

Review

# Applications of Natural Coumarins in Bone and Joint Diseases: Pharmacological Mechanisms and Advances in Clinical Research

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## Abstract

This review examines the applications of natural coumarins in the context of bone and joint disorders. It provides comprehensive coverage of their fundamental pharmacology, the epidemiology and pathological mechanisms of osteoarticular diseases, as well as the pharmacological actions, clinical applications, future prospects, and ongoing challenges associated with these compounds. Through systematic analysis of relevant studies, we summarize the basic characteristics of natural coumarins, including their chemical structures, metabolic pathways, and pharmacological activities. We also examine their mechanisms of action in conditions such as osteoarthritis, rheumatoid arthritis, and osteoporosis, with a particular emphasis on their anti-inflammatory and cartilage-repairing properties. Furthermore, we summarize current clinical trials, formulation development, delivery strategies, and safety evaluations. We also explore potential new indications, combination therapies, and personalized treatment approaches. Several controversies and ongoing challenges are discussed, including variability in dose-dependent efficacy, divergent therapeutic outcomes across diseases, and concerns regarding long-term safety. This review aims to provide a foundational reference for further research and clinical application of natural coumarins in the treatment of bone and joint disorders.

**Keywords:** natural coumarins; bone and joint disorders; pharmacological mechanism; clinical application; potential for medicinal use

## 1. Introduction

Coumarins are a class of lactones characterized by an  $\alpha$ -benzopyranone core that is biosynthesized from *cis*-*o*-hydroxycinnamic acid. First isolated by Friedrich Vogel in the eighteenth century, coumarin derivatives have attracted increasing attention since the 1960s owing to their low molecular weight, favorable bioavailability, and diverse pharmacological activities, notably anti-inflammatory and antineoplastic effects. By 1996, at least 1300 coumarin derivatives had been identified [1]. These secondary metabolites are widely distributed in plants—particularly within the Apiaceae, Rutaceae, Fabaceae and Asteraceae families—and to a lesser extent among microbial secondary metabolites [2,3]. Structurally, coumarins share a benzopyranone scaffold and are classified according to the nature and position of substituents into simple coumarins, furanocoumarins, pyranocoumarins, and other subclasses. Many coumarins display diverse biological activities, including anti-inflammatory [4], antiviral, antibacterial [5], anticoagulant, anticancer [6], anti-asthmatic [7], anti-osteoporotic [8] and neuroprotective effects [9]. Moreover, coumarins serve as privileged scaffolds in drug discovery, providing natural product templates for medicinal chemistry. This is exemplified by the anticoagulant warfarin, a coumarin derivative.

Osteoarticular disorders encompass a spectrum of pathological conditions of bone and joint arising from degeneration, inflammation, trauma, metabolic disturbances, infection or neoplasia. These conditions include rheumatoid arthritis (RA), osteoarthritis (OA), bursitis, synovitis, cervical and lumbar spondylopathies, adhesive capsulitis, osteophyte formation, avascular necrosis of the femoral head, and degenerative joint disease [10]. They are highly prevalent in middle-aged and older adults, and are characterized primarily by chronic pain and loss of function [11]. Progressive disease often leads to joint stiffness and deformity, resulting in substantial functional impairment and representing a major cause of disability in the elderly. The demographic shift towards an aging population has led to a rising incidence of osteoarticular disease, with associated increases in social and healthcare burdens [12].

Consequently, the identification of effective strategies for the prevention and treatment of osteoarticular disease remains a key priority in geriatric medicine. Traditional Chinese medicine (TCM) has a long history of treating joint disorders and has yielded numerous empirically effective formulas [13]. Contemporary pharmacological investigations indicate that natural coumarins in medicinal herbs contribute substantially to therapeutic effects by mediating anti-inflammatory actions, inhibition of ferroptosis, sup-



pression of pyroptosis, as well as other protective mechanisms [14]. Coumarins therefore hold considerable promise for the management of osteoarticular disease. Our research group has also identified several coumarin derivatives from traditional medicines that have drug development potential [15,16].

For this review, we performed a literature search using the keywords “coumarin”, “osteoarthritis”, “rheumatoid arthritis”, and “osteoporosis” across the Web of Science (<https://webofscience.clarivate.cn>) and China National Knowledge Infrastructure (<https://www.cnki.net>). Literature from the SCIE and Chinese Core Journal Directory within the past five years was included. Irrelevant topics (such as non-disease research), low-quality research, conference abstracts and preprints were excluded. Records retrieved from these databases were manually screened to remove duplicates and irrelevant works. Based on classical TCM texts and the primary literature, all studies meeting our inclusion criteria were analyzed and synthesized. This review summarizes current advances in the study of natural coumarins for the treatment of osteoarticular disease, with the aim of informing future efforts to exploit their therapeutic potential.

## 2. Literature Review

### 2.1 Fundamental Structural Characteristics and Pharmacological Activities of Natural Coumarins

The basic structural framework of coumarin compounds consists of a benzopyrone nucleus [17]. These compounds occur predominantly in dicotyledonous plants either as aglycones or glycosides [18], and are found in traditional Chinese medicinal herbs such as *Psoralea corylifolia*, *Peucedanum praeruptorum*, *Angelica pubescens*, *Angelica dahurica*, *Fraxinus rhynchophylla*, *Cnidium monnieri*, and *Artemisia capillaris* [19]. Chemical modifications to the core structure can significantly influence their pharmacological properties [20]. Based on their chemical structures, coumarins derived from medicinal plants can be categorized into four classes: simple coumarins, furanocoumarins, pyranocoumarins, and other structural variants.

Simple coumarins are defined by the presence of substituents solely on the benzene ring, without the formation of additional furan or pyran rings at the 6- or 8-position relative to the 7-hydroxy group. Oxygen-containing functional groups—such as hydroxyl or methoxy groups—are frequently observed at the C-7 position, although substitutions may also occur at other sites. Representative examples include umbelliferone from *Peucedanum praeruptorum*, and daphnetin from *Daphne odora*.

Complex coumarins arise through the annulation of heterocyclic rings onto the benzene moiety of simple coumarins, leading to the formation of furanocoumarins and pyranocoumarins. Furanocoumarins typically result

from cyclization of an isopentenyl group with an adjacent phenolic hydroxyl on the coumarin backbone. These are further subdivided into 6,7-furanocoumarins (linear type, e.g., psoralen from *Psoralea corylifolia*), and 7,8-furanocoumarins (angular type, e.g., angelicin).

Pyranocoumarins are characterized by a 2,2-dimethylpyran ring formed via cyclization between an isopentenyl group at C-6 or C-8 and a neighboring phenolic hydroxyl. These are also classified into linear (6,7-) and angular (7,8-) types, exemplified by xanthyletin from *Zanthoxylum simulans* and seselin from *Seseli indicum*, as well as decursin and decursinol from *Peucedanum praeruptorum*.

Beyond these three primary categories, natural coumarins also exhibit additional structural diversity. For example, dicoumarols represent dimers of coumarin units, which may involve linkages between simple coumarins or between pyranocoumarins in linear-linear or linear-angular configurations. Isocoumarins are isomers of coumarins and occur predominantly in plants as dihydroisocoumarin derivatives. Furthermore, certain coumarins carry substituents at the C-3 or C-4 position of the  $\alpha$ -pyrone ring, commonly including phenyl, hydroxyl, or isopentenyl groups. Table 1 (Ref. [21–65]) summarizes common coumarin compounds found in TCM, together with their botanical sources.

Natural coumarins exhibit a broad spectrum of pharmacological activities, including anti-inflammatory, antioxidant, antitumor, antibacterial, and antiviral effects [66]. In terms of anti-inflammatory activity, numerous coumarins act by suppressing key inflammatory signaling pathways. For instance, umbelliferone and methyl galate have been shown to significantly inhibit the secretion of nitric oxide (NO) and prostaglandin E2 (PGE2) in lipopolysaccharide (LPS)-stimulated macrophages, concurrently downregulating the expression of inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2), thereby exerting anti-inflammatory effects [67]. Common mechanisms underlying these anti-inflammatory actions are summarized in Table 2 (Ref. [21,27,44,49,50,66,68,69]) and Fig. 1.

The antioxidant properties of coumarins are primarily mediated through free radical scavenging and modulation of the antioxidant enzyme system. Notably, 7-hydroxycoumarin demonstrates considerable antioxidant capacity by effectively neutralizing DPPH (1,1-diphenyl-2-picrylhydrazyl) radicals. This activity is closely associated with the presence of hydroxyl groups in its structure [70]. Furthermore, coumarins have diverse biological activities, such as antioxidant and anticoagulant properties. For example, coumarin and several of its derivatives (e.g., esculetin, scoparone, and 4-methylumbelliferone) were evaluated for their impact on carbon tetrachloride-induced hyperlipidemia in rats. Structural variations among these compounds were found to confer distinct modulatory ef-

