

Review

# Applications and Possible Mechanisms of ULK4 in Brain: Implications for Neuropsychiatric Disorders

Wen Luo<sup>1,\*</sup>, Jisheng Wang<sup>1</sup>, Jing Chen<sup>1</sup><sup>1</sup>Department of Clinical Pharmacy, The Third Hospital of Mianyang, Sichuan Mental Health Center, 621000 Mianyang, Sichuan, China\*Correspondence: [qq.xiao.wen@163.com](mailto:qq.xiao.wen@163.com) (Wen Luo)

Academic Editor: Maarten Van den Buuse

Submitted: 24 November 2024 Revised: 15 February 2025 Accepted: 21 February 2025 Published: 21 May 2025

## Abstract

**Background:** Neuropsychiatric disorders make up 14% of the global disease burden and are the leading cause of disability from non-communicable diseases worldwide. The primary treatment for these disorders is drug therapy. Nonetheless, these therapies do not work completely for most patients, and even with attempts to create novel drugs, no medication has been confirmed as safe and effective for treating neuropsychiatric disorders. Recent studies have emphasized the role of gene therapy in neuropsychiatric disorders. Unc-51-like kinase (ULK) has connections to central nervous system functions and disorders, but the role of ULK4 is less well understood than other members of that family. **Methods:** The PubMed database was searched for articles regarding ULK4 in neuropsychiatric disorders and neurodevelopment with no restriction on publication date. **Results:** ULK4 is believed to function as a pseudokinase, potentially acting as a scaffold to connect kinases or other enzymes with their substrates or to manage the subcellular location of interacting proteins in different biological processes, abnormal low expression of which may increase the risk of neuropsychiatric disorders. **Conclusions:** This review updates the latest evidence on the roles of ULK4 in brain development and neuronal function, and highlights some controversies and uncertainties in the current research on ULK4. This review offers perspectives on the continuous development and design of drugs targeting ULK4, supporting possibilities for their future clinical application.

**Keywords:** ULK4; neuropsychiatric disorders; neurodevelopment; neuromodulation; pseudokinase

## 1. Introduction

Neurological and psychiatric disorders pose a significant public health challenge globally, affecting one in eight individuals with mental disorders or one in four individuals with neurological disorders at some stage in their lives [1,2]. Neuropsychiatric conditions are complex illnesses with vague neurobiological roots, and they are all too common yet profoundly destructive [3,4]. Despite years of research, progress in developing central nervous system (CNS) disease treatments has been slow, and small-molecule drugs continue to be the mainstay in clinical practice. An additional limitation of these therapies is that they do not cure the disease but only alleviate symptoms to slow its progression.

There is overwhelming evidence that neurological and psychiatric disorders have a developmental origin with a strong genetic underpinning [5]. Brain development disturbances may cause cerebral deformities and functional impairments, leading to developmental neuropathology [6]. The neuronal connectivity and cellular architecture of the brain are believed to be disturbed by these developmental insults. The rapid advancement of genome-engineering technologies has ushered in a new era for enhancing brain disease treatments through gene therapy [7–9]. Genome-wide association studies (GWAS), databases such as the Cancer Genome Atlas (TCGA) database and the Gene Expression Omnibus (GEO) database, and ribonucleic acid

(RNA) sequencing all connect Unc-51-like kinase (*ULK*) 4 to the CNS in processes like developmental/language delay and intellectual disability [10], mental disorders [11], ischemic stroke [12]. Outside the CNS, they have been found to be involved in hypertension [13,14], mucociliary clearance impaired [15], sporadic thoracic aortic dissection [16], formation of bumps on the legs of pigs [17], cancer [18,19], and age-related cataract [20].

Gene therapy provides unique advantages over traditional small-molecule therapies, such as the ability to control genes that encode proteins and achieve high target specificity [21,22]. Significantly, there has been a consistent increase in research on gene therapy for CNS diseases in recent decades, as indicated by the swift growth in clinical trials and publications. Gene therapies are still in their infancy, and currently, there is no approved gene therapy for CNS diseases available in clinical settings. In this paper, we build on existing literature by examining the association of *ULK4* with brain development and neuropsychiatric disorders, update the latest evidence with regard to *ULK4*, point out some controversies and uncertainty in the current research, and highlight its promising clinical application as a potential biomarker for the treatment of brain diseases [23].



## 2. Overview of *ULK4*

### 2.1 Data Source and Search Strategy

We searched PubMed using the search terms “Unc-51 like kinase 4”, “unc-51-like kinase 4”, and “ULK4” with no time restrictions. Inclusion criteria included articles on ULK4 in neuropsychiatric disorders, neurodevelopment, and structure and characteristics, as well as articles on its involvement in the peripheral system (published 2020 and beyond), the rest were excluded. Table 1 (Ref. [10–12,24–44]) summarizes the research on ULK4 in neuropsychiatric disorders and neurodevelopment.

### 2.2 Pseudokinases

The ULK family in mammals consists of four kinases, namely ULK1, ULK2, ULK3, and serine/threonine kinase 36 (STK36), along with one pseudokinase, ULK4. In ULKs, the kinase domains are situated at the N terminals, and the C-terminals contain protein interaction motifs crucial for substrate recruitment. Although ULK1 and ULK2 share only 29% sequence identity with ULK4 pseudokinase domain (PD), their structures superimposed well [45,46]. All enzymes are considered crucial for development, especially for neurological development. Pseudokinases lack catalytically active kinase domains and primarily signal through their function as allosteric regulators or scaffolds [47]. Some pseudokinases serve as adenosine triphosphate (ATP) sensors, which respond to ATP binding without transferring phosphate [48–50].

### 2.3 Structure and Characteristics

ULK4 is a pseudokinase because it lacks the catalytic residues required for its kinase function. The connection of ULK4 to multiple human disorders underscores its developmental role as a kinase scaffold protein that lacks catalytic activity. The *ULK4* gene’s single-nucleotide polymorphisms affect not only the CNS but also the peripheral immune system. Nevertheless, the mechanism by which *ULK4* affects brain applications and mechanisms remains a mystery.

ULK4 has been discovered to have no apparent phosphotransfer activity, but it can bind ATP more effectively than other known pseudokinases [24–26]. Furthermore, a unique mechanism is discovered where ULK4 can bind ATP without the need for magnesium ( $Mg^{2+}$ ), but it binds less potently when  $Mg^{2+}$  is present [24,26].

### 2.4 Interaction Partners of *ULK4*

ULK4 interacts with neighboring centrosomal proteins, such as centriolar coiled-coil protein 110 (CCP110), centrosome and spindle pole-associated protein 1 (CSPP1), and kinesin family member 1B (KIF1B) [26]. Microtubule-associated proteins, such as rho-associated coiled-coil containing protein kinase (ROCK) 1, ROCK2, and protein tyrosine phosphatase non-receptor type 14 (PTPN14), co-migrate with ULK4 [25,26]. Additionally, the ULK fam-

ily member STK36, like ULK4 [27], which plays a crucial role in regulating the hedgehog pathway [28,51,52], was found to be a high-confidence proximity interactor [25,26]. However, how the pseudosubstrate binds to ULK4 pseudokinase, and whether ULK4 acts as an allosteric regulator of canonical kinases, are poorly understood.

## 3. Role of *ULK4* in Normal CNS Function

### 3.1 Role in Neurodevelopment

Neurogenesis during embryonic and adult stages is essential for the formation and functioning of the brain. Disturbed neural proliferation is widely associated with brain disorders, such as schizophrenia and autism which are linked to *ULK4* [10,29]. There is genetic proof that *ULK4* is deleted in 1.2 of 1000 patients who have clinical signs like developmental or language delays and intellectual disabilities [30].

