subtype in all stages of MS and are closely associated with axonal damage [20,93].

### 8.2 Helper CD4<sup>+</sup> T Cells

Helper T lymphocytes (Th cells) express CD4 on their surface and are activated upon recognising antigens presented on MHC class II molecules. Naive CD4<sup>+</sup> T cells can differentiate into various effector subsets (Th1, Th2, Th9, Th17) or Tregs (Fig. 4). Th1 and Th17 cells mediate responses against intra- and extracellular pathogens but can contribute to autoimmune pathology if dysregulated. In contrast, Tregs suppress excessive immune activation and help maintain immune tolerance [94]. The subsets are classified based on their cytokine profiles and immunologic

functions. Th1 cells mainly produce IFN- $\gamma$  and IL-2. Th2 cells secrete IL-4, IL-5, IL-6, and IL-10. Th0 cells produce a combination of type 1 and type 2 cytokines. Th3 cells produce transforming growth factor-beta (TGF- $\beta$ ), while Tr1 cells primarily produce IL-10 [95]. Th17 cells produce IL-17 and other proinflammatory cytokines. Their differentiation from Th0 cells is driven by IL-23, which is secreted by innate immune cells. Th17 cells also secrete IL-21, which contributes to the activation of NK cells [96].

### 8.3 Th1 Cells

Th1 cells exert their effects by activating CD8<sup>+</sup> cytotoxic T cells, NK cells, and macrophages by producing IFN- $\gamma$ , IL-2, and other cytokines such as IL-12 and IL-18 (Fig. 5). IFN- $\gamma$  activates the cerebral endothelium and promotes the expression of adhesion molecules on BBB endothelial cells [31,97]. APC-derived IL-12 supports Th1 differentiation while inhibiting Th2 polarization. Pathologically, excessive Th1 activity contributes to type IV hypersensitivity reactions characterized by granuloma formation, as observed in MS lesions [16,93]. Th1 hyperactivation is implicated in chronic inflammation and tissue injury in MS [98].

### 8.4 Th2 Cells

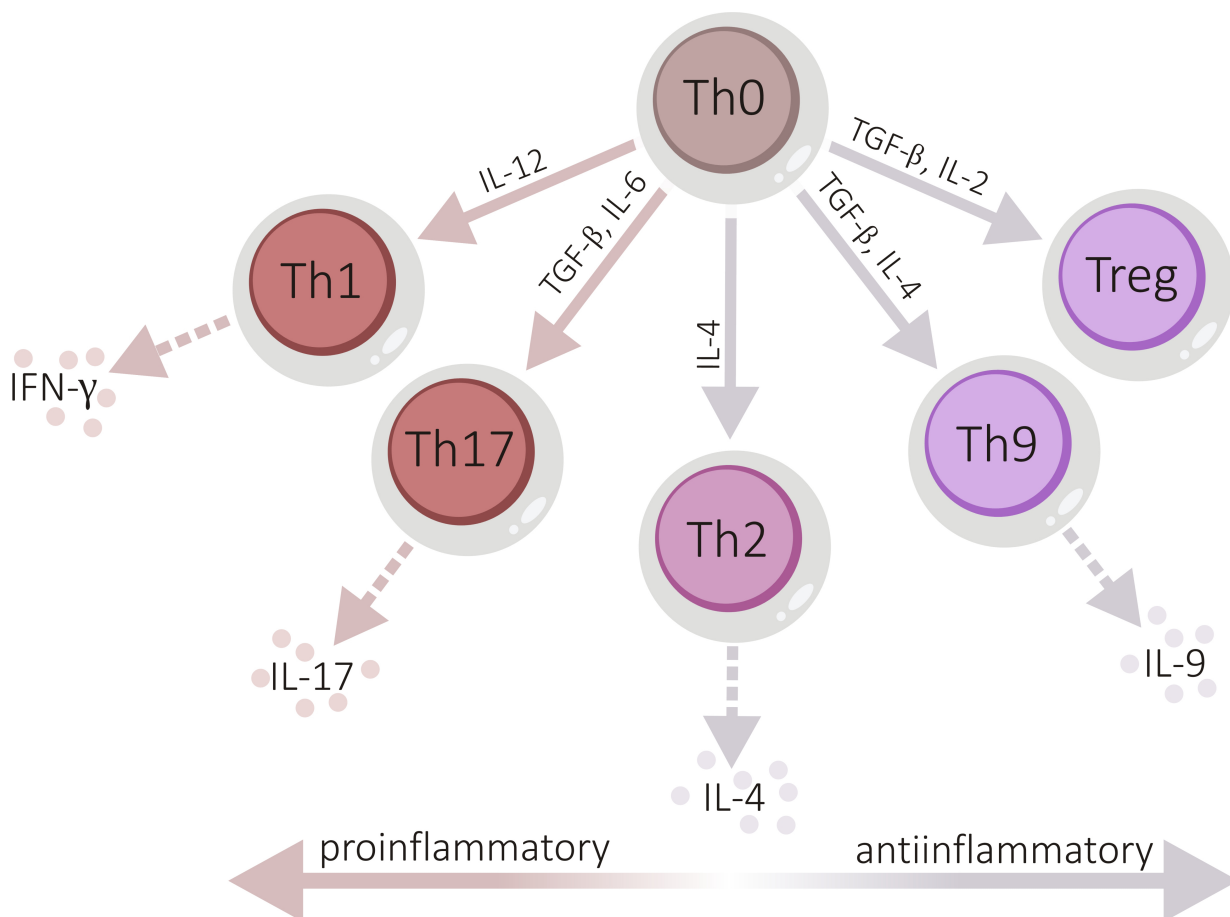
Th2 cells are primarily involved in immune responses against extracellular parasites and allergens. They produce IL-4, IL-5, IL-6, IL-9, and IL-10. Their differentiation is promoted by the presence of IL-4 during antigen presentation on MHC class II molecules [99]. In tertiary lymphoid follicles of the CNS, Th2 cells assist in fully activating antigen-stimulated B cells [100]. Nevertheless, Th2 cells generally exert anti-inflammatory effects and can counterbalance Th1-mediated inflammation.

### 8.5 Th9 Cells

Th9 cells participate in allergic inflammation and immunity against intestinal parasites. They produce IL-9 and, to a lesser extent, IL-10 and IL-21. IL-9 regulates the balance between Th17 and Tregs. Th9 cell differentiation requires a specific cytokine environment, particularly the presence of both TGF- $\beta$  and IL-4 [101]. IL-9 has neuroprotective effects and may counteract inflammatory synaptopathy in EAE, suggesting a potential anti-inflammatory role in MS [102].

### 8.6 Th17 Cells

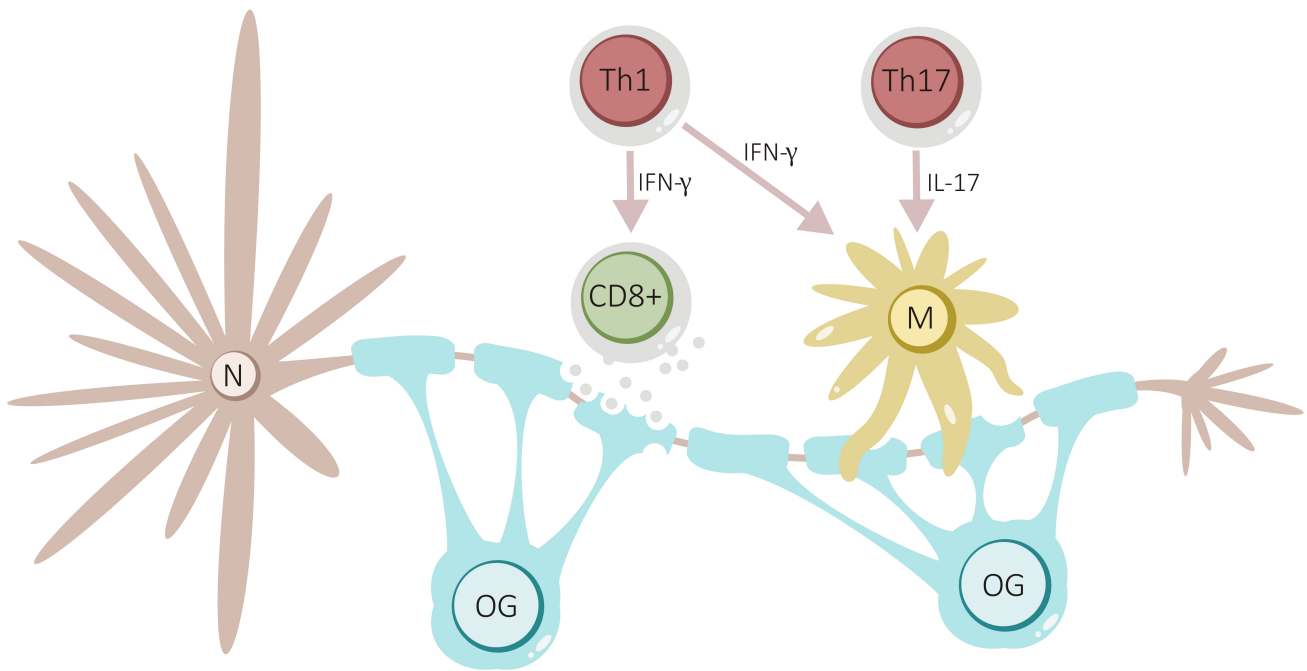
Th17 cells are defined by their secretion of IL-17A, IL-17F, IL-21, IL-22, TNF- $\alpha$ , and low levels of IFN- $\gamma$  (Fig. 5) [103]. Their differentiation requires TGF- $\beta$  in combination with either IL-6 or IL-23 [104]. TGF- $\beta$  supports immune tolerance and promotes Tregs differentiation at high concentrations, whereas lower concentrations in the presence of IL-6 favour Th17 induction [105]. IL-6 thus plays a critical role in regulating the Th17-Tregs balance.



**Fig. 4. Differentiation of CD4<sup>+</sup> T cell subsets based on cytokine milieu.** Naïve Th0 lymphocytes differentiate into various CD4<sup>+</sup> T cell subtypes depending on the surrounding cytokines. Th1 and Th17 cells promote inflammation via the secretion of IFN- $\gamma$  and IL-17, respectively. Th9 cells may contribute to inflammation, while regulatory T cells (Treg) exert anti-inflammatory effects. Th2 lymphocytes have a more ambivalent role—they aid in B cell maturation in tertiary follicles but are generally anti-inflammatory. TGF- $\beta$ , transforming growth factor-beta; Th, T helper.

Th17 cells are highly proinflammatory. They promote the recruitment of neutrophils and macrophages, drive the expansion of dendritic and T cells, and amplify the inflammatory response through positive feedback loops. Th17 cells also induce the production of cytokines, chemokines, antimicrobial peptides, and matrix metalloproteinases [106]. They facilitate CD4<sup>+</sup> T-cell migration across the BBB by disrupting tight junctions via IL-17 and synergise with IFN- $\gamma$  to induce ICAM-1 expression on endothelial cells [107]. Alongside Th1 cells, Th17 cells are the principal proinflammatory T-cell subtype in MS. However, recent evidence reveals substantial heterogeneity within Th17 populations, with different subsets playing opposing roles in disease progression. Single-cell RNA sequencing studies have identified functionally distinct Th17 subpopulations in both blood and CSF of MS patients [108, 109]. Pathogenic Th17.1 cells (CCR6<sup>+</sup>CXCR3<sup>+</sup>CCR4<sup>-</sup>), characterized by co-expression of IFN- $\gamma$  and GM-CSF, are enriched in the CSF and correlate with disease activity, whereas non-pathogenic Th17 cells expressing IL-10

may exert regulatory functions [110]. Furthermore, tissue-resident memory T cells (TRM) with both CD4<sup>+</sup> and CD8<sup>+</sup> phenotypes have been identified in the CSF compartment, suggesting that CNS-resident T cell populations contribute to compartmentalized inflammation independently of peripheral immune cell trafficking [111]. Spatial transcriptomics approaches have further refined our understanding of T-cell organization within MS lesions, revealing distinct tissue niches at the inflamed rim of chronic active lesions where T cells interact with activated microglia and astrocytes [112]. Complementary studies have identified spatially resolved gene signatures that distinguish lesion progression trajectories, with implications for understanding how local T-cell responses evolve during disease chronicity [113]. These findings underscore the complexity of T-cell-mediated pathology in MS and highlight the need for subset-specific therapeutic approaches.