**Table 1. Common coumarin compounds in traditional Chinese medicine.**

No.	Type	English name	Molecular formula	Source (Botanical origin)	Ref.	
1	Simple Coumarin	Umbelliferone	C <sub>9</sub> H <sub>9</sub> O <sub>3</sub>	<i>Peucedanum praeruptorum</i>	[21]	
2		Daphnetin	C <sub>9</sub> H <sub>6</sub> O <sub>4</sub>	<i>Daphne genkwa</i>	[22,23]	
3		Scoparone	C <sub>11</sub> H <sub>10</sub> O <sub>4</sub>	<i>Artemisia scoparia</i>	[24]	
4		Osthole	C <sub>15</sub> H <sub>16</sub> O <sub>3</sub>	<i>Cnidium monnieri</i>	[25–27]	
5		Fraxetin	C <sub>10</sub> H <sub>8</sub> O <sub>5</sub>	<i>Fraxinus rhynchophylla</i>	[28]	
6		Esculetin	C <sub>9</sub> H <sub>6</sub> O <sub>4</sub>	<i>Fraxinus rhynchophylla</i>	[29]	
7		Scopoletin	C <sub>10</sub> H <sub>8</sub> O <sub>4</sub>	<i>Angelica dahurica</i>	[30]	
8	Pyranocoumarin	Xanthoxyletin	C <sub>15</sub> H <sub>14</sub> O <sub>4</sub>	<i>Zanthoxylum americanum</i>	[31]	
9		Xanthyletin	C <sub>14</sub> H <sub>12</sub> O <sub>3</sub>	<i>Zanthoxylum bungeanum</i>	[32]	
10		Luvangetin	C <sub>15</sub> H <sub>14</sub> O <sub>4</sub>	<i>Toddalia asiatica</i>	[33]	
11		Decursin	C <sub>19</sub> H <sub>20</sub> O <sub>5</sub>	<i>Angelica decursiva</i>	[34]	
12		Decursinol	C <sub>14</sub> H <sub>14</sub> O <sub>4</sub>	<i>Angelica decursiva</i>	[35]	
13		Seselin	C <sub>14</sub> H <sub>12</sub> O <sub>3</sub>	<i>Seseli indicum</i>	[36]	
14		5-Hydroxyseselin	C <sub>14</sub> H <sub>12</sub> O <sub>4</sub>	<i>Seseli indicum</i>	[37]	
15		5-Methoxyseselin	C <sub>15</sub> H <sub>14</sub> O <sub>4</sub>	<i>Seseli indicum</i>	[38]	
16		Praeruptorin A	C <sub>21</sub> H <sub>22</sub> O <sub>7</sub>	<i>Peucedanum praeruptorum</i>	[39]	
17		Praeruptorin B	C <sub>24</sub> H <sub>26</sub> O <sub>7</sub>	<i>Peucedanum praeruptorum</i>	[40]	
18		Praeruptorin C	C <sub>24</sub> H <sub>28</sub> O <sub>7</sub>	<i>Peucedanum praeruptorum</i>	[41]	
19		Furanocoumarin	Psoralen	C <sub>11</sub> H <sub>6</sub> O <sub>3</sub>	<i>Psoralea corylifolia</i>	[42–44]
20	Imperatorin		C <sub>16</sub> H <sub>14</sub> O <sub>4</sub>	<i>Angelica dahurica</i>	[45]	
21	Isoimperatorin		C <sub>16</sub> H <sub>14</sub> O <sub>4</sub>	<i>Angelica dahurica</i>	[46]	
22	Xanthotoxin		C <sub>12</sub> H <sub>8</sub> O <sub>4</sub>	<i>Angelica pubescens</i>	[47]	
23	Bergapten		C <sub>12</sub> H <sub>8</sub> O <sub>4</sub>	<i>Citrus medica</i>	[48,49]	
24	Phellopterin		C <sub>17</sub> H <sub>16</sub> O <sub>5</sub>	<i>Angelica dahurica</i>	[50]	
25	Byakangelicol		C <sub>17</sub> H <sub>16</sub> O <sub>6</sub>	<i>Angelica dahurica</i>	[51]	
26	Cnidicin		C <sub>21</sub> H <sub>22</sub> O <sub>5</sub>	<i>Angelica sinensis</i>	[52]	
27	Oxypeucedanin		C <sub>16</sub> H <sub>14</sub> O <sub>5</sub>	<i>Peucedanum praeruptorum</i>	[53]	
28	Isopsoralen		C <sub>11</sub> H <sub>6</sub> O <sub>3</sub>	<i>Psoralea corylifolia</i>	[54]	
29	Isobergapten		C <sub>12</sub> H <sub>8</sub> O <sub>4</sub>	<i>Notopterygium incisum</i>	[55]	
30	Pimpinellin		C <sub>13</sub> H <sub>10</sub> O <sub>5</sub>	<i>Heracleum hemsleyanum</i>	[56]	
31	Angenomalin		C <sub>14</sub> H <sub>12</sub> O <sub>3</sub>	<i>Angelica dahurica</i> cv.	[57]	
32	Columbianadin		C <sub>19</sub> H <sub>20</sub> O <sub>5</sub>	<i>Angelica sinensis</i>	[58]	
33	Other Coumarins		Euphorbetin	C <sub>18</sub> H <sub>10</sub> O <sub>8</sub>	<i>Euphorbia lathyris</i>	[59]
34			Daphnoretin	C <sub>19</sub> H <sub>12</sub> O <sub>7</sub>	<i>Stellera chamaejasme</i>	[60]
35			Dicoumarol	C <sub>19</sub> H <sub>12</sub> O <sub>6</sub>	<i>Medicago sativa</i>	[61]
36		Capillarin	C <sub>13</sub> H <sub>10</sub> O <sub>2</sub>	<i>Artemisia capillaris</i>	[62]	
37		Agrimolide	C <sub>18</sub> H <sub>18</sub> O <sub>5</sub>	<i>Agrimonia pilosa</i>	[63]	
38		Plumbapin	C <sub>11</sub> H <sub>8</sub> O <sub>3</sub>	<i>Plumbago zeylanica</i>	[64]	
39		Pachyrrhizin	C <sub>19</sub> H <sub>12</sub> O <sub>6</sub>	<i>Pachyrrhizus erosus</i>	[65]	

fects on the serum lipid profile, underscoring the importance of chemical structure in determining biological activity [71].

With regard to antitumor applications, certain coumarin derivatives can induce apoptosis, suppress proliferation, and inhibit the metastasis of tumor cells. Specifically, some natural coumarins isolated from plants have been reported to modulate the expression of cyclins and apoptosis-related proteins, thereby promoting cancer cell apoptosis and impairing migration and invasive capacities [72]. A series of furo [3,2-c] coumarin derivatives were synthesized and assessed for their anti-proliferative

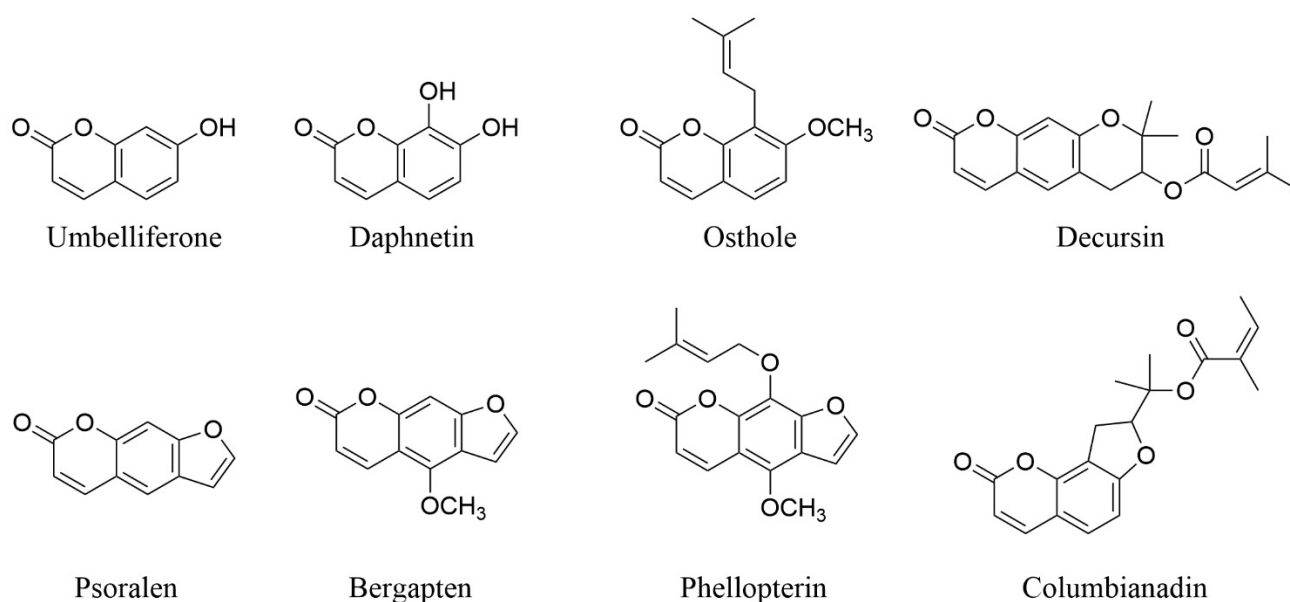
activity against MCF-7 breast cancer and HCT-15 colon cancer cell lines. Among these, compounds 1b and 1d exhibited potent activity, and UV-Vis spectroscopic studies demonstrated binding affinity toward DNA and bovine serum albumin (BSA) [73].

Natural coumarins also demonstrate notable antibacterial and antiviral activities through mechanisms such as the disruption of bacterial cell membranes and inhibition of viral replication, highlighting potential applications in anti-infective therapeutics [74]. Several coumarin derivatives that inhibit a range of microorganisms have been identified. For example, novel Cu(II) complexes synthesized

**Table 2. Anti-inflammatory effect and mechanism of coumarins.**

Chemical compound	Test subject	Mechanism of action	Reference
Decursin	Mouse chondrocytes	Inhibits IL-1 $\beta$ -mediated abnormal activation of the PI3K/AKT/NF- $\kappa$ B axis	[66]
Phellopterin	Human immortalized keratinocytes (HaCaT)	Upregulates SIRT1 and downregulates ICAM-1	[50]
Osthole	Human keratinocyte cell line (HaCaT)	Modulates secretion of IL-1, TNF- $\alpha$ , CCL2/MCP-1, and CCL5/RANTES; inhibits TLR2 and COX-2 expression	[27]
Umbelliferone	Arthritic rat model	Mediates MAPK signaling via the NF- $\kappa$ B pathway	[21]
Daphnetin	Inflammatory pain mouse model	Negatively regulates the spinal NF- $\kappa$ B pathway and preferentially activates the Nrf2/HO-1 signaling pathway	[68]
Bergapten	Mouse monocyte-macrophage leukemia cells (RAW264.7)	Inhibits JAK/STAT activation and ROS production	[49]
Columbianadin	Human peripheral blood mononuclear cells (hPBMCs)	Suppresses LPS-induced production of inflammatory cytokines and downregulates the NOD1/NF- $\kappa$ B pathway	[69]
Psoralen	Mouse monocyte-macrophage leukemia cells (RAW264.7)	Inhibits NF- $\kappa$ B activation and MAPKs phosphorylation	[44]

ROS, reactive oxygen species.

**Fig. 1. Representative coumarins with anti-inflammatory effects.**

with 6-acetyl-7-hydroxy-4-methylcoumarin (HL1) and 8-acetyl-7-hydroxy-4-methylcoumarin (HL2) show enhanced antibacterial and cytotoxic activities compared to their parent ligands. Notably, the complex Cu(HL2)<sub>2</sub>·0.5H<sub>2</sub>O exhibited antifungal efficacy comparable to that of commercially available fluconazole [75].

## 2.2 Pharmacological Actions of Coumarins in Common Bone and Joint Disorders

### 2.2.1 Pathological Mechanisms of Osteoarthritis and Therapeutic Effects of Coumarin Compounds

OA is a prevalent chronic and degenerative joint disorder [10]. Medical expenses related to OA account for 1–2.5% of gross domestic product (GDP) in high-income countries of Europe and North America [76,77], with joint

replacement procedures constituting a major proportion of these costs. As global demographic aging continues to increase, OA is projected to become one of the most common diseases worldwide [78]. The pathophysiological process of OA involves alterations in multiple joint tissues, including articular cartilage, synovium, and subchondral bone. Inflammation plays a central role in the initiation and progression of OA. The release of various cytokines such as interleukin-1 $\beta$  (IL-1 $\beta$ ) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) activates a cascade of signaling pathways in chondrocytes, leading to degradation of the cartilage extracellular matrix (ECM). For instance, IL-1 $\beta$  stimulates chondrocytes to express matrix metalloproteinases (MMPs), including MMP-1, MMP-3, and MMP-13. These degrade type

II collagen and proteoglycans in the cartilage, thereby disrupting its structural integrity [79]. Concurrently, increased chondrocyte apoptosis represents another key pathological feature of OA. Studies of cartilage tissues from OA patients have demonstrated upregulation of apoptosis-related proteins, such as members of the caspase family. This leads to a reduction in the number of chondrocytes and impairs normal cartilage metabolism and repair mechanisms [80].

