

Review

The Role of the Ganglion Cell Layer as an OCT Biomarker in Neurodegenerative Diseases

Francesco Ruggeri^{1,†}, Daniele Fumi^{1,†}, Lorena Bassis¹, Mariachiara Di Pippo¹,
Solmaz Abdolrahimzadeh^{1,2,*}¹Ophthalmology Unit, Neurosciences, Mental Health, and Sense Organs (NESMOS) Department, Faculty of Medicine and Psychology, University of Rome Sapienza, 00189 Rome, Italy²Ophthalmology Unit, Department of Surgical Sciences, St. Andrea Hospital, 00189 Rome, Italy*Correspondence: solmaz.abdolrahimzadeh@uniroma1.it (Solmaz Abdolrahimzadeh)

†These authors contributed equally.

Academic Editor: Bettina Platt

Submitted: 7 August 2024 Revised: 26 November 2024 Accepted: 4 December 2024 Published: 23 May 2025

Abstract

Optical coherence tomography (OCT) is a non-invasive imaging technique in the field of ophthalmology that has been increasingly recognized for its capability to identify potential biomarkers in neurodegenerative processes. While the retinal nerve fiber layer (RNFL) has been vastly explored, this review focuses on the ganglion cell layer (GCL), highlighting its relevance and potential advantages in the diagnostic approach and monitoring of neurodegenerative conditions. In the present review we explore the role of GCL changes detected by OCT in Alzheimer's disease (AD), Parkinson's disease (PD), and multiple sclerosis (MS). We focus on these conditions due to their prevalence and substantial social impact among neurodegenerative diseases. We summarize key findings on the changes in the GCL and their correlations with disease progression and severity. Moreover, we highlight GCL measurements in the context of a multidisciplinary diagnostic approach, and their potential in adapting tailored therapeutic strategies in neurodegenerative disease management. Challenges such as methodological variability in OCT measurements, automatic instrumental output parameters, the limitations of GCL as a standalone diagnostic tool, and the impact of systemic and ocular factors are discussed. Finally, we propose that forthcoming advancements in OCT technology, integration with other biomarkers, and longitudinal studies will likely further enhance the understanding of GCL changes over time.

Keywords: ganglion cell layer; spectral domain optical coherence tomography; Alzheimer disease; Parkinson's disease; multiple sclerosis; neurodegenerative diseases

1. Introduction

Retinal tissue and the optic nerve share an embryological origin with the central nervous system, as they both derive from the neuroectoderm [1–9]. Given the similarities in structure and characteristics, there is a growing interest in utilizing retinal *in vivo* imaging as a potential diagnostic and monitoring tool for neurological conditions, particularly neurodegenerative diseases [1–9].

Optical coherence tomography (OCT) delivers rapid, non-invasive, cost-effective, and accessible imaging with near-histological precision, facilitating the distinct identification of individual retinal layers. OCT evaluation of the inner retinal structures, such as the retinal nerve fiber layer (RNFL), inner plexiform layer (IPL), and ganglion cell layer (GCL), is valuable in evaluating neurodegenerative diseases [1,6–13] (Fig. 1, Ref. [14]).

Traditionally, the diagnosis and follow-up of neurodegenerative pathology often involves costly and time-consuming neuroimaging examinations such as magnetic resonance imaging (MRI) and positron emission tomography (PET) scans. In contrast, *in vivo* retinal imaging through OCT offers a quicker and more widely available

alternative. Moreover, OCT can capture micrometric variations that are not discernible with conventional neuroimaging techniques [1–3,10,13]. Therefore, a significant quantity of publications have recently emerged, exploring the relationship between retinal imaging and traditional biomarkers of neurodegenerative diseases. While it is not anticipated that retinal imaging will supplant neuroimaging as the main diagnostic tool for neurodegenerative processes, nor serve as a standalone diagnostic criterion, its potential to efficiently deliver further insights is undeniable [1–3,6–9,12,13,15]. The integration of OCT could prove instrumental during the diagnostic and screening phases—owing to its rapidity and non-invasiveness—and throughout the monitoring phases of disease, significantly enhancing patient care quality [1–3,6–9,12,13,15].

The three innermost layers of the retina are the RNFL, the GCL and the IPL, also known as the ganglion cell complex (GCC) [1,10]. The RNFL constitutes the axons, the GCL contains the cell bodies of the inner retina's third neuron, making it a critical indicator of neuronal loss, and the IPL contains the ganglion cell dendrites primarily consisting of synaptic connections [11]. The RNFL has demon-



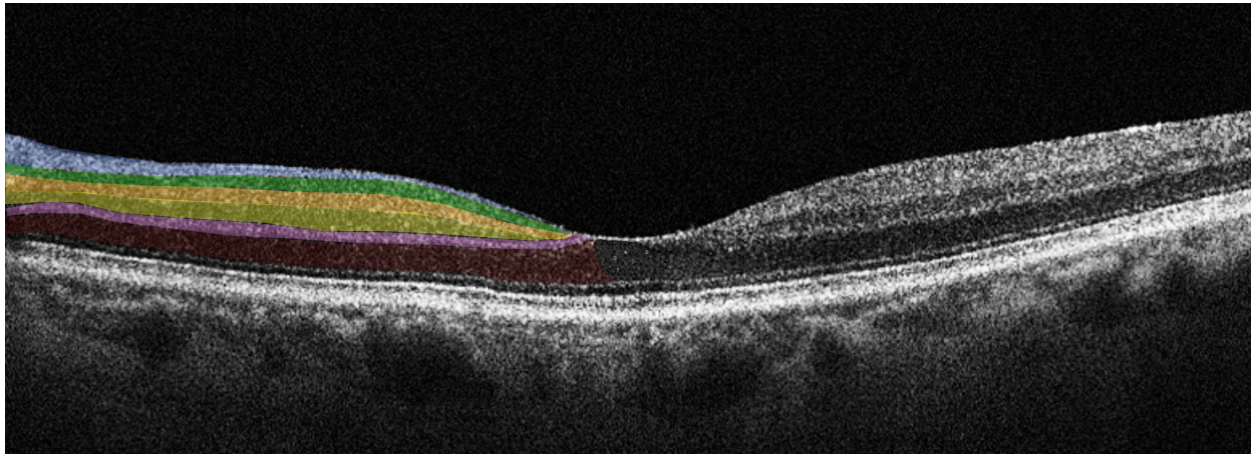


Fig. 1. Retinal layers optical coherence tomography (OCT) depiction. Horizontal spectral domain optical coherence tomography (SD-OCT) cross-sectional scan passing through the fovea. The various retinal layers are shown with added color overlays on the left side as follows: blue: retinal nerve fiber layer; green: ganglion cell layer; orange: inner plexiform layer; yellow: inner nuclear layer; pink: outer plexiform layer; red: outer nuclear layer. The right side shows original SD-OCT output in scales of grey. From Di Pippo M, Fragiotta S, Di Staso F, Scuderi L, Abdolrahimzadeh S. The Role of Alpha-Synuclein Deposits in Parkinson's Disease: A Focus on the Human Retina. *International Journal of Molecular Sciences*. 2023; 24: 4391. <https://doi.org/10.3390/ijms24054391> [14].

strated a strong correlation with disease markers and has been extensively studied [4,5,16–23], but the presence of large retinal vessels in the peripapillary region can complicate RNFL measurement, as these vessels may obscure or alter the apparent thickness of the RNFL [24]. In contrast, the GCL is not affected by vascular structures, potentially offering more consistent measurements [25]. While this paper focuses on the GCL, including the IPL may be beneficial for understanding trans-synaptic degeneration and other neurodegenerative mechanisms by offering a more comprehensive view of retinal layer changes. Nevertheless, analysis of the GCL reflects its prominence in the existing literature and ensures consistency in synthesizing findings [26,27].

In this review, we aim to summarize the existing literature on the role of the GCL as a biomarker in Alzheimer's disease (AD), Parkinson's disease (PD), and multiple sclerosis (MS), highlighting the GCL as a valuable biomarker across neurodegenerative diseases. In AD the thinning of the GCL correlates with cognitive decline, suggesting its potential as a biomarker for integrating diagnosis and monitoring disease [28]. In PD, GCL thinning is associated with both motor and non-motor symptoms, reflecting broader central nervous system involvement [29]. In MS, GCL thinning correlates with disease activity and progression, particularly in cases of optic neuritis [30]. When integrated appropriately into a diagnostic workflow, OCT imaging of the GCL can significantly enhance the diagnostic process and longitudinal neurodegenerative disease evaluation.

2. The Ganglion Cell Layer as a Biomarker

The complexity and frequently invasive nature of traditional diagnostic methods in the diagnosis and follow-up

of disease progression have posed significant challenges in the management of neurodegenerative conditions. Given the crucial need for early diagnostic approaches, there is an ongoing imperative to develop and enhance techniques for detecting initial pathological signs while ensuring they are minimally invasive. Recent advances in medical imaging have highlighted the role of retinal imaging methods as a potential tool in the diagnostic workup of neurodegenerative diseases [2,3,14]. The retina, an accessible extension of the central nervous system, shares many anatomical and physiological characteristics with brain tissue [28,31]. This has led to the hypothesis that changes occurring in central degenerative diseases might be reflected, or even anticipated, at the retina [4,5,16,17,32]. The retinal GCL and the RNFL contain the neuronal body and axons that connect the photoreceptors with the pathways of vision in the central nervous system. These structures can be analyzed using OCT, which enables *in vivo* assessment of the GCL, IPL, and RNFL thickness [18]. The basis for GCL evaluation as a valid biomarker lies in the fact that retinal ganglion cells (RGC) are neurons located in the GCL [1,11,33–38]. This leads to a strong likelihood that, neurodegenerative changes affecting the retina would have an impact on this specific layer [26]. Indeed, it has been extensively reported that cognitive impairment is related to thinning of the neuronal retinal layers, specifically impacting the RGCs observed through retinal layer segmentation provided by OCT software [18,19,33–35]. Thinning of the GCL has been linked to cognitive decline in AD, motor and non-motor symptoms in PD, and disease activity in MS [29,30,33].