**Fig. 5. Role of cellular immunity to demyelination and axonal loss.** Th1 cells, through IFN- $\gamma$ , promote activation of CD8<sup>+</sup> cytotoxic T lymphocytes, while Th17 cells release IL-17, which stimulates macrophages and microglia. CD8<sup>+</sup> T cells induce apoptosis of neural cells, contributing to axonal damage. Macrophages and activated microglia act as phagocytes, clearing myelin debris and further exacerbating demyelination. Together, these cellular mechanisms drive both inflammatory demyelination and axonal degeneration. CD8<sup>+</sup>, cytotoxic CD8<sup>+</sup> T lymphocyte; M, macrophage and microglia; OG, oligodendrocyte forming the myelin sheath; N, neuron.

### 8.7 Regulatory T Cells

Tregs play a crucial role in maintaining immune homeostasis by suppressing other immune cells' activation, proliferation, and effector functions. They prevent the emergence of autoreactive T and B cells. Tregs are classified into thymus-derived (tTregs) and peripherally induced (pTregs) subtypes. tTregs arise in the thymus upon recognizing self-antigens presented via MHC class II, requiring CD28 co-stimulation. pTregs develop in peripheral tissues from naive CD4<sup>+</sup> T-cells under the influence of IL-2 and TGF- $\beta$ . In contrast, IL-6 inhibits Tregs differentiation [102,114,115]. Tregs are identified by the expression of CD25 and the transcription factor Forkhead box P3 (Foxp3). tTregs are Foxp3<sup>+</sup>; pTregs also express Foxp3, although there are subsets such as Tr1 (Foxp3<sup>-</sup>, IL-10-producing) and Th3 (TGF- $\beta$ -producing) cells within the broader regulatory population [116]. tTregs suppress autoreactive T cells that have escaped thymic deletion, while pTregs act to control antigen-specific effector T-cell responses in the periphery; their functions are complementary [72]. Flow cytometric studies have documented both quantitative and qualitative Treg abnormalities in MS. Resting Tregs (CD45RA<sup>+</sup>Foxp3<sup>+</sup>) are significantly reduced in both untreated and treated MS patients compared to healthy donors, while activated Tregs (CD45RA<sup>-</sup>Foxp3<sup>high</sup>) are paradoxically increased [117]. In healthy donors, approx-

imately 70–80% of CD4<sup>+</sup>CD25<sup>high</sup> cells express Foxp3, whereas this proportion is consistently lower in untreated RRMS patients [118]. Furthermore, Foxp3 mean fluorescence intensity (MFI) per cell is reduced in RRMS compared to secondary progressive MS and healthy controls, indicating diminished Foxp3 protein expression at the single-cell level [118]. Functionally, CFSE-based suppression assays have demonstrated that the *in vitro* suppressive capacity of CD4<sup>+</sup>CD25<sup>high</sup> Tregs correlates positively with Foxp3 expression levels, with RRMS patients showing impaired suppression compared to controls [118–120]. The stability of Foxp3 expression depends on epigenetic modifications at the Treg-specific demethylated region (TSDR/CNS2) within the Foxp3 locus. Demethylation of this conserved enhancer element is a hallmark of thymus-derived Tregs and is required for stable Foxp3 expression and sustained suppressive function [121,122]. Although direct TSDR methylation data from MS cohorts remain limited, recent mechanistic studies in inducible Tregs show that the clustered regularly interspaced short palindromic repeats - ten-eleven translocation 1 (CRISPR-TET1)-mediated Foxp3-TSDR demethylation (in combination with the signal transducer and activator of transcription 6 (STAT6) inhibition/deficiency) increases Foxp3 stability and suppressive function, supporting the idea that insufficient TSDR demethylation may contribute to Treg instability—potentially relevant to autoimmune set-

tings [123]. Notably, IFN- $\gamma$ -producing Foxp3<sup>+</sup> T cells—indicative of Treg plasticity toward a Th1-like phenotype—have been identified in RRMS patients, further suggesting epigenetic dysregulation of the Treg lineage [124]. Functionally, Treg deficiency or dysfunction facilitates Th17 expansion, thereby amplifying the proinflammatory milieu. During the recovery phase in MS, Tregs constitute up to one-third of CNS-infiltrating CD4<sup>+</sup> cells [125,126]. Their presence is essential for tissue repair and disease remission, while their absence impairs recovery [127].

These insights into T-cell biology not only deepen our understanding of MS immunopathogenesis but also clarify the rationale behind many current and emerging treatment approaches.

## 9. T-Cell-Mediated Mechanisms of Action of Disease-Modifying Drugs

T lymphocytes represent critical therapeutic targets in MS for several fundamental reasons. First, they initiate and orchestrate CNS inflammation—autoreactive T cells must be activated in the periphery and migrate across the BBB before any significant demyelination occurs, providing an upstream intervention point [128]. Second, unlike microglia or other CNS-resident cells, circulating T cells are readily accessible to therapeutic agents, eliminating the need for BBB penetration. Third, T cell responses demonstrate remarkable plasticity—pathogenic Th1/Th17 cells can be shifted toward regulatory phenotypes, potentially restoring immune tolerance without permanent immunosuppression [127]. Fourth, the diversity of T-cell trafficking mechanisms offers multiple therapeutic intervention points: preventing activation (teriflunomide), blocking adhesion molecules (natalizumab), sequestering in lymph nodes (S1P modulators), or depleting specific subsets (alemtuzumab), each with distinct safety-efficacy profiles that enable personalised treatment selection [116].

Although not all DMTs for MS treatment were specifically designed to target T lymphocytes, many exert substantial effects on T-cell-mediated immune mechanisms (Table 1, (Ref. [4,29,129–138]) and Table 2, (Ref. [139–143])). These diverse approaches—from cytokine modulation to lymphocyte depletion—ultimately converge on disrupting autoreactive T-cell function, though with varying efficacy depending on disease stage and individual patient characteristics. Importantly, the clinical efficacy of these T cell-targeting mechanisms varies dramatically depending on disease stage. The therapeutic landscape reveals a critical disconnect between efficacy in relapsing versus progressive disease phases [144]. While therapies highly effective in RRMS often show minimal benefit in progressive MS, this reflects fundamental differences in how T cells contribute to pathology at different stages [145]. In RRMS, where peripheral T-cell infiltration drives acute inflammation, treatments blocking CNS entry (natalizumab, S1P modulators) or depleting circulating lymphocytes (alem-

tuzumab, anti-CD20 antibodies) demonstrate robust efficacy. However, once compartmentalized inflammation establishes behind a closed blood-brain barrier in progressive MS, these same mechanisms fail to address ongoing neurodegeneration [146]. Only therapies with direct CNS penetration or effects on resident immune cells show promise in progressive disease—explaining why siponimod (which crosses the BBB and affects CNS S1P receptors) and potentially brain-penetrant Bruton's tyrosine kinase (BTK) inhibitors demonstrate efficacy where purely peripheral immunomodulation fails [147,148].

### 9.1 Therapies Modulating Cytokine Profiles and T-Cell Polarization

First-generation DMTs primarily function by shifting T cell responses from a proinflammatory to a regulatory phenotype. Interferon-beta (IFN- $\beta$ ) exerts broad immunomodulatory effects, enhancing production of anti-inflammatory cytokines, including IL-10, while suppressing proinflammatory cytokines such as IL-17, thereby shifting the balance towards Th2 cells [129]. IFN- $\beta$  reduces antigen presentation, inhibits T-cell proliferation, and strengthens antiviral defence mechanisms, for instance by inducing the antiviral Mx protein. Similarly, glatiramer acetate, a synthetic mixture of peptides that mimics the immunodominant sequences of myelin basic protein, shifts the immune response from a pro-inflammatory Th1 phenotype to an anti-inflammatory Th2 profile. This transition is accompanied by a reduction in the production of cytokines such as IL-2 and IL-12, and an increase in the release of IL-4, IL-10, and IL-1 [4]. Glatiramer acetate also binds to T-cell receptors (TCRs) on autoreactive T-cells, thereby preventing the need for co-stimulatory signals, which leads to functional anergy or apoptosis. Both IFN- $\beta$  and glatiramer acetate impede lymphocyte migration across the BBB and increase the number and function of Tregs. Fumarates represent an oral alternative for cytokine modulation, working through activation of the transcription nuclear factor erythroid 2-related factor 2 (Nrf2), which mediates antioxidative, anti-inflammatory, cytoprotective, and neuroprotective effects [130]. In the CNS, Nrf2 is expressed in neurons, astrocytes, and microglial cells, where it limits neuroinflammation and microglial activation. Regarding T cell effects, fumarates reduce the frequency and function of proinflammatory Th1 and Th17 cells, promoting a shift toward Tregs. They downregulate production of proinflammatory cytokines such as IL-17 and IFN- $\gamma$ , enhance IL-10 secretion, and reduce the expression of adhesion molecules and lymphocyte migration across the BBB [131]. This dual action on both peripheral immunity and CNS-resident cells distinguishes fumarates from purely immunomodulatory therapies.

**Table 1. DMTs and Mechanism of Action.**

Disease modifying treatments (DMTs)	Route of administration	Mechanism of action
Glatiramer acetate [4]	subcutaneous	shift the immune response from a proinflammatory Th1 phenotype to an anti-inflammatory Th2 profile, reduce production of cytokines IL-2 and IL-12, increase release of IL-4, IL-10, and IL-1, bind to T-cell receptors (TCRs) on autoreactive T-cells without co-stimulatory signals, impede lymphocyte migration across the BBB and increase the number and function of Tregs
Interferon-beta 1a and 1b [129]	subcutaneous, intramuscular	enhance the production of anti-inflammatory cytokines (IL-10), suppress proinflammatory cytokines (IL-17), reduce antigen presentation and inhibit T-cell proliferation, strengthen antiviral defence mechanisms, impede lymphocyte migration across the BBB, increase the number and function of Tregs
Fumarates (dimethyl fumarate, diroximel fumarate) [130,131]	oral	activation of the transcription Nrf2 - induce the transcription of antioxidant response genes - antioxidative, anti-inflammatory, cytoprotective, and neuroprotective effects - limit neuroinflammation and microglial activation, immunomodulatory effects on T lymphocytes (reduce the frequency and function of proinflammatory Th1 and Th17 cells and promote a shift toward Tregs), downregulate the production of proinflammatory cytokines (IL-17 and IFN- $\gamma$ ), enhance IL-10 secretion, reduce the expression of adhesion molecules and lymphocyte migration across the BBB
Teriflunomide [132]	oral	reversibly inhibit dihydroorotate dehydrogenase, reduce proliferation of activated T and B cells impact the CD8 <sup>+</sup> T-cell compartment, suppress the production of proinflammatory cytokines (TNF- $\alpha$ and IFN- $\gamma$ )
S1P modulators (fingolimod, ozanimod, ponesimod, siponimod) [134]	oral	bind to S1P receptors on lymphocytes - leading to their internalization and functional inactivation, prevent lymphocyte egress from lymph nodes, decrease peripheral lymphocytes, involve in reparative processes in CNS
Natalizumab [133]	subcutaneous, intravenous	a monoclonal antibody targeting VLA-4, prevent interaction with VCAM-1 on endothelial cells of the BBB, block the migration of T-cells across the BBB almost entirely
Cladribine [138]	oral	a purine nucleoside analogue - increase deoxycytidine kinase expression in lymphocytes and lead to lymphocyte apoptosis
Alemtuzumab [137]	intravenous	a monoclonal antibody targeting CD52 (a surface protein expressed on T and B cells), induce profound lymphocyte depletion, subsequent repopulation favour regulatory and tolerogenic immune subsets, promoting a shift toward immune tolerance
anti-CD20 monoclonal antibodies (rituximab, ocrelizumab, ofatumumab, ublituximab) [29,135,136]	subcutaneous, intravenous	a monoclonal antibody targeting CD20 (a surface protein expressed on pre-B and mature B cells), induce complement-mediated lysis of B cells, indirectly reduce T-cell autoreactivity and proliferation, dampen the proinflammatory cytokine milieu, increase in the proportion of naive CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells, decrease in the proportion of T cells producing IFN- $\gamma$ , decrease in the percentage of T cells with measurable expression of CD20

S1P, sphingosine-1-phosphate; Nrf2, nuclear factor erythroid 2-related factor 2; TNF- $\alpha$ , tumor necrosis factor alpha; VLA-4, very late antigen-4; DMT, disease modifying therapies.