Isofraxidin has been found to inhibit IL-1 $\beta$ -induced inflammatory responses in human OA chondrocytes by suppressing the production of NO and prostaglandin E<sub>2</sub> (PGE<sub>2</sub>). This downregulates the expression of inflammatory proteins such as COX-2, iNOS, MMP-1, MMP-3, and MMP-13, and simultaneously increases the levels of aggrecan and type II collagen. Such anti-inflammatory activity is mediated through suppression of the NF- $\kappa$ B signaling pathway [81]. Given the limited self-repair capacity of articular cartilage following injury, natural coumarins exhibit beneficial effects in promoting cartilage regeneration. Coumestrol counteracts the catabolic effects induced by IL-1 $\beta$  in primary rat chondrocytes and articular cartilage by inhibiting proteoglycan loss and the expression of matrix-degrading enzymes. It also reduces the production of inflammatory cytokines, thereby protecting and facilitating the repair of cartilage [82]. A hyaluronic acid-triethylene glycol-coumarin-based hydrogel was shown to promote early-stage cartilage repair in a minipig model. This hydrogel exhibited excellent biocompatibility and injectability, allowed *in situ* photo-crosslinking, and provided a favorable microenvironment for cartilage regeneration. It significantly improved the macroscopic and histological scores of the repaired tissue and enhanced chondrogenic parameters, including defect structure, repair tissue surface, and subchondral bone restoration [83]. These findings suggest that natural coumarins and their formulations may be effective strategies for facilitating the repair of articular cartilage. Furthermore, bergapten was shown to alleviate OA by activating the ANP32A/ATM signaling pathway. This subsequently inhibits the expression of IL-1 $\beta$ -induced inflammatory cytokines and mediators, maintains the chondrocyte phenotype, and promotes secretion of cartilage-specific ECM components, thereby attenuating the progression of OA [84]. Similarly, osthole exerted chondroprotective and anti-inflammatory effects in a monosodium iodoacetate (MIA)-induced mouse model of OA by downregulating COX-2 and RUNX2 expression and inhibiting the activation of NF- $\kappa$ B and HIF-2 $\alpha$  signaling pathways, thereby reducing expression of MMP-13, syndecan IV, and ADAMTS-5 [85]. Collectively, these studies indicate that natural coumarins can mitigate OA through multiple anti-inflammatory and cartilage-protective mechanisms, thus offering novel insights for OA treatment.

## 2.2.2 Pathological Mechanisms of Rheumatoid Arthritis and Therapeutic Effects of Coumarin Compounds

RA is a chronic and progressive systemic autoimmune disorder characterized primarily by inflammation of the synovial membrane within joints. The global prevalence of RA ranges from 0.18% to 1.07%, while the reported prevalence in mainland China is 0.42%, corresponding to an affected total population of approximately 5 million individuals [86]. The pathogenesis of RA involves complex interactions among genetic, environmental, and immunological factors [87]. Genetic susceptibility plays a critical role, particularly alleles of the human leukocyte antigen (*HLA*) genes, such as specific variants of *HLA-DRB1* (e.g., shared epitope). These alleles influence antigen presentation and T-cell activation, thereby increasing disease risk. RA shows familial aggregation, with first-degree relatives of RA patients having a three-fold higher risk of developing the disease. Moreover, the concordance rate among monozygotic twins ranges from 12–15%, underscoring the contribution of genetic factors. Immunological dysregulation is central to RA pathology. Abnormal activation of CD4<sup>+</sup> T helper cells leads to the secretion of proinflammatory cytokines, including TNF- $\alpha$  and interleukin-1 (IL-1), which promote synovitis and joint tissue damage. CD8<sup>+</sup> T cells may also contribute to inflammatory processes. Concurrently, B-cell hyperactivity results in the production of autoantibodies such as rheumatoid factor (RF) and anti-cyclic citrullinated peptide (anti-CCP) antibodies. These autoantibodies form immune complexes that activate the complement system, exacerbating inflammatory injury. Furthermore, a dysregulated cytokine network—characterized by excessive production of TNF- $\alpha$ , IL-1, and IL-6—creates a positive feedback loop that perpetuates and amplifies the inflammatory response. Prolonged inflammation leads to synovial hyperplasia, formation of pannus tissue, and infiltration of immune cells and inflammatory mediators into the joint space. These processes contribute to the destruction of articular cartilage and subchondral bone. Over time, persistent joint damage, along with muscle atrophy and spasm around affected joints, results in deformities and functional impairment. In summary, RA arises from the interplay of genetic predisposition, environmental triggers (e.g., infections, smoking), and immune dysfunction. Early diagnosis and comprehensive treatment are essential to control disease activity and prevent structural joint damage [88].

Coumarin derivatives exert multifaceted effects in the pathological process of RA. For instance, 8-methoxychromen-2-one (MCO), isolated from *Ruta graveolens*, significantly reduces the arthritis index and clinical scores in a rat model of collagen-induced arthritis (CIA), while also mitigating joint damage and reducing plasma levels of TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and nitric oxide [89]. This therapeutic effect was associated with the suppression of cytokine production and NF- $\kappa$ B activation in LPS-stimulated J774 cells. Furthermore, certain coumarins

modulate osteoclast differentiation and function, thereby influencing RA progression. Coumarin compounds have been shown to inhibit the expression of osteoclast-specific genes (e.g., TRAP, CTSK, and MMP-9) as well as the activation of key transcription factors (e.g., NFATc1 and *c-Fos*) and signaling pathways including NF- $\kappa$ B and MAPKs. Consequently, these actions attenuate bone resorption and alleviate bone destruction in RA [90]. Multiple studies have also demonstrated that natural coumarins can effectively reduce the expression of relevant inflammatory mediators. For instance, esculetin, a natural coumarin derivative, reduces plasma levels of leukotriene B4 (LTB4) in an adjuvant-induced arthritis model in rats. LTB4 is one of the most potent chemotactic factors promoting neutrophil migration into the joints. Elevated serum levels of LTB4 in RA patients correlate with disease severity, suggesting that esculetin may be a potential therapeutic agent for RA [91]. Similarly, 6,7-dihydroxy-3-[3',4'-methylenedioxyphenyl]-coumarin (3-PD-5), both in its free and liposomal forms, inhibits immune complex-stimulated elastase release and ROS generation in neutrophils obtained from healthy donors, as well as from RA patients in remission (i-RA) or with active disease (a-RA). Furthermore, 3-PD-5 suppresses the release of neutrophil extracellular traps (NETs) and impairs neutrophil chemotaxis, demonstrating its regulatory effects on immune cell activation and function [92]. On the other hand, certain coumarins influence cytokine secretion pathways. Psoralidin, for example, inhibits LPS-induced expression of inflammatory cytokines such as TNF- $\alpha$  and IL-6, and modulates the RANKL/OPG ratio, thereby attenuating both inflammatory responses and bone resorption [93]. Some coumarins also exert immunomodulatory effects by regulating key immune signaling pathways. Notopterol, for instance, ameliorates RA pathology by inhibiting the JAK/STAT signaling pathway, thereby reducing the production of inflammatory cytokines and chemokines. These findings suggest that coumarins may regulate the immune system through multiple mechanisms, highlighting their important role in immunomodulation during RA progression [94].

With regard to apoptosis, the combination of daphnetin with Bcl2-targeted small interfering RNA (si-Bcl2) was shown to modulate the expression of anti-apoptotic genes in fibroblast-like synoviocytes (FLS) derived from CIA rats. Both si-Bcl2 and daphnetin alone downregulated the mRNA and protein expression of Bcl2, and reduced the mRNA level of signal transducer and activator of transcription 3 (STAT3). Their combined application resulted in a stronger pro-apoptotic effect on RA FLS, providing a theoretical and experimental foundation for novel RA treatments based on daphnetin-based siRNA combination therapies [95]. Furthermore, 7-hydroxycoumarin (7-HC) was shown to inhibit proliferation and induce apoptosis of FLS in the CIA rat model by suppressing the Wnt/ $\beta$ -

catenin signaling pathway. This led to a reduction in the severity of CIA, as manifested by decreased paw swelling, lower arthritis indices, mitigated joint damage, and reduced production of IL-1 $\beta$ , IL-6, and TNF- $\alpha$  in both serum and synovial tissue [96].

Coumarins have also been shown to modulate osteoclast differentiation and function. For instance, cinnamoyloxy-mammeisin (CNM) inhibited macrophage colony-stimulating factor (M-CSF)- and receptor activator of nuclear factor kappa-B ligand (RANKL)-induced osteoclast differentiation, as well as the expression of osteoclast-specific markers, thereby reducing bone resorption and alleviating bone destruction in RA [97].

### 2.2.3 Pathological Mechanisms of Osteoporosis and Therapeutic Effects of Coumarin Compounds

Osteoporosis (OP) is a common bone disorder, particularly prevalent among postmenopausal women and the elderly [98]. The maintenance of bone homeostasis relies on a delicate balance between bone formation mediated by osteoblasts, and bone resorption facilitated by osteoclasts. The pathophysiology of OP is characterized by an imbalance between these processes, wherein bone resorption exceeds bone formation, leading to reduced bone mass and the deterioration of bone microarchitecture [99]. Osteoblasts promote the differentiation of osteoclast precursors into mature osteoclasts by secreting M-CSF and RANKL, thereby accelerating bone resorption [100]. Concurrently, osteoblasts also produce osteoprotegerin (OPG), which competitively binds to RANKL and inhibits osteoclast differentiation and maturation [101]. Consequently, the RANKL/OPG ratio serves as a critical indicator of bone resorption activity [102]. Aging as well as declining estrogen levels also contribute to a state of chronic, low-grade inflammation. Proinflammatory cytokines such as TNF- $\alpha$ , interleukins (ILs), and prostaglandin E2 upregulate the expression of M-CSF and RANKL, thereby exacerbating bone resorption and suppressing bone formation [103]. Additionally, the accumulation of reactive oxygen species (ROS) and diminished antioxidant capacity can induce osteoblast apoptosis, further reducing bone formation [104]. Disruptions in calcium and phosphorus metabolism, such as vitamin D deficiency, may also exacerbate bone loss through secondary hyperparathyroidism [105].