3. GCL Changes in Alzheimer's Disease

AD is a neurodegenerative disease that represents the predominant cause of dementia. The condition primarily starts with episodic memory deficits, and as the disease progresses, patients experience difficulty in routine activity, cognitive status changes, and visual alterations, eventually leading to brain atrophy [39]. Historically, AD was diagnosed at autopsy, while the clinical diagnosis was considered as “probable AD”. Over time, the definition of AD has broadened, encompassing both a prototypical clinical syndrome lacking neuropathological confirmation and neurologically-defined AD [18,40].

Research on AD benefits from the ongoing development of biomarkers indicative of critical neuropathological changes such as β -amyloid ($A\beta$) deposition, pathological tau, and neurodegeneration, where a definitive diagnosis currently relies on invasive procedures that aim to identify these deposits [18,41]. Additionally, diagnostic methods like MRI, PET, and cerebrospinal fluid (CSF) for $A\beta$ and tau detection, along with serological exams for genetic markers, have proven valuable [42]. Morinaga *et al.* [43] reported that CSF biomarkers were most sensitive at Clinical Dementia Rating (CDR) 0.5 (90.0%), while fluorodeoxyglucose-PET (FDG-PET) (96.7%) and CSF biomarkers (95.5%) were highly sensitive at CDR 1. At CDR 2, all tools showed consistently high detection rates. Despite their diagnostic utility, the disadvantages of these methods are the high costs, invasive nature, and variable sensitivity and specificity. Therefore, AD diagnosis is substantially clinical [18,42], predominantly through anamnestic and clinical neurological assessment and evaluation of cognitive status. The latter is through applications such as the Mini-Mental State Examination (MMSE) and the CDR scale, which can be time-consuming and challenging to carry out accurately, even by specialists [18,40,42].

The pathological finding in the retina of AD patients is the deposit of $A\beta$ plaques in the GCC [18]. AD patients often report blurred vision and visual symptoms, including alteration in spatial contrast sensitivity, motion perception, and color discrimination [44–47]. These conditions were thought to be secondary to impairment in the visual cortex or degeneration in higher cortical areas [48]. As more data has been gathered through the years and technology has advanced it has become clear that the locus of the pathology is not cortical but rather at a pre-cortical level. Iseri *et al.* [49] reported physiological visual evoked potential (VEP) responses, suggesting that although there is RNFL loss, there is no contextual involvement of the primary cortical region. Blanks *et al.* [34] identified a retinal correlate for AD through histological examination. They reported a 25% loss of GCL neurons in the foveal/parafoveal area. This was the rationale for implementing OCT documentation for AD [49]. In a recent study, Kao *et al.* [18] analyzed 156 patients, dividing them into two groups based on their MMSE scores. One group included patients with mild AD,

who had MMSE scores ranging from 21 to 26, while the other group included those with moderate AD, with scores from 14 to 20. These authors found that the ganglion cell-inner plexiform layer (GC-IPL) was particularly vulnerable to damage in AD, showing a reduction in overall thickness [18]. This pattern aligned with similar findings in other studies [49–51]. The thinning of the retinal layers was particularly evident in the 1–3 mm ring of the macula due to the distribution of the RGCs, the density of which is higher in the central retina, diminishing centrifugally [18].

OCT evaluation could be useful in the diagnostic phase and in monitoring disease activity. Although not well established, Kao *et al.* [18] hypothesized that there could be a correlation between disease severity, evaluated through cognitive function impairment, and macular thickness as they found that retinal thickness correlated with clinical disease severity measured with both MMSE and CDR but better aligned with the CDR scores.

Wisely *et al.* [52], based on OCT and optical coherence tomography angiography (OCTA), developed a convolutional neural network (CNN) to evaluate AD using multimodal retinal images. These authors analyzed images from 284 eyes of 159 subjects [52]. The CNN model successfully distinguished AD from healthy controls. The GC-IPL maps were identified as the most reliable value in discerning between AD subjects and healthy controls, achieving a high predictive accuracy (area under the curve (AUC) of 0.809). The integration of multiple imaging modalities and patient data further enhanced the model's performance (AUC of 0.836), showing the potential of retinal imaging using OCT scans as a rapid and non-invasive method for early AD monitoring [52]. Santangelo *et al.* [53] recently studied a total of 137 patients: 43 with AD, 37 with mild cognitive impairment (MCI), and 57 healthy controls assessed through OCT examination to evaluate the RNFL, GCL, and IPL. The authors found that the reduction in macular GCL thickness, serving as a biomarker, was more specific for AD, while the peripapillary RNFL was significantly reduced in thickness both globally and in the superior quadrant in MCI and in AD, with no major differences between the two, thus, making GCL a better fit as a diagnostic biomarker [53]. The authors, however, did not report on axial eye length measurement, which can affect retinal layer thickness values. Thinning of the GCL in AD was also observed predominantly in the central and temporal regions of the retina, aligning with the distribution of RGCs [54].

Wang *et al.* [55] conducted a cross-sectional study including 159 patients with AD and 299 healthy control subjects in order to assess the diagnostic role of a machine learning (ML) model. The authors found a positive correlation of GCL thickness with CSF $A\beta_{42}/A\beta_{40}$ and a negative correlation with CSF p-tau levels. IPL thickness showed a positive correlation with CSF $A\beta_{42}$ levels and a negative correlation with CSF t-tau levels. The authors hypothesized that retinal $A\beta$ and tau levels could reflect brain levels

[55]. However, these authors did not carry out a longitudinal study in order to assess the progression of neurodegeneration and reported that patients with more advanced stage AD (CDR = 3) were not included in the study based on a lack of patient cooperation necessary for OCT imaging and neurophysiological examinations. Indeed, while the GCL offers a promising window as an effective biomarker in AD progression, findings must be interpreted with caution, and further longitudinal studies are necessary to better define the utility of GCL measurements in the broader diagnostic and monitoring strategies for AD.

The diagnosis of AD is predominantly clinical, given the well-documented limitations of the methods required for definitive instrumental and laboratory diagnosis [18,41,42]. In this context, OCT analysis could serve as a valuable tool in managing the disease. Several studies have shown that a reduction in GCL thickness may serve as a marker not only to differentiate between healthy and affected individuals [52,53] but also to distinguish between different stages of the disease [49,56]. A limitation lies in the nonspecific nature of this finding, which can be addressed through a comprehensive patient evaluation. Thus, retinal thickness analysis should not be considered a standalone metric but one of several parameters to observe in a global patient approach.

4. GCL Changes in Parkinson's Disease

PD is a progressive neurodegenerative disorder characterized by degeneration of dopaminergic neurons in the substantia nigra, and abnormal protein aggregates (mainly alpha-synuclein) known as Lewy bodies [57,58]. The abnormal aggregates cause progressive neuronal loss and, consequently, decreased dopamine, resulting in a multitude of motor and non-motor symptoms [59,60]. The etiology of PD is not completely understood, and only a small percentage of defined monogenic mutations have been shown [61]. Therefore, most forms are idiopathic, and recognizing characteristic symptoms, even in the early stages, is crucial for the diagnosis. However, PD presents with a wide range of clinical features, including motor and non-motor symptoms, reflecting its heterogeneous nature [62]. While the classical motor triad of tremor, rigidity, and bradykinesia is often used for diagnosis, non-motor symptoms such as cognitive decline, sleep disturbances, autonomic dysfunction, and impaired olfaction frequently precede motor symptoms by years. The variability in symptom presentation and progression has led to subclassifications, including tremor-dominant and non-tremor-dominant PD, each with distinct clinical features and treatment responses. This diversity underscores the complexity of diagnosing and managing PD [59,62]. Given the multiplicity of possible clinical presentations, misdiagnoses often occur, leading to delays in the therapy and management of these patients.

Documented retinal changes in PD patients were extensively studied through histopathological investigations

on both human and animal models. These studies suggested that retinal damage and subsequent layer atrophy primarily arose from deposits of α -synuclein (α S) [63] and depletion of levodopa [64]. The presence of α S was observed as abnormal deposits in the retina in numerous animal studies [65–67]. However, although there is evidence of the presence of α S and beta-synuclein in the IPL, this does not appear to be the only layer where these deposits can be found, and it is not clear whether the IPL represents the first layer in which these lesions appear; the exact location and progression over time of these deposits are still not completely defined [68]. α S is believed to influence neurotransmission among photoreceptors, amacrine cells, and bipolar cells [69]. Furthermore, the presence of intracytoplasmic inclusions of α S positively correlated with similar ubiquitin inclusions, allowing further parallels between the eye and the brain [70]. According to Archibald *et al.* [71], the impact of these α S deposits includes direct damage to retinal layers and, together with dopaminergic deficits, leads to significant remodeling of amacrine cell connectivity circuits and reduced trophic support at the retinal level. These effects collectively contribute to RGC dystrophy. Furthermore, histological studies on PD patients showed the presence of a phosphorylated form of α S, denominated p-syn, within the RNFL, GCL, and inner layers of the IPL [63,72–74]. Notable are intraneuronal globular Lewy bodies within the inner nuclear layer (INL), diffuse α S in the IPL, and both intra- and extracellular inclusions of α S in the GCL [73]. These inclusions within the GCL seemed to result from overexpression of the α S gene or increased endosome/exosome-mediated transport by the INL and IPL [74] (Fig. 2, Ref. [14]). The evidence of these accumulations within the RGCs, particularly related to p-syn, enabled to demonstrate a strong correlation between the density of these accumulations and the extent of cerebral pathology in PD patients, affecting both motor scores and disease stages [63]. This suggested a synchronized progression of pathology between the brain and retina, supporting the use of retinal imaging as a potential biomarker [63]. Among the various studies [5,12,13,16,17,32,58] undertaken on this biomarker, the primary focus was on elucidating the correlation between GCL and PD, establishing an association between these variables.