The majority of mammalian cortical neurogenesis typically occurs in the embryonic stage, leaving a limited number of neural stem cells (NSCs) for adult neurogenesis [31]. These cells either divide slowly or remain inactive until they are needed for postnatal oligodendrogenesis [53,54] and adult neurogenesis [55–57], and generate different layers of cortical neurons in a cell-cycle-dependent manner [58–60]. Thus, cell-cycle regulation is essential during cortical lamination. *ULK4* is expressed alongside markers for cell proliferation and markers for neural stem/progenitor cell, showing the greatest abundance during the gap 2/mitosis (G2/M) phases [30,32]. Consistent with *ULK4* expression being specific to the cell cycle, Liu and colleagues noted that disruption of *ULK4* resulted in a thinner cortex [31]. A notable co-localization of anti-*ULK4* and Ki67 was detected in the neonatal subventricular zone (SVZ) region, but *ULK4* expression is greatly diminished in nearby mature neuron areas [30]; this also reinforces the idea that *ULK4* is needed not just for embryonic neurogenesis but also for the preservation of the NSC pool [31]. Whole-genome RNA sequencing identifies deregulation of gene expression in 19,652 in *ULK4* mutants [30]. Analyses of pathways revealed a strong link between *ULK4* targets and the proliferation of neural precursor cells as well as the cell cycle [31]. The absence of *ULK4* in human neuroblastoma cells disturbed microtubule composition, leading to reduced neuritogenesis and cell motility [31]. Disassembled  $\alpha$ -tubulin reduces acetylation, leading to decreased dendrite length, reduced branching, and impaired radial migration of cortical neurons [61,62]. In the embryonic brain, silencing *ULK4* led to a decrease in acetylated  $\alpha$ -tubulin and affected both radial migration and proliferation of NSCs [33].

Although the specific molecular pathways have not been fully determined, *ULK4* hypomorph mice exhibited the following phenotypes: predisposed mental disorders and neurodevelopmental diseases [23,63,64]; dysregulated inhibitory pathways [49]; and decreased white-matter inte-

**Table 1. Summarized literature on Unc-51-like kinase (ULK) 4 in neuropsychiatric disorders and neurodevelopment.**

Disease	Model	ULK4 expression	Association of ULK4 with neurological condition	Ref.
	Patients	Decreased	ULK4 CNVs abnormalities are associated with neuropsychiatric features and obesity	[10]
Mental disorders	Patients	No change	Genetic variation in ULK4 may increase the vulnerability to mental disorders by modulating the extended reward system function	[11]
Ischemic stroke	Patients	No change	ULK4 as the shared risk SNPs between obesity and ischemic stroke	[12]
			ULK4 structure and characteristics	[24]
			ULK4 structure and characteristics	[25]
			ULK4 structure and characteristics	[26]
	Lrp2 mutant mice on FVB/N background	Increased	ULK4 as microtubule-associated proteins, is positive regulator of SHH signaling, and as components of primary cilia in the neuroepithelium	[27]
	NIH3T3 cells/HEK293T cells	No change	ULK4 acts in conjunction with STK36 to promote hedgehog pathway activation	[28]
Schizophrenia	Patients/ULK4 knockdown SH-SY5Y cells/ULK4 <sup>-/-</sup> mice	Decreased	ULK4 CNVs abnormalities are associated with schizophrenia	[29]
			ULK4 deletion reduce neurite extension and impaired neurite branching, cell motility	
			ULK4 deletion reduce p-ERK, p-JNK, increase p-PKC	
Neurodevelopment	Patients/ULK4 <sup>tm1a/tm1a</sup> mice	Decreased	ULK4 deletion lead to defects of corpus callosum	[30]
			ULK4 deletion lead to severe language delay and learning difficulties	
			ULK4 deletion reduce cerebral cortex	
Neurodevelopment	Disrupted ULK4 newborn mice	Decreased	ULK4 as regulator of cell cycle and NSC proliferation	
			ULK4 deletion reduce NSC pool	[31]
	Xenopus laevis	No change	ULK4 deletion disturb cell cycle pathways	
Neurodevelopment	ULK4 Knockdown mouse/Primary cultured mouse cortical neurons ULK4 knockdown	Decreased	ULK4 co-expressed with the neural stem/progenitor cell marker	[32]
			ULK4 deletion decrease corticogenesis	[33]
Neurodevelopment	cultured mouse cortical neurons ULK4 knockdown	Decreased	ULK4 deletion decrease cell proliferation and deficits in radial migration and neurite ramification	
			ULK4 deletion decrease acetylation of $\alpha$ -tubulin	
Myelination	ULK4 <sup>tm1a/tm1a</sup> mice	Decreased	ULK4 deletion reduce myelin, which may help to explain severe learning difficulty and language delay	[34]
Ciliopathy	ULK4 <sup>tm/Lex</sup> mouse	Decreased	ULK4 deletion downregulate oligodendrocyte-related genes	
			ULK4 deletion lead to congenital hydrocephalus	[35]
Ciliopathy	ULK4 <sup>tm1a/tm1a</sup> mice	Decreased	ULK4 deletion reduced/disorganize cilia with abnormal axonemes	[36]
Ciliopathy	ULK4 <sup>tm1a/tm1a</sup> mice	Decreased	ULK4 deletion lead to hydrocephalus and functionally impairs the CSF circulation	
			ULK4 deletion increase FOXJ1 ciliogenesis gene	

Table 1. Continued.

Disease	Model	ULK4 expression	Association of ULK4 with neurological condition	Ref.
Ciliopathy	Primary cultured mouse ependymal cells ULK4 deletion	Decreased	ULK4 deletion lead to structural defects of the flagellum cytoskeleton	[37]
		No change	ULK4 interacts with STK36	
Ciliopathy	Yeast/HEK293T cells/ ULK4 <sup>tm1a/tm1a</sup> mice	No change	ULK4 interacts with STK36	[38]
		Decreased	ULK4 deletion lead to dysfunctional CSF flow and defects of motile cilia	
Ciliopathy	HEK293T cells/Depleted ULK4 by RNA interference in NIH3T3 cells	No change	ULK4 deletion increase FOXJ1 ciliogenesis gene	[39]
		Decreased	ULK4 forms a complex with STK36 and GLI2 ULK4 deletion decrease the association between STK36 and GLI2 ULK4 deletion decrease STK36 ciliary localization, GLI2 phosphorylation, and hedgehog pathway activation	
Schizophrenia	Patients	No change	ULK4 SNPs variants may be schizophrenia etiology	[40]
Schizophrenia	ULK4 <sup>tm1a/tm1a</sup> mice/ Nestin-Cre: ULK4 <sup>f/f</sup> mice	Decreased	ULK4 deletion lead to growth retardation	[41]
			ULK4 deletion reduce cerebral cortex development Cortical neuron migration is not affected in ULK4 deletion	
Schizophrenia	Emx1-Cre: ULK4 <sup>lox/lox</sup> mice/ SH-SY5Y cells transfected with pLex-Flag-ULK4 plasmid	Decreased	ULK4 deletion lead to deficits in the spatial and working memories and sensorimotor gating	[42]
		Increased	ULK4 deletion reduce levels of p-AKT and p-GSK3 $\alpha/\beta$ ULK4 overexpression lead to a significant enrichment of PP2A/PP1 $\alpha$ ULK4 deletion lead to anxiety-like behavior	[43]
Anxiety disorder	ULK4 <sup>+tm1a</sup> mice	Decreased	ULK4 deletion dysregulate hypo-anxious and hyper-anxious genes ULK4 deletion reduce GAD67 interneurons in the hippocampus and basolateral amygdala ULK4 deletion reduce of GABAergic neuronal subtypes	
Cerebral ischemia-reperfusion injury	MCAO/R rats/Primary cortical neurons subjected to OGD/R	Decreased	ULK4 overexpression reduce apoptosis, neurological deficit scores, cerebral infarct volume, and histopathological damage ULK4 overexpression increase PI3K p110 $\alpha$ , p-AKT, p-GSK3 $\beta$	[44]

CNV, copy number variation; SNP, single nucleotide polymorphism; SHH, sonic hedgehog; Lrp2, low-density lipoprotein receptor-related protein 2; STK36, serine/threonine kinase 36; ERK, extracellular signal-regulated kinase; PKC, protein kinase C; JNK, c-Jun N-terminal kinase; NSC, neural stem cell; CSF, cerebrospinal fluid; FOXJ1, forkhead box j1; GLI2, GLI family zinc finger 2; AKT, protein kinase B; GSK3, glycogen synthase kinase 3; PP2A, protein phosphatases 2A; PP1 $\alpha$ , protein phosphatases 1 $\alpha$ ; GABA,  $\gamma$ -aminobutyric acid; MCAO/R, middle cerebral artery occlusion/reperfusion; OGD/R, oxygen-glucose deprivation/reperfusion; PI3K, phosphatidylinositol 3-kinase; GAD, glutamic acid decarboxylase; FVB/N, friend virus B/NIH.

grity [34]. Moreover, *ULK4* modulates the extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinase (JNK), and protein kinase C (PKC) signaling pathways, which are involved in managing the neuronal cytoskeleton often linked to schizophrenia neuropathology [29]. *ULK4* is a key regulator of cell cycle and NSC proliferation involved in modulating wingless-int (Wnt) signaling pathways [30], and Wnt signaling is well-recognized for its role in neural development [65], and mental disorders [66].