Natural coumarin compounds demonstrate considerable potential for the prevention and treatment of OP. For example, the XLGB-B fraction, isolated from the TCM formulation Xianlinggubao (XLGB) and known to contain multiple compounds including coumarins, effectively prevents bone loss in ovariectomized mice. XLGB-B improves bone quality, microstructure, and strength, while also modulating bone turnover markers [106]. Furthermore, *in vitro* studies have shown that osthole and imperatorin stimulate the proliferation of MCF-7 cells and upregulate the mRNA levels of estrogen receptor-regulated genes, including estro-

gen receptor- $\alpha$  ( $Er\alpha$ ), progesterone receptor ( $PR$ ), and pre-senilin 2 ( $PS2$ ). Additionally, they significantly enhance alkaline phosphatase ( $ALP$ ) activity in Saos-2 osteoblastic cells, suggesting these compounds may promote osteoblast activity via estrogen-like effects or estrogen receptor pathways, highlighting their potential therapeutic value in OP [107]. Moreover, coumarin constituents from *Psoralea corylifolia*, such as psoralen and isopsoralen, have been shown to modulate the functions of both osteoblasts and osteoclasts. They promote bone formation while inhibiting bone resorption, thereby ameliorating the pathological condition of OP [108]. Natural coumarins act on multiple signaling pathways, including  $NF-\kappa B$  and  $MAPK$ . For example, umbelliferone inhibits  $RANKL$ -induced osteoclast differentiation and bone resorption by suppressing the  $Akt-c-Fos-NFATc1$  signaling axis, thereby attenuating inflammatory bone loss [109].

### 2.3 Clinical Applications of Coumarin in Bone and Joint Diseases

Bone and joint diseases including OA, RA and OP represent major global public health challenges [110]. There is an urgent need to develop safe and effective therapeutics for these conditions. Herbal formulas containing multiple ingredients are commonly prescribed to patients in TCM practice. These formulas often include herbs rich in coumarin.

#### 2.3.1 Clinical Applications of Coumarin-Rich Traditional Chinese Medicinal Formulas

Coumarin-containing TCM formulas have long been employed in clinical practice, and contemporary investigations have provided experimental and clinical support for the therapeutic contributions of coumarins. For example, Duhuo Jisheng Tang (DJT), with Duhuo as the monarch herb, contains multiple coumarin constituents including dihydroxypeucedanin, angeloylcolumbianetin, and osthol derivatives. DJT is used clinically to dispel wind-damp, relieve arthralgia, and tonify the liver and kidneys—actions that correspond conceptually to the TCM pattern of “deficiency underlying excess” in osteoarticular disorders. A randomized controlled trial allocated patients with knee OA to hyaluronate, DJT, or combination therapy [111]. After 8 weeks, the combination group had significantly lower scores on the Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC), lower TCM syndrome scores, and lower post-treatment serum  $TNF-\alpha$  and  $IL-1$  levels compared with the hyaluronate or DJT monotherapy groups ( $p < 0.05$ ). Moreover, the combination regimen was well tolerated and clinically effective. Another study demonstrated that DJT combined with methotrexate improved clinical symptom scores and reduced serum  $CRP$ ,  $ESR$ ,  $RF$ ,  $TNF-\alpha$ ,  $IL-6$ , and  $IL-1$  levels, thereby enhancing outcomes in patients with RA [112].

Jitongning Tablet (JTNT), a TCM formula for ankylosing spondylitis (AS), has progressed to a phase II clinical trial (NCT03932019). AS is an inflammatory spondyloarthropathy related to RA. UPLC-Q-TOF-MS profiling of JTNT metabolites in rat biofluids (plasma, urine and brain tissue), together with network pharmacology and molecular docking analyses, have implicated the regulation of inflammatory and immunerelated genes as key mechanisms of JTNT action. Moreover, coumarins were predicted to be among the principal active constituents. Such “constituent–metabolite–target” multidimensional investigations provide a mechanistic foundation for elucidating the role of coumarins within complex formulas. They also guide subsequent targeted isolation and development of active compounds [113].

Several proprietary Chinese medicines derived from traditional formulas owe part of their efficacy to coumarin-bearing herbs. Xianling Gubao capsule, developed by Professor Shi Guangda from an ethnomedicinal Miao prescription and produced using modern pharmaceutical techniques [114], contains Epimedium, *Psoralea* and *Dipsacus* among its principal ingredients. *Psoralea* is rich in psoralen and isopsoralen, while *Dipsacus* also contains coumarin derivatives. The Clinical Practice Guideline for Chinese Patent Medicines in the Treatment of Osteoporosis [115] indicates that Xianling Gubao combined with conventional anti-OP therapy (bisphosphonates) significantly improves lumbar and femoral neck bone mineral density. Additionally, it has clinical relevance for preventing and treating osteoporotic vertebral compression fractures and femoral neck fractures. In a comparative clinical study, Fu and Shi [116] evaluated oral calcium carbonate with vitamin D3 plus intravenous zoledronic acid versus the same regimen combined with Xianling Gubao. The combined treatment group demonstrated a superior overall response rate of 92.68% vs. 75.61%.

Collectively, these findings indicate that coumarins are not only active ingredients of traditional TCM, but also bioactive substances that can undergo modern scientific evaluation and inform precision clinical use and drug development. This body of evidence forms an integrated “clinic–mechanism–practice” loop and exemplifies a pragmatic pathway for the modernization of TCM.

#### 2.3.2 Combination Therapy and Personalized Treatment

Combination therapy is an important strategy to increase efficacy while reducing single-agent doses and adverse effects. Coumarins demonstrate substantial potential for synergistic interactions with conventional therapeutics.

Synergy with immunosuppressive approaches: studies have shown that daphnetin combined with Bcl-2-targeting small interfering RNA (siRNA) markedly enhanced the apoptosis of pathological synovial cells in CIA rats. This result suggests a novel “drug–gene” combinatorial paradigm for RA therapy, whereby coumarins paired

with nucleic acid-based agents can eliminate pathogenic cell populations more effectively [117].

**Synergy with natural products:** a synergistic effect of imperatorin and  $\beta$ sitosterol in RA was predicted by integrative computational pharmacology. Subsequent *in vivo* experiments confirmed this combination resulted in significant amelioration of arthritic signs in CIA rats, whereas monotherapy with either compound was less effective. Mechanistic analyses indicated the synergy involved co-regulation of targets such as lymphotoxin alpha (*LTA*), Homo sapiens CD83 molecule (*CD83*) and sterol regulatory element binding transcription factor 1 (*SREBF1*), as well as modulation of multiple canonical signaling pathways [118]. As multifunctional, multitargeted constituents within TCM, natural coumarins offer distinctive advantages for the treatment of osteoarticular diseases through network-level modulation. Elucidation of their mechanisms using modern analytical, pharmacological and formulation technologies, together with rigorous clinical evaluation, will facilitate their translation as safe and effective therapeutics that bridge traditional medicine and contemporary pharmacotherapy. This should both enhance the modernization of TCM and contribute Eastern perspectives to global precision medicine.

The therapeutic efficacy of natural coumarins varies across different bone and joint disorders. As previously discussed, multiple natural coumarins demonstrate beneficial effects in OA by inhibiting inflammatory responses and modulating chondrocyte metabolism. In contrast, although certain coumarin compounds can promote bone formation and suppress bone resorption in OP, their mechanisms and effectiveness may differ significantly depending on structural variations. Furthermore, research on natural coumarins remains relatively limited for other bone and joint diseases, such as RA and AS, and their efficacy in these conditions is not yet well-established. These variations in treatment outcomes may be attributed to the complex and diverse pathophysiological mechanisms underlying different disorders. Each disease involves distinct cell types, signaling pathways, and tissue injury mechanisms, leading to differences in the targets and effects of natural coumarins. Therefore, a deeper understanding of the mechanisms of action of natural coumarins in various bone and joint diseases, along with the optimization of treatment strategies for specific conditions, is essential for maximizing their therapeutic potential.

Personalized medicine is the future goal of clinical practice, with the application of natural coumarins in the management of bone and joint disorders holding considerable promise. Given the variations in genetic background, disease status, and lifestyle among individuals, patient responses to natural coumarin-based therapies may differ significantly. For example, genetic polymorphisms influence the drug metabolism and efficacy of coumarin-derived oral anticoagulants. Polymorphisms in Cytochrome P450 pro-

teins 2C9 (*CYP2C9*) and vitamin K epoxide reductase complex subunit 1 (*VKORC1*) are strongly associated with warfarin dosing requirements, such that patients carrying certain mutant alleles often require lower doses to avoid adverse effects such as bleeding [119]. Therefore, genetic testing and biomarker assessments can help to characterize a patient's genetic profile in clinical practice, enabling the design of individualized treatment regimens involving natural coumarins. This includes the selection of appropriate drug formulations, determination of optimal dosages, and adjustment of dosing frequency. Additionally, comorbidities and concomitant medications must also be considered in order to comprehensively evaluate the risk-benefit profile of treatment, thereby achieving precision therapy and improving both therapeutic outcomes and quality of life. Personalized treatment is increasingly recognized as essential in the context of RA, and natural coumarins may play a valuable role. Due to patient heterogeneity in RA pathogenesis, disease progression, and treatment response, personalized strategies aim to tailor therapies based on individual characteristics.

At the genetic level, polymorphisms in genes encoding drug-metabolizing enzymes, such as cytochrome P450 isoforms, may influence the efficacy and safety of coumarins by altering their pharmacokinetics. Genetic profiling can provide guidance for dose individualization and rational combination therapies. From a clinical perspective, markers of disease activity such as CRP and erythrocyte sedimentation rate (ESR), together with joint damage assessments, can inform coumarin dosing and formulation selection. Patients with mild disease may benefit from low-dose coumarins combined with gentle therapeutic regimens, whereas those with severe inflammation may require higher doses, or combination with more potent agents. Furthermore, the presence of comorbidities such as cardiovascular disease or diabetes should guide the choice of coumarin compounds or delivery systems to minimize adverse impacts on concurrent conditions. Through this integrated approach, personalized treatment with natural coumarins can be optimized to enhance therapeutic efficacy and improve patient quality of life. The dose-response relationship of natural coumarins remains a subject of ongoing debate. Various studies have demonstrated that different doses of coumarins can lead to distinct pharmacological outcomes. For instance, coumarin and its derivatives exerted differential effects on the serum lipid profile in experimental rat models. At a specific dosage, 4-methylumbelliferone significantly reduced triglyceride and very-low-density lipoprotein cholesterol levels in rats with carbon tetrachloride-induced hyperlipidemia, but failed to restore total cholesterol levels. In contrast, umbelliferone and scoparone were effective in preventing the reduction of high-density lipoprotein cholesterol in the same model of lipid dysregulation [71].