While a broader literature concerning other OCT-related PD biomarkers exists [60], the GCL received comparatively less attention, but its increasing potential future significance has made it one of the main biomarkers in the diagnostic and therapeutic evaluation of this neurodegenerative disorder. Numerous studies identified a global thinning of the RNFL and GCL in PD patients [75–80]. The thickness reduction at these levels appeared to correlate with disease severity and duration, providing a rationale for OCT as a follow-up method [81]. In a longitudinal study including 129 PD patients, Garcia-Martin *et al.* [51] demonstrated RNFL and GCL thinning to be progressive. This

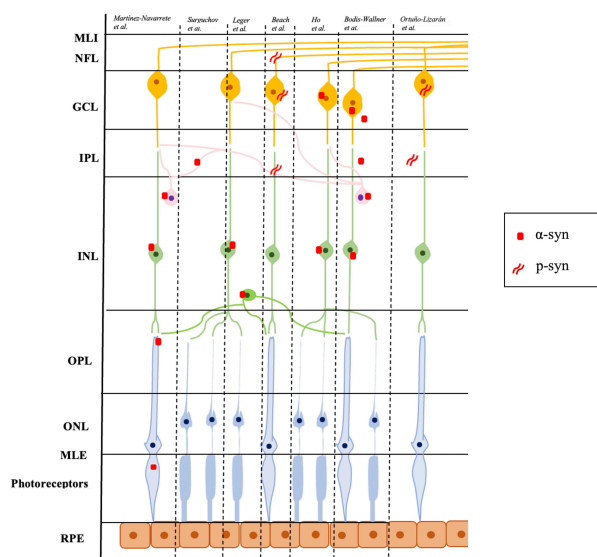


Fig. 2. Schematic representation of the main localization of α -syn and p-syn in retinal tissue. α -syn, α -synuclein; p-syn, phosphorylated α -synuclein; MLI, internal limiting membrane; NFL, nerve fiber layer; GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer; MLE, external limiting membrane; RPE, retinal pigment epithelium. From Di Pippo M, Fragiotta S, Di Staso F, Scuderi L, Abdolrahimzadeh S. The Role of Alpha-Synuclein Deposits in Parkinson's Disease: A Focus on the Human Retina. *International Journal of Molecular Sciences*. 2023; 24: 4391. [14].

study was among the first to suggest that the observed retinal thinning could be due to axonal loss resulting from PD lesions within the posterior visual pathways and the direct degeneration of RGCs. Modestino *et al.* [82] and Bayhan *et al.* [83] used the Hoehn-Yahr scale (the scoring system used to evaluate the extent of clinical disability), the Unified Parkinson's Disease Rating Scale (UPDRS) total, and motor score as indicators of disease severity. These authors examined the correlation between average GCC thickness and disease severity and introduced novel parameters such as the ganglion cell volume (GLV) measuring the average volume loss across the entire GCC and focal loss volume (FLV) detecting focal losses adjusting for absolute changes similar to corrected pattern standard deviation [83]. This enabled the precise measurement of GCC volume loss, drawing parallels to traditional visual field assessments. These findings revealed significant differences in the GLV and FLV between PD patients, particularly in evaluating the eye ipsilateral to the body's most affected side and control subjects [83]. This asymmetry supported the theory of uneven neurodegeneration in PD, impacting both the substantia nigra and the eye [83]. Notably, this study also highlighted that GCC thinning was more pronounced in the eye opposite to the body's most affected side, suggesting a neurodegenerative spread pattern. Despite the small sample size,

which might limit general conclusions, these observations underscored the potential of OCT in the diagnostic workup, evaluating the efficacy of new treatments, and monitoring disease progression [83].

Murueta-Goyena *et al.* [84] elucidated that diminished parafoveal thickness, particularly within the 1–3 mm ring of the GC-IPL, was directly correlated with a higher risk of cognitive decline progression over a 3-year period. However, the authors highlighted that this correlation was likely attributable to the reliance of a significant portion of cognitive assessments on visual input, which may consequently be compromised. Indeed, the authors did not report a relationship between GC-IPL thinning and a decline in UPDRS motor score, suggesting divergent underlying mechanisms for these respective impairments [84].

Sari *et al.* [85], in a longitudinal analysis demonstrated that disease duration correlated significantly with a progressive reduction in thickness in the GC-IPL that was more evident in the inferior and inferotemporal sectors. These findings suggested an evolving nature of the neurodegeneration process and its reflection in a retinal setting, providing the rationale for OCT implementation in the management of PD. Other studies highlighted the importance of the inferior sector in correlation with disease severity. Živković *et al.* [86], besides identifying a clear statistical relevance in the association between PD and thinning of the GC-IPL in all sectors, reported that the areas most associated with disease duration were the inferotemporal and inferior sector of the GC-IPL, and those most associated with disease severity were the inferior and inferonasal sectors. Similarly, as previously observed, Sari *et al.* [85] highlighted the stronger correlation between disease duration and inferior GC-IPL thinning. A key study on the relationship between retinal atrophy and disease severity was reported in the meta-analysis by Ahn *et al.* [87], in which the authors reported that thinning of inferior GCL correlated with the Hoehn-Yahr disease severity scale. This study was focused on *de novo*, drug-naive PD patients and showed that inner retinal layer thinning, particularly in the inferior sector of the GCL, strongly correlated with Hoehn and Yahr staging of PD, dopamine transporter (DAT) loss in the substantia nigra, and microperimetry damage [87]. According to these authors, the inferior localization of this thinning was likely due to the loss being focal rather than diffuse in the early stages [87]. Lee *et al.* [88] demonstrated significant thinning across the GCC, with marked variations between different sectors. The authors showed that the parafoveal region exhibited the most substantial thinning, correlating with reduced visual acuity. The inferior and inferotemporal sectors of the GCC were particularly affected, suggesting a differential vulnerability among RGCs based on their location. Furthermore, the extent of thinning was linked to both the severity and duration of PD, highlighting the potential of sector-specific GCC thinning as a valuable biomarker in disease progression [88].

However, these retinal changes suffer from limited specificity, as similar findings do indeed appear in other neurodegenerative conditions [54,89,90]. There have been a few studies in which no direct correlation between the thickness of the GCL in PD was found [91,92]. The significance of the correlation between GCL/GCL-IPL/GCC and PD has emerged relatively recently, but it is increasingly gaining attention due to the potential applications of this parameter in disease management. It holds promise for facilitating novel assessments of ocular pathogenesis [93] in this disease and providing new insights into the visual mechanisms underlying cognitive impairment [88] and their association with disease severity and duration. Particularly noteworthy is the necessity to establish a parallelism between retinal-brain damage in preclinical PD patients to conduct more extensive studies investigating correlations with disease duration, severity, and cognitive decline [94]. Further research is warranted to validate OCT as a diagnostic tool in PD; as imaging technology advances, retinal imaging could potentially be included in the comprehensive assessment of PD and increase our understanding of this neurodegenerative disorder.

5. GCL Changes in Multiple Sclerosis

MS is a chronic neurodegenerative demyelinating condition of the central nervous system, causing a plethora of symptoms varying from cognitive and motor impairment to visual alterations. It typically manifests between the ages of 20 and 30 and is more prevalent in females, affecting nearly two females for every male [95]. Diagnosis is confirmed via the McDonald criteria, which integrate clinical, imaging, and laboratory findings but are only applicable in the advanced stages of the disease [96].

One frequent acute manifestation of MS is optic neuritis (ON), which can affect the entire visual pathway from the retina to the cortex [97–99]. Acute ON is the initial symptom in about 20% of MS patients, and symptomatic ON attacks occur in 30–70% of patients throughout the disease course [97–99]. Axonal loss in the optic nerve and retina can even be detected in patients without a history of ON, indicating the sensitivity of these structures to MS pathology [98]. Acute inflammation in ON can cause axonal damage at the retrobulbar level, possibly spreading to RGCs [100]. Histopathological and imaging studies showed that when ON is present, retrograde axonal degeneration of the RGCs can determine residual optic nerve head pallor and thinning of the RNFL [101,102]. This degeneration results in pronounced GCL, IPL, and RNFL thinning. Özbilen *et al.* [103] found consistent GCL thinning across all macular sectors, significantly more pronounced in MS patients with ON. This was confirmed in a systematic review and meta-analysis by Britze *et al.* [104]. These authors suggested that this thinning may result from retrograde transsynaptic degeneration of RGCs and their axons, triggered by MS lesions in the posterior visual pathways or by subclinical episodes of ON [104].

Despite the absence of previous ON episodes, GCL thickness evaluation is reported to be more sensitive in detecting damage with respect to peripapillary RNFL analysis, particularly in patients with relapsing-remitting MS (RRMS). Unlike peripapillary RNFL analysis, which may underestimate damage when measured soon after an acute ON episode due to swelling, macular GCL thickness measurements remained unaffected by optic nerve edema, offering more accurate assessments of structural changes early in the disease course [103].