### 3.2 Role in Ciliopathy

Hydrocephalus is a serious neurological condition marked by an accumulation of cerebrospinal fluid in the brain and enlargement of the brain's ventricles [67,68]. The complexity of hydrocephalus pathology is evident, with growing evidence indicating that a primary cause is the malfunction of motile cilia [69–73]. In a mouse knockout and phenotyping study, *ULK4* was found among a group of genes that, when disrupted, led to hydrocephalus [35]. The *ULK4* hypomorph mutant in mice leads to impaired ependymal cell differentiation and disrupted cilia function [36]. *ULK4*-deficient mice (*ULK4*<sup>tm1a/tm1a</sup>) exhibit progressive hydrocephalus and stunted growth, with the mutants not surviving beyond 3 weeks [35,36]. RNA-sequencing analysis revealed that a number of genes associated with ciliogenesis were dysregulated in *ULK4*<sup>tm1a/tm1a</sup>, including the key cilia gene forkhead box j1 (*FOXJ1*) [36], underscoring the crucial role of *ULK4* in motile cilia. In *ULK4*<sup>tm1a/tm1a</sup> mice, the dynein axonemal light chain 1 (*DNALI1*) and tubulin tyrosine ligase like 6 (*TTL6*) are reduced by 10% and 49%, respectively, affecting outer and inner dynein arms [74,75], though the molecular mechanisms remain mostly unclear.

*ULK4* may homodimerize with the help of *STK36* in mammals [37–39]. Continued investigation found that mutations in *STK36* and *ULK4* in mice resulted in hydrocephalus and other phenotypes related to ciliary defects and, as such, effectively phenocopied each other, producing growth retardation, early mortality, ventriculomegaly, underdeveloped or deficient ependymal cells, reduced or dysfunctional or disorganized cilia, disordered axoneme structure, and misaligned base body (BB) structure [37,38]. The lack of *STK36* also affects the expression of *FOXJ1* and various other genes related to ciliogenesis [38]. Earlier studies indicated that mammalian *STK36* is found in the motile cilium [76], and recent findings have shown that *ULK4* is also present in the motile cilia of ependymal cells [37]. From these findings, we suggest that *ULK4* and *STK36* directly interact to jointly regulate ciliogenesis and ciliary function.

### 3.3 Role in Myelination

Myelin is crucial for high-speed and accurate electrical-impulse conduction in axons, and also manages the synchronization of impulse transmission between spatially separated cortical regions deemed critical for

perception and cognitive function [77]. Hypomyelination frequently occurs in neurodegenerative and neurodevelopmental disorders. Children experiencing developmental delays exhibited a notable decrease in myelinated tissue, with 19.8% brain volume compared to 21.4% in controls ( $p < 0.01$ ) [78]. *ULK4* mutants have a 50% reduction in myelin, indicating hypomyelination, which is considerably higher than the 1.6% reduction in children with developmental delays [34,78]. This may help explain the severe language delay and learning difficulty in some *ULK4* patients [30]. The expression of genes 2',3'-cyclic nucleotide 3' phosphodiesterase (*CNP*), erb-B2 receptor tyrosine kinase 3 (*ERBB3*), gelsolin (*GSN*), mal, T cell differentiation protein (*MAL*), brain enriched myelin associated protein 1 (*BCAS1*) in *ULK4* hypomorph mice was assessed, indicating that all genes with reduced expression were linked to myelin and schizophrenia [34,79,80].

Oligodendrocytes are responsible for producing myelin, making contact with multiple neighboring axons, and enveloping short axonal segments [81,82]. Deficiency in *ULK4* results in a notable downregulation of essential oligodendrogenic transcription factors, which are critical for the maintenance of oligodendrocyte progenitor cells and myelinating oligodendrocyte differentiation and maturation [34].

## 4. Role of *ULK4* in Neuropsychiatric Disorders

Neuropsychiatric disorders, which have complex pathogenesis, are leading causes of disability [83,84]. Despite the efforts to elucidate their mechanisms or etiologies, they remain elusive and not yet clarified. Identification of specific gene-expression signatures has contributed to understanding the relationship between genes and brain health, and genetic risk for neuropsychiatric disorders [85]. Here, we discuss the potential effects of *ULK4* on neuropsychiatric disorders, including anxiety disorders, schizophrenia, and cerebral ischemia-reperfusion injury (CIRI).

### 4.1 Schizophrenia

Damage to synaptic connections is believed to affect neurodevelopment and brain function negatively, whether occurring alone or in combination, thereby increasing the predisposition to schizophrenia in adults [86]. *ULK4* copy-number variations are found in several mental illnesses, including schizophrenia [29]. A test of association using two independent schizophrenia case-control cohorts of north Indian ethnicity reveals that *ULK4* was found to alter the neurodevelopmental expression [40]. Further evidence has been found, in humans, that genetic variation in *ULK4* increases vulnerability to mental disorders by modulating the extended reward system function [11]. The results offer compelling support for *ULK4* as a novel rare risk factor for schizophrenia [29,33,41]. Genetic-linkage analyses have

identified a strong co-segregation of protein kinase B (*AKT*) gene (*AKT1*) haplotypes with schizophrenia [87]. Many downstream targets of AKT, including glycogen synthase kinase 3 (GSK3), may contribute to schizophrenia [42,88].

According to recent findings, *ULK4*<sup>flox/flox</sup> conditional knockout (CKO) mice demonstrate cognitive dysfunction and a decrease in the phosphorylation of AKT and GSK3 [42]. As a pseudokinase, ULK4 generally executes its biological roles by recruiting other active kinases for allosteric regulation or by acting as scaffolds for substrate binding within signaling pathways [89]. The analysis of the proteome indicates that protein phosphatases (PP) 2A and 1 $\alpha$ , located in the signaling pathways upstream of AKT, can physically associate with ULK4 without changing their phosphorylation status [42]. Hence, we propose that the impaired AKT-GSK3 signaling cascade increases the likelihood of *ULK4* CKO mice developing the core traits of schizophrenia.

Despite showing impairments in spatial and working memory, and sensorimotor gating, CKO mice do not exhibit anxiety-like behavior changes, which contradicts the report that *ULK4*<sup>+/<sup>tm1a</sup></sup> mice display an anxiety-like behavioral phenotype [43]. The following factors might account for the varying results: (a) *ULK4* is deleted only in the excitatory neurons of the cerebral cortex and hippocampus in CKO mice, but was only reduced by half in the whole brains of *ULK4*<sup>+/<sup>tm1a</sup></sup> mice; (b) impaired  $\gamma$ -aminobutyric acid (GABA)ergic signaling might be responsible for the anxiety-like behaviors in *ULK4*<sup>+/<sup>tm1a</sup></sup> mice, whereas it might be unaffected or less affected in CKO mice.

Another question on the matter of debate is the radial migration of pyramidal neurons. The recent study on schizophrenia involving *ULK4* found that the radial migration of pyramidal neurons is not apparent in *ULK4* deletion [41], which was contrary to the findings of the previous study [33]. We hypothesized that the inconsistency might be due to the hydrocephalus. Hydrocephalus may either have “add-on” effects or may “mask” the genuine migration delay through the resultant thinning of the individual sublayers.