In the context of bone and joint disorders, the optimal therapeutic dosage for many coumarins has yet to be conclusively established. Some studies suggest that increasing the dose may enhance efficacy, but this could also increase the risk of adverse reactions. For example, excessively high doses of coumarin-based anticoagulants increase the likelihood of serious adverse events such as bleeding, while insufficient dosing may fail to achieve the desired anticoagulant effect. Therefore, defining the optimal dosage range for natural coumarins in various bone and joint diseases, where efficacy is balanced with safety, represents a significant challenge in current research. Further clinical studies and robust data are essential to resolve these controversies and establish evidence-based dosing guidelines.

### 2.3.3 Current Limitations and Future Directions in Natural Coumarin Research

The application of natural coumarins in RA faces several controversies and challenges. From a clinical perspective, only a limited number of high-quality trials have investigated coumarins in RA patients. Large-scale, multi-center validations of their efficacy and safety are still lacking. Although coumarin derivatives have shown promising therapeutic effects in animal models, interspecies differences complicate the extrapolation of these results to humans. Additionally, potential drug–drug interactions between coumarins and conventional RA treatments—such as disease-modifying antirheumatic drugs (DMARDs) and NSAIDs—are not yet fully understood, which may impact the safety and efficacy of combination therapies. Moreover, while some mechanistic studies have identified molecular targets and signaling pathways through which coumarins may act, the complex pathophysiology of RA raises questions about their precise mechanisms of action *in vivo* and the possibility of undiscovered side effects. Challenges also remain in the extraction, purification, and formulation of natural coumarins, including how to ensure batch-to-batch consistency, improve bioavailability, and achieve stable and reproducible product quality.

Future research on coumarins in RA should prioritize several key areas. First, well-designed clinical studies are essential. Large-scale, randomized, multi-center controlled trials are needed to rigorously evaluate the efficacy and safety of coumarins in RA patients and to define optimal treatment protocols across different patient subpopulations. Second, further investigation into the mechanisms of action of coumarins is warranted. Despite current insights into the targets and pathways of coumarins, the complexity of RA pathogenesis necessitates deeper exploration of how they interact within biological networks and with endogenous molecules. This should provide a more robust theoretical foundation for drug development. In terms of pharmaceutical innovation, continued efforts should be made to optimize coumarin structures through computer-aided drug design and synthetic modification, leading to the development

of derivatives with enhanced potency and reduced toxicity. The development of novel delivery systems, such as smart nanocarriers, should also be pursued to improve targeting and bioavailability. Finally, the integration of multi-omics technologies (e.g., genomics, proteomics, metabolomics) will help to elucidate the systemic effects and influencing factors of coumarin therapy *in vivo*, supporting the advancement of personalized treatment strategies.

## 2.4 Development of New Coumarin Drugs

### 2.4.1 Exploration of Potential New Indications for Natural Coumarins

The application of coumarins in RA research has a long history, and several coumarin-containing plants have been used in the treatment of RA and related conditions for many years. For instance, *Saussurea involucreata* has been used in traditional Uyghur, Mongolian, Kazakh, and Chinese medicine for its purported effects in promoting blood circulation, dispelling cold-dampness, and reducing inflammation. With advances in modern science and technology, research on coumarins has progressively expanded. Initial studies were primarily focused on the phytochemical analysis of coumarin-rich plants, leading to the identification of coumarin structures and their natural occurrence. Subsequent investigations delved into their biological activities, revealing that coumarins possess anti-inflammatory, antioxidant, and immunomodulatory properties, all of which are highly relevant to the pathological mechanisms of RA. This prompted further exploration of their therapeutic potential in RA.

The use of coumarins for the treatment of osteoarticular diseases is similar to that of other compounds. Although a variety of components play a role in the treatment of these diseases, their mechanisms and focus differ. For example, the flavonoids Epimedium and daidzein exert estrogen-like effects, promote osteogenesis, inhibit osteoclastogenesis, have antioxidant and anti-inflammatory properties, promote chondrocyte proliferation, and inhibit matrix degradation [120,121]. Alkaloid Sinomenin and Matrine strongly inhibit the inflammatory response and suppress chondrocyte apoptosis and osteoclast formation [122,123]. Polyphenolics such as Resveratrol and Curcumin have strong antioxidant and anti-inflammatory properties, regulate chondrocyte metabolism, and exert multi-target actions and cardiovascular protection [124,125]. Coumarin compounds such as Daphnetin and Bergapten can promote osteoblast differentiation and inhibit osteoclast activity, while also being anti-inflammatory via the regulation of NF- $\kappa$ B and other pathways [126,127].

Beyond their applications in OA and OP, natural coumarins exhibit potential for a range of new therapeutic indications. Due to their broad pharmacological spectrum including anti-inflammatory and antioxidant properties, they may offer therapeutic benefits for other bone and joint disorders associated with inflammation and oxidative

stress. For example, in RA, where inflammatory responses are central to disease progression, natural coumarins may alleviate symptoms by inhibiting key inflammatory signaling pathways, similar to their mechanism in OA [128].

Structural modification of natural coumarins is an important strategy for enhancing their therapeutic efficacy in the treatment of bone and joint disorders. Interactions with biological targets can be optimized by altering the coumarin scaffold, thereby improving pharmacological activity. For example, a series of 3-(4-aminophenyl)coumarin derivatives were synthesized and subjected to activity screening. Among these, compound 5e demonstrated the strongest inhibitory activity against the proliferation of fibroblast-like synoviocytes (FLS), along with suppression of RA-related cytokines including IL-1, IL-6, and TNF- $\alpha$ . Preliminary mechanistic studies indicate this compound inhibits the activation of both the NF- $\kappa$ B and MAPK signaling pathways. Its anti-inflammatory efficacy was further validated in a rat model of joint inflammation [129].

A coumarin compound isolated from *Glycine tabacina*, dolichosin A, was identified as a potential therapeutic agent for RA by integrating network pharmacology and experimental validation. Network pharmacology predictions suggest its involvement in the PI3K/AKT and MAPK signaling pathways. Experimental results confirmed that dolichosin A significantly inhibits IL-1 $\beta$ -induced inflammation in SW982 human synovial cells and suppresses RANKL-induced osteoclastogenesis. These findings provide novel targets and strategic directions for the structural optimization and improved efficacy of coumarin-based compounds [130].

Another active research area is the development of novel coumarin-based therapeutics, including structural modification of natural coumarins to enhance efficacy and reduce toxicity and side effects. Additionally, combination therapies involving coumarins and other drugs are being explored to leverage synergistic effects and improve treatment outcomes. The design of advanced coumarin delivery systems is also a focus, with efforts directed toward enhancing target specificity and bioavailability while minimizing systemic adverse effects. Examples include smart drug delivery platforms that respond to pathological features of RA, such as the inflammatory microenvironment or metabolic abnormalities, enabling precise and controlled drug release.

The development of advanced formulations of natural coumarins is critical for enhancing their bioavailability and therapeutic efficacy. Several innovative delivery systems have been investigated, including nanoparticle-based approaches. A nanoparticle formulation effectively increased the Bax/Bcl-2 expression ratio and induced apoptosis, suggesting it can substantially improve the therapeutic potential of natural coumarins [131].

Similarly, a micellar system based on poly(ethylene glycol)-block-poly( $\epsilon$ -caprolactone) (PCL-PEG) polymers was developed for the targeted delivery of dexamethasone

(Dex). These micelles were prepared via thin-film dispersion and self-assembly, then administered intravenously to rats with adjuvant-induced arthritis. At a low dose of 0.8 mg/kg, the micelles exhibited prolonged circulation time and preferential accumulation in inflamed joints. They effectively reduced joint swelling, bone erosion, and the expression of inflammatory cytokines, while having only moderate adverse effects on body weight, lymphocyte count, and blood glucose levels, along with weak activation of the complement system. This strategy offers a safe and effective approach for low-dose glucocorticoid therapy in inflammatory diseases, as well as being a promising model for the delivery of coumarin-based drugs [132].

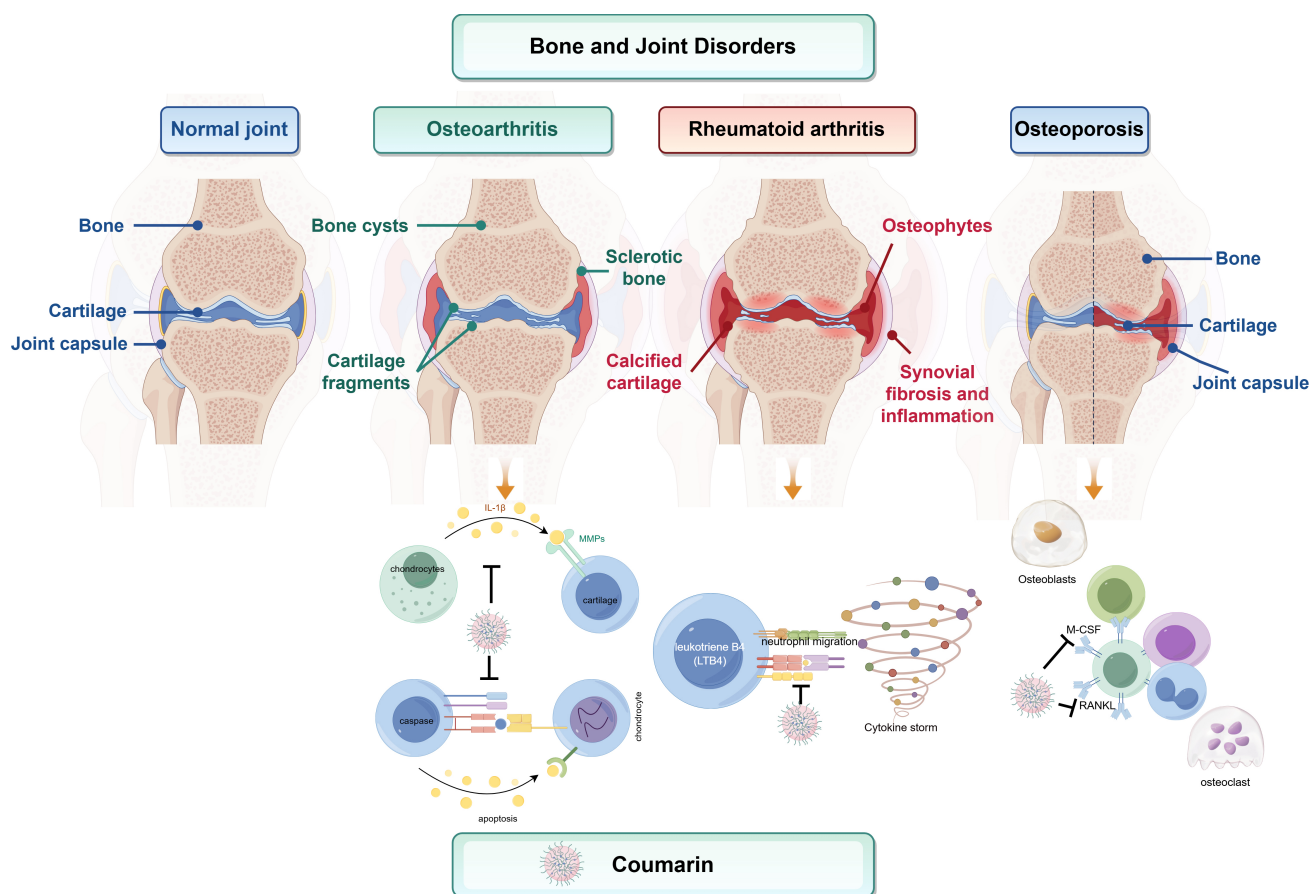
A metabolically bioresponsive hydrogel was also designed for the controlled release of psoralen in the treatment of RA. This enables on-demand release of psoralen and oxygen in response to inflammatory stimuli, thereby modulating homeostasis and metabolic disorders in the hypoxic arthritic microenvironment. Such pathology-responsive delivery systems represent a novel strategy for coumarin administration, with the potential to enhance treatment efficacy while minimizing systemic side effects [133].