Relapse episodes and inflammation in MS can result in extensive neurodegeneration. Neuronal and axonal damage are common outcomes, with damage occurring through mechanisms such as acute axonal injury, Wallerian degeneration (anterograde), or retrograde degeneration [105]. These factors underscore the importance of monitoring structural changes in the retina, particularly the GCL, to understand and track the progression of MS [103]. Furthermore, RNFL thickness is not as well correlated with visual dysfunction, while macular GCL-IPL thickness correlates with visual acuity, low-contrast letter acuity, visual field mean deviation, vision-related quality of life, and disability [104]. This makes GCL-IPL thinning a valuable marker of disease and visual function in MS patients, surpassing traditional RNFL measurements in predictive reliability and utility [106].

Research by Lee *et al.* [107] demonstrated that baseline and six-month follow-up measurements of RNFL and GCL-IPL thickness can effectively predict the likelihood of progression in disability based on Expanded Disability Status Scale (EDSS)-plus score at one-year follow-up. The EDSS-plus system is a tool designed to evaluate the functional systems of the central nervous system, using a score from 0 to 10, where the lowest represents a normal status and the highest death from MS [108]. Therefore, understanding and monitoring GCL-IPL thickness through OCT may serve as a critical prognostic tool for assessing the progression of MS. However, further longitudinal studies are required to provide a deeper and more precise understanding of GCL reliability and validity as a biomarker in monitoring disease progression over time.

The reduction in GCL thickness observed in MS appears to have a well-defined cause, primarily linked to episodes of ON, whether clinically evident or subclinical and thus harder to detect. While this explanation holds true, it is equally valid that changes in the GCL have been observed in patients without documented episodes of ON [104]. This has been particularly noted in patients with RRMS, where the thinning seems to be associated with the pro-inflammatory characteristic of this disease, leading to axonal injury and degeneration, both anterograde (Wallerian) and retrograde [107]. Furthermore, the extent of thinning appears to correlate with differences in disease progression [107].

Table 1. Key optical coherence tomography findings examining the ganglion cell layer in AD, PD, and MS, detailing findings, sample sizes, methods, significant correlations with disease markers, and limitations.

Study	Disease focus	Main findings on GCL	Sample size	OCT model used	Significant correlations	Limitations
Kao <i>et al.</i> , 2023 [18]	AD	Thinning of GC-IPL particularly vulnerable to AD	156	Spectralis OCT (Heidelberg Engineering, Heidelberg, Germany)	Thinning correlated with MMSE scores	Limited longitudinal data
Wisely <i>et al.</i> , 2022 [52]	AD	Developed a CNN using OCT to detect AD with high accuracy	284	Zeiss Cirrus HD-5000 Spectral-Domain OCT with AngioPlex OCT Angiography (Carl Zeiss Meditec, Dublin, CA, USA)	GC-IPL maps most useful; AUC 0.836 with full data model	Cross-sectional study
Santangelo <i>et al.</i> , 2020 [53]	AD	GCL thinning specific for AD compared to MCI and controls	137	Spectralis OCT (Heidelberg Engineering, Heidelberg, Germany)	GCL thinning more pronounced in AD; aligned with CDR	Axial eye length not reported
Querques <i>et al.</i> , 2019 [109]	AD	Thinning of the GCL	56	Spectralis high-resolution imaging (HRA) + OCT device (Heidelberg Engineering, Heidelberg, Germany)	Thinning is mainly confined to the central and temporal regions	Employment of a single time point for each patient
Garcia-Martin <i>et al.</i> , 2014 [51]	PD	Progressive thinning of RNFL and GCL	129	Spectralis OCT (Heidelberg Engineering, Heidelberg, Germany)	Thinning correlated with disease progression	Only early-stage PD; limited to a specific cohort
Murueta-Goyena <i>et al.</i> 2021 [84]	PD	Diminished parafoveal thickness linked with cognitive decline	89	OCT Angiography Module (Heidelberg Engineering, Germany)	Correlation between parafoveal GCL-IPL thinning and cognitive decline	Correlation with motor symptoms not reported
Sari <i>et al.</i> 2015 [85]	PD	Progressive reduction in GC-IPL thickness, especially in inferior and inferotemporal sectors	108	Cirrus high-definition (HD)-OCT Model 4000 (Carl Zeiss Meditec, Dublin, CA, USA)	GC-IPL thinning correlated with disease duration and severity	Focus on specific sectors
Živković <i>et al.</i> 2017 [86]	PD	Thinning of GC-IPL in all sectors, particularly in inferotemporal and inferior sectors	92	Cirrus spectral domain (SD)-OCT device 4000 (software version 6.0, Carl Zeiss Meditec, Dublin, CA, USA)	Strong correlation between GC-IPL thinning and disease severity and duration	Cross-sectional analysis
Knier <i>et al.</i> , 2017 [110]	MS	Low GC-IPL volumes	312	Not applicable	Association between low GC-IPL volumes and increased intrathecal B-cell frequencies and intrathecal IgG synthesis	Observational cohort study

AD, Alzheimer disease; GC-IPL, ganglion cell-inner plexiform layer; OCT, optical coherence tomography; MMSE, Mini Mental Status Examination; CNN, convolutional neural network; MCI, mild cognitive impairment; GCL, ganglion cell layer; CDR, Clinical Dementia Rating; PD, Parkinson's Disease; RNFL, retinal nerve fibers layer; MS, multiple sclerosis; AUC, area under the curve; IgG, immunoglobulin G.

6. Retinal Changes Detected by OCT in AD, PD, and MS

Detailed findings on GCL and GC-IPL thinning in AD, PD, and MS, emphasizing the diagnostic value of OCT in neurodegenerative diseases, are shown in Table 1 (Ref. [18,51–53,84–86,109,110]). Studies underscore the specificity of GCL thinning in distinguishing between AD and other conditions like MCI and controls. This specificity is supported by findings such as those reported by Santangelo *et al.* [53], who demonstrated a strong alignment between GCL thinning and CDR stages, underscoring its association with disease severity. Complementing these results, Kao *et al.* [18] found significant correlations between GCL thinning and MMSE scores, although their findings were limited by cross-sectional analysis. Further studies, such as those by Querques *et al.* [109], localized GCL thinning predominantly to the central and temporal macular regions, emphasizing the structural specificity of these changes. Together, these findings point to GCL thinning as a promising biomarker for both early detection and disease progression monitoring in AD [18,52,53,109]. GCL thinning in PD shows a progressive pattern, with studies identifying strong correlations with motor and cognitive impairments. Research by Garcia-Martin *et al.* [51] and Živković *et al.* [86] highlights the inferotemporal and inferior sectors as key regions of thinning, correlating closely with disease duration and severity. Similarly, Murueta-Goyena *et al.* [84] linked parafoveal GCL thinning to cognitive decline, reflecting PD's dual motor and cognitive dysfunctions. The consistent evidence of focal thinning in PD further supports the relevance of GCL as a biomarker for disease monitoring [51,84–86]. In MS, GCL thinning has been strongly associated with markers of neuroinflammation and disease progression. Knier *et al.* [110] demonstrated correlations between GCL thinning, intrathecal immunoglobulin G (IgG) synthesis, and disability scores, emphasizing the link between retinal structural changes and central nervous system inflammation.

These studies demonstrate the potential of OCT in capturing disease-specific patterns of GCL thinning across neurodegenerative conditions [18,53,84,109,110], and consistent correlations with disease severity, cognitive impairment, and functional outcomes emphasize its relevance in the diagnostic work-up and monitoring of disease.

7. Additional OCT Parameters in Neurodegenerative Research

OCT technology advancements have significantly expanded its capacity to assess additional parameters beyond retinal layer thickness, including volume, cell density, and structural integrity. These parameters offer a more comprehensive understanding of neuronal health, enhancing the utility of OCT in the assessment of neurodegenerative diseases. For example, GCL volume measurements provide a three-dimensional evaluation that captures neuronal loss

more holistically than thickness alone. Studies such as that by Al-Hawasi *et al.* [111] demonstrated that GCL volume changes can detect subtle retinal alterations associated with neurodegeneration, offering potential as early biomarkers, particularly in AD. Their findings highlight how volumetric metrics enhance the sensitivity of OCT in capturing retinal changes that might otherwise go unnoticed in traditional two-dimensional thickness assessments.

Furthermore, combining OCT with adaptive optics (AO) has enabled the visualization and quantification of individual retinal cells, offering indirect measures of cell density [112]. This technology allows researchers to gain novel insight into neuronal loss at a cellular level, particularly in neurodegenerative diseases such as AD. For instance, Soltanian-Zadeh *et al.* [113] utilized AO-OCT to segment and quantify ganglion cell density, demonstrating its potential as a diagnostic tool in assessing neuronal damage.

In addition to volume and density, OCT has proven useful in assessing the structural integrity of retinal layers by detecting microstructural abnormalities in retinal architecture. This application is particularly relevant in MS, where disruptions in retinal layer structure correlate with disease activity and progression. Saidha *et al.* [106] demonstrated that OCT can identify subtle microstructural damage within the GCL and other layers, highlighting its ability to act as a biomarker for neurodegeneration. Furthermore, Sharma *et al.* [114] reviewed the broader applications of OCT in neurodegenerative diseases, emphasizing its role in assessing trans-synaptic degeneration and visual pathway abnormalities.