#### 4.2 Anxiety Disorder

Various studies have highlighted an imbalance between excitation and inhibition as a frequent mechanism in numerous neurodevelopmental disorders [90–92]. In line with the behavioral alterations, genes associated with hypo-anxiety, ATPase Na<sup>+</sup>/K<sup>+</sup> Transporting Subunit Alpha 2 (*ATP1A2*), pleiotrophin (*PTN*), and midkine (*MDK*), were downregulated, whereas genes linked to hyper-anxiety, glutamate ionotropic receptor AMPA type subunit 1 (*GRI1A1*), neuropeptide Y receptor Y2 (*NPY2R*), protein tyrosine phosphatase receptor type A (*PTPRA*), were significantly upregulated in *ULK4*<sup>+/<sup>tm1a</sup></sup> mice [43,93]. Deleting *ULK4* causes changes of GABAergic neurons, which may be a major factor reinforcing anxiety. In this case, not only

were glutamic acid decarboxylase (GAD) 67 neurons of the amygdala and hippocampus reduced significantly, but there was also a subtype-specific reduction of GABA interneurons [43]. In addition, *ULK4* deletion resulted in disruption of postsynaptic GABAergic signaling transmission, notably with significant upregulation of gamma-aminobutyric acid type A receptor subunit alpha (*GABRA1*), *GABRA3*, *GABRA4*, *GABRA5*, and gamma-aminobutyric acid type A receptor subunit beta (*GABRB3*) [43]. However, it remains uncertain if *ULK4* deficiency has any effect on the formation or function of GABAergic neurons.

#### 4.3 CIRI

Evidence shows that schizophrenia is positively correlated with increased mortality in ischemic stroke patients [94,95]. Antipsychotic medication use has been shown to reduce mortality rates in ischemic heart disease and stroke [96]. Mice experiencing experimentally induced CIRI demonstrated impairments in learning and the development of new motor skills [97], and rats also displayed anxiety-like behaviors after similar trauma [98]. The question of whether CIRI influences the expression of the schizophrenia-susceptibility gene *ULK4*, thereby contributing to the injuries and diseases mentioned, remains unanswered. Whether *ULK4* acts protectively or detrimentally in CIRI is uncertain. Using an oxygen-glucose deprivation/reperfusion (OGD/R) model *in vitro* and a middle cerebral artery occlusion/reperfusion (MCAO/R) rat model *in vivo*, we show a decline in *ULK4* expression in the cortex of MCAO/R rats, and primary cortical neurons in the OGD/R [44]. Overexpression of *ULK4* inhibited apoptosis and improved a series of indicators of cerebral ischemia-reperfusion injury, including neurological deficit scores, infarct volume, and histopathological damage [44]. Through the application of network medicine analysis, our investigation has established that ischemic stroke is prominently linked to the phosphatidylinositol 3-kinase (PI3K)-AKT-GSK3 $\beta$  pathway, with the *AKT1* gene and the *GSK3 $\beta$*  gene playing crucial roles (still ongoing). Our team found that *ULK4* plays a role in regulating the PI3K-AKT-GSK3 $\beta$  pathway, which provides neuroprotection against CIRI [44]. Alternatively, increased levels of PI3K p110 $\alpha$  proteins were induced by *ULK4* in MCAO/R rats [44] and OGD/R-treated pheochromocytoma cell line 12 (PC12) cells, along with an observed increase in the colocalization of PI3K p110 $\alpha$  and PI3K p85 $\alpha$  (still ongoing). The catalytic subunit p110 $\alpha$  of PI3K in mammals is inactive when alone and needs to be expressed with the regulatory subunit p85 $\alpha$  to become active [99]. Upon exposure to external injury signals, the protein might become non-functional and unstable, necessitating p85 to activate PI3K by attaching to the N-terminus of p110 via its Src homology 2 (SH2) domain, which then triggers downstream signaling. Therefore, we theorized that *ULK4* supports the interaction between PI3K p110 $\alpha$  and PI3K p85 $\alpha$ , lead-

ing to PI3K activation. Whether ULK4 directly interacts with PI3K p110 $\alpha$ , activates it via PI3K p85 $\alpha$ , or influences it indirectly through other elements, remains to be confirmed. Consistent with our current understanding of *ULK4*, the latest research confirmed the close relationship between *ULK4* and ischemic stroke through the summary-data-based Mendelian randomization method [12].

Although studies suggest that *ULK4* is associated with neurodegenerative disorders, direct evidence for this role is currently lacking, such as animal experiments or clinical evidence. *ULK4* (also known as family with sequence similarity 7A, *FAM7A*) gene fuses with cholinergic receptor nicotinic Alpha 7 subunit (*CHRNA7*) gene to form the *CHRNA7* (exons 5–10) and *FAM7A* (exons A–E) fusion (*CHRFAM7A*) gene [100]. Gene *CHRFAM7A* has been associated with Alzheimer's disease risk in GWAS [101], which confirms the indirect relationship between *ULK4* and Alzheimer's disease. However, the direct evidence for the involvement of *ULK4* in Multiple sclerosis, Alzheimer's disease, Parkinson's disease, and aging has not been retrieved. This may be an interesting direction for research, we will further connect *ULK4* to neurodegenerative diseases and aging in future studies.

## 5. Perspective and Conclusion

Deletion of *ULK4* results in a thinner cortex, and a negative influence on neurite branching and neuronal motility. Brain-specific deletion of *ULK4* leads to the malfunction of motile cilia, thereby resulting in congenital hydrocephalus. Severe language delay and learning difficulty are observed in the majority of *ULK4*-mutant children. *ULK4*, on the other hand, is closely associated with neuropsychiatric disorders. Susceptibility to schizophrenia is increased by *ULK4* deletion. Deleting *ULK4* causes changes in GABAergic neurons, which may be a major factor in reinforcing anxiety. Conversely, overexpression of *ULK4* inhibits apoptosis, subsequently alleviating CIRI. The role of *ULK4* in neurodevelopment and neuropsychiatric disorders is well established in overall association (Graphic abstract). Nevertheless, partial behavioral phenotypes and specific mechanisms are still controversial, such as anxiety-like behavior and radial migration of pyramidal neurons. This review summarizes the most recent research updates in the neuropsychiatric disorders and neurodevelopment concerning *ULK4*, and covers a larger coverage, such as CIRI, and interaction partners of *ULK4*, which further demonstrates the positive effects of *ULK4* on neuropsychiatric disorders. In addition to this, this review highlights current inconsistencies in *ULK4* research while explaining possible reasons for these discrepancies.

*ULK4* is a gene that has not been extensively researched in association with neuropsychiatric disorders. Present findings somewhat elucidate the role and mechanism of *ULK4* in brain function, holding promise for the development of new treatments for neuropsychiatric diseases.

Given the nature of *ULK4*, further research should focus on identifying its role in conditions, including hemorrhagic stroke, depression, and comorbid neurological conditions, such as Alzheimer's disease, epilepsy, Multiple sclerosis, and Parkinson's disease. Although the results are promising, the therapeutic effectiveness of *ULK4* manipulation for the aforementioned neuropsychiatric disorders relies on studies conducted on animals and cells. A future comprehensive study of the *ULK4* mechanisms and functions in neuropsychiatric disorders will aid in converting pharmacological studies into clinical drug development, offering new clinical treatment alternatives.

## Abbreviations

ULK, unc-51-like kinase; CNS, central nervous system; TCGA, the cancer genome atlas; GEO, gene expression omnibus; STK36, serine/threonine kinase 36; ATP, adenosine triphosphate; CCP110, centriolar coiled-coil protein 110; CSPP1, centrosome and spindle pole-associated protein 1; KIF1B, kinesin family member 1B; ROCK, rho-associated coiled-coil containing protein kinase; PTPN14, protein tyrosine phosphatase non-receptor type 14; NSC, neural stem cell; ERK, extracellular signal-regulated kinase; JNK, c-Jun N-terminal kinase; PKC, protein kinase C; FOXJ1, forkhead box j1; RNA, ribonucleic acid; DNAL1, dynein axonemal light chain 1; TLL6, tubulin tyrosine ligase like 6; CNP, 2',3'-cyclic nucleotide 3' phosphodiesterase; ERBB3, erb-B2 receptor tyrosine kinase 3; GSN, gelsolin; MAL, mal, T cell differentiation protein; BCAS1, brain enriched myelin associated protein 1; AKT, protein kinase B; GSK3, glycogen synthase kinase 3; CKO, conditional knockout; GABA,  $\gamma$ -aminobutyric acid; ATP1A2, ATPase Na<sup>+</sup>/K<sup>+</sup> transporting Subunit Alpha 2; PTN, pleiotrophin; MDK, midkine; GRIA1, glutamate ionotropic receptor AMPA type subunit 1; NPY2R, neuropeptide Y receptor Y2; PTPRA, protein tyrosine phosphatase receptor type A; GABRA, gamma-aminobutyric acid type A receptor subunit alpha; GABRB, gamma-aminobutyric acid type A receptor subunit beta; CIRI, cerebral ischemia-reperfusion injury; OGD/R, oxygen-glucose deprivation/reperfusion; MCAO/R, middle cerebral artery occlusion/reperfusion; PI3K, phosphatidylinositol 3-kinase; PC12, phaeochromocytoma cell line 12; SH2, Src homology 2; FAM7A, family with sequence similarity 7A; CHRNA7, cholinergic receptor nicotinic Alpha 7 subunit; CHRFAM7A, CHRNA7 (exons 5-10) and FAM7A (exons A-E) fusion; GWAS, genome-wide association studies; SVZ, subventricular zone; BB, base body; Mg, magnesium; G2/M, gap 2/mitosis; Wnt, wingless-int; PP2A, protein phosphatases 2A; PP1 $\alpha$ , protein phosphatases 1 $\alpha$ ; GAD, glutamic acid decarboxylase.