**Table 2. Emerging Therapies and Mechanism of Action.**

Emerging therapies	Route of administration	Mechanism of action
BTK inhibitors (fenebrutinib, tolebrutinib, remibrutinib, orlabrutinib, evobrutinib) [139]	oral	limit B-cell and microglial activation, suppress proinflammatory cytokines (IFN- $\gamma$ and TNF- $\alpha$ ), reduce CD4 <sup>+</sup> T-cell differentiation into pathogenic subsets, suppress proinflammatory macrophages
anti-CD40 ligand (frexalimab) [140,141]	subcutaneous, intravenous	a monoclonal antibody targeting CD40 ligand (expressed predominantly on antigen-activated T-cells and, to a lesser extent, on B cells, mast cells, eosinophils, NK cells, and other immune cells) - modulate both adaptive and innate immune responses
CAR T-cell therapy [142]	intravenous	<i>ex vivo</i> genetic modification of T cells to target CD19 <sup>+</sup> B cells - eliminate autoreactive B-cell clones
anti-CD3 monoclonal antibody (foralumab) [143]	intranasal	promoting regulatory T cell (Treg) expansion and suppressing effector T cell activation without causing lymphocyte depletion

BTK, Bruton's tyrosine kinase; CAR, chimeric antigen receptor.

### 9.2 Therapies With Antiproliferative Effects

Teriflunomide selectively and reversibly inhibits dihydroorotate dehydrogenase, a key mitochondrial enzyme essential for de novo pyrimidine synthesis, leading to reduced proliferation of activated T and B cells. Teriflunomide specifically impacts the CD8<sup>+</sup> T-cell compartment by reducing homeostatic proliferation and suppressing production of proinflammatory cytokines such as TNF- $\alpha$  and IFN- $\gamma$  [132].

### 9.3 Therapies Blocking Lymphocyte Migration

A third therapeutic strategy involves preventing T-cell entry into the CNS. Natalizumab, a monoclonal antibody targeting VLA-4, blocks the migration of T cells across the BBB almost entirely. VLA-4 is highly expressed in central memory and effector memory T cells involved in MS pathogenesis. By binding to VLA-4, natalizumab prevents its interaction with VCAM-1 on endothelial cells of the BBB [133]. This leads to profound suppression of CNS inflammation but carries a risk of reactivation of latent infections, most notably John Cunningham virus (JCV) and the development of progressive multifocal leukoencephalopathy (PML) [149]. PML risk is stratified by three major factors: anti-JCV antibody serostatus and index level, prior immunosuppressant use, and treatment duration beyond 24 months [150,151]. In high-risk patients (JCV-seropositive with an index >0.9 and prior immunosuppression), PML incidence may reach 1% or higher [152]. Extended interval dosing protocols have been implemented to mitigate this risk while maintaining efficacy, with retrospective data suggesting up to 80–90% risk reduction compared to standard dosing [153]. S1P modulators represent an alternative approach to limiting CNS infiltration. These agents bind to S1P receptors on lymphocytes, leading to their internalisation and functional inactivation. This prevents lymphocyte egress from lymph nodes, decreasing peripheral T-cell counts [134]. Notably, S1P receptors are abundantly expressed on CNS-resident cells, including oligodendrocytes, astrocytes, and microglia, where they mediate neuropro-

protective and potentially remyelinating effects. Siponimod, a selective S1P1/S1P5 modulator, demonstrated efficacy in secondary progressive MS (EXPAND trial), suggesting that direct CNS effects beyond lymphocyte sequestration contribute to its therapeutic benefit. These CNS effects may include promotion of oligodendrocyte survival, reduction of astrogliosis, and modulation of microglial activation [146,154].

### 9.4 Therapies Depleting Lymphocyte Populations

Several DMTs work by selectively or broadly depleting lymphocytes, followed by immune reconstitution. Anti-CD20 monoclonal antibodies (rituximab, ocrelizumab, ofatumumab, ublituximab) primarily target B cells but have significant indirect effects on T-cell function. By depleting B cells, these therapies reduce antigen presentation to T cells and alter the cytokine milieu, indirectly reducing T-cell autoreactivity and proliferation [135]. *In vitro* studies confirm that rituximab suppresses T-cell proliferation and dampens the proinflammatory cytokine environment [29]. Ocrelizumab, in a study of patients with PPMS, reduced the number of naive and memory B cells while increasing the relative proportion of plasmablasts. There was also an increase in the proportion of naive CD4<sup>+</sup> and CD8<sup>+</sup> T cells and a decrease in the proportion of T cells producing IFN- $\gamma$  and a decrease in the percentage of T cells with measurable expression of CD20 [136]. More aggressive depletion strategies include alemtuzumab, a monoclonal antibody that targets CD52, which is expressed on both T and B cells. It induces profound lymphocyte depletion, followed by repopulation that favours regulatory and tolerogenic immune subsets, promoting a shift toward immune tolerance [137]. Cladribine, a purine nucleoside analogue, selectively accumulates in lymphocytes where it disrupts DNA synthesis and repair, inhibits ribonucleotide reductase, alters endonuclease activity, and ultimately leads to lymphocyte apoptosis [138]. Mitoxantrone, a cytotoxic agent, suppresses the proliferation of T cells, B cells, and macrophages, inhibits antigen presentation, reduces proin-

flammatory cytokine secretion, enhances suppressor T-cell activity, and interferes with macrophage-mediated myelin degradation [155].

### 9.5 Emerging Therapies

Current therapies for MS reduce both relapses and relapse-associated worsening of disability, which is thought to be mainly related to the transient infiltration of peripheral immune cells into the CNS. However, approved therapies are less effective in slowing the accumulation of disability in MS patients, in part due to their lack of relevant effects on inflammation in CNS compartments, where B cells and microglia are considered key and which is thought to be the cause of disability [156]. Phase-3 trials have demonstrated efficacy in progressive disease for ocrelizumab (primary progressive MS) and siponimod (secondary progressive MS); beyond these settings, effects on progression remain limited or uncertain [147,157].

BTK inhibitors aim to modulate both B-cell receptor signalling and myeloid/microglial activation, to address peripheral as well as compartmentalized inflammation. CNS exposure varies by molecule, and agents with demonstrable brain penetration are theoretically better positioned to influence microglia-associated pathology [139]. Phase 3 results have been mixed: in non-relapsing SPMS (HERCULES), investigators reported a reduction in 6-month confirmed disability progression with tolebrutinib, whereas in relapsing MS (GEMINI-1/-2), the primary endpoint on annualised relapse rate versus teriflunomide was not met, despite reported signals on disability-worsening metrics [158,159]. Conversely, evobrutinib did not show superiority on relapse outcomes in phase 3 despite earlier biomarker signals [160]. Safety profiles differ across the class; early transaminase elevations have been observed with some BTK inhibitors, so early liver-function monitoring is advisable, while longer-term risks (e.g., infection, malignancy) require further clarification [139]. Interpretation of these findings is limited by heterogeneous trial designs, relatively short follow-up periods, and the need for longer-term safety surveillance, particularly regarding hepatotoxicity and potential infectious complications. Overall, BTK inhibition remains promising but unfortunately still unproven.

The CD40-CD40L pathway represents an attractive target given its central role in T-B cell crosstalk. However, first-generation anti-CD40L antibodies were abandoned due to thromboembolic complications arising from platelet activation [161]. Frexalimab, engineered without Fc effector function, appears to have overcome this safety hurdle—no thromboembolic events were reported in phase 2 trials [140]. Nevertheless, completely blocking this fundamental co-stimulatory pathway raises theoretical concerns about the potential for opportunistic infections and impaired vaccine responses [162]. In the phase 2 trial, frexalimab demonstrated robust efficacy with an 89% reduction in new gadolinium-enhancing T1 lesions at week 12

and sustained disease control through 48 weeks, with 96% of high-dose recipients remaining free of new active lesions [140]. The drug progressively reduced plasma neurofilament light chain levels, reaching 41% reduction by week 48, suggesting potential neuroprotective effects beyond the suppression of acute inflammation [140]. The ongoing phase 3, which includes trials in both relapsing MS (NCT06141473) and non-relapsing secondary progressive MS (NCT06141486), will be critical in determining whether CD40L blockade can address compartmentalized inflammation and impact long-term disability progression where other therapies have failed [140,141].

Chimeric antigen receptor (CAR)-T cell therapy represents the most aggressive immune reconstitution strategy currently under investigation, involving *ex vivo* genetic modification of T cells to target CD19<sup>+</sup> B cells. While this approach has revolutionised the treatment of hematologic malignancies, its application in MS remains experimental. Fischbach *et al.* [142] reported initial cases in progressive MS demonstrating CSF penetration and intrathecal antibody reduction, notably, in these initial cases, without the immune effector cell-associated neurotoxicity syndrome (ICANS) that has complicated oncologic applications. Recent data presented at American Academy of Neurology (AAN) annual meeting 2025 from a small cohort of four patients with treatment-refractory secondary progressive MS showed robust CAR-T expansion in both peripheral blood and CSF, successful elimination of CD19<sup>+</sup> B-lymphocytes, and reduction in oligoclonal bands [163,164]. Although preliminary data suggest a more favourable safety profile than anticipated—with no ICANS observed despite CSF penetration and only mild cytokine release syndrome in one patient—expected cytopenias, including lymphopenia, neutropenia, and hypogammaglobulinemia, occurred [165]. However, mainly due to the limited number of cohort studies so far, these findings are preliminary, require confirmation in larger trials, and the long-term safety and efficacy of this approach remain to be established. Also, significant barriers to wider implementation remain, including the complex manufacturing process that requires apheresis and *ex vivo* cell modification, the need for lymphodepleting chemotherapy, the unknown long-term consequences of complete B-cell elimination, and questions about efficacy in progressive MS, where CNS-compartmentalized inflammation may persist despite peripheral B-cell depletion. Current phase 1 trials (NCT06451159, NCT06138132) are appropriately restricted to patients with highly active, treatment-refractory disease where conventional therapies, including anti-CD20 antibodies, have failed, and the risk-benefit ratio may justify this experimental approach.

Beyond aggressive B-cell depletion strategies, novel approaches targeting T-cell modulation are emerging. Foralumab, an intranasal anti-CD3 monoclonal antibody, represents a fundamentally different strategy—promoting regulatory T cell (Treg) expansion and suppressing effec-

tor T cell activation without causing lymphocyte depletion. By targeting nasal mucosal lymphoid tissue, this approach avoids systemic immunosuppression while potentially modulating CNS inflammation. A phase 2a trial (NCT06292923) in non-active secondary progressive MS is evaluating changes in microglial activity via the translocator protein-positron emission tomography (TSPO-PET) imaging, with preliminary data suggesting reduced fatigue and decreased microglial inflammation without neurological worsening [143]. This non-depleting approach may offer a safer alternative for patients where aggressive immunosuppression poses unacceptable risks.

## 10. Future Directions and Outstanding Questions

Despite decades of research, fundamental questions about the pathogenesis of MS remain unanswered. Perhaps the most striking paradox lies in the cellular hierarchy of the disease. B-cell depletion therapies demonstrate superior clinical efficacy, despite CD8<sup>+</sup> T cells being the predominant infiltrating population in MS lesions, which challenges our basic understanding of which cells truly drive the disease. Recent evidence suggests B cells may control myeloid cell responses through oxidative phosphorylation-regulated mechanisms, positioning them as potential master regulators rather than secondary players [83]. Yet anti-CD20 therapies, while effectively reducing relapses, fail to resolve chronic active lesions with paramagnetic rim lesions even after two years of treatment, highlighting a disconnect between clinical benefit and persistent tissue pathology [91]. Elucidating the mechanistic basis of this therapeutic paradox—why B-cell depletion confers clinical benefit despite CD8<sup>+</sup> T cell predominance in lesions—represents a key research priority [112,113]. Future studies employing spatial transcriptomics and longitudinal sampling may clarify whether B cells serve as upstream orchestrators of T-cell responses or operate through parallel pathogenic pathways.

The epidemiology of MS presents equally puzzling questions. Recent genetic studies revealed that MS risk alleles emerged in steppe pastoralist populations and spread with their migrations, yet the disease shows dramatic geographic variation that genetics alone cannot explain [166]. The role of EBV remains particularly enigmatic—while nearly universal in MS patients, the mechanistic link between infection and disease onset remains elusive. Recent serological studies have identified specific antibody signatures against the EBV peptidome that precede MS onset by years [46], and T-cell responses to EBV antigens show heightened cross-reactivity with CNS proteins in MS patients [167,168]. The discovery that up to 47% of expanded CSF T-cell clones specifically target EBV-infected B cells suggests direct pathogenic mechanisms beyond traditional molecular mimicry [50], yet this cannot explain why most EBV-infected individuals never develop MS. Prospective cohorts tracking EBV-specific T-cell responses before MS

onset, combined with deep immunophenotyping, may identify the additional host or environmental factors required for disease initiation [46,167].