These advancements underscore the potential of OCT to move beyond simple thickness measurements, enabling a more comprehensive analysis of retinal health. Incorporating metrics such as GCL volume, cell density, and structural integrity into routine OCT assessments may significantly enhance diagnostic accuracy and disease monitoring in neurodegenerative conditions [106,111–114]. These developments expand the scope of OCT in clinical and research settings and provide a promising foundation for more precise and personalized approaches to understanding and managing neurodegeneration.

8. Conclusions

This review highlights the potential of the GCL as a biomarker in AD, PD, and MS. Data in the literature show that OCT imaging provides evidence of the relationships between GCL thinning and disease progression. This suggests the value of implementing OCT imaging in research and in the clinical setting owing to the advantages of performing a rapid, non-invasive, and cost-effective examination to monitor disease activity. However, the use of GCL as a standalone diagnostic tool is currently limited by variability in measurement techniques owing to the use of different OCT devices and systemic and ocular factors

that affect the specificity and sensitivity of this biomarker. Additionally, the limitations of the existing studies in the literature, such as small sample sizes and the predominantly cross-sectional nature of research, do not provide information regarding GCL changes over time. In order to enhance OCT technology and improve diagnostic accuracy, future research should prioritize the standardization of imaging protocols across devices and explore the development of higher-resolution OCT platforms. Emerging innovations, such as AO and deep learning algorithms, could improve both the sensitivity and precision of GCL measurements. Advanced image analysis techniques, such as automated segmentation and three-dimensional mapping of retinal structures, may further refine diagnostic workflows. Integrating GCL measurements with molecular biomarkers, such as neurofilament light chain, A β , and tau proteins, could provide a more comprehensive understanding of neurodegenerative processes. These biomarkers, in combination with OCT findings, could improve the diagnostic and follow-up process by correlating structural changes in the retina with biochemical markers of neurodegeneration. Additionally, OCT could complement neurological imaging, such as PET and high-field MRI, possibly linking retinal changes to central nervous system degeneration. Future studies should focus on validating these integrative methods through large-scale, longitudinal research, ultimately enhancing patient outcomes in neurodegenerative diseases.

Author Contributions

FR, MDP and SA designed the research study. FR, DF and LB performed the literature research. MDP and SA provided help and advice on reviewing and editing the original manuscript. FR, DF and LB analyzed and synthesized the literature. All authors wrote the manuscript and contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

We would like to express our gratitude to the editor and the peer reviewers for their opinions and suggestions.

Funding

This research received no external funding.

Conflict of Interest

The authors declare no conflict of interest.

References

[1] Jones-Odeh E, Hammond CJ. How strong is the relationship between glaucoma, the retinal nerve fibre layer, and neurodegen-

erative diseases such as Alzheimer's disease and multiple sclerosis? *Eye*. 2015; 29: 1270–1284. <https://doi.org/10.1038/eye.2015.158>.

- [2] Trebbastoni A, D'Antonio F, Bruscolini A, Marcelli M, Cerecero M, Campanelli A, *et al.* Retinal nerve fibre layer thickness changes in Alzheimer's disease: Results from a 12-month prospective case series. *Neuroscience Letters*. 2016; 629: 165–170. <https://doi.org/10.1016/j.neulet.2016.07.006>.
- [3] Alves JN, Westner BU, Højlund A, Weil RS, Dalal SS. Structural and functional changes in the retina in Parkinson's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2023; 94: 448–456. <https://doi.org/10.1136/jnnp-2022-329342>.
- [4] London A, Benhar I, Schwartz M. The retina as a window to the brain—from eye research to CNS disorders. *Nature Reviews. Neurology*. 2013; 9: 44–53. <https://doi.org/10.1038/nrneuro.2012.227>.
- [5] Srinivasan S, Efron N. Optical coherence tomography in the investigation of systemic neurologic disease. *Clinical & Experimental Optometry*. 2019; 102: 309–319. <https://doi.org/10.1111/coxo.12858>.
- [6] Kromer R, Serbecic N, Hausner L, Froelich L, Aboul-Enein F, Beutelspacher SC. Detection of Retinal Nerve Fiber Layer Defects in Alzheimer's Disease Using SD-OCT. *Frontiers in Psychiatry*. 2014; 5: 22. <https://doi.org/10.3389/fpsy.2014.00022>.
- [7] Lu Y, Li Z, Zhang X, Ming B, Jia J, Wang R, *et al.* Retinal nerve fiber layer structure abnormalities in early Alzheimer's disease: evidence in optical coherence tomography. *Neuroscience Letters*. 2010; 480: 69–72. <https://doi.org/10.1016/j.neulet.2010.06.006>.
- [8] Sepulcre J, Murie-Fernandez M, Salinas-Alaman A, García-Layana A, Bejarano B, Villoslada P. Diagnostic accuracy of retinal abnormalities in predicting disease activity in MS. *Neurology*. 2007; 68: 1488–1494. <https://doi.org/10.1212/01.wnl.0000260612.51849.ed>.
- [9] Wong TY, Klein R, Sharrett AR, Nieto FJ, Boland LL, Couper DJ, *et al.* Retinal microvascular abnormalities and cognitive impairment in middle-aged persons: the Atherosclerosis Risk in Communities Study. *Stroke*. 2002; 33: 1487–1492. <https://doi.org/10.1161/01.str.0000016789.56668.43>.
- [10] Kashani AH, Chen CL, Gahm JK, Zheng F, Richter GM, Rosenfeld PJ, *et al.* Optical coherence tomography angiography: A comprehensive review of current methods and clinical applications. *Progress in Retinal and Eye Research*. 2017; 60: 66–100. <https://doi.org/10.1016/j.preteyeres.2017.07.002>.
- [11] Mei X, Qiu C, Zhou Q, Chen Z, Chen Y, Xu Z, *et al.* Changes in retinal multilayer thickness and vascular network of patients with Alzheimer's disease. *Biomedical Engineering Online*. 2021; 20: 97. <https://doi.org/10.1186/s12938-021-00931-2>.
- [12] Spaide RF, Koizumi H, Pozzoni MC. Enhanced depth imaging spectral-domain optical coherence tomography. *American Journal of Ophthalmology*. 2008; 146: 496–500. <https://doi.org/10.1016/j.ajo.2008.05.032>.
- [13] Spaide RF, Fujimoto JG, Waheed NK, Sadda SR, Staurenghi G. Optical coherence tomography angiography. *Progress in Retinal and Eye Research*. 2018; 64: 1–55. <https://doi.org/10.1016/j.preteyeres.2017.11.003>.
- [14] Di Pippo M, Fragiotta S, Di Staso F, Scuderi L, Abdollahimzadeh S. The Role of Alpha-Synuclein Deposits in Parkinson's Disease: A Focus on the Human Retina. *International Journal of Molecular Sciences*. 2023; 24: 4391. <https://doi.org/10.3390/ijms24054391>.
- [15] Cameron JR, Megaw RD, Tatham AJ, McGrory S, MacGillivray TJ, Doubal FN, *et al.* Lateral thinking - Interocular symmetry and asymmetry in neurovascular patterning, in health and disease. *Progress in Retinal and Eye Research*. 2017; 59: 131–157. <https://doi.org/10.1016/j.preteyeres.2017.04.003>.