## Availability of Data and Materials

The data used in this study are included in the included articles and Table 1.

## Author Contributions

WL organized, designed, and wrote the manuscript. JW and JC performed the literature searches and revised the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

## Ethics Approval and Consent to Participate

Not applicable.

## Acknowledgment

Not applicable.

## Funding

Supported by Health Commission of Sichuan Province Medical Science and Technology Program (24WSXT027).

## Conflict of Interest

The authors declare no conflict of interest.

## References

- [1] World Health Organization. Mental Disorders. 2022. Available at: <https://www.who.int/news-room/fact-sheets/detail/mental-disorders> (Accessed: 22 December 2022).
- [2] GBD 2016 Neurology Collaborators. Global, regional, and national burden of neurological disorders, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *The Lancet. Neurology*. 2019; 18: 459–480. [https://doi.org/10.1016/S1474-4422\(18\)30499-X](https://doi.org/10.1016/S1474-4422(18)30499-X).
- [3] Bray NJ, O'Donovan MC. The genetics of neuropsychiatric disorders. *Brain and Neuroscience Advances*. 2019; 2: 2398212818799271. <https://doi.org/10.1177/2398212818799271>.
- [4] Rihal V, Khan H, Kaur A, Singh TG, Abdel-Daim MM. Therapeutic and mechanistic intervention of vitamin D in neuropsychiatric disorders. *Psychiatry Research*. 2022; 317: 114782. <https://doi.org/10.1016/j.psychres.2022.114782>.
- [5] Christova T, Ho SK, Liu Y, Gill M, Attisano L. LTK and ALK promote neuronal polarity and cortical migration by inhibiting IGF1R activity. *EMBO Reports*. 2023; 24: e56937. <https://doi.org/10.15252/embr.202356937>.
- [6] Vinsland E, Baskaran P, Mihaylov SR, Hobbs C, Wood H, Bouybayoune I, *et al.* The zinc finger/RING domain protein Unkempt regulates cognitive flexibility. *Scientific Reports*. 2021; 11: 16299. <https://doi.org/10.1038/s41598-021-95286-y>.
- [7] Huang W, Liang Y, Sang C, Mei C, Li X, Chen T. Therapeutic nanosystems co-deliver anticancer drugs and oncogene siRNA to achieve synergetic precise cancer chemo-gene therapy. *Journal of Materials Chemistry. B*. 2018; 6: 3013–3022. <https://doi.org/10.1039/c8tb00004b>.
- [8] Jin Z, Piao L, Sun G, Lv C, Jing Y, Jin R. Dual functional nanoparticles efficiently across the blood-brain barrier to combat glioblastoma via simultaneously inhibit the PI3K pathway and NKG2A axis. *Journal of Drug Targeting*. 2021; 29: 323–335. <https://doi.org/10.1080/1061186X.2020.1841214>.
- [9] Meyer AH, Feldsien TM, Mezler M, Untucht C, Venugopalan R, Lefebvre DR. Novel Developments to Enable Treatment of CNS diseases with targeted drug delivery. *Pharmaceutics*. 2023; 15: 1100. <https://doi.org/10.3390/pharmaceutics15041100>.
- [10] Tassano E, Uccella S, Giacomini T, Striano P, Severino M, Porta S, *et al.* Intragenic Microdeletion of ULK4 and Partial Microduplication of BRWD3 in Siblings with Neuropsychiatric Features and Obesity. *Cytogenetic and Genome Research*. 2018; 156: 14–21. <https://doi.org/10.1159/000491871>.
- [11] Treutlein J, Löhlein S, Einkenel KE, Picotin R, Diekhof EK, Gruber O. Association of Unc-51-like Kinase 4 (ULK4) with the reactivity of the extended reward system in response to conditioned stimuli. *The World Journal of Biological Psychiatry: The Official Journal of the World Federation of Societies of Biological Psychiatry*. 2024; 25: 443–450. <https://doi.org/10.1080/15622975.2024.2393381>.
- [12] Yang R, Zhang T, Han F. Disentangling the genetic overlap between ischemic stroke and obesity. *Diabetology & Metabolic Syndrome*. 2024; 16: 314. <https://doi.org/10.1186/s13098-024-01555-x>.
- [13] Chai T, Tian M, Yang X, Qiu Z, Lin X, Chen L. Association of Circulating Cathepsin B Levels with blood pressure and aortic dilation. *Frontiers in Cardiovascular Medicine*. 2022; 9: 762468. <https://doi.org/10.3389/fcvm.2022.762468>.
- [14] Nandakumar P, Lee D, Hoffmann TJ, Ehret GB, Arking D, Ranatunga D, *et al.* Analysis of putative cis-regulatory elements regulating blood pressure variation. *Human Molecular Genetics*. 2020; 29: 1922–1932. <https://doi.org/10.1093/hmg/ddaa098>.
- [15] Tanaka Y, Fujisawa T, Yazawa S, Ohta I, Takaku Y, Ito M, *et al.* Obesity impairs ciliary function and mucociliary clearance in the murine airway epithelium. *American Journal of Physiology. Lung Cellular and Molecular Physiology*. 2024; 327: L406–L414. <https://doi.org/10.1152/ajplung.00114.2024>.
- [16] Huang L, Tang J, Lin L, Wang R, Chen F, Wei Y, *et al.* Association of genetic variants in ULK4 with the age of first onset of type B aortic dissection. *Frontiers in Genetics*. 2022; 13: 956866. <https://doi.org/10.3389/fgene.2022.956866>.
- [17] Bakoev S, Getmantseva L, Kolosova M, Bakoev F, Kolosov A, Romanets E, *et al.* Identifying Significant SNPs of the total number of piglets born and their relationship with leg bumps in pigs. *Biology*. 2024; 13: 1034. <https://doi.org/10.3390/biolog13121034>.
- [18] Wang M, Jing J, Li H, Liu J, Yuan Y, Sun L. The expression characteristics and prognostic roles of autophagy-related genes in gastric cancer. *Peer J*. 2021; 9: e10814. <https://doi.org/10.7717/peerj.10814>.
- [19] Clavero E, Sanchez-Maldonado JM, Macaudo A, Ter Horst R, Sampaio-Marques B, Jurczynszyn A, *et al.* Polymorphisms within autophagy-related genes as susceptibility biomarkers for multiple myeloma: a meta-analysis of three large cohorts and functional characterization. *International Journal of Molecular Sciences*. 2023; 24: 8500. <https://doi.org/10.3390/ijms24108500>.
- [20] Jee D, Kang S, Huang S, Park S. Polygenetic-Risk Scores Related to Crystallin Metabolism Are Associated with Age-Related Cataract Formation and Interact with Hyperglycemia, Hypertension, Western-Style Diet, and Na Intake. *Nutrients*. 2020; 12: 3534. <https://doi.org/10.3390/nu12113534>.
- [21] Setten RL, Rossi JJ, Han SP. The current state and future directions of RNAi-based therapeutics. *Nature Reviews. Drug Discovery*. 2019; 18: 421–446. <https://doi.org/10.1038/s41573-019-0017-4>.
- [22] Roy SK, Srivastava S, Hancock A, Shrivastava A, Morvant J, Shankar S, *et al.* Inhibition of ribosome assembly factor PNO1 by CRISPR/Cas9 technique suppresses lung adenocarcinoma and Notch pathway: Clinical application. *Journal of Cellular and Molecular Medicine*. 2023; 27: 365–378. <https://doi.org/10.1111/jcmm.17657>.
- [23] Luo S, Zheng N, Lang B. ULK4 in neurodevelopmental and