Despite recognising MS as a disease continuum with overlapping inflammatory and neurodegenerative processes from onset, predicting individual disease trajectories remains impossible. The concept of PIRA, now recognised to occur even in early “stable” disease, reveals that neurodegeneration proceeds despite apparent clinical stability [169]. Furthermore, the plasticity of T-cell phenotypes within the CNS compartment during disease progression remains poorly understood - particularly how effector T cells may transition between Th1/Th17/Treg phenotypes, acquire tissue-resident memory characteristics, or shift functional programmes in response to the changing CNS microenvironment [170]. Current therapies effectively suppress acute inflammation but fail to address this insidious progression, with no biomarkers able to predict which patients will experience rapid disability accumulation versus those who remain stable for decades [171]. These observations raise the hypothesis that MS may comprise pathogenically distinct subtypes requiring tailored therapeutic approaches. Single-nucleus RNA sequencing has already demonstrated that progressive MS patients can be stratified into distinct groups based on white matter glial responses, with potential implications for treatment selection [172]. Testing this hypothesis will require integrating clinical, imaging, and molecular biomarkers, including neurofilament light chain trajectories, paramagnetic rim lesion burden, and CSF immune profiling, to stratify patients and predict treatment response [173]. Such precision medicine strategies, if validated in prospective trials, could transform MS management from empirical escalation to mechanism-based therapy selection.

## 11. Conclusion

Multiple sclerosis is a complex immune-mediated disease of the central nervous system, characterized by both inflammatory and neurodegenerative components. Among immune players, T lymphocytes—particularly CD4<sup>+</sup> Th1 and Th17, as well as cytotoxic CD8<sup>+</sup> subsets—are key drivers of demyelination, axonal damage, and BBB disruption. Conversely, Tregs play a protective role, and their dysfunction or insufficiency may exacerbate disease progression. Accumulating evidence highlights that genetic and epigenetic factors, along with environmental triggers such as EBV, smoking, vitamin D deficiency, obesity, and gut dysbiosis, modulate T-cell responses and influence MS susceptibility and course. These findings have expanded our understanding of disease mechanisms and opened avenues for targeted therapeutic interventions. DMTs modulate T-cell function through diverse mechanisms, from altering cytokine profiles (e.g., interferons, glatiramer acetate), inhibiting lymphocyte trafficking (e.g., natalizumab, S1P modulators), to inducing immune reconstitution (e.g.,

cladribine, alemtuzumab) or targeting B cell-T cell interactions (e.g., anti-CD20 antibodies). Recent agents such as fumarates act both peripherally and within the CNS, modulating T-cell subsets and promoting regulatory phenotypes via oxidative stress pathways. Novel targets are under active investigation: BTK inhibitors have shown mixed phase III results, CD40L blockade is in phase III development, and CAR T-cell therapy remains in early-phase trials. Whether these approaches can durably restore immune tolerance and impact long-term outcomes (e.g., sustained neurofilament light chain reduction, disability progression, chronic active lesion resolution) requires further validation. A deeper understanding of T-cell subpopulations and their molecular regulation can guide the development of personalised treatment strategies in MS.

## Abbreviations

AAN, American Academy of Neurology; APCs, antigen-presenting cells; AHR, aryl hydrocarbon receptor; BBB, blood-brain barrier; Bregs, regulatory B cells; BTK, Bruton's tyrosine kinase; CCR6, CC chemokine receptor 6; CRISPR-TET1, clustered regularly interspaced short palindromic repeats - ten-eleven translocation 1; CNS, central nervous system; CSF, cerebrospinal fluid; DAMP, damage associated molecular pattern; DMTs, disease modifying therapies; EAE, experimental autoimmune encephalomyelitis; EBNA1, EBV nuclear antigen 1; EBV, Epstein-Barr virus; Foxp3, Forkhead box protein 3; GM-CSF, granulocyte-macrophage colony – stimulating factor; GWAS, genome-wide association studies; HLA, human leukocyte antigen; ICANS, immune effector cell-associated neurotoxicity syndrome; Ig, immunoglobulin; IL, interleukin; IFN- $\gamma$ , interferon gamma; IFN- $\beta$ , interferon beta; JCV, John Cunningham virus; LFA-1, lymphocyte function-associated antigen 1; MAG, myelin associated glycoprotein; MBP, myelin basic protein; MFI, mean fluorescence intensity; MHC, major histocompatibility complex; miR, microRNA; MOG, myelin oligodendrocyte glycoprotein; MRI, magnetic resonance imaging; MS, multiple sclerosis; Nrf2, nuclear factor erythroid 2–related factor 2; OCBs, oligoclonal bands; OR, odds ratio; PAMP, pathogen associated molecular pattern; PLP, proteolipid protein; PML, progressive multifocal leukoencephalopathy; PP, primary progressive; PRRs, pattern-recognition receptors; PIRA, progression independent of relapse activity; pTreg, peripheral Treg; RR, relapsing-remitting; SCFA, short-chain fatty acid; SEL, slowly expanding lesions; SP, secondary progressive; STAT6, signal transducer and activator of transcription 6; SIP, sphingosine-1-phosphate; TCR, T-cell receptor; TGF $\beta$ , transforming growth factor beta; Th, helper T cells; TNF- $\alpha$ , tumor necrosis factor alpha; TLR, toll-like receptor; Tregs, regulatory T cells; TRM, tissue-resident memory T cells; Tr1, type 1 regulatory T cells; TSDR, Treg-specific demethylated region; TSPO-PET, translocator protein-positron emission tomog-

raphy; tTreg, thymic Treg; VLA-4, very late antigen-4; RIS, radiologically isolated syndrome; CAR, chimeric antigen receptor; ICAM-1, intercellular adhesion molecule 1; VCAM-1, vascular cell adhesion molecule 1.

## Author Contributions

PP performed the literature search, wrote the manuscript, and approved the final version. JV, MP prepared the figures, revised the manuscript, and approved the final version. DS, IM and MV performed the literature search, contributed to conception and manuscript editing, scientific and English revision, supervision of the work, and approved the final version. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

## Ethics Approval and Consent to Participate

Not applicable.

## Acknowledgment

Not applicable.

## Funding

This work was supported by a Czech Ministry of Health project - Conceptual Development of Research Organization - [University Hospital in Pilsen - FNPI, 00669806], by the National Health and Medical Research Council grant NU23-05-00462, by the General University Hospital in Prague project [grant number MH CZ-DRO-VFN64165] and the Charles University: Cooperatio Program in Neuroscience.

## Conflict of Interest

PP received compensations for travel, speaker honoraria and consultant fees from Biogen Idec, Novartis, Merck Serono, Roche, Sanofi Genzyme. MP received compensations for travel, and/or speaker honoraria, and/or consultant fees from Biogen, Novartis, Merck Serono, Sanofi, Roche, Janssen and Teva. MV received compensation for travel and/or speaker honoraria and/or consultant fees from Biogen, Merck Serono, Novartis, Roche, Sanofi, Teva, and Janssen-Cilag. DS has received financial support for conference travel and/or speaker honoraria from Novartis, Biogen, Merck, Teva, Janssen-Cilag, and Roche. None of the other authors has any conflict of interest to disclose.

## References

- [1] GBD 2016 Multiple Sclerosis Collaborators. Global, regional, and national burden of multiple sclerosis 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. *The Lancet. Neurology*. 2019; 18: 269–285. [https://doi.org/10.1016/S1474-4422\(18\)30443-5](https://doi.org/10.1016/S1474-4422(18)30443-5).
- [2] Prosperini L, Lucchini M, Ruggieri S, Tortorella C, Haggiag S, Mirabella M, *et al.* Shift of multiple sclerosis onset towards older