- [16] Pless ML. The eye as a window to the brain. *Annals of the Academy of Medicine, Singapore*. 2023; 52: 60–61. <https://doi.org/10.47102/annals-acadmedsg.202317>.
- [17] Wang F, Zhong W, Yang Q, Zhao W, Liu X, Rao B, *et al.* Distribution and synaptic organization of substance P-like immunoreactive neurons in the mouse retina. *Brain Structure & Function*. 2023; 228: 1703–1724. <https://doi.org/10.1007/s00429-023-02688-x>.
- [18] Kao CC, Hsieh HM, Chang YC, Chu HC, Yang YH, Sheu SJ. Optical Coherence Tomography Assessment of Macular Thickness in Alzheimer's Dementia with Different Neuropsychological Severities. *Journal of Personalized Medicine*. 2023; 13: 1118. <https://doi.org/10.3390/jpm13071118>.
- [19] Kim JI, Kang BH. Decreased retinal thickness in patients with Alzheimer's disease is correlated with disease severity. *PLoS ONE*. 2019; 14: e0224180. <https://doi.org/10.1371/journal.pone.0224180>.
- [20] Sánchez-Puebla L, de Hoz R, Salobar-García E, Arias-Vázquez A, González-Jiménez M, Ramírez AI, *et al.* Age-Related Retinal Layer Thickness Changes Measured by OCT in *APP^{NL-F/NL-F}* Mice: Implications for Alzheimer's Disease. *International Journal of Molecular Sciences*. 2024; 25: 8221. <https://doi.org/10.3390/ijms25158221>.
- [21] Manchanda R, Samanta R, Narayan ML, Kumar M, Tiwari A, Agarwal A, *et al.* Connecting the Dots: Exploring the Relationship between Optical Coherence Tomography and ^{99m}Tc-TRODAT-1 SPECT Parameters in Parkinson's Disease. *Annals of Indian Academy of Neurology*. 2024; 27: 188–195. https://doi.org/10.4103/aian.aian_31_24.
- [22] Kılıç OHT, Bayram ZN, Kiyat P, Karti O, Aral A, Munis ND, *et al.* Examination of optical coherence tomography findings in patients with pregabalin use disorder. *PeerJ*. 2024; 12: e18395. <https://doi.org/10.7717/peerj.18395>.
- [23] Allen A, Robbins CB, Joseph S, Hemesat A, Kundu A, Ma JP, *et al.* Angioarchitectural alterations in the retina and choroid in frontotemporal dementia. *PLoS ONE*. 2024; 19: e0312118. <https://doi.org/10.1371/journal.pone.0312118>.
- [24] Chen CL, Zhang A, Bojikian KD, Wen JC, Zhang Q, Xin C, *et al.* Peripapillary Retinal Nerve Fiber Layer Vascular Microcirculation in Glaucoma Using Optical Coherence Tomography-Based Microangiography. *Investigative Ophthalmology & Visual Science*. 2016; 57: OCT475–OCT485. <https://doi.org/10.1167/iovs.15-18909>.
- [25] Hirasawa K, Yamaguchi J, Nagano K, Kanno J, Kasahara M, Shoji N. Degree of loss in the tissue thickness, microvascular density, specific perimetry and standard perimetry in early glaucoma. *BMJ Open Ophthalmology*. 2023; 8: e001256. <https://doi.org/10.1136/bmjophth-2023-001256>.
- [26] El-Kattan MM, Esmat SM, Esmail EH, Deraz HA, Ismail RS. Optical coherence tomography in patients with Parkinson's disease. *The Egyptian Journal of Neurology, Psychiatry and Neurosurgery*. 2022; 58: 21. <https://doi.org/10.1186/s41983-021-00421-1>.
- [27] López-de-Eguileta A, Cerveró A, Ruiz de Sabando A, Sánchez-Juan P, Casado A. Ganglion Cell Layer Thinning in Alzheimer's Disease. *Medicina*. 2020; 56: 553. <https://doi.org/10.3390/medicina56100553>.
- [28] López-de-Eguileta A, Lage C, López-García S, Pozueta A, García-Martínez M, Kazimierczak M, *et al.* Ganglion cell layer thinning in prodromal Alzheimer's disease defined by amyloid PET. *Alzheimer's & Dementia*. 2019; 5: 570–578. <https://doi.org/10.1016/j.trci.2019.08.008>.
- [29] Chorostecki J, Seraji-Bozorgzad N, Shah A, Bao F, Bao G, George E, *et al.* Characterization of retinal architecture in Parkinson's disease. *Journal of the Neurological Sciences*. 2015; 355: 44–48. <https://doi.org/10.1016/j.jns.2015.05.007>.
- [30] Pilat A, McLean RJ, Proudlock FA, Maconachie GDE, Sheth V, Rajabally YA, *et al.* In Vivo Morphology of the Optic Nerve and Retina in Patients With Parkinson's Disease. *Investigative Ophthalmology & Visual Science*. 2016; 57: 4420–4427. <https://doi.org/10.1167/iovs.16-20020>.
- [31] Trost A, Lange S, Schroedl F, Bruckner D, Motloch KA, Bogner B, *et al.* Brain and Retinal Pericytes: Origin, Function and Role. *Frontiers in Cellular Neuroscience*. 2016; 10: 20. <https://doi.org/10.3389/fncel.2016.00020>.
- [32] Abdolrahimzadeh S, Felli L, Plateroti AM, Perdicchi A, Conestabile MT, Recupero SM. Spectral Domain Optical Coherence Tomography Evidence of Retinal Nerve Fiber Layer and Ganglion Cell Loss in Adult Patients with Neurofibromatosis Type 1. *Retina*. 2016; 36: 75–81. <https://doi.org/10.1097/IAE.0000000000000650>.
- [33] López-de-Eguileta A, López-García S, Lage C, Pozueta A, García-Martínez M, Kazimierczak M, *et al.* The retinal ganglion cell layer reflects neurodegenerative changes in cognitively unimpaired individuals. *Alzheimer's Research & Therapy*. 2022; 14: 57. <https://doi.org/10.1186/s13195-022-00998-6>.
- [34] Blanks JC, Torigoe Y, Hinton DR, Blanks RH. Retinal pathology in Alzheimer's disease. I. Ganglion cell loss in foveal/parafoveal retina. *Neurobiology of Aging*. 1996; 17: 377–384. [https://doi.org/10.1016/0197-4580\(96\)00010-3](https://doi.org/10.1016/0197-4580(96)00010-3).
- [35] Blanks JC, Hinton DR, Sadun AA, Miller CA. Retinal ganglion cell degeneration in Alzheimer's disease. *Brain Research*. 1989; 501: 364–372. [https://doi.org/10.1016/0006-8993\(89\)90653-7](https://doi.org/10.1016/0006-8993(89)90653-7).
- [36] Hart de Ruyter FJ, Morrema THJ, den Haan J, Netherlands Brain Bank, Twisk JWR, de Boer JF, *et al.* Phosphorylated tau in the retina correlates with tau pathology in the brain in Alzheimer's disease and primary tauopathies. *Acta Neuropathologica*. 2023; 145: 197–218. <https://doi.org/10.1007/s00401-022-02525-1>.
- [37] Davis MR, Robinson E, Koronyo Y, Salobar-García E, Rentsendorj A, Gaire BP, *et al.* Retinal ganglion cell vulnerability to pathogenic tau in Alzheimer's disease. *bioRxiv*. 2024. (preprint) <https://doi.org/10.1101/2024.09.17.613293>.
- [38] Yu DY, Cringle SJ, Balaratnasingam C, Morgan WH, Yu PK, Su EN. Retinal ganglion cells: Energetics, compartmentation, axonal transport, cytoskeletons and vulnerability. *Progress in Retinal and Eye Research*. 2013; 36: 217–246. <https://doi.org/10.1016/j.preteyeres.2013.07.001>.
- [39] Pini L, Pievani M, Bocchetta M, Altomare D, Bosco P, Cavado E, *et al.* Brain atrophy in Alzheimer's Disease and aging. *Ageing Research Reviews*. 2016; 30: 25–48. <https://doi.org/10.1016/j.arr.2016.01.002>.
- [40] Blennow K, de Leon MJ, Zetterberg H. Alzheimer's disease. *Lancet*. 2006; 368: 387–403. [https://doi.org/10.1016/S0140-6736\(06\)69113-7](https://doi.org/10.1016/S0140-6736(06)69113-7).
- [41] Jack CR, Jr, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, *et al.* NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimer's & Dementia*. 2018; 14: 535–562. <https://doi.org/10.1016/j.jalz.2018.02.018>.
- [42] Sabbagh MN, Lue LF, Fayard D, Shi J. Increasing Precision of Clinical Diagnosis of Alzheimer's Disease Using a Combined Algorithm Incorporating Clinical and Novel Biomarker Data. *Neurology and Therapy*. 2017; 6: 83–95. <https://doi.org/10.1007/s40120-017-0069-5>.
- [43] Morinaga A, Ono K, Ikeda T, Ikeda Y, Shima K, Noguchi-Shinohara M, *et al.* A comparison of the diagnostic sensitivity of MRI, CBF-SPECT, FDG-PET and cerebrospinal fluid biomarkers for detecting Alzheimer's disease in a memory clinic. *Dementia and Geriatric Cognitive Disorders*. 2010; 30: 285–292. <https://doi.org/10.1159/000320265>.
- [44] Salamone G, Di Lorenzo C, Mosti S, Lupo F, Cravello L, Palmer K, *et al.* Color discrimination performance in patients with