- neuropsychiatric disorders. *Frontiers in Cell and Developmental Biology*. 2022; 10: 873706. <https://doi.org/10.3389/fcell.2022.873706>.
- [24] Khamrui S, Ung PMU, Secor C, Schlessinger A, Lazarus MB. High-resolution structure and inhibition of the schizophrenia-linked pseudokinase ULK4. *Journal of the American Chemical Society*. 2020; 142: 33–37. <https://doi.org/10.1021/jacs.9b10458>.
- [25] Eysers PA. Marveling at the Incredible ULK4. *Structure*. 2020; 28: 1181–1183. <https://doi.org/10.1016/j.str.2020.10.005>.
- [26] Preuss F, Chatterjee D, Mathea S, Shrestha S, St-Germain J, Saha M, *et al*. Nucleotide binding, evolutionary insights, and interaction partners of the pseudokinase Unc-51-like kinase 4. *Structure*. 2020; 28: 1184–1196.e6. <https://doi.org/10.1016/j.str.2020.07.016>.
- [27] Mecklenburg N, Kowalczyk I, Witte F, Görne J, Laier A, Mamo TM, *et al*. Identification of disease-relevant modulators of the SHH pathway in the developing brain. *Development*. 2021; 148: dev199307. <https://doi.org/10.1242/dev.199307>.
- [28] Zhou M, Han Y, Jiang J. Phosphorylation-induced SUMOylation promotes Ulk4 condensation at ciliary tip to transduce Hedgehog signal. *BiorXiv: The Preprint Server for Biology*. 2024. <https://doi.org/10.1101/2024.09.19.613872>. (preprint)
- [29] Lang B, Pu J, Hunter I, Liu M, Martín-Granados C, Reilly TJ, *et al*. Recurrent deletions of ULK4 in schizophrenia: a gene crucial for neurogenesis and neuronal motility. *Journal of Cell Science*. 2014; 127: 630–640. <https://doi.org/10.1242/jcs.137604>.
- [30] Liu M, Guan Z, Shen Q, Flinter F, Domínguez L, Ahn JW, *et al*. Ulk4 regulates neural stem cell pool. *Stem Cells*. 2016; 34: 2318–2331. <https://doi.org/10.1002/stem.2423>.
- [31] Liu M, Xu P, O'Brien T, Shen S. Multiple roles of Ulk4 in neurogenesis and brain function. *Neurogenesis (Austin, Tex.)*. 2017; 4: e1313646. <https://doi.org/10.1080/23262133.2017.1313646>.
- [32] Domínguez L, Schlosser G, Shen S. Expression of a novel serine/threonine kinase gene, Ulk4, in neural progenitors during *Xenopus laevis* forebrain development. *Neuroscience*. 2015; 290: 61–79. <https://doi.org/10.1016/j.neuroscience.2014.12.060>.
- [33] Lang B, Zhang L, Jiang G, Hu L, Lan W, Zhao L, *et al*. Control of cortex development by ULK4, a rare risk gene for mental disorders including schizophrenia. *Scientific Reports*. 2016; 6: 31126. <https://doi.org/10.1038/srep31126>.
- [34] Liu M, Xu P, Guan Z, Qian X, Dockery P, Fitzgerald U, *et al*. Ulk4 deficiency leads to hypomyelination in mice. *Glia*. 2018; 66: 175–190. <https://doi.org/10.1002/glia.23236>.
- [35] Vogel P, Read RW, Hansen GM, Payne BJ, Small D, Sands AT, *et al*. Congenital hydrocephalus in genetically engineered mice. *Veterinary Pathology*. 2012; 49: 166–181. <https://doi.org/10.1177/0300985811415708>.
- [36] Liu M, Guan Z, Shen Q, Lalor P, Fitzgerald U, O'Brien T, *et al*. Ulk4 is essential for ciliogenesis and CSF flow. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*. 2016; 36: 7589–7600. <https://doi.org/10.1523/JNEUROSCI.0621-16.2016>.
- [37] McCoy CJ, Paupelin-Vaucelle H, Gorilak P, Beneke T, Varga V, Gluenz E. ULK4 and Fused/STK36 interact to mediate assembly of a motile flagellum. *Molecular Biology of the Cell*. 2023; 34: ar66. <https://doi.org/10.1091/mbc.E22-06-0222>.
- [38] Zhang H, Yang M, Zhang J, Li L, Guan T, Liu J, *et al*. The putative protein kinase Stk36 is essential for ciliogenesis and CSF flow by associating with Ulk4. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2023; 37: e23138. <https://doi.org/10.1096/fj.202300481R>.
- [39] Zhou M, Han Y, Jiang J. Ulk4 promotes Shh signaling by regulating Stk36 ciliary localization and Gli2 phosphorylation. *ELife*. 2023; 12: RP88637. <https://doi.org/10.7554/eLife.88637>.
- [40] Bhattacharyya U, Deshpande SN, Bhatia T, Thelma BK. Revisiting schizophrenia from an evolutionary perspective: an association study of recent evolutionary markers and schizophrenia. *Schizophrenia Bulletin*. 2021; 47: 827–836. <https://doi.org/10.1093/schbul/sbaa179>.
- [41] Hu L, Chen Y, Yang CP, Huang Y, Song NN, Chen JY, *et al*. Ulk4, a Newly Discovered Susceptibility Gene for Schizophrenia, Regulates Corticogenesis in Mice. *Frontiers in Cell and Developmental Biology*. 2021; 9: 645368. <https://doi.org/10.3389/fcell.2021.645368>.
- [42] Hu L, Zhou BY, Yang CP, Lu DY, Tao YC, Chen L, *et al*. Deletion of Schizophrenia Susceptibility Gene Ulk4 leads to abnormal cognitive behaviors via Akt-GSK-3 signaling pathway in mice. *Schizophrenia bulletin*. 2022; 48: 804–813. <https://doi.org/10.1093/schbul/sbac040>.
- [43] Liu M, Fitzgibbon M, Wang Y, Reilly J, Qian X, O'Brien T, *et al*. Ulk4 regulates GABAergic signaling and anxiety-related behavior. *Translational Psychiatry*. 2018; 8: 43. <https://doi.org/10.1038/s41398-017-0091-5>.
- [44] Luo W, Yang J. Schizophrenia predisposition gene Unc-51-like kinase 4 for the improvement of cerebral ischemia/reperfusion injury. *Molecular Biology Reports*. 2022; 49: 2933–2943. <https://doi.org/10.1007/s11033-021-07108-z>.
- [45] Chaikuad A, Koschade SE, Stolz A, Zivkovic K, Pohl C, Shaid S, *et al*. Conservation of structure, function and inhibitor binding in UNC-51-like kinase 1 and 2 (ULK1/2). *The Biochemical Journal*. 2019; 476: 875–887. <https://doi.org/10.1042/BCJ20190038>.
- [46] Lazarus MB, Novotny CJ, Shokat KM. Structure of the human autophagy initiating kinase ULK1 in complex with potent inhibitors. *ACS Chemical Biology*. 2015; 10: 257–261. <https://doi.org/10.1021/cb500835z>.
- [47] Boudeau J, Miranda-Saavedra D, Barton GJ, Alessi DR. Emerging roles of pseudokinases. *Trends in Cell Biology*. 2006; 16: 443–452. <https://doi.org/10.1016/j.tcb.2006.07.003>.
- [48] Origlia N, Bonadonna C, Rosellini A, Leznik E, Arancio O, Yan SS, *et al*. Microglial receptor for advanced glycation end product-dependent signal pathway drives beta-amyloid-induced synaptic depression and long-term depression impairment in entorhinal cortex. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*. 2010; 30: 11414–11425. <https://doi.org/10.1523/JNEUROSCI.2127-10.2010>.
- [49] Zeqiraj E, van Aalten DM. Pseudokinases-remnants of evolution or key allosteric regulators. *Current Opinion in Structural Biology*. 2010; 20: 772–781. <https://doi.org/10.1016/j.sbi.2010.10.001>.
- [50] Kannan N, Taylor SS. Rethinking pseudokinases. *Cell*. 2008; 133: 204–205. <https://doi.org/10.1016/j.cell.2008.04.005>.
- [51] Han Y, Wang B, Cho YS, Zhu J, Wu J, Chen Y, *et al*. Phosphorylation of Ci/Gli by fused family kinases promotes hedgehog signaling. *Developmental Cell*. 2019; 50: 610–626.e4. <https://doi.org/10.1016/j.devcel.2019.06.008>.
- [52] Murone M, Luoh SM, Stone D, Li W, Gurney A, Armanini M, *et al*. Gli regulation by the opposing activities of fused and suppressor of fused. *Nature Cell Biology*. 2000; 2: 310–312. <https://doi.org/10.1038/35010610>.
- [53] Rowitch DH, Kriegstein AR. Developmental genetics of vertebrate glial-cell specification. *Nature*. 2010; 468: 214–222. <https://doi.org/10.1038/nature09611>.
- [54] Negintaji K, Ghanbari A, Frozanfar M, Jafarinia M, Zibara K. Pregnenolone enhances the proliferation of mouse neural stem cells and promotes oligodendrogenesis, together with Sox10, and neurogenesis, along with Notch1 and Pax6. *Neurochemistry International*. 2023; 163: 105489. <https://doi.org/10.1016/j.neui>