- age. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2022; 93: 1137–1139. <https://doi.org/10.1136/jnnp-2022-329049>.
- [3] Portaccio E, Magyari M, Havrdova EK, Ruet A, Brochet B, Scalfari A, *et al.* Multiple sclerosis: emerging epidemiological trends and redefining the clinical course. *The Lancet Regional Health. Europe*. 2024; 44: 100977. <https://doi.org/10.1016/j.lanepe.2024.100977>.
- [4] Jakimovski D, Bittner S, Zivadinov R, Morrow SA, Benedict RH, Zipp F, *et al.* Multiple sclerosis. *Lancet (London, England)*. 2024; 403: 183–202. [https://doi.org/10.1016/S0140-6736\(23\)01473-3](https://doi.org/10.1016/S0140-6736(23)01473-3).
- [5] Walton C, King R, Rechtman L, Kaye W, Leray E, Marrie RA, *et al.* Rising prevalence of multiple sclerosis worldwide: Insights from the Atlas of MS, third edition. *Multiple Sclerosis (Houndmills, Basingstoke, England)*. 2020; 26: 1816–1821. <https://doi.org/10.1177/1352458520970841>.
- [6] Binder MD, Nwoke EC, Morwath E, Dwyer C, Li V, Xavier A, *et al.* HLA-DRB1\*15:01 and the *MERTK* Gene Interact to Selectively Influence the Profile of *MERTK*-Expressing Monocytes in Both Health and MS. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2024; 11: e200190. <https://doi.org/10.1212/NXI.0000000000200190>.
- [7] Giovannoni G, Vanderdonck P, Hartung HP, Lassmann H, Comi G. EBV and multiple sclerosis: Setting the research agenda. *Multiple Sclerosis and Related Disorders*. 2022; 67: 104158. <https://doi.org/10.1016/j.msard.2022.104158>.
- [8] Munger KL, Zhang SM, O'Reilly E, Hernán MA, Olek MJ, Willett WC, *et al.* Vitamin D intake and incidence of multiple sclerosis. *Neurology*. 2004; 62: 60–65. <https://doi.org/10.1212/01.wnl.0000101723.79681.38>.
- [9] Manouchehrinia A, Huang J, Hillert J, Alfredsson L, Olsson T, Kockum I, *et al.* Smoking Attributable Risk in Multiple Sclerosis. *Frontiers in Immunology*. 2022; 13: 840158. <https://doi.org/10.3389/fimmu.2022.840158>.
- [10] Jiang X, Olsson T, Hillert J, Kockum I, Alfredsson L. Stressful life events are associated with the risk of multiple sclerosis. *European Journal of Neurology*. 2020; 27: 2539–2548. <https://doi.org/10.1111/ene.14458>.
- [11] Jangi S, Gandhi R, Cox LM, Li N, von Glehn F, Yan R, *et al.* Alterations of the human gut microbiome in multiple sclerosis. *Nature Communications*. 2016; 7: 12015. <https://doi.org/10.1038/ncomms12015>.
- [12] Carotenuto A, Wilson H, Giordano B, Caminiti SP, Chappell Z, Williams SCR, *et al.* Impaired connectivity within neuro-modulatory networks in multiple sclerosis and clinical implications. *Journal of Neurology*. 2020; 267: 2042–2053. <https://doi.org/10.1007/s00415-020-09806-3>.
- [13] Mistry N, Hobart J, Rog D, Muhlert N, Mathews J, Baker D, *et al.* Reconciling lesions, relapses and smouldering associated worsening: A unifying model for multiple sclerosis pathogenesis. *Multiple Sclerosis and Related Disorders*. 2024; 88: 105706. <https://doi.org/10.1016/j.msard.2024.105706>.
- [14] Absinta M, Sati P, Masuzzo F, Nair G, Sethi V, Kolb H, *et al.* Association of Chronic Active Multiple Sclerosis Lesions With Disability In Vivo. *JAMA Neurology*. 2019; 76: 1474–1483. <https://doi.org/10.1001/jamaneurol.2019.2399>.
- [15] Šťastná D, Menkyová I, Horáková D. Progresivní roztroušená skleróza ve světle nových poznatků. *Česká a Slovenská Neurologie a Neurochirurgie*. 2023; 86: 10–17. <https://doi.org/10.48095/cccsnn202310>.
- [16] Yusuf-Makagiansar H, Anderson ME, Yakovleva TV, Murray JS, Siahaan TJ. Inhibition of LFA-1/ICAM-1 and VLA-4/VCAM-1 as a therapeutic approach to inflammation and autoimmune diseases. *Medicinal Research Reviews*. 2002; 22: 146–167. <https://doi.org/10.1002/med.10001>.
- [17] Meitei HT, Jadhav N, Lal G. CCR6-CCL20 axis as a therapeutic target for autoimmune diseases. *Autoimmunity Reviews*. 2021; 20: 102846. <https://doi.org/10.1016/j.autrev.2021.102846>.
- [18] Broux B, Zandee S, Gowing E, Charabati M, Lécuyer MA, Tastet O, *et al.* Interleukin-26, preferentially produced by T<sub>H</sub>17 lymphocytes, regulates CNS barrier function. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2020; 7: e870. <https://doi.org/10.1212/NXI.0000000000000870>.
- [19] Mohammadinasr M, Montazersaheb S, Ayromlou H, Hosseini V, Molavi O, Hejazi MS. Exosome Content-Mediated Signaling Pathways in Multiple Sclerosis. *Molecular Neurobiology*. 2024; 61: 5404–5417. <https://doi.org/10.1007/s12035-023-03862-2>.
- [20] Hauser SL, Bhan AK, Gilles F, Kemp M, Kerr C, Weiner HL. Immunohistochemical analysis of the cellular infiltrate in multiple sclerosis lesions. *Annals of Neurology*. 1986; 19: 578–587. <https://doi.org/10.1002/ana.410190610>.
- [21] Moll NM, Rietsch AM, Thomas S, Ransohoff AJ, Lee JC, Fox R, *et al.* Multiple sclerosis normal-appearing white matter: pathology-imaging correlations. *Annals of Neurology*. 2011; 70: 764–773. <https://doi.org/10.1002/ana.22521>.
- [22] Baecher-Allan C, Kaskow BJ, Weiner HL. Multiple Sclerosis: Mechanisms and Immunotherapy. *Neuron*. 2018; 97: 742–768. <https://doi.org/10.1016/j.neuron.2018.01.021>.
- [23] Traugott U, Reinherz EL, Raine CS. Multiple sclerosis. Distribution of T cells, T cell subsets and Ia-positive macrophages in lesions of different ages. *Journal of Neuroimmunology*. 1983; 4: 201–221. [https://doi.org/10.1016/0165-5728\(83\)90036-x](https://doi.org/10.1016/0165-5728(83)90036-x).
- [24] Kutzelnigg A, Lucchinetti CF, Stadelmann C, Brück W, Rauschka H, Bergmann M, *et al.* Cortical demyelination and diffuse white matter injury in multiple sclerosis. *Brain: a Journal of Neurology*. 2005; 128: 2705–2712. <https://doi.org/10.1093/brain/awh641>.
- [25] Beecham AH, Patsopoulos NA, Xifara DK, Davis MF, Kempinen A, Cotsapas C, *et al.* Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis. *Nature Genetics*. 2013; 45: 1353–1360. <https://doi.org/10.1038/ng.2770>.
- [26] International Multiple Sclerosis Genetics Consortium. Multiple sclerosis genomic map implicates peripheral immune cells and microglia in susceptibility. *Science (New York, N.Y.)*. 2019; 365: eaav7188. <https://doi.org/10.1126/science.aav7188>.
- [27] Chitnis T. The role of CD4 T cells in the pathogenesis of multiple sclerosis. *International Review of Neurobiology*. 2007; 79: 43–72. [https://doi.org/10.1016/S0074-7742\(07\)79003-7](https://doi.org/10.1016/S0074-7742(07)79003-7).
- [28] Hollenbach JA, Oksenberg JR. The immunogenetics of multiple sclerosis: A comprehensive review. *Journal of Autoimmunity*. 2015; 64: 13–25. <https://doi.org/10.1016/j.jaut.2015.06.010>.
- [29] Jelcic I, Al Nimer F, Wang J, Lentsch V, Planas R, Jelcic I, *et al.* Memory B Cells Activate Brain-Homing, Autoreactive CD4<sup>+</sup> T Cells in Multiple Sclerosis. *Cell*. 2018; 175: 85–100.e23. <https://doi.org/10.1016/j.cell.2018.08.011>.
- [30] Maciak K, Dziedzic A, Miller E, Saluk-Bijak J. miR-155 as an Important Regulator of Multiple Sclerosis Pathogenesis. A Review. *International Journal of Molecular Sciences*. 2021; 22: 4332. <https://doi.org/10.3390/ijms22094332>.
- [31] Zhang J, Cheng Y, Cui W, Li M, Li B, Guo L. MicroRNA-155 modulates Th1 and Th17 cell differentiation and is associated with multiple sclerosis and experimental autoimmune encephalomyelitis. *Journal of Neuroimmunology*. 2014; 266: 56–63. <https://doi.org/10.1016/j.jneuroim.2013.09.019>.
- [32] Setiadi AF, Abbas AR, Jeet S, Wong K, Bischof A, Peng I, *et al.* IL-17A is associated with the breakdown of the blood-brain barrier in relapsing-remitting multiple sclerosis. *Journal of Neuroimmunology*. 2019; 332: 147–154. <https://doi.org/10.1016/j.jneuroim.2019.04.011>.
- [33] Munk Nielsen N, Corn G, Frisch M, Stenager E, Koch-Henriksen N, Wohlfahrt J, *et al.* Multiple sclerosis among first-

- and second-generation immigrants in Denmark: a population-based cohort study. *Brain: a Journal of Neurology*. 2019; 142: 1587–1597. <https://doi.org/10.1093/brain/awz088>.
- [34] Willer CJ, Dymnt DA, Risch NJ, Sadovnick AD, Ebers GC, Canadian Collaborative Study Group. Twin concordance and sibling recurrence rates in multiple sclerosis. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100: 12877–12882. <https://doi.org/10.1073/pnas.1932604100>.
- [35] Palacios N, Alonso A, Brønnum-Hansen H, Ascherio A. Smoking and increased risk of multiple sclerosis: parallel trends in the sex ratio reinforce the evidence. *Annals of Epidemiology*. 2011; 21: 536–542. <https://doi.org/10.1016/j.annepidem.2011.03.001>.
- [36] Alrouji M, Manouchehrinia A, Gran B, Constantinescu CS. Effects of cigarette smoke on immunity, neuroinflammation and multiple sclerosis. *Journal of Neuroimmunology*. 2019; 329: 24–34. <https://doi.org/10.1016/j.jneuroim.2018.10.004>.
- [37] Helm EY, Zhou L. Transcriptional regulation of innate lymphoid cells and T cells by aryl hydrocarbon receptor. *Frontiers in Immunology*. 2023; 14: 1056267. <https://doi.org/10.3389/fimmu.2023.1056267>.
- [38] Saaoud F, Shao Y, Cornwell W, Wang H, Rogers TJ, Yang X. Cigarette Smoke Modulates Inflammation and Immunity via Reactive Oxygen Species-Regulated Trained Immunity and Trained Tolerance Mechanisms. *Antioxidants & Redox Signaling*. 2023; 38: 1041–1069. <https://doi.org/10.1089/ars.2022.0087>.
- [39] Carlberg C, Mycko MP. Linking Mechanisms of Vitamin D Signaling with Multiple Sclerosis. *Cells*. 2023; 12: 2391. <https://doi.org/10.3390/cells12192391>.
- [40] Killick J, Hay J, Morandi E, Vermeren S, Kari S, Angles T, *et al.* Vitamin D/CD46 Crosstalk in Human T Cells in Multiple Sclerosis. *Frontiers in Immunology*. 2020; 11: 598727. <https://doi.org/10.3389/fimmu.2020.598727>.
- [41] Fetahu IS, Höbaus J, Kállay E. Vitamin D and the epigenome. *Frontiers in Physiology*. 2014; 5: 164. <https://doi.org/10.3389/fphys.2014.00164>.
- [42] Kowalówka M, Górna I, Karaźniewicz-Lada M, Kusiak D, Przysławski J, Drzymała-Czyż S. Nutri-Epigenetic Regulation of Vitamin D-Impact on Metabolism and Biological Functions: Narrative Review. *Metabolites*. 2025; 15: 436. <https://doi.org/10.3390/metabo15070436>.
- [43] Vachova M, Stastna D, Mazouchova A, Hruskova P, Uher T, Preiningerova Lizrova J, *et al.* From sunlight to MS fight: impact of vitamin D levels on multiple sclerosis activity. *Neurological Sciences: Official Journal of the Italian Neurological Society and of the Italian Society of Clinical Neurophysiology*. 2025; 47: 38. <https://doi.org/10.1007/s10072-025-08729-z>.
- [44] Handel AE, Williamson AJ, Disanto G, Handunnetthi L, Giovannoni G, Ramagopalan SV. An updated meta-analysis of risk of multiple sclerosis following infectious mononucleosis. *PloS One*. 2010; 5: e12496. <https://doi.org/10.1371/journal.pone.0012496>.
- [45] Loosen SH, Doege C, Meuth SG, Luedde T, Kostev K, Roderburg C. Infectious mononucleosis is associated with an increased incidence of multiple sclerosis: Results from a cohort study of 32,116 outpatients in Germany. *Frontiers in Immunology*. 2022; 13: 937583. <https://doi.org/10.3389/fimmu.2022.937583>.
- [46] Bjornevik K, Cortese M, Healy BC, Kuhle J, Mina MJ, Leng Y, *et al.* Longitudinal analysis reveals high prevalence of Epstein-Barr virus associated with multiple sclerosis. *Science (New York, N.Y.)*. 2022; 375: 296–301. <https://doi.org/10.1126/science.abj8222>.
- [47] Lünemann JD, Edwards N, Muraro PA, Hayashi S, Cohen JL, Münz C, *et al.* Increased frequency and broadened specificity of latent EBV nuclear antigen-1-specific T cells in multiple sclerosis. *Brain: a Journal of Neurology*. 2006; 129: 1493–1506. <https://doi.org/10.1093/brain/awl067>.
- [48] Schneider-Hohendorf T, Wunsch C, Falk S, Raposo C, Rubelt F, Mirebrahim H, *et al.* Broader anti-EBV TCR repertoire in multiple sclerosis: disease specificity and treatment modulation. *Brain: a Journal of Neurology*. 2025; 148: 933–940. <https://doi.org/10.1093/brain/awae244>.
- [49] Mohammadinasr M, Montazersaheb S, Hosseini V, Kahroba H, Talebi M, Molavi O, *et al.* Epstein-Barr virus-encoded BART9 and BART15 miRNAs are elevated in exosomes of cerebrospinal fluid from relapsing-remitting multiple sclerosis patients. *Cytokine*. 2024; 179: 156624. <https://doi.org/10.1016/j.cyto.2024.156624>.
- [50] Gottlieb A, Pham HPT, Saltarrelli JG, Lindsey JW. Expanded T lymphocytes in the cerebrospinal fluid of multiple sclerosis patients are specific for Epstein-Barr-virus-infected B cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2024; 121: e2315857121. <https://doi.org/10.1073/pnas.2315857121>.
- [51] Stampanoni Bassi M, Iezzi E, Buttari F, Gilio L, Simonelli I, Carbone F, *et al.* Obesity worsens central inflammation and disability in multiple sclerosis. *Multiple Sclerosis (Houndmills, Basingstoke, England)*. 2020; 26: 1237–1246. <https://doi.org/10.1177/1352458519853473>.
- [52] Cox LM, Maghzi AH, Liu S, Tankou SK, Dhang FH, Willocq V, *et al.* Gut Microbiome in Progressive Multiple Sclerosis. *Annals of Neurology*. 2021; 89: 1195–1211. <https://doi.org/10.1002/ana.26084>.
- [53] Chen J, Chia N, Kalari KR, Yao JZ, Novotna M, Paz Soldan MM, *et al.* Multiple sclerosis patients have a distinct gut microbiota compared to healthy controls. *Scientific Reports*. 2016; 6: 28484. <https://doi.org/10.1038/srep28484>.
- [54] Hu M, Alashkar Alhamwe B, Santner-Nanan B, Mieth S, Harb H, Renz H, *et al.* Short-Chain Fatty Acids Augment Differentiation and Function of Human Induced Regulatory T Cells. *International Journal of Molecular Sciences*. 2022; 23: 5740. <https://doi.org/10.3390/ijms23105740>.
- [55] Duscha A, Gisevius B, Hirschberg S, Yissachar N, Stangl GI, Dawin E, *et al.* Propionic Acid Shapes the Multiple Sclerosis Disease Course by an Immunomodulatory Mechanism. *Cell*. 2020; 180: 1067–1080.e16. <https://doi.org/10.1016/j.cell.2020.02.035>.
- [56] Sun M, Wu W, Chen L, Yang W, Huang X, Ma C, *et al.* Microbiota-derived short-chain fatty acids promote Th1 cell IL-10 production to maintain intestinal homeostasis. *Nature Communications*. 2018; 9: 3555. <https://doi.org/10.1038/s41467-018-05901-2>.
- [57] Nouri M, Bredberg A, Weström B, Lavasani S. Intestinal barrier dysfunction develops at the onset of experimental autoimmune encephalomyelitis, and can be induced by adoptive transfer of auto-reactive T cells. *PloS One*. 2014; 9: e106335. <https://doi.org/10.1371/journal.pone.0106335>.
- [58] Pröbstel AK, Zhou X, Baumann R, Wischniewski S, Kutza M, Rojas OL, *et al.* Gut microbiota-specific IgA<sup>+</sup> B cells traffic to the CNS in active multiple sclerosis. *Science Immunology*. 2020; 5: eabc7191. <https://doi.org/10.1126/sciimmunol.abc7191>.
- [59] Takewaki D, Suda W, Sato W, Takayasu L, Kumar N, Kimura K, *et al.* Alterations of the gut ecological and functional microenvironment in different stages of multiple sclerosis. *Proceedings of the National Academy of Sciences of the United States of America*. 2020; 117: 22402–22412. <https://doi.org/10.1073/pnas.2011703117>.
- [60] Preiningerova JL, Jiraskova Zakostelska Z, Srinivasan A, Ticha V, Kovarova I, Kleinova P, *et al.* Multiple Sclerosis and Microbiome. *Biomolecules*. 2022; 12: 433. <https://doi.org/10.3390/bi12040433>.

om12030433.