#### 2.4.2 Metabolic Pathways and Bioavailability of Natural Coumarins

The administration route significantly influences the absorption and distribution of natural coumarins. For example, some orally administered coumarin compounds may be affected by gastric acid, digestive enzymes, and other gastrointestinal factors, resulting in reduced bioavailability. Further investigation of appropriate delivery routes and formulations—such as lipid-based carriers (e.g., liposomes), microspheres, or transdermal delivery systems—holds promise for improving their pharmacokinetic profile and therapeutic efficacy. However, factors such as drug-carrier interactions and formulation stability must be carefully evaluated to ensure safety and effectiveness.

After entering the systemic circulation, natural coumarins are primarily metabolized in the liver through the action of various enzymes, with the cytochrome P450 (CYP) superfamily playing a central role. For example, coumarin is metabolized in both rat and human liver microsomes, with CYP1A2 catalyzing its conversion to *o*-hydroxyphenylacetic acid, and CYP2A6 mediating 7-hydroxylation to form 7-hydroxycoumarin. Notably, human hepatocytes have an approximately 10-fold higher metabolic capacity for the 7-hydroxylation of coumarin compared to rat hepatocytes [134].

The bioavailability of coumarin compounds is influenced by multiple factors, with their chemical structure playing a critical role. Specific substituents can affect gastrointestinal absorption, plasma protein binding, and metabolic conversion. Physiological factors such as age, sex, genetic background and disease status also contribute to interindividual variability. For example, genetic poly-



**Fig. 2. Mechanism diagram of bone and joint diseases.** This schematic compares normal joint architecture with pathological features of three major bone and joint disorders: osteoarthritis (OA), rheumatoid arthritis (RA), and osteoporosis (OP). It further outlines how natural coumarins intervene in disease-specific mechanisms. In OA, coumarins inhibit IL-1 $\beta$ -driven inflammatory cascades and chondrocyte apoptosis, thereby preserving cartilage integrity. In RA, they suppress leukotriene B4 (LTB4)-mediated neutrophil recruitment, preventing subsequent cytokine amplification and joint damage. In OP, coumarins restore bone homeostasis by blunting osteoclast differentiation via downregulation of RANKL and M-CSF signaling, while supporting osteoblast function. Arrows denote activation or promotion; flat ends represent inhibition. IL-1 $\beta$ , Interleukin-1 $\beta$ ; M-CSF, Macrophage Colony Stimulating Factor; RANKL, Receptor Activator of Nuclear Factor- $\kappa$ B Ligand. Figure created with Figdraw (ID: RPPIU42477, <https://www.figdraw.com/static/index.html>).

morphisms in CYP enzymes are associated with differences in the rate of coumarin metabolism, thereby altering bioavailability. Drug–drug interactions must also be considered, as the co-administration of coumarins with agents that induce or inhibit CYP activity can modify metabolic pathways and rates, ultimately affecting the bioavailability of coumarin [135].

### 2.5 Safety and Evaluation of Adverse Effects of Natural Coumarins

The safety profile of natural coumarins is a critical consideration for their clinical application. Several studies have conducted preliminary safety assessments. For example, toxicogenomic and toxicological analyses of Soulatrolide and Mammaea (A/BA + A/BB) coumarins extracted from *Haploclathra paniculata* (Brazilian plant) found that short-term oral administration (100 mg/kg/day for one week) induced changes in hepatic gene expression, primar-

ily involving drug-metabolizing enzymes. However, no histopathological damage was observed in the liver, kidneys, or spleen, suggesting a favorable short-term safety profile [136].

*Saussurea involucrata*, a traditional herbal medicine containing coumarins among other constituents, is commonly used in the treatment of RA and other conditions. Although it demonstrates various beneficial biological activities, precise quality control and comprehensive toxicological studies are essential to ensure consistency and safety in clinical applications [137].

Studies on *Angelica pubescentis radix* (AP) have revealed that its coumarin components exhibit a wide range of pharmacological effects. However, due to the complexity and diversity of its chemical composition and mechanisms, standardized criteria for evaluating its quality and efficacy are still lacking. Further research is needed to elucidate the structure–activity relationships among its constituents, as

well as potential synergistic or antagonistic effects. Pharmacokinetic and toxicity studies in humans are also needed to ensure safe application [138].

*Ardisia gigantifolia* Stapf is used in traditional medicine for the management of RA, with clinical observations showing no significant toxic side effects. This provides supportive evidence, albeit preliminary, for the safety of coumarin-containing preparations in RA treatment. Nonetheless, detailed mechanistic toxicological studies are required to confirm the safety of individual active compounds [139].

Several studies have reported potential adverse effects associated with coumarin derivatives. Long-term use of oral coumarin-based anticoagulants, such as warfarin, has been linked to an increased risk of OP and fractures. Although the evidence remains controversial, this potential risk may be clinically significant for chronic users [140].

Long-term follow-up of patients using coumarin drugs has revealed serious adverse events, including gastrointestinal bleeding and intracranial hemorrhage. Furthermore, the impact of prolonged natural coumarin consumption on hepatic and renal functions requires further evaluation. While some studies indicate a favorable safety profile for certain natural coumarins within specific dosage ranges and treatment durations, the potential risks associated with extended use necessitate additional clinical research and vigilant monitoring to ensure patient safety.

Additionally, some coumarin compounds may exert embryotoxic effects at high doses. For example, several 4-phenyldihydrocoumarin derivatives demonstrated higher embryotoxicity in a zebrafish model than coumarin itself [141]. Thorough safety assessment and close monitoring of potential adverse reactions are therefore imperative when using natural coumarins in therapeutic applications.

### 3. Conclusions

In summary, natural coumarins represent a promising class of bioactive compounds with multi-target therapeutic potential in bone and joint disorders. Their broad anti-inflammatory, chondroprotective, and bone-modulating properties are underpinned by diverse molecular mechanisms, supporting their use in conditions such as OA, RA, and OP (Fig. 2). However, challenges remain in optimizing bioavailability, ensuring long-term safety, and validating efficacy through rigorous clinical trials.

This review analyzed and summarized the literature and research data on the chemical composition and pharmacological effects of natural coumarins, their relevance to clinical practice, efficacy in traditional medicine, and metabolism of circulating components. Future research should focus on achieving structural optimization and developing smart delivery systems and personalized treatment strategies. This should enable the translation of these natural agents into clinically viable therapies, while providing

guidance for further discoveries of the potential of natural coumarins as drugs and treatments.

### Abbreviations

AAA, abdominal aortic aneurysm; ALP, alkaline phosphatase; anti-CCP, anti-cyclic citrullinated peptide; ALT, alanine transaminase; AST, aspartate transaminase; AS, ankylosing spondylitis; AVF, arteriovenous fistula; CCL2, motif chemokine ligand 2; CCL5, motif chemokine ligand 5; CNM, cinnamoyloxy-mammeisin; COX-2, cyclooxygenase-2; CRP, C-reactive protein; CRRT, continuous renal replacement therapy; CTSK, Recombinant Human Cathepsin K; CT, computed tomography; CYP2C9, Cytochrome P450 proteins 2C9; CIA, collagen-induced arthritis; DBIL, direct bilirubin; 7-HC, 7-hydroxycoumarin; OA, osteoarthritis; DPPH, 1,1-diphenyl-2-picrylhydrazyl; Dex, dexamethasone; DMARDs, disease-modifying antirheumatic drugs; ESR, erythrocyte sedimentation rate; FLS, fibroblast-like synoviocytes; GDP, gross domestic product; HLA, human leukocyte antigen; ICAM-1, intercellular cell adhesion molecule-1; ICU, intensive care unit; IL-1 interleukin-1; IL-1 $\beta$ , interleukin-1 $\beta$ ; ILs, interleukins; IVC, inferior vena cava; JTNT, Jitongning Tablet; LPS, lipopolysaccharide; LTB4, levels of leukotriene B4; MAE, microwave-assisted extraction; MCO, methoxychromen-2-one; MCP-1, monocyte chemoattractant protein-1; M-CSF, macrophage colony-stimulating factor; MAPKs, mitogen-activated protein kinases; RUNX2, Runt-related transcription factor 2; MIA, monosodium iodoacetate; MMPs, matrix metalloproteinases; MMP-9, Matrix metalloproteinase 9; NETs, neutrophil extracellular traps; NFATc1, Recombinant Nuclear Factor Of Activated T-Cells, Cytoplasmic 1; NO, nitric oxide; NSAIDs, Non-steroidal anti-inflammatory drugs; NF- $\kappa$ B, nuclear factor kappa-B; OP, osteoporosis; OPG, osteoprotegerin; PGE<sub>2</sub>, prostaglandin E2; PHPLC, performance liquid chromatography; PR, progesterone receptor; SIRT1, sirtuin1; RA, Rheumatoid arthritis; RANKL, receptor activator of nuclear factor kappa-B ligand; RANTES, T cell expressed and secreted; RF, rheumatoid factor; ROS, reactive oxygen species; RP-MPLC, reversed-phase medium-pressure liquid chromatography; STAT3, signal transducer and activator of transcription 3; TBIL, total bilirubin; TLR2, Toll-like receptor 2; TRAP, Tartrate - Resistant Acid Phosphatase; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; XLGB, Xianlinggubao; VKORC1, vitamin K epoxide reductase complex subunit 1.