nt.2023.105489.

- [55] Furutachi S, Miya H, Watanabe T, Kawai H, Yamasaki N, Harada Y, *et al.* Slowly dividing neural progenitors are an embryonic origin of adult neural stem cells. *Nature Neuroscience*. 2015; 18: 657–665. <https://doi.org/10.1038/nn.3989>.
- [56] Fuentealba LC, Rompani SB, Parraguez JI, Obernier K, Romero R, Cepko CL, *et al.* Embryonic Origin of Postnatal Neural Stem Cells. *Cell*. 2015; 161: 1644–1655. <https://doi.org/10.1016/j.cell.2015.05.041>.
- [57] Chaker Z, Segalada C, Kretz JA, Acar IE, Delgado AC, Crotet V, *et al.* Pregnancy-responsive pools of adult neural stem cells for transient neurogenesis in mothers. *Science*. 2023; 382: 958–963. <https://doi.org/10.1126/science.abo5199>.
- [58] Shinawi M, Liu P, Kang SH, Shen J, Belmont JW, Scott DA, *et al.* Recurrent reciprocal 16p11.2 rearrangements associated with global developmental delay, behavioural problems, dysmorphism, epilepsy, and abnormal head size. *Journal of Medical Genetics*. 2010; 47: 332–341. <https://doi.org/10.1136/jmg.2009.073015>.
- [59] Kunoh S, Nakashima H, Nakashima K. Epigenetic regulation of neural stem cells in developmental and adult stages. *Epigenomes*. 2024; 8: 22. <https://doi.org/10.3390/epigenomes8020022>.
- [60] Neaverson A, Andersson MHL, Arshad OA, Foulser L, Goodwin-Trotman M, Hunter A, *et al.* Differentiation of human induced pluripotent stem cells into cortical neural stem cells. *Frontiers in Cell and Developmental Biology*. 2022; 10: 1023340. <https://doi.org/10.3389/fcell.2022.1023340>.
- [61] Li L, Wei D, Wang Q, Pan J, Liu R, Zhang X, *et al.* MEC-17 deficiency leads to reduced  $\alpha$ -tubulin acetylation and impaired migration of cortical neurons. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*. 2012; 32: 12673–12683. <https://doi.org/10.1523/JNEUROSCI.0016-12.2012>.
- [62] Wei D, Gao N, Li L, Zhu JX, Diao L, Huang J, *et al.*  $\alpha$ -Tubulin acetylation restricts axon overbranching by dampening microtubule plus-end dynamics in neurons. *Cerebral Cortex*. 2018; 28: 3332–3346. <https://doi.org/10.1093/cercor/bhx225>.
- [63] Lewis DA, Hashimoto T, Volk DW. Cortical inhibitory neurons and schizophrenia. *Nature Reviews Neuroscience*. 2005; 6: 312–324. <https://doi.org/10.1038/nrn1648>.
- [64] Peters BD, Karlsgodt KH. White matter development in the early stages of psychosis. *Schizophrenia Research*. 2015; 161: 61–69. <https://doi.org/10.1016/j.schres.2014.05.021>.
- [65] Mulligan KA, Cheyette BN. Wnt signaling in vertebrate neural development and function. *Journal of Neuroimmune Pharmacology: The Official Journal of the Society on Neuroimmune Pharmacology*. 2012; 7: 774–787. <https://doi.org/10.1007/s11481-012-9404-x>.
- [66] Bem J, Brożko N, Chakraborty C, Lipiec MA, Koziński K, Nagalski A, *et al.* Wnt/ $\beta$ -catenin signaling in brain development and mental disorders: keeping TCF7L2 in mind. *FEBS Letters*. 2019; 593: 1654–1674. <https://doi.org/10.1002/1873-3468.13502>.
- [67] Tully HM, Dobyns WB. Infantile hydrocephalus: a review of epidemiology, classification and causes. *European Journal of Medical Genetics*. 2014; 57: 359–368. <https://doi.org/10.1016/j.ejmg.2014.06.002>.
- [68] Kahle KT, Kulkarni AV, Limbrick DD Jr, Warf BC. Hydrocephalus in children. *Lancet*. 2016; 387: 788–799. [https://doi.org/10.1016/S0140-6736\(15\)60694-8](https://doi.org/10.1016/S0140-6736(15)60694-8).
- [69] Jiang Z, Zhou J, Qin X, Zheng H, Gao B, Liu X, *et al.* MT1-MMP deficiency leads to defective ependymal cell maturation, impaired ciliogenesis, and hydrocephalus. *JCI Insight*. 2020; 5: e132782. <https://doi.org/10.1172/jci.insight.132782>.
- [70] Ohata S, Nakatani J, Herranz-Pérez V, Cheng J, Belinson H, Inubushi T, *et al.* Loss of Dishevelleds disrupts planar polarity in ependymal motile cilia and results in hydrocephalus. *Neuron*. 2014; 83: 558–571. <https://doi.org/10.1016/j.neuron.2014.06.022>.
- [71] Shukla S, Haenold R, Urbánek P, Frappart L, Monajembashi S, Grigaravicius P, *et al.* TRIP6 functions in brain ciliogenesis. *Nature Communications*. 2021; 12: 5887. <https://doi.org/10.1038/s41467-021-26057-6>.
- [72] Guerra MM, Henzi R, Ortloff A, Lichtin N, Vio K, Jiménez AJ, *et al.* Cell junction pathology of neural stem cells is associated with ventricular zone disruption, hydrocephalus, and abnormal neurogenesis. *Journal of Neuropathology and Experimental Neurology*. 2015; 74: 653–671. <https://doi.org/10.1097/NEU.0000000000000203>.
- [73] Kumar V, Umair Z, Kumar S, Goutam RS, Park S, Kim J. The regulatory roles of motile cilia in CSF circulation and hydrocephalus. *Fluids and Barriers of the CNS*. 2021; 18: 31. <https://doi.org/10.1186/s12987-021-00265-0>.
- [74] Mazor M, Alkrinawi S, Chalifa-Caspi V, Manor E, Sheffield VC, Aviram M, *et al.* Primary ciliary dyskinesia caused by homozygous mutation in DNAL1, encoding dynein light chain 1. *American Journal of Human Genetics*. 2011; 88: 599–607. <https://doi.org/10.1016/j.ajhg.2011.03.018>.
- [75] Suryavanshi S, Eddé B, Fox LA, Guerrero S, Hard R, Hennessey T, *et al.* Tubulin glutamylation regulates ciliary motility by altering inner dynein arm activity. *Current Biology*. 2010; 20: 435–440. <https://doi.org/10.1016/j.cub.2009.12.062>.
- [76] Nozawa YI, Yao E, Lin C, Yang JH, Wilson CW, Gacayan R, *et al.* Fused (Stk36) is a ciliary protein required for central pair assembly and motile cilia orientation in the mammalian oviduct. *Developmental Dynamics: An Official Publication of the American Association of Anatomists*. 2013; 242: 1307–1319. <https://doi.org/10.1002/dvdy.24024>.
- [77] Fields RD. White matter in learning, cognition and psychiatric disorders. *Trends in Neurosciences*. 2008; 31: 361–370. <https://doi.org/10.1016/j.tins.2008.04.001>.
- [78] Pujol J, López-Sala A, Sebastián-Gallés N, Deus J, Cardoner N, Soriano-Mas C, *et al.* Delayed myelination in children with developmental delay detected by volumetric MRI. *NeuroImage*. 2004; 22: 897–903. <https://doi.org/10.1016/j.neuroimage.2004.01.029>.
- [79] Jungerius BJ, Hoogendoorn ML, Bakker SC, Van't Slot R, Bardeol AF, Ophoff RA, *et al.* An association screen of myelin-related genes implicates the chromosome 22q11 PIK4CA gene in schizophrenia. *Molecular Psychiatry*. 2008; 13: 1060–1068. <https://doi.org/10.1038/sj.mp.4002080>.
- [80] Ishimoto T, Ninomiya K, Inoue R, Koike M, Uchiyama Y, Mori H. Mice lacking BCAS1, a novel myelin-associated protein, display hypomyelination, schizophrenia-like abnormal behaviors, and upregulation of inflammatory genes in the brain. *Glia*. 2017; 65: 727–739. <https://doi.org/10.1002/glia.23129>.
- [81] Baumann N, Pham-Dinh D. Biology of oligodendrocyte and myelin in the mammalian central nervous system. *Physiological Reviews*. 2001; 81: 871–927. <https://doi.org/10.1152/physrev.2001.81.2.871>.
- [82] Ferreira RS, Ribeiro PR, Silva JHCE, Hoppe JB, de Almeida MMA, de Lima Ferreira BC, *et al.* Amburana caerensis seed extract stimulates astrocyte glutamate homeostatic mechanisms in hippocampal brain slices and protects oligodendrocytes against ischemia. *BMC Complementary Medicine and Therapies*. 2023; 23: 154. <https://doi.org/10.1186/s12906-023-03959-0>.
- [83] GBD 2019 Mental Disorders Collaborators. Global, regional, and national burden of 12 mental disorders in 204 countries and territories, 1990–2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet Psychiatry*. 2022; 9: 137–150. [https://doi.org/10.1016/S2215-0366\(21\)00395-3](https://doi.org/10.1016/S2215-0366(21)00395-3).