- [61] Krejssek J. Defensive and damaging inflammation: basic characteristics. *Vnitřní lékařství*. 2019; 65: 76–80. <https://doi.org/10.36290/vnl.2019.019>. (In Czech)
- [62] Svobodová M, Ščouráč P. Role specifické buněčné imunity v patogenezi roztroušené sklerózy se zaměřením na Th17 a Treg lymfocyty. *Česká a Slovenská Neurologie a Neurochirurgie*. 2017; 80: 173–179. <https://doi.org/10.14735/amcsnn2017173>. (In Czech)
- [63] Bravo GÁ, Cedeño RR, Casadevall MP, Ramió-Torrentà L. Sphingosine-1-Phosphate (S1P) and S1P Signaling Pathway Modulators, from Current Insights to Future Perspectives. *Cells*. 2022; 11: 2058. <https://doi.org/10.3390/cells11132058>.
- [64] Dhaiban S, Al-Ani M, Elemam NM, Maghazachi AA. Targeting Chemokines and Chemokine Receptors in Multiple Sclerosis and Experimental Autoimmune Encephalomyelitis. *Journal of Inflammation Research*. 2020; 13: 619–633. <https://doi.org/10.2147/JIR.S270872>.
- [65] Mastorakos P, McGavern D. The anatomy and immunology of vasculature in the central nervous system. *Science Immunology*. 2019; 4: eaav0492. <https://doi.org/10.1126/sciimmunol.av0492>.
- [66] Alter A, Duddy M, Hebert S, Biernacki K, Prat A, Antel JP, *et al*. Determinants of human B cell migration across brain endothelial cells. *Journal of Immunology (Baltimore, Md.: 1950)*. 2003; 170: 4497–4505. <https://doi.org/10.4049/jimmunol.170.9.4497>.
- [67] Hartung HP, Rieckmann P. Pathogenesis of immune-mediated demyelination in the CNS. *Journal of Neural Transmission. Supplementum*. 1997; 50: 173–181. [https://doi.org/10.1007/978-3-7091-6842-4\\_17](https://doi.org/10.1007/978-3-7091-6842-4_17).
- [68] Machado-Santos J, Saji E, Tröschler AR, Paunovic M, Liblau R, Gabrieli G, *et al*. The compartmentalized inflammatory response in the multiple sclerosis brain is composed of tissue-resident CD8<sup>+</sup> T lymphocytes and B cells. *Brain: a Journal of Neurology*. 2018; 141: 2066–2082. <https://doi.org/10.1093/brain/awy151>.
- [69] Molnarfi N, Schulze-Topphoff U, Weber MS, Patarroyo JC, Prod'homme T, Varrin-Doyer M, *et al*. MHC class II-dependent B cell APC function is required for induction of CNS autoimmunity independent of myelin-specific antibodies. *The Journal of Experimental Medicine*. 2013; 210: 2921–2937. <https://doi.org/10.1084/jem.20130699>.
- [70] Li R, Rezk A, Miyazaki Y, Hilgenberg E, Touil H, Shen P, *et al*. Proinflammatory GM-CSF-producing B cells in multiple sclerosis and B cell depletion therapy. *Science Translational Medicine*. 2015; 7: 310ra166. <https://doi.org/10.1126/scitranslmed.aab4176>.
- [71] Villar LM, Masterman T, Casanova B, Gómez-Rial J, Espiño M, Sádaba MC, *et al*. CSF oligoclonal band patterns reveal disease heterogeneity in multiple sclerosis. *Journal of Neuroimmunology*. 2009; 211: 101–104. <https://doi.org/10.1016/j.jneuroim.2009.03.003>.
- [72] Ireland S, Monson N. Potential impact of B cells on T cell function in multiple sclerosis. *Multiple Sclerosis International*. 2011; 2011: 423971. <https://doi.org/10.1155/2011/423971>.
- [73] Pukoli D, Vécsei L. Smouldering Lesion in MS: Microglia, Lymphocytes and Pathobiochemical Mechanisms. *International Journal of Molecular Sciences*. 2023; 24: 12631. <https://doi.org/10.3390/ijms241612631>.
- [74] Kuhlmann T, Lassmann H, Brück W. Diagnosis of inflammatory demyelination in biopsy specimens: a practical approach. *Acta Neuropathologica*. 2008; 115: 275–287. <https://doi.org/10.1007/s00401-007-0320-8>.
- [75] Van Kaer L, Postoak JL, Wang C, Yang G, Wu L. Innate, innate-like and adaptive lymphocytes in the pathogenesis of MS and EAE. *Cellular & Molecular Immunology*. 2019; 16: 531–539. <https://doi.org/10.1038/s41423-019-0221-5>.
- [76] Frischer JM, Weigand SD, Guo Y, Kale N, Parisi JE, Pirko I, *et al*. Clinical and pathological insights into the dynamic nature of the white matter multiple sclerosis plaque. *Annals of Neurology*. 2015; 78: 710–721. <https://doi.org/10.1002/ana.24497>.
- [77] Howell OW, Reeves CA, Nicholas R, Carassiti D, Radotra B, Gentleman SM, *et al*. Meningeal inflammation is widespread and linked to cortical pathology in multiple sclerosis. *Brain: a Journal of Neurology*. 2011; 134: 2755–2771. <https://doi.org/10.1093/brain/awr182>.
- [78] Barros C, Alberro A, Fernandes A. Microglia and Immune cells interactions in multiple sclerosis cognitive impairment: a post-mortem study. *Journal of Neuroinflammation*. 2024; 21: 332. <https://doi.org/10.1186/s12974-024-03326-x>.
- [79] Calabrese M, Preziosa P, Scafari A, Colato E, Marastoni D, Absinta M, *et al*. Determinants and Biomarkers of Progression Independent of Relapses in Multiple Sclerosis. *Annals of Neurology*. 2024; 96: 1–20. <https://doi.org/10.1002/ana.26913>.
- [80] Dong Y, Yong VW. When encephalitogenic T cells collaborate with microglia in multiple sclerosis. *Nature Reviews. Neurology*. 2019; 15: 704–717. <https://doi.org/10.1038/s41582-019-0253-6>.
- [81] Guerrero BL, Sicotte NL. Microglia in Multiple Sclerosis: Friend or Foe? *Frontiers in Immunology*. 2020; 11: 374. <https://doi.org/10.3389/fimmu.2020.00374>.
- [82] Absinta M, Maric D, Gharagozloo M, Garton T, Smith MD, Jin J, *et al*. A lymphocyte-microglia-astrocyte axis in chronic active multiple sclerosis. *Nature*. 2021; 597: 709–714. <https://doi.org/10.1038/s41586-021-03892-7>.
- [83] Li R, Lei Y, Rezk A, Diego A Espinoza, Wang J, Feng H, *et al*. Oxidative phosphorylation regulates B cell effector cytokines and promotes inflammation in multiple sclerosis. *Science Immunology*. 2024; 9: eadk0865. <https://doi.org/10.1126/sciimmunol.adk0865>.
- [84] Kumar G, Axtell RC. Dual Role of B Cells in Multiple Sclerosis. *International Journal of Molecular Sciences*. 2023; 24: 2336. <https://doi.org/10.3390/ijms24032336>.
- [85] Li R, Patterson KR, Bar-Or A. Reassessing B cell contributions in multiple sclerosis. *Nature Immunology*. 2018; 19: 696–707. <https://doi.org/10.1038/s41590-018-0135-x>.
- [86] Barr TA, Shen P, Brown S, Lampropoulou V, Roch T, Lawrie S, *et al*. B cell depletion therapy ameliorates autoimmune disease through ablation of IL-6-producing B cells. *The Journal of Experimental Medicine*. 2012; 209: 1001–1010. <https://doi.org/10.1084/jem.20111675>.
- [87] Tzartos JS, Craner MJ, Friese MA, Jakobsen KB, Newcombe J, Esiri MM, *et al*. IL-21 and IL-21 receptor expression in lymphocytes and neurons in multiple sclerosis brain. *The American Journal of Pathology*. 2011; 178: 794–802. <https://doi.org/10.1016/j.ajpath.2010.10.043>.
- [88] Jackson SW, Jacobs HM, Arkatkar T, Dam EM, Scharping NE, Kolhatkar NS, *et al*. B cell IFN- $\gamma$  receptor signaling promotes autoimmune germinal centers via cell-intrinsic induction of BCL-6. *The Journal of Experimental Medicine*. 2016; 213: 733–750. <https://doi.org/10.1084/jem.20151724>.
- [89] Bsibsi M, Peferoen LAN, Holtman IR, Nacken PJ, Gerritsen WH, Witte ME, *et al*. Demyelination during multiple sclerosis is associated with combined activation of microglia/macrophages by IFN- $\gamma$  and alpha B-crystallin. *Acta Neuropathologica*. 2014; 128: 215–229. <https://doi.org/10.1007/s00401-014-1317-8>.
- [90] van Langelaar J, Rijvers L, Janssen M, Wierenga-Wolf AF, Melief MJ, Siepmann TA, *et al*. Induction of brain-infiltrating Tbet-expressing B cells in multiple sclerosis. *Annals of Neurology*. 2019; 86: 264–278. <https://doi.org/10.1002/ana.25508>.
- [91] Maggi P, Bulcke CV, Pedrini E, Bugli C, Sellimi A, Wynen M, *et al*. B cell depletion therapy does not resolve chronic active