### Author Contributions

Conceptualization, CH and ZQL; methodology, WJW and JHM; software, LW; data curation, LW and XDZ; writing—original draft preparation, JHM; writing—review and editing, JHM; visualization, WJW; supervision, CH; project administration, CH; funding acquisition, CH. All

authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All data reported in this paper will also be shared by the lead contact upon request. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

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## Conflict of Interest

The authors declare no conflict of interest.

## Declaration of AI and AI-Assisted Technologies in the Writing Process

During the preparation of this work the authors used DeepSeek-V3 in order to check spell and grammar. After using this tool, the authors reviewed and edited the content as needed and takes full responsibility for the content of the publication.

## References

- [1] Koul B, Taak P, Kumar A, Kumar A, Sanyal I. Genus *Psoralea*: A review of the traditional and modern uses, phytochemistry and pharmacology. *Journal of Ethnopharmacology*. 2019; 232: 201–226. <https://doi.org/10.1016/j.jep.2018.11.036>.
- [2] Stringlis IA, de Jonge R, Pieterse CMJ. The Age of Coumarins in Plant-Microbe Interactions. *Plant & Cell Physiology*. 2019; 60: 1405–1419. <https://doi.org/10.1093/pcp/pcz076>.
- [3] Costa TM, Tavares LBB, de Oliveira D. Fungi as a source of natural coumarins production. *Applied Microbiology and Biotechnology*. 2016; 100: 6571–6584. <https://doi.org/10.1007/s00253-016-7660-z>.
- [4] Jayakumar T, Huang CJ, Yen TL, Hsia CW, Sheu JR, Bhavan PS, *et al.* Activation of Nrf2 by Esculetin Mitigates Inflammatory Responses through Suppression of NF- $\kappa$ B Signaling Cascade in RAW 264.7 Cells. *Molecules (Basel, Switzerland)*. 2022; 27: 5143. <https://doi.org/10.3390/molecules27165143>.
- [5] Min JS, Jin YH, Kwon S. Auraptene Has Antiviral Activity against Human Coronavirus OC43 in MRC-5 Cells. *Nutrients*. 2023; 15: 2960. <https://doi.org/10.3390/nu15132960>.
- [6] Golfakhrabadi F, Abdollahi M, Ardakani MRS, Saeidnia S, Akbarzadeh T, Ahmadabadi AN, *et al.* Anticoagulant activity of isolated coumarins (suberosin and suberenol) and toxicity evaluation of *Ferulago carduchorum* in rats. *Pharmaceutical Biology*. 2014; 52: 1335–1340. <https://doi.org/10.3109/13880209.2014.892140>.
- [7] Wang P, Fan Z, Wei W, Yang C, Wang Y, Shen X, *et al.* Biosynthesis of the Plant Coumarin Osthole by Engineered *Saccharomyces cerevisiae*. *ACS Synthetic Biology*. 2023; 12: 2455–2462. <https://doi.org/10.1021/acssynbio.3c00321>.
- [8] Chopra B, Dhingra AK, Dhar KL. *Psoralea corylifolia* L. (Buguchi) - folklore to modern evidence: review. *Fitoterapia*. 2013; 90: 44–56. <https://doi.org/10.1016/j.fitote.2013.06.016>.
- [9] He W, Chen W, Zhou Y, Tian Y, Liao F. Xanthotoxol exerts neuroprotective effects via suppression of the inflammatory response in a rat model of focal cerebral ischemia. *Cellular and Molecular Neurobiology*. 2013; 33: 715–722. <https://doi.org/10.1007/s10571-013-9939-2>.
- [10] Abramoff B, Caldera FE. Osteoarthritis: Pathology, Diagnosis, and Treatment Options. *The Medical Clinics of North America*. 2020; 104: 293–311. <https://doi.org/10.1016/j.mcna.2019.10.007>.
- [11] Barnett R. Osteoarthritis. *Lancet (London, England)*. 2018; 391: 1985. [https://doi.org/10.1016/S0140-6736\(18\)31064-X](https://doi.org/10.1016/S0140-6736(18)31064-X).
- [12] Courties A, Kouki I, Soliman N, Mathieu S, Sellam J. Osteoarthritis year in review 2024: Epidemiology and therapy. *Osteoarthritis and Cartilage*. 2024; 32: 1397–1404. <https://doi.org/10.1016/j.joca.2024.07.014>.
- [13] Zeng X, Tian X. 2024 Chinese guidelines for the diagnosis and treatment of rheumatoid arthritis. *Zhonghua Nei Ke Za Zhi*. 2024; 63: 1059–1077. <https://doi.org/10.3760/cma.j.cn.112138-20240531-00360>. (In Chinese)
- [14] Han S, Kong L, Li W, Deng Y, Meng H, Wang Z. Research progress on the intervention of traditional Chinese medicine monomers and compounds in pain-related signaling pathways of knee osteoarthritis. *Chinese Journal of Osteoporosis*. 2023; 29: 1490–1495. (In Chinese)
- [15] YANG X, Yao X, Huang C. Identifying rheumatoid arthritis-related genes and evaluating the intervention of compound Q-1 based on transcriptomics combined with m6A screening. *Science, Technology, and Engineering*. 2025; 25: 9734–9739. (In Chinese)
- [16] Ma J, Wang W, Zhang X, Huang C, Dong M. Crystal structure of 5-(2,5-dimethoxyphenyl)-8-methoxy-2-oxo-2H-chromene-6-carbaldehyde, C<sub>19</sub>H<sub>16</sub>O<sub>6</sub>. *Zeitschrift für Kristallographie - New Crystal Structures*. 2026; 241: 57–58. <https://doi.org/10.1515/ncrs-2025-0402>.
- [17] Zhao Y, Zhao Y, Zhang M, Luo L, Yang Y. Research on the Physiological and Pharmacological Activities of Coumarin Compounds. *Shandong Chemical Industry*. 2021: 30–33. <https://doi.org/10.3969/j.issn.1008-021X.2021.04.012>. (In Chinese)
- [18] Yang L, Yao X, Ding W. Study on the antibacterial activity of coumarin compounds. *Natural Product Research and Development*. 2018; 30: 332–338. <https://doi.org/10.16333/j.1001-6880.2018.2.026>. (In Chinese)
- [19] Huang S, Zhou Q, Luo T, Xie W. Research progress on the anti-tumor mechanism of coumarin compounds. *Shanghai Pharmaceutical*. 2022; 43: 70–74.
- [20] Annunziata F, Pinna C, Dallavalle S, Tamborini L, Pinto A. An Overview of Coumarin as a Versatile and Readily Accessible Scaffold with Broad-Ranging Biological Activities. *International Journal of Molecular Sciences*. 2020; 21: 4618. <https://doi.org/10.3390/ijms21134618>.
- [21] Ouyang L, Dan Y, Shao Z, Yang S, Yang C, Liu G, *et al.* Effect of umbelliferone on adjuvant-induced arthritis in rats by MAPK/NF- $\kappa$ B pathway. *Drug Design, Development and Therapy*. 2019; 13: 1163–1170. <https://doi.org/10.2147/DDDT.S190155>.
- [22] Fan X, Xie M, Zhao F, Li J, Fan C, Zheng H, *et al.* Daphnetin triggers ROS-induced cell death and induces cytoprotective autophagy by modulating the AMPK/Akt/mTOR pathway in ovarian cancer. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2021; 82: 153465. <https://doi.org/10.1016/j.phymed.2021.153465>.
- [23] Hang S, Wu W, Wang Y, Sheng R, Fang Y, Guo R. Daphnetin, a Coumarin in Genus *Stellera Chamaejasme* Linn: Chem-