- [84] Global Research on Developmental Disabilities Collaborators. Developmental disabilities among children younger than 5 years in 195 countries and territories, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. *The Lancet. Global Health*. 2018; 6: e1100–e1121. [https://doi.org/10.1016/S2214-109X\(18\)30309-7](https://doi.org/10.1016/S2214-109X(18)30309-7).
- [85] Power RA, Tansey KE, Buttenschön HN, Cohen-Woods S, Bigdeli T, Hall LS, *et al*. Genome-wide association for major depression through age at onset stratification: major depressive disorder working group of the psychiatric genomics consortium. *Biological Psychiatry*. 2017; 81: 325–335. <https://doi.org/10.1016/j.biopsych.2016.05.010>.
- [86] Insel TR. Rethinking schizophrenia. *Nature*. 2010; 468: 187–193. <https://doi.org/10.1038/nature09552>.
- [87] Chadha R, Meador-Woodruff JH. Downregulated AKT-mTOR signaling pathway proteins in dorsolateral prefrontal cortex in Schizophrenia. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*. 2020; 45: 1059–1067. <https://doi.org/10.1038/s41386-020-0614-2>.
- [88] Matsuda S, Ikeda Y, Murakami M, Nakagawa Y, Tsuji A, Kitagishi Y. Roles of PI3K/AKT/GSK3 Pathway involved in psychiatric illnesses. *Diseases*. 2019; 7: 22. <https://doi.org/10.3390/diseases7010022>.
- [89] Jacobsen AV, Murphy JM. The secret life of kinases: insights into non-catalytic signalling functions from pseudokinases. *Biochemical Society Transactions*. 2017; 45: 665–681. <https://doi.org/10.1042/BST20160331>.
- [90] Selten M, van Bokhoven H, Nadif Kasri N. Inhibitory control of the excitatory/inhibitory balance in psychiatric disorders. *F1000Research*. 2018; 7: 23. <https://doi.org/10.12688/f1000research.12155.1>.
- [91] Lopatina OL, Malinovskaya NA, Komleva YK, Gorina YV, Shuvaev AN, Olovyannikova RY, *et al*. Excitation/inhibition imbalance and impaired neurogenesis in neurodevelopmental and neurodegenerative disorders. *Reviews in the Neurosciences*. 2019; 30: 807–820. <https://doi.org/10.1515/revneuro-2019-0014>.
- [92] Jia DW, Vogels TP, Costa RP. Developmental depression-to-facilitation shift controls excitation-inhibition balance. *Communications Biology*. 2022; 5: 873. <https://doi.org/10.1038/s42003-022-03801-2>.
- [93] Viggiano A, Cacciola G, Widmer DA, Viggiano D. Anxiety as a neurodevelopmental disorder in a neuronal subpopulation: Evidence from gene expression data. *Psychiatry Research*. 2015; 228: 729–740. <https://doi.org/10.1016/j.psychres.2015.05.032>.
- [94] Yung NCL, Wong CSM, Chan JKN, Or PCF, Chen EYH, Chang WC. Mortality in patients with schizophrenia admitted for incident ischemic stroke: A population-based cohort study. *European Neuropsychopharmacology: The Journal of the European College of Neuropsychopharmacology*. 2020; 31: 152–157. <https://doi.org/10.1016/j.euroneuro.2019.12.107>.
- [95] Chu RST, Chong RCH, Chang DHH, Shan Leung AL, Chan JKN, Wong CSM, *et al*. The risk of stroke and post-stroke mortality in people with schizophrenia: A systematic review and meta-analysis study. *Psychiatry Research*. 2024; 332: 115713. <https://doi.org/10.1016/j.psychres.2024.115713>.
- [96] Oh J, Nam H, Park S, Chae JH, Kim TS. Decreased cardiovascular death in schizophrenia patients treated with antipsychotics: A Korean national cohort study. *Schizophrenia Research*. 2021; 228: 417–424. <https://doi.org/10.1016/j.schres.2021.01.006>.
- [97] Linden J, Van de Beeck L, Plumier JC, Ferrara A. Procedural learning as a measure of functional impairment in a mouse model of ischemic stroke. *Behavioural Brain Research*. 2016; 307: 35–45. <https://doi.org/10.1016/j.bbr.2016.03.032>.
- [98] Fatemi I, Saeed-Askari P, Hakimzadeh E, Kaeidi A, Esmail-Moghaddam S, Pak-Hashemi M, *et al*. Long-term metformin therapy improves neurobehavioral functions and antioxidative activity after cerebral ischemia/reperfusion injury in rats. *Brain Research Bulletin*. 2020; 163: 65–71. <https://doi.org/10.1016/j.brainresbull.2020.07.015>.
- [99] Li X, Ma X, Miao Y, Zhang J, Xi B, Li W, *et al*. Duvelisib attenuates bleomycin-induced pulmonary fibrosis via inhibiting the PI3K/Akt/mTOR signalling pathway. *Journal of Cellular and Molecular Medicine*. 2023; 27: 422–434. <https://doi.org/10.1111/jcmm.17665>.
- [100] Riley B, Williamson M, Collier D, Wilkie H, Makoff A. A 3-Mb map of a large Segmental duplication overlapping the alpha7-nicotinic acetylcholine receptor gene (CHRNA7) at human 15q13-q14. *Genomics*. 2002; 79: 197–209. <https://doi.org/10.1006/geno.2002.6694>.
- [101] Szigeti K, Ihnatovych I, Birkaya B, Chen Z, Ouf A, Indurthi DC, *et al*. CHRFAM7A: A human specific fusion gene, accounts for the translational gap for cholinergic strategies in Alzheimer’s disease. *EBioMedicine*. 2020; 59: 102892. <https://doi.org/10.1016/j.ebiom.2020.102892>.