- Alzheimer's disease. *Dementia and Geriatric Cognitive Disorders*. 2009; 27: 501–507. <https://doi.org/10.1159/000218366>.
- [45] Cormack FK, Tovee M, Ballard C. Contrast sensitivity and visual acuity in patients with Alzheimer's disease. *International Journal of Geriatric Psychiatry*. 2000; 15: 614–620. [https://doi.org/10.1002/1099-1166\(200007\)15:7<614::aid-gps153>3.0.co;2-0](https://doi.org/10.1002/1099-1166(200007)15:7<614::aid-gps153>3.0.co;2-0).
- [46] Kavcic V, Vaughn W, Duffy CJ. Distinct visual motion processing impairments in aging and Alzheimer's disease. *Vision Research*. 2011; 51: 386–395. <https://doi.org/10.1016/j.visres.2010.12.004>.
- [47] Gilmore GC, Wenk HE, Naylor LA, Koss E. Motion perception and Alzheimer's disease. *Journal of Gerontology*. 1994; 49: P52–P57. <https://doi.org/10.1093/geronj/49.2.p52>.
- [48] Kirby E, Bandelow S, Hogervorst E. Visual impairment in Alzheimer's disease: a critical review. *Journal of Alzheimer's Disease*. 2010; 21: 15–34. <https://doi.org/10.3233/JAD-2010-080785>.
- [49] Iseri PK, Altınış O, Tokay T, Yüksel N. Relationship between cognitive impairment and retinal morphological and visual functional abnormalities in Alzheimer disease. *Journal of Neuro-Ophthalmology*. 2006; 26: 18–24. <https://doi.org/10.1097/01.wno.0000204645.56873.26>.
- [50] Ito Y, Sasaki M, Takahashi H, Nozaki S, Matsuguma S, Motomura K, *et al.* Quantitative Assessment of the Retina Using OCT and Associations with Cognitive Function. *Ophthalmology*. 2020; 127: 107–118. <https://doi.org/10.1016/j.ophtha.2019.05.021>.
- [51] García-Martin E, Larrosa JM, Polo V, Satue M, Marques ML, Alarcia R, *et al.* Distribution of retinal layer atrophy in patients with Parkinson disease and association with disease severity and duration. *American Journal of Ophthalmology*. 2014; 157: 470–478.e2. <https://doi.org/10.1016/j.ajo.2013.09.028>.
- [52] Wisely CE, Wang D, Henaio R, Grewal DS, Thompson AC, Robbins CB, *et al.* Convolutional neural network to identify symptomatic Alzheimer's disease using multimodal retinal imaging. *The British Journal of Ophthalmology*. 2022; 106: 388–395. <https://doi.org/10.1136/bjophthalmol-2020-317659>.
- [53] Santangelo R, Huang SC, Bernasconi MP, Falautano M, Comi G, Magnani G, *et al.* Neuro-Retina Might Reflect Alzheimer's Disease Stage. *Journal of Alzheimer's Disease*. 2020; 77: 1455–1468. <https://doi.org/10.3233/JAD-200043>.
- [54] Cipollini V, Abdolrahimzadeh S, Troili F, De Carolis A, Calafiore S, Scuderi L, *et al.* Neurocognitive Assessment and Retinal Thickness Alterations in Alzheimer Disease: Is There a Correlation? *Journal of Neuro-Ophthalmology*. 2020; 40: 370–377. <https://doi.org/10.1097/WNO.0000000000000831>.
- [55] Wang X, Jiao B, Liu H, Wang Y, Hao X, Zhu Y, *et al.* Machine learning based on Optical Coherence Tomography images as a diagnostic tool for Alzheimer's disease. *CNS Neuroscience & Therapeutics*. 2022; 28: 2206–2217. <https://doi.org/10.1111/cns.13963>.
- [56] Sáez ME, García-Sánchez A, de Rojas I, Alarcón-Martín E, Martínez J, Cano A, *et al.* Genome-wide association study and polygenic risk scores of retinal thickness across the cognitive continuum: data from the NORFACE cohort. *Alzheimer's Research & Therapy*. 2024; 16: 38. <https://doi.org/10.1186/s13195-024-01398-8>.
- [57] Lees AJ, Hardy J, Revesz T. Parkinson's disease. *Lancet*. 2009; 373: 2055–2066. [https://doi.org/10.1016/S0140-6736\(09\)60492-X](https://doi.org/10.1016/S0140-6736(09)60492-X).
- [58] Tolosa E, Garrido A, Scholz SW, Poewe W. Challenges in the diagnosis of Parkinson's disease. *The Lancet. Neurology*. 2021; 20: 385–397. [https://doi.org/10.1016/S1474-4422\(21\)00030-2](https://doi.org/10.1016/S1474-4422(21)00030-2).
- [59] Jankovic J. Parkinson's disease: clinical features and diagnosis. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2008; 79: 368–376. <https://doi.org/10.1136/jnnp.2007.131045>.
- [60] Chaudhuri KR, Odin P, Antonini A, Martínez-Martin P. Parkinson's disease: the non-motor issues. *Parkinsonism & Related Disorders*. 2011; 17: 717–723. <https://doi.org/10.1016/j.parkreldis.2011.02.018>.
- [61] Jia F, Fellner A, Kumar KR. Monogenic Parkinson's Disease: Genotype, Phenotype, Pathophysiology, and Genetic Testing. *Genes*. 2022; 13: 471. <https://doi.org/10.3390/genes13030471>.
- [62] Kouli A, Torsney KM, Kuan W-L. Parkinson's Disease: Etiology, Neuropathology, and Pathogenesis. In Stoker TB, Greenland JC (eds) *Parkinson's Disease: Pathogenesis and Clinical Aspects*. Codon Publications: Brisbane (AU). 2018.
- [63] Ortuño-Lizarán I, Beach TG, Serrano GE, Walker DG, Adler CH, Cuenca N. Phosphorylated α -synuclein in the retina is a biomarker of Parkinson's disease pathology severity. *Movement Disorders*. 2018; 33: 1315–1324. <https://doi.org/10.1002/mds.27392>.
- [64] Harnois C, Di Paolo T. Decreased dopamine in the retinas of patients with Parkinson's disease. *Investigative Ophthalmology & Visual Science*. 1990; 31: 2473–2475.
- [65] Veys L, Vandenabeele M, Ortuño-Lizarán I, Baekelandt V, Cuenca N, Moons L, *et al.* Retinal α -synuclein deposits in Parkinson's disease patients and animal models. *Acta Neuropathologica*. 2019; 137: 379–395. <https://doi.org/10.1007/s00401-018-01956-z>.
- [66] Eschbach J, Danzer KM. α -Synuclein in Parkinson's disease: pathogenic function and translation into animal models. *Neuro-Degenerative Diseases*. 2014; 14: 1–17. <https://doi.org/10.1159/000354615>.
- [67] Giráldez-Pérez R, Antolín-Vallespín M, Muñoz M, Sánchez-Capelo A. Models of α -synuclein aggregation in Parkinson's disease. *Acta Neuropathologica Communications*. 2014; 2: 176. <https://doi.org/10.1186/s40478-014-0176-9>.
- [68] Surguchov A, McMahan B, Masliah E, Surgucheva I. Synucleins in ocular tissues. *Journal of Neuroscience Research*. 2001; 65: 68–77. <https://doi.org/10.1002/jnr.1129>.
- [69] Martínez-Navarrete GC, Martín-Nieto J, Esteve-Rudd J, Angulo A, Cuenca N. Alpha synuclein gene expression profile in the retina of vertebrates. *Molecular Vision*. 2007; 13: 949–961.
- [70] Leger F, Fernagut PO, Canron MH, Léoni S, Vital C, Tison F, *et al.* Protein aggregation in the aging retina. *Journal of Neuro-pathology and Experimental Neurology*. 2011; 70: 63–68. <https://doi.org/10.1097/NEN.0b013e31820376cc>.
- [71] Archibald NK, Clarke MP, Mosimann UP, Burn DJ. The retina in Parkinson's disease. *Brain*. 2009; 132: 1128–1145. <https://doi.org/10.1093/brain/awp068>.
- [72] Beach TG, Carew J, Serrano G, Adler CH, Shill HA, Sue LI, *et al.* Phosphorylated α -synuclein-immunoreactive retinal neuronal elements in Parkinson's disease subjects. *Neuroscience Letters*. 2014; 571: 34–38. <https://doi.org/10.1016/j.neulet.2014.04.027>.
- [73] Peng X, Tehranian R, Dietrich P, Stefanis L, Perez RG. Alpha-synuclein activation of protein phosphatase 2A reduces tyrosine hydroxylase phosphorylation in dopaminergic cells. *Journal of Cell Science*. 2005; 118: 3523–3530. <https://doi.org/10.1242/jcs.02481>.
- [74] Bodis-Wollner I, Kozłowski PB, Glazman S, Miri S. α -synuclein in the inner retina in parkinson disease. *Annals of Neurology*. 2014; 75: 964–966. <https://doi.org/10.1002/ana.24182>.
- [75] Huang L, Wang C, Wang W, Wang Y, Zhang R. The specific pattern of retinal nerve fiber layer thinning in Parkinson's disease: a systematic review and meta-analysis. *Journal of Neurology*. 2021; 268: 4023–4032. <https://doi.org/10.1007/s00415-020-10094-0>.
- [76] Chang Z, Xie F, Li H, Yuan F, Zeng L, Shi L, *et al.* Retinal Nerve Fiber Layer Thickness and Associations With Cognitive Impair-