- multiple sclerosis lesions. *EBioMedicine*. 2023; 94: 104701. <https://doi.org/10.1016/j.ebiom.2023.104701>.
- [92] Kuhlmann T, Moccia M, Coetzee T, Cohen JA, Correale J, Graves J, *et al.* Multiple sclerosis progression: time for a new mechanism-driven framework. *The Lancet. Neurology*. 2023; 22: 78–88. [https://doi.org/10.1016/S1474-4422\(22\)00289-7](https://doi.org/10.1016/S1474-4422(22)00289-7).
- [93] Lassmann H, Ransohoff RM. The CD4-Th1 model for multiple sclerosis: a crucial re-appraisal. *Trends in Immunology*. 2004; 25: 132–137. <https://doi.org/10.1016/j.it.2004.01.007>.
- [94] Denic A, Wootla B, Rodriguez M. CD8(+) T cells in multiple sclerosis. *Expert Opinion on Therapeutic Targets*. 2013; 17: 1053–1066. <https://doi.org/10.1517/14728222.2013.815726>.
- [95] Chiarini M, Serana F, Zanotti C, Capra R, Rasia S, Rotoli M, *et al.* Modulation of the central memory and Tr1-like regulatory T cells in multiple sclerosis patients responsive to interferon-beta therapy. *Multiple Sclerosis (Houndmills, Basingstoke, England)*. 2012; 18: 788–798. <https://doi.org/10.1177/1352458511427720>.
- [96] Lovett-Racke AE, Yang Y, Racke MK. Th1 versus Th17: are T cell cytokines relevant in multiple sclerosis? *Biochimica et Biophysica Acta*. 2011; 1812: 246–251. <https://doi.org/10.1016/j.bbdis.2010.05.012>.
- [97] Rostami A, Ciric B. Role of Th17 cells in the pathogenesis of CNS inflammatory demyelination. *Journal of the Neurological Sciences*. 2013; 333: 76–87. <https://doi.org/10.1016/j.jns.2013.03.002>.
- [98] Nyirenda MH, Morandi E, Vinkemeier U, Constantin-Teodosiu D, Drinkwater S, Mee M, *et al.* TLR2 stimulation regulates the balance between regulatory T cell and Th17 function: a novel mechanism of reduced regulatory T cell function in multiple sclerosis. *Journal of Immunology (Baltimore, Md.: 1950)*. 2015; 194: 5761–5774. <https://doi.org/10.4049/jimmunol.1400472>.
- [99] Maier E, Duschl A, Horejs-Hoeck J. STAT6-dependent and -independent mechanisms in Th2 polarization. *European Journal of Immunology*. 2012; 42: 2827–2833. <https://doi.org/10.1002/eji.201242433>.
- [100] Šťastná D, Seňavová J, Horáková D. Terapie roztroušené sklerózy zaměřená na B-lymfocyty: od teorie po dekádu ocrelizumabu v praxi. *Neurologie Pro Praxi*. 2024; 25: 124–129. <https://doi.org/10.36290/neu.2024.011>.
- [101] Perumal NB, Kaplan MH. Regulating IL9 transcription in T helper cells. *Trends in Immunology*. 2011; 32: 146–150. <https://doi.org/10.1016/j.it.2011.01.006>.
- [102] Guadalupi L, Vanni V, Balletta S, Caioli S, De Vito F, Fresegna D, *et al.* Interleukin-9 protects from microglia- and TNF-mediated synaptotoxicity in experimental multiple sclerosis. *Journal of Neuroinflammation*. 2024; 21: 128. <https://doi.org/10.1186/s12974-024-03120-9>.
- [103] Dargahi N, Katsara M, Tselios T, Androutsou ME, de Courten M, Matsoukas J, *et al.* Multiple Sclerosis: Immunopathology and Treatment Update. *Brain Sciences*. 2017; 7: 78. <https://doi.org/10.3390/brainsci7070078>.
- [104] Wu X, Tian J, Wang S. Insight Into Non-Pathogenic Th17 Cells in Autoimmune Diseases. *Frontiers in Immunology*. 2018; 9: 1112. <https://doi.org/10.3389/fimmu.2018.01112>.
- [105] Wang J, Zhao X, Wan YY. Intricacies of TGF- $\beta$  signaling in Treg and Th17 cell biology. *Cellular & Molecular Immunology*. 2023; 20: 1002–1022. <https://doi.org/10.1038/s41423-023-01036-7>.
- [106] Zambrano-Zaragoza JF, Romo-Martínez EJ, Durán-Avelar MDJ, García-Magallanes N, Vibanco-Pérez N. Th17 cells in autoimmune and infectious diseases. *International Journal of Inflammation*. 2014; 2014: 651503. <https://doi.org/10.1155/2014/651503>.
- [107] Starr TK, Jameson SC, Hogquist KA. Positive and negative selection of T cells. *Annual Review of Immunology*. 2003; 21: 139–176. <https://doi.org/10.1146/annurev.immunol.21.120601.141107>.
- [108] Cantoni C, Smirnov RA, Firulyova M, Andhey PS, Bradstreet TR, Esaulova E, *et al.* A single-cell compendium of human cerebrospinal fluid identifies disease-associated immune cell populations. *The Journal of Clinical Investigation*. 2025; 135: e177793. <https://doi.org/10.1172/JCI177793>.
- [109] Straeten F, Zhu J, Börsch AL, Zhang B, Li K, Lu IN, *et al.* Integrated single-cell transcriptomics of cerebrospinal fluid cells in treatment-naïve multiple sclerosis. *Journal of Neuroinflammation*. 2022; 19: 306. <https://doi.org/10.1186/s12974-022-02667-9>.
- [110] Ostkamp P, Deffner M, Schulte-Mecklenbeck A, Wunsch C, Lu IN, Wu GF, *et al.* A single-cell analysis framework allows for characterization of CSF leukocytes and their tissue of origin in multiple sclerosis. *Science Translational Medicine*. 2022; 14: eadc9778. <https://doi.org/10.1126/scitranslmed.adc9778>.
- [111] Schnell A, Littman DR, Kuchroo VK. T<sub>H</sub>17 cell heterogeneity and its role in tissue inflammation. *Nature Immunology*. 2023; 24: 19–29. <https://doi.org/10.1038/s41590-022-01387-9>.
- [112] Lerma-Martin C, Badia-I-Mompel P, Ramirez Flores RO, Sekol P, Schäfer PSL, Riedl CJ, *et al.* Cell type mapping reveals tissue niches and interactions in subcortical multiple sclerosis lesions. *Nature Neuroscience*. 2024; 27: 2354–2365. <https://doi.org/10.1038/s41593-024-01796-z>.
- [113] Alsema AM, Wijering MHC, Miedema A, Kotah JM, Koster M, Rijnsburger M, *et al.* Spatially resolved gene signatures of white matter lesion progression in multiple sclerosis. *Nature Neuroscience*. 2024; 27: 2341–2353. <https://doi.org/10.1038/s41593-024-01765-6>.
- [114] Yshii L, Pasciuto E, Bielefeld P, Mascali L, Lemaitre P, Marino M, *et al.* Astrocyte-targeted gene delivery of interleukin 2 specifically increases brain-resident regulatory T cell numbers and protects against pathological neuroinflammation. *Nature Immunology*. 2022; 23: 878–891. <https://doi.org/10.1038/s41590-022-01208-z>.
- [115] Korn T, Bettelli E, Gao W, Awasthi A, Jäger A, Strom TB, *et al.* IL-21 initiates an alternative pathway to induce proinflammatory T(H)17 cells. *Nature*. 2007; 448: 484–487. <https://doi.org/10.1038/nature05970>.
- [116] Bar-Or A, Li R. Cellular immunology of relapsing multiple sclerosis: interactions, checks, and balances. *The Lancet. Neurology*. 2021; 20: 470–483. [https://doi.org/10.1016/S1474-4422\(21\)00063-6](https://doi.org/10.1016/S1474-4422(21)00063-6).
- [117] Verma ND, Lam AD, Chiu C, Tran GT, Hall BM, Hodgkinson SJ. Multiple sclerosis patients have reduced resting and increased activated CD4<sup>+</sup>CD25<sup>+</sup>FOXP3<sup>+</sup>T regulatory cells. *Scientific Reports*. 2021; 11: 10476. <https://doi.org/10.1038/s41598-021-88448-5>.
- [118] Sambucci M, Gargano F, De Rosa V, De Bardi M, Picozza M, Placido R, *et al.* FoxP3 isoforms and PD-1 expression by T regulatory cells in multiple sclerosis. *Scientific Reports*. 2018; 8: 3674. <https://doi.org/10.1038/s41598-018-21861-5>.
- [119] Venken K, Hellings N, Thewissen M, Somers V, Hensen K, Rummens JL, *et al.* Compromised CD4<sup>+</sup> CD25(high) regulatory T-cell function in patients with relapsing-remitting multiple sclerosis is correlated with a reduced frequency of FOXP3-positive cells and reduced FOXP3 expression at the single-cell level. *Immunology*. 2008; 123: 79–89. <https://doi.org/10.1111/j.1365-2567.2007.02690.x>.
- [120] Danikowski KM, Jayaraman S, Prabhakar BS. Regulatory T cells in multiple sclerosis and myasthenia gravis. *Journal of Neuroinflammation*. 2017; 14: 117. <https://doi.org/10.1186/s12974-017-0892-8>.
- [121] Yue Y, Ren Y, Lu C, Li P, Zhang G. Epigenetic regulation of human FOXP3<sup>+</sup> Tregs: from homeostasis maintenance to

- pathogen defense. *Frontiers in Immunology*. 2024; 15: 1444533. <https://doi.org/10.3389/fimmu.2024.1444533>.
- [122] Gootjes C, Zwaginga JJ, Roep BO, Nikolic T. Defining Human Regulatory T Cells beyond FOXP3: The Need to Combine Phenotype with Function. *Cells*. 2024; 13: 941. <https://doi.org/10.3390/cells13110941>.
- [123] Arroyo-Olarte RD, Flores-Castelán JC, Armas-López L, Escobedo G, Terrazas LI, Ávila-Moreno F, *et al.* Targeted Demethylation of FOXP3-TSDR Enhances the Suppressive Capacity of STAT6-deficient Inducible T Regulatory Cells. *Inflammation*. 2024; 47: 2159–2172. <https://doi.org/10.1007/s10753-024-02031-4>.
- [124] Dominguez-Villar M, Baecher-Allan CM, Hafler DA. Identification of T helper type 1-like, Foxp3+ regulatory T cells in human autoimmune disease. *Nature Medicine*. 2011; 17: 673–675. <https://doi.org/10.1038/nm.2389>.
- [125] Koutouros M, Berer K, Kawakami N, Wekerle H, Krishnamoorthy G. Treg cells mediate recovery from EAE by controlling effector T cell proliferation and motility in the CNS. *Acta Neuropathologica Communications*. 2014; 2: 163. <https://doi.org/10.1186/s40478-014-0163-1>.
- [126] O'Connor RA, Anderton SM. Foxp3+ regulatory T cells in the control of experimental CNS autoimmune disease. *Journal of Neuroimmunology*. 2008; 193: 1–11. <https://doi.org/10.1016/j.jneuroim.2007.11.016>.
- [127] Dombrowski Y, O'Hagan T, Dittmer M, Penalva R, Mayoral SR, Bankhead P, *et al.* Regulatory T cells promote myelin regeneration in the central nervous system. *Nature Neuroscience*. 2017; 20: 674–680. <https://doi.org/10.1038/nn.4528>.
- [128] Dendrou CA, Fugger L, Friese MA. Immunopathology of multiple sclerosis. *Nature Reviews. Immunology*. 2015; 15: 545–558. <https://doi.org/10.1038/nri3871>.
- [129] Jakimovski D, Kolb C, Ramanathan M, Zivadinov R, Weinstock-Guttman B. Interferon  $\beta$  for Multiple Sclerosis. *Cold Spring Harbor Perspectives in Medicine*. 2018; 8: a032003. <https://doi.org/10.1101/cshperspect.a032003>.
- [130] Bresciani G, Manai F, Davinelli S, Tucci P, Saso L, Amadio M. Novel potential pharmacological applications of dimethyl fumarate—an overview and update. *Frontiers in Pharmacology*. 2023; 14: 1264842. <https://doi.org/10.3389/fphar.2023.1264842>.
- [131] Diebold M, Sievers C, Bantug G, Sanderson N, Kappos L, Kuhle J, *et al.* Dimethyl fumarate influences innate and adaptive immunity in multiple sclerosis. *Journal of Autoimmunity*. 2018; 86: 39–50. <https://doi.org/10.1016/j.jaut.2017.09.009>.
- [132] Tilly G, Cadoux M, Garcia A, Morille J, Wiertlewski S, Pecqueur C, *et al.* Teriflunomide Treatment of Multiple Sclerosis Selectively Modulates CD8 Memory T Cells. *Frontiers in Immunology*. 2021; 12: 730342. <https://doi.org/10.3389/fimmu.2021.730342>.
- [133] Polman CH, O'Connor PW, Havrdova E, Hutchinson M, Kappos L, Miller DH, *et al.* A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *The New England Journal of Medicine*. 2006; 354: 899–910. <https://doi.org/10.1056/NEJMoa044397>.
- [134] Bordet R, Camu W, De Zeze J, Laplaud DA, Ouallet JC, Thouvenot E. Mechanism of action of s1p receptor modulators in multiple sclerosis: The double requirement. *Revue Neurologique*. 2020; 176: 100–112. <https://doi.org/10.1016/j.neurol.2019.02.007>.
- [135] von Essen MR, Hansen RH, Højgaard C, Ammitzbøll C, Wiendl H, Sellebjerg F. Ofatumumab Modulates Inflammatory T Cell Responses and Migratory Potential in Patients With Multiple Sclerosis. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2022; 9: e200004. <https://doi.org/10.1212/NXI.000000000200004>.
- [136] Fernández-Velasco JI, Kuhle J, Monreal E, Meca-Lallana V, Meca-Lallana J, Izquierdo G, *et al.* Effect of Ocrelizumab in Blood Leukocytes of Patients With Primary Progressive MS. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2021; 8: e940. <https://doi.org/10.1212/NXI.0000000000000940>.
- [137] Evan JR, Bozkurt SB, Thomas NC, Bagnato F. Alemtuzumab for the treatment of multiple sclerosis. *Expert Opinion on Biological Therapy*. 2018; 18: 323–334. <https://doi.org/10.1080/14712598.2018.1425388>.
- [138] Jacobs BM, Ammoscato F, Giovannoni G, Baker D, Schmierer K. Cladribine: mechanisms and mysteries in multiple sclerosis. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2018; 89: 1266–1271. <https://doi.org/10.1136/jnnp-2017-317411>.
- [139] Krämer J, Bar-Or A, Turner TJ, Wiendl H. Bruton tyrosine kinase inhibitors for multiple sclerosis. *Nature Reviews. Neurology*. 2023; 19: 289–304. <https://doi.org/10.1038/s41582-023-00800-7>.
- [140] Vermersch P, Granziera C, Mao-Draayer Y, Cutter G, Kalbus O, Staikov I, *et al.* Inhibition of CD40L with Frexalimab in Multiple Sclerosis. *The New England Journal of Medicine*. 2024; 390: 589–600. <https://doi.org/10.1056/NEJMoa2309439>.
- [141] Vermersch P, Wagner D, Mars LT, Noelle R, Giovannoni G. Inhibiting CD40 Ligand in Multiple Sclerosis: A Review of Emerging Therapeutic Potential. *Current Treatment Options in Neurology*. 2025; 27: 7. <https://doi.org/10.1007/s11940-024-00818-2>.
- [142] Fischbach F, Richter J, Pfeffer LK, Fehse B, Berger SC, Reinhardt S, *et al.* CD19-targeted chimeric antigen receptor T cell therapy in two patients with multiple sclerosis. *Med (New York, N.Y.)*. 2024; 5: 550–558.e2. <https://doi.org/10.1016/j.medj.2024.03.002>.
- [143] Tiziana Life Sciences. 2025 (online). Available at: <https://www.tizianalifesciences.com> (Accessed: 25 October 2025).
- [144] Lublin FD, Häring DA, Ganjgahi H, Ocampo A, Hatami F, Čuklina J, *et al.* How patients with multiple sclerosis acquire disability. *Brain: a Journal of Neurology*. 2022; 145: 3147–3161. <https://doi.org/10.1093/brain/awac016>.
- [145] Faissner S, Gold R. Progressive multiple sclerosis: latest therapeutic developments and future directions. *Therapeutic Advances in Neurological Disorders*. 2019; 12: 1756286419878323. <https://doi.org/10.1177/1756286419878323>.
- [146] Lassmann H. Multiple Sclerosis Pathology. *Cold Spring Harbor Perspectives in Medicine*. 2018; 8: a028936. <https://doi.org/10.1101/cshperspect.a028936>.
- [147] Kappos L, Bar-Or A, Cree BAC, Fox RJ, Giovannoni G, Gold R, *et al.* Siponimod versus placebo in secondary progressive multiple sclerosis (EXPAND): a double-blind, randomised, phase 3 study. *Lancet (London, England)*. 2018; 391: 1263–1273. [https://doi.org/10.1016/S0140-6736\(18\)30475-6](https://doi.org/10.1016/S0140-6736(18)30475-6).
- [148] Reich DS, Arnold DL, Vermersch P, Bar-Or A, Fox RJ, Matta A, *et al.* Safety and efficacy of tolebrutinib, an oral brain-penetrant BTK inhibitor, in relapsing multiple sclerosis: a phase 2b, randomised, double-blind, placebo-controlled trial. *The Lancet. Neurology*. 2021; 20: 729–738. [https://doi.org/10.1016/S1474-4422\(21\)00237-4](https://doi.org/10.1016/S1474-4422(21)00237-4).
- [149] Wollebo HS, White MK, Gordon J, Berger JR, Khalili K. Persistence and pathogenesis of the neurotropic polyomavirus JC. *Annals of Neurology*. 2015; 77: 560–570. <https://doi.org/10.1002/ana.24371>.
- [150] Plavina T, Subramanyam M, Bloomgren G, Richman S, Pace A, Lee S, *et al.* Anti-JC virus antibody levels in serum or plasma further define risk of natalizumab-associated progressive multifocal leukoencephalopathy. *Annals of Neurology*. 2014; 76: 802–812. <https://doi.org/10.1002/ana.24286>.
- [151] Ho PR, Koendgen H, Campbell N, Haddock B, Richman S,