- ment in Parkinson's Disease. *Frontiers in Aging Neuroscience*. 2022; 14: 832768. <https://doi.org/10.3389/fnagi.2022.832768>.
- [77] Lee JY, Kim JM, Ahn J, Kim HJ, Jeon BS, Kim TW. Retinal nerve fiber layer thickness and visual hallucinations in Parkinson's Disease. *Movement Disorders*. 2014; 29: 61–67. <https://doi.org/10.1002/mds.25543>.
- [78] Yang ZJ, Wei J, Mao CJ, Zhang JR, Chen J, Ji XY, *et al*. Retinal nerve fiber layer thinning: a window into rapid eye movement sleep behavior disorders in Parkinson's disease. *Sleep & Breathing*. 2016; 20: 1285–1292. <https://doi.org/10.1007/s11325-016-1366-4>.
- [79] Suciú VI, Suciú CI, Nicoară SD, Perju-Dumbravă L. Circumpapillary Retinal Nerve Fiber Layer OCT Imaging in a Parkinson's Disease Cohort-A Multidisciplinary Approach in a Clinical Research Hospital. *Journal of Personalized Medicine*. 2022; 12: 80. <https://doi.org/10.3390/jpm12010080>.
- [80] Inzelberg R, Ramirez JA, Nisipeanu P, Ophir A. Retinal nerve fiber layer thinning in Parkinson disease. *Vision Research*. 2004; 44: 2793–2797. <https://doi.org/10.1016/j.visres.2004.06.009>.
- [81] Satue M, Obis J, Rodrigo MJ, Otin S, Fuertes MI, Vilades E, *et al*. Optical Coherence Tomography as a Biomarker for Diagnosis, Progression, and Prognosis of Neurodegenerative Diseases. *Journal of Ophthalmology*. 2016; 2016: 8503859. <https://doi.org/10.1155/2016/8503859>.
- [82] Modestino EJ, Reinhofer A, Blum K, Amenechi C, O'Toole P, Hoehn and Yahr staging of Parkinson's disease in relation to neuropsychological measures. *Frontiers in Bioscience (Landmark Edition)*. 2018; 23: 1370–1379. <https://doi.org/10.2741/4649>.
- [83] Bayhan HA, Aslan Bayhan S, Tanık N, Gürdal C. The association of spectral-domain optical coherence tomography determined ganglion cell complex parameters and disease severity in Parkinson's disease. *Current Eye Research*. 2014; 39: 1117–1122. <https://doi.org/10.3109/02713683.2014.894080>.
- [84] Murueta-Goyena A, Del Pino R, Galdós M, Arana B, Acera M, Carmona-Abellán M, *et al*. Retinal Thickness Predicts the Risk of Cognitive Decline in Parkinson Disease. *Annals of Neurology*. 2021; 89: 165–176. <https://doi.org/10.1002/ana.25944>.
- [85] Sari ES, Koc R, Yazıcı A, Sahin G, Ermiş SS. Ganglion cell-inner plexiform layer thickness in patients with Parkinson disease and association with disease severity and duration. *Journal of Neuro-Ophthalmology*. 2015; 35: 117–121. <https://doi.org/10.1097/WNO.0000000000000203>.
- [86] Živković M, Dayanir V, Stamenović J, Ljubisavljević S, Pražić A, Zlatanović M, *et al*. Retinal ganglion cell/inner plexiform layer thickness in patients with Parkinson's disease. *Folia Neuropathologica*. 2017; 55: 168–173. <https://doi.org/10.5114/fn.2017.68584>.
- [87] Ahn J, Lee JY, Kim TW, Yoon EJ, Oh S, Kim YK, *et al*. Retinal thinning associates with nigral dopaminergic loss in de novo Parkinson disease. *Neurology*. 2018; 91: e1003–e1012. <https://doi.org/10.1212/WNL.0000000000006157>.
- [88] Lee YW, Lim MN, Lee JY, Yoo YJ. Central retina thickness measured with spectral-domain optical coherence tomography in Parkinson disease: A meta-analysis. *Medicine*. 2023; 102: e35354. <https://doi.org/10.1097/MD.00000000000035354>.
- [89] Mwanza JC, Oakley JD, Budenz DL, Chang RT, Knight OJ, Feuer WJ. Macular ganglion cell-inner plexiform layer: automated detection and thickness reproducibility with spectral domain-optical coherence tomography in glaucoma. *Investigative Ophthalmology & Visual Science*. 2011; 52: 8323–8329. <https://doi.org/10.1167/iovs.11-7962>.
- [90] Hajee ME, March WF, Lazzaro DR, Wolintz AH, Shrier EM, Glazman S, *et al*. Inner retinal layer thinning in Parkinson disease. *Archives of Ophthalmology*. 2009; 127: 737–741. <https://doi.org/10.1001/archophthalmol.2009.106>.
- [91] Matlach J, Wagner M, Malzahn U, Schmidtmann I, Steigerwald F, Musacchio T, *et al*. Retinal changes in Parkinson's disease and glaucoma. *Parkinsonism & Related Disorders*. 2018; 56: 41–46. <https://doi.org/10.1016/j.parkreldis.2018.06.016>.
- [92] Kaur M, Saxena R, Singh D, Behari M, Sharma P, Menon V. Correlation Between Structural and Functional Retinal Changes in Parkinson Disease. *Journal of Neuro-Ophthalmology*. 2015; 35: 254–258. <https://doi.org/10.1097/WNO.0000000000000240>.
- [93] Rascunà C, Russo A, Terravecchia C, Castellino N, Avitabile T, Bonfiglio V, *et al*. Retinal Thickness and Microvascular Pattern in Early Parkinson's Disease. *Frontiers in Neurology*. 2020; 11: 533375. <https://doi.org/10.3389/fneur.2020.533375>.
- [94] Zhou WC, Tao JX, Li J. Optical coherence tomography measurements as potential imaging biomarkers for Parkinson's disease: A systematic review and meta-analysis. *European Journal of Neurology*. 2021; 28: 763–774. <https://doi.org/10.1111/ene.14613>.
- [95] Goldenberg MM. Multiple sclerosis review. P & T: a Peer-reviewed Journal for Formulary Management. 2012; 37: 175–184.
- [96] Thompson AJ, Banwell BL, Barkhof F, Carroll WM, Coetzee T, Comi G, *et al*. Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. *The Lancet. Neurology*. 2018; 17: 162–173. [https://doi.org/10.1016/S1474-4422\(17\)30470-2](https://doi.org/10.1016/S1474-4422(17)30470-2).
- [97] Katz B. The dyschromatopsia of optic neuritis: a descriptive analysis of data from the optic neuritis treatment trial. *Transactions of the American Ophthalmological Society*. 1995; 93: 685–708.
- [98] Castro SMC, Damasceno A, Damasceno BP, Vasconcellos JPD, Reis F, Iyeyasu JN, *et al*. Visual pathway abnormalities were found in most multiple sclerosis patients despite history of previous optic neuritis. *Arquivos De Neuro-Psiquiatria*. 2013; 71: 437–441. <https://doi.org/10.1590/0004-282X20130058>.
- [99] Shaygannejad V, Golabchi K, Dehghani A, Ashtari F, Haghighi S, Mirzendehtdel M, *et al*. Color blindness among multiple sclerosis patients in Isfahan. *Journal of Research in Medical Sciences*. 2012; 17: 254–257.
- [100] Garrett B, Dmytriw AA, Maxner C. Acute optic neuritis in multiple sclerosis. *CMAJ: Canadian Medical Association Journal*. 2016; 188: E199. <https://doi.org/10.1503/cmaj.150811>.
- [101] Green AJ, McQuaid S, Hauser SL, Allen IV, Lyness R. Ocular pathology in multiple sclerosis: retinal atrophy and inflammation irrespective of disease duration. *Brain*. 2010; 133: 1591–1601. <https://doi.org/10.1093/brain/awq080>.
- [102] Syc SB, Saidha S, Newsome SD, Ratchford JN, Levy M, Ford E, *et al*. Optical coherence tomography segmentation reveals ganglion cell layer pathology after optic neuritis. *Brain*. 2012; 135: 521–533. <https://doi.org/10.1093/brain/awr264>.
- [103] Özbilen KT, Gündüz T, Kartal SNÇ, Ceylan NA, Eraksoy M, Kürtüncü M. Detailed Evaluation of Macular Ganglion Cell Complex in Patients with Multiple Sclerosis. *Noro Psikiyatri Arşivi*. 2021; 58: 176–183. <https://doi.org/10.29399/npa.27531>.
- [104] Britze J, Pihl-Jensen G, Frederiksen JL. Retinal ganglion cell analysis in multiple sclerosis and optic neuritis: a systematic review and meta-analysis. *Journal of Neurology*. 2017; 264: 1837–1853. <https://doi.org/10.1007/s00415-017-8531-y>.
- [105] Singh S, Dallenga T, Winkler A, Roemer S, Maruschak B, Siebert H, *et al*. Relationship of acute axonal damage, Wallerian degeneration, and clinical disability in multiple sclerosis. *Journal of Neuroinflammation*. 2017; 14: 57. <https://doi.org/10.1186/s12974-017-0831-8>.
- [106] Saidha S, Syc SB, Durbin MK, Eckstein C, Oakley JD, Meyer SA, *et al*. Visual dysfunction in multiple sclerosis correlates better with optical coherence tomography derived estimates of macular ganglion cell layer thickness than peripapillary retinal nerve fiber layer thickness. *Multiple Sclerosis*. 2011; 17: 1449–1463.

<https://doi.org/10.1177/1352458511418630>.

- [107] Lee JY, Taghian K, Petratos S. Axonal degeneration in multiple sclerosis: can we predict and prevent permanent disability? *Acta Neuropathologica Communications*. 2014; 2: 97. <https://doi.org/10.1186/s40478-014-0097-7>.
- [108] Meyer-Moock S, Feng YS, Maeurer M, Dippel FW, Kohlmann T. Systematic literature review and validity evaluation of the Expanded Disability Status Scale (EDSS) and the Multiple Sclerosis Functional Composite (MSFC) in patients with multiple sclerosis. *BMC Neurology*. 2014; 14: 58. <https://doi.org/10.1186/1471-2377-14-58>.
- [109] Querques G, Borrelli E, Sacconi R, De Vitis L, Leocani L, Santangelo R, *et al.* Functional and morphological changes of the retinal vessels in Alzheimer's disease and mild cognitive impairment. *Scientific Reports*. 2019; 9: 63. <https://doi.org/10.1038/s41598-018-37271-6>.
- [110] Knier B, Leppenhetier G, Wetzlmair C, Aly L, Hoshi MM, Pernpeintner V, *et al.* Association of Retinal Architecture, Intrathecal Immunity, and Clinical Course in Multiple Sclerosis. *JAMA Neurology*. 2017; 74: 847–856. <https://doi.org/10.1001/jamaneurol.2017.0377>.
- [111] Al-Hawasi A, Lagali N. Retinal ganglion cell layer thickness and volume measured by OCT changes with age, sex, and axial length in a healthy population. *BMC Ophthalmology*. 2022; 22: 278. <https://doi.org/10.1186/s12886-022-02488-7>.
- [112] Liu Z, Saeedi O, Zhang F, Villanueva R, Asanad S, Agrawal A, *et al.* Quantification of Retinal Ganglion Cell Morphology in Human Glaucomatous Eyes. *Investigative Ophthalmology & Visual Science*. 2021; 62: 34. <https://doi.org/10.1167/iovs.62.3.34>.
- [113] Soltanian-Zadeh S, Kurokawa K, Liu Z, Zhang F, Saeedi O, Hammer DX, *et al.* Weakly supervised individual ganglion cell segmentation from adaptive optics OCT images for glaucomatous damage assessment. *Optica*. 2021; 8: 642–651. <https://doi.org/10.1364/optica.418274>.
- [114] Sharma S, Chitranshi N, Wall RV, Basavarajappa D, Gupta V, Mirzaei M, *et al.* Trans-synaptic degeneration in the visual pathway: Neural connectivity, pathophysiology, and clinical implications in neurodegenerative disorders. *Survey of Ophthalmology*. 2022; 67: 411–426. <https://doi.org/10.1016/j.survophthal.2021.06.001>.