- Chang I. Risk of natalizumab-associated progressive multifocal leukoencephalopathy in patients with multiple sclerosis: a retrospective analysis of data from four clinical studies. *The Lancet. Neurology*. 2017; 16: 925–933. [https://doi.org/10.1016/S1474-4422\(17\)30282-X](https://doi.org/10.1016/S1474-4422(17)30282-X).
- [152] Sørensen PS, Bertolotto A, Edan G, Giovannoni G, Gold R, Havrdova E, *et al.* Risk stratification for progressive multifocal leukoencephalopathy in patients treated with natalizumab. *Multiple Sclerosis (Houndmills, Basingstoke, England)*. 2012; 18: 143–152. <https://doi.org/10.1177/1352458511435105>.
- [153] Ryerson LZ, Foley J, Chang I, Kister I, Cutter G, Metzger RR, *et al.* Risk of natalizumab-associated PML in patients with MS is reduced with extended interval dosing. *Neurology*. 2019; 93: e1452–e1462. <https://doi.org/10.1212/WNL.00000000000008243>.
- [154] Benedict RHB, Tomic D, Cree BA, Fox R, Giovannoni G, Bar-Or A, *et al.* Siponimod and Cognition in Secondary Progressive Multiple Sclerosis: EXPAND Secondary Analyses. *Neurology*. 2021; 96: e376–e386. <https://doi.org/10.1212/WNL.00000000000011275>.
- [155] Almeida D, Pinho R, Correia V, Soares J, Bastos MDL, Carvalho F, *et al.* Mitoxantrone is More Toxic than Doxorubicin in SH-SY5Y Human Cells: A ‘Chemobrain’ In Vitro Study. *Pharmaceuticals (Basel, Switzerland)*. 2018; 11: 41. <https://doi.org/10.3390/ph11020041>.
- [156] Healy LM, Stratton JA, Kuhlmann T, Antel J. The role of glial cells in multiple sclerosis disease progression. *Nature Reviews. Neurology*. 2022; 18: 237–248. <https://doi.org/10.1038/s41582-022-00624-x>.
- [157] Montalban X, Hauser SL, Kappos L, Arnold DL, Bar-Or A, Comi G, *et al.* Ocrelizumab versus Placebo in Primary Progressive Multiple Sclerosis. *The New England Journal of Medicine*. 2017; 376: 209–220. <https://doi.org/10.1056/NEJMoa1606468>.
- [158] Fox RJ, Bar-Or A, Traboulsee A, Oreja-Guevara C, Giovannoni G, Vermersch P, *et al.* Efficacy and safety of tolebrutinib versus placebo in non-relapsing secondary progressive multiple sclerosis: Results from the phase 3 HERCULES trial. In 40th Congress of the European Committee for Treatment and Research in Multiple Sclerosis. Denmark. Copenhagen. 2024.
- [159] Oh J, Arnold DL, Cree BA, Ionete C, Kim HJ, Sormani MP, *et al.* Efficacy and safety of tolebrutinib versus teriflunomide in relapsing multiple sclerosis: Results from the phase 3 GEMINI 1 and 2 trials. In 40th Congress of the European Committee for Treatment and Research in Multiple Sclerosis. Denmark. Copenhagen. 2024.
- [160] Montalban X, Vermersch P, Arnold DL, Bar-Or A, Cree BAC, Cross AH, *et al.* Safety and efficacy of evobrutinib in relapsing multiple sclerosis (evolutionRMS1 and evolutionRMS2): two multicentre, randomised, double-blind, active-controlled, phase 3 trials. *The Lancet. Neurology*. 2024; 23: 1119–1132. [https://doi.org/10.1016/S1474-4422\(24\)00328-4](https://doi.org/10.1016/S1474-4422(24)00328-4).
- [161] Boumpas DT, Furie R, Manzi S, Illei GG, Wallace DJ, Balow JE, *et al.* A short course of BG9588 (anti-CD40 ligand antibody) improves serologic activity and decreases hematuria in patients with proliferative lupus glomerulonephritis. *Arthritis and Rheumatism*. 2003; 48: 719–727. <https://doi.org/10.1002/art.10856>.
- [162] Hassan SB, Sørensen JF, Olsen BN, Pedersen AE. Anti-CD40-mediated cancer immunotherapy: an update of recent and ongoing clinical trials. *Immunopharmacology and Immunotoxicology*. 2014; 36: 96–104. <https://doi.org/10.3109/08923973.2014.890626>.
- [163] Gupta S, Seshadri M, Lincoln R, Okiniskhi N, Shum N, Shenoy S, *et al.* An Investigator Initiated Study of KYV-101, a CD19 CAR T Cell Therapy, in Participants with Treatment Refractory Progressive Multiple Sclerosis (S3.002). *Neurology*. 2025; 104: 4110. <https://doi.org/10.1212/WNL.000000000000211457>.
- [164] Dunn J, Galetta K, Tomczak A, Timmons G, Ton-Nu C, Sumera J, *et al.* A Phase 1, Open-label, Single Center Study of KYV-101, an Autologous Fully human Anti-CD19 Chimeric Antigen Receptor T Cell (CD19 CAR T) Therapy, in Subjects with Non-relapsing and Forms of Multiple Sclerosis (S3.001). *Neurology*. 2025; 104: 5371. <https://doi.org/10.1212/WNL.000000000000212317>.
- [165] Richter J, Fischbach F, Pfeffer LK, Fehse B, Berger SC, Reinhardt S, *et al.* CD19-Directed CAR T Cell Therapy in 4 Patients with Refractory Multiple Sclerosis. *Blood*. 2024; 144: 2073. <https://doi.org/10.1182/blood-2024-205103>.
- [166] Barrie W, Yang Y, Irving-Pease EK, Atfield KE, Scorrano G, Jensen LT, *et al.* Elevated genetic risk for multiple sclerosis emerged in steppe pastoralist populations. *Nature*. 2024; 625: 321–328. <https://doi.org/10.1038/s41586-023-06618-z>.
- [167] Behrens M, Comabella M, Lünemann JD. EBV-specific T-cell immunity: relevance for multiple sclerosis. *Frontiers in Immunology*. 2024; 15: 1509927. <https://doi.org/10.3389/fimmu.2024.1509927>.
- [168] Thomas OG, Haigh TA, Croom-Carter D, Leese A, Van Wijck Y, Douglas MR, *et al.* Heightened Epstein-Barr virus immunity and potential cross-reactivities in multiple sclerosis. *PLoS Pathogens*. 2024; 20: e1012177. <https://doi.org/10.1371/journal.ppat.1012177>.
- [169] Portaccio E, Bellinva A, Fonderico M, Pastò L, Razzolini L, Totaro R, *et al.* Progression is independent of relapse activity in early multiple sclerosis: a real-life cohort study. *Brain: a Journal of Neurology*. 2022; 145: 2796–2805. <https://doi.org/10.1093/brain/awac111>.
- [170] Kaufmann M, Evans H, Schaupp AL, Engler JB, Kaur G, Willing A, *et al.* Identifying CNS-colonizing T cells as potential therapeutic targets to prevent progression of multiple sclerosis. *Med (New York, N.Y.)*. 2021; 2: 296–312.e8. <https://doi.org/10.1016/j.medj.2021.01.006>.
- [171] Abdelhak A, Benkert P, Schaedelin S, Boscardin WJ, Cordano C, Oechtering J, *et al.* Neurofilament Light Chain Elevation and Disability Progression in Multiple Sclerosis. *JAMA Neurology*. 2023; 80: 1317–1325. <https://doi.org/10.1001/jamaneurol.2023.3997>.
- [172] Macnair W, Calini D, Agirre E, Bryois J, Jäkel S, Smith RS, *et al.* snRNA-seq stratifies multiple sclerosis patients into distinct white matter glial responses. *Neuron*. 2025; 113: 396–410.e9. <https://doi.org/10.1016/j.neuron.2024.11.016>.
- [173] Oh J, Giacomini PS, Yong VW, Costello F, Blanchette F, Freedman MS. From progression to progress: The future of multiple sclerosis. *Journal of Central Nervous System Disease*. 2024; 16: 11795735241249693. <https://doi.org/10.1177/11795735241249693>.