- istry, Bioactivity and Therapeutic Potential. *Chemistry & Biodiversity*. 2022; 19: e202200261. <https://doi.org/10.1002/cbdv.202200261>.
- [24] Huang P, Yang L, Liu T, Jiang Y, Chen Z, Song H, *et al*. Scoparone alleviates nonalcoholic fatty liver disease by modulating the PPAR $\alpha$  signaling pathway. *European Journal of Pharmacology*. 2024; 984: 177033. <https://doi.org/10.1016/j.ejphar.2024.177033>.
- [25] Sumorek-Wiadro J, Zajac A, Bądziul D, Langner E, Skalicka-Woźniak K, Maciejczyk A, *et al*. Coumarins modulate the anti-glioma properties of temozolomide. *European Journal of Pharmacology*. 2020; 881: 173207. <https://doi.org/10.1016/j.ejphar.2020.173207>.
- [26] Yin S, Liu H, Wang J, Feng S, Chen Y, Shang Y, *et al*. Osthole Induces Apoptosis and Inhibits Proliferation, Invasion, and Migration of Human Cervical Carcinoma HeLa Cells. Evidence-based Complementary and Alternative Medicine: ECAM. 2021; 2021: 8885093. <https://doi.org/10.1155/2021/8885093>.
- [27] Kordulewska N, Topa J, Cieślińska A, Jarmołowska B. Osthole Regulates Secretion of Pro-Inflammatory Cytokines and Expression of *TLR2* and *NF- $\kappa$ B* in Normal Human Keratinocytes and Fibroblasts. *Journal of Inflammation Research*. 2022; 15: 1501–1519. <https://doi.org/10.2147/JIR.S349216>.
- [28] Yuan Z, Yu S, Su D, Gao Y, Zheng F, Yan P, *et al*. Fraxetin attenuates DNA damage and inflammation in cisplatin-induced nephrotoxicity via FoxO1 activation. *International Immunopharmacology*. 2025; 147: 114010. <https://doi.org/10.1016/j.intimp.2024.114010>.
- [29] Choi YJ, Lee CM, Park SH, Nam MJ. Esculetin induces cell cycle arrest and apoptosis in human colon cancer LoVo cells. *Environmental Toxicology*. 2019; 34: 1129–1136. <https://doi.org/10.1002/tox.22815>.
- [30] Sotoudeheian M, Mirahmadi SMS, Pirhayati M, Farahmandian N, Azarbad R, Toroudi HP. Targeting SIRT1 by Scopoletin to Inhibit XBB.1.5 COVID-19 Life Cycle. *Current Reviews in Clinical and Experimental Pharmacology*. 2025; 20: 4–13. <https://doi.org/10.2174/0127724328281178240225082456>.
- [31] Zhang X, Li L, Wu Y. Xanthoxyletin blocks the RANK/RANKL signaling pathway to suppress the growth of human pancreatic cancer cells. *Acta Pharmaceutica (Zagreb, Croatia)*. 2023; 73: 503–513. <https://doi.org/10.2478/acph-2023-0024>.
- [32] Cazal CDM, Forim MR, Terezan AP, Matos AP, Cunha GOS, da Silva MFDGF, *et al*. Development of Xanthyletin-Loaded Nanoparticles for the Control of *Leucoagaricus gongylophorus*. *Molecules (Basel, Switzerland)*. 2025; 30: 2469. <https://doi.org/10.3390/molecules30112469>.
- [33] Chen J, Zhou Y, Liu D, Lu X, Chen H, Huang M, *et al*. Discovery and Development of Luvangetin from *Zanthoxylum avicennae* as a New Fungicide Candidate for *Fusarium verticillioides*. *Journal of Agricultural and Food Chemistry*. 2024; 72: 8550–8568. <https://doi.org/10.1021/acs.jafc.3c09513>.
- [34] Muralikrishnan A, Sekar M, Kumarasamy V, Gan SH, Ravi S, Subramaniyan V, *et al*. Chemistry, Pharmacology and Therapeutic Potential of Decursin: A Promising Natural Lead for New Drug Discovery and Development. *Drug Design, Development and Therapy*. 2024; 18: 3741–3763. <https://doi.org/10.2147/DDDT.S476279>.
- [35] Kim SE, Lee JE, Han YH, Lee SI, Kim DK, Park SR, *et al*. Decursinol from *Angelica gigas* Nakai enhances endometrial receptivity during implantation. *BMC Complementary Medicine and Therapies*. 2020; 20: 36. <https://doi.org/10.1186/s12906-020-2822-z>.
- [36] Wang Y, Yu Y, Hou YP, Mao XW, Liu ZL, Cui J, *et al*. Crucial Role of the Ca<sup>2+</sup>/CN Signaling Pathway in the Antifungal Activity of Seselin against *Botrytis cinerea*. *Journal of Agricultural and Food Chemistry*. 2023; 71: 9772–9781. <https://doi.org/10.1021/acs.jafc.3c01474>.
- [37] Melliou E, Magiatis P, Mitaku S, Skaltsounis AL, Chinou E, Chinou I. Natural and synthetic 2,2-dimethylpyranocoumarins with antibacterial activity. *Journal of Natural Products*. 2005; 68: 78–82. <https://doi.org/10.1021/np0497447>.
- [38] Cao JL, Shen SL, Yang P, Qu J. A catalyst-free one-pot construction of skeletons of 5-methoxyseselin and alloxanthoxyletin in water. *Organic Letters*. 2013; 15: 3856–3859. <https://doi.org/10.1021/ol401581p>.
- [39] Yu CL, Yu YL, Yang SF, Hsu CE, Lin CL, Hsieh YH, *et al*. Praeruptorin A reduces metastasis of human hepatocellular carcinoma cells by targeting ERK/MMP1 signaling pathway. *Environmental Toxicology*. 2021; 36: 540–549. <https://doi.org/10.1002/tox.23059>.
- [40] Xu K, Chen Z, Hou J, Dong C, Shi C, Gao L, *et al*. Praeruptorin B inhibits osteoclastogenesis by targeting GSTP1 and impacting on the S-glutathionylation of IKK $\beta$ . *Biomedicine & Pharmacotherapy = Biomedicine & Pharmacotherapie*. 2022; 154: 113529. <https://doi.org/10.1016/j.biopha.2022.113529>.
- [41] Liu CM, Shen HT, Lin YA, Yu YL, Chen YS, Liu CJ, *et al*. Antiproliferative and Antimetastatic Effects of Praeruptorin C on Human Non-Small Cell Lung Cancer Through Inactivating ERK/CTSD Signalling Pathways. *Molecules (Basel, Switzerland)*. 2020; 25: 1625. <https://doi.org/10.3390/molecules25071625>.
- [42] Wu Y, Zhang YZ, Li MJ, Yang WQ, Cheng LF. The *In Vitro* Effect of Psoralen on Glioma Based on Network Pharmacology and Potential Target Research. Evidence-based Complementary and Alternative Medicine: ECAM. 2022; 2022: 1952891. <https://doi.org/10.1155/2022/1952891>.
- [43] Li S, Tu H. Psoralen inhibits the proliferation and promotes apoptosis through endoplasmic reticulum stress in human osteosarcoma cells. *Folia Histochemica et Cytobiologica*. 2022; 60: 101–109. <https://doi.org/10.5603/FHC.a2022.0010>.
- [44] Lee Y, Hyun CG. Anti-Inflammatory Effects of Psoralen Derivatives on RAW264.7 Cells via Regulation of the NF- $\kappa$ B and MAPK Signaling Pathways. *International Journal of Molecular Sciences*. 2022; 23: 5813. <https://doi.org/10.3390/ijms23105813>.
- [45] Lv M, Xu Q, Zhang B, Yang Z, Xie J, Guo J, *et al*. Imperatorin induces autophagy and G0/G1 phase arrest via PTEN-P13K-AKT-mTOR/p21 signaling pathway in human osteosarcoma cells in vitro and in vivo. *Cancer Cell International*. 2021; 21: 689. <https://doi.org/10.1186/s12935-021-02397-7>.
- [46] Fan L, Li Z, Gao L, Zhang N, Chang W. Isoimperatorin alleviates lipopolysaccharide-induced periodontitis by downregulating ERK1/2 and NF- $\kappa$ B pathways. *Open Life Sciences*. 2023; 18: 20220541. <https://doi.org/10.1515/biol-2022-0541>.
- [47] Wu A, Lu J, Zhong G, Lu L, Qu Y, Zhang C. Xanthotoxin (8-methoxypsoralen): A review of its chemistry, pharmacology, pharmacokinetics, and toxicity. *Phytotherapy Research: PTR*. 2022; 36: 3805–3832. <https://doi.org/10.1002/ptr.7577>.
- [48] Lin CP, Lin CS, Lin HH, Li KT, Kao SH, Tsao SM. Bergapten induces G1 arrest and pro-apoptotic cascade in colorectal cancer cells associating with p53/p21/PTEN axis. *Environmental Toxicology*. 2019; 34: 303–311. <https://doi.org/10.1002/tox.22685>.
- [49] Zhou Y, Wang J, Yang W, Qi X, Lan L, Luo L, *et al*. Bergapten prevents lipopolysaccharide-induced inflammation in RAW264.7 cells through suppressing JAK/STAT activation and ROS production and increases the survival rate of mice after LPS challenge. *International Immunopharmacology*. 2017; 48: 159–168. <https://doi.org/10.1016/j.intimp.2017.04.026>.
- [50] Zou J, Duan Y, Wang Y, Liu A, Chen Y, Guo D, *et al*. Phelopterin cream exerts an anti-inflammatory effect that facilitates diabetes-associated cutaneous wound healing via SIRT1. *Phytomedicine: International Journal of Phytotherapy and Phy-*

- topharmacology. 2022; 107: 154447. <https://doi.org/10.1016/j.phymed.2022.154447>.
- [51] Guo A, Lin J, Zhong P, Chen J, Wang L, Lin X, *et al.* Phellopterin attenuates ovarian cancer proliferation and chemoresistance by inhibiting the PU.1/CLEC5A/PI3K-AKT feedback loop. *Toxicology and Applied Pharmacology*. 2023; 477: 116691. <https://doi.org/10.1016/j.taap.2023.116691>.
- [52] Iwai K, Kon T, Fujita Y, Abe H, Honma H, Kawasumi N, *et al.* Genetic diversity of viruses infecting cnidium plants (*Cnidium officinale*) in Japan. *Virusdisease*. 2023; 34: 431–439. <https://doi.org/10.1007/s13337-023-00835-w>.
- [53] Mottaghipisheh J. Oxypeucedanin: Chemotaxonomy, Isolation, and Bioactivities. *Plants (Basel, Switzerland)*. 2021; 10: 1577. <https://doi.org/10.3390/plants10081577>.
- [54] Zhan W, Ruan B, Dong H, Wang C, Wu S, Yu H, *et al.* Isoporsalen suppresses receptor activator of nuclear factor kappa- $\beta$  ligand-induced osteoclastogenesis by inhibiting the NF- $\kappa$ B signaling. *PeerJ*. 2023; 11: e14560. <https://doi.org/10.7717/peerj.14560>.
- [55] O'Neill T, Johnson JA, Webster D, Gray CA. The Canadian medicinal plant *Heracleum maximum* contains antimycobacterial diynes and furanocoumarins. *Journal of Ethnopharmacology*. 2013; 147: 232–237. <https://doi.org/10.1016/j.jep.2013.03.009>.
- [56] Yang L, Du M, Liu K, Wang P, Zhu J, Li F, *et al.* Pimpinellin ameliorates macrophage inflammation by promoting RNF146-mediated PARP1 ubiquitination. *Phytotherapy Research: PTR*. 2024; 38: 1783–1798. <https://doi.org/10.1002/ptr.8135>.
- [57] Yamaguchi S, Muro S, Kobayashi M, Miyazawa M, Hirai Y. Absolute structures of some naturally occurring isopropenyldihydrobenzofurans, remirol, remiridiol, angenomalin, and isoangenomalin. *The Journal of Organic Chemistry*. 2003; 68: 6274–6278. <https://doi.org/10.1021/jo034396j>.
- [58] Han Y, Liu C, Chen S, Sun H, Jia Z, Shi J, *et al.* Columbianadin ameliorates rheumatoid arthritis by attenuating synoviocyte hyperplasia through targeted vimentin to inhibit the VAV2/Rac-1 signaling pathway. *Journal of Advanced Research*. 2025; 74: 609–620. <https://doi.org/10.1016/j.jare.2024.09.030>.
- [59] Mesas C, Garcés V, Martínez R, Ortiz R, Doello K, Dominguez-Vera JM, *et al.* Colon cancer therapy with calcium phosphate nanoparticles loading bioactive compounds from *Euphorbia lathyris*: In vitro and in vivo assay. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*. 2022; 155: 113723. <https://doi.org/10.1016/j.biopha.2022.113723>.
- [60] Xie Q, Fan X, Han Y, Wu BX, Zhu B. Daphnoretin Arrests the Cell Cycle and Induces Apoptosis in Human Breast Cancer Cells. *Journal of Natural Products*. 2022; 85: 2332–2339. <https://doi.org/10.1021/acs.jnatprod.2c00504>.
- [61] Sun C, Zhao W, Wang X, Sun Y, Chen X. A pharmacological review of dicoumarol: An old natural anticoagulant agent. *Pharmacological Research*. 2020; 160: 105193. <https://doi.org/10.1016/j.phrs.2020.105193>.
- [62] Gillis-Germitsch N, Müller S, Gori F, Schnyder M. *Capillaria boehmi* (syn. *Eucoleus boehmi*): Challenging treatment of a rarely diagnosed nasal nematode in dogs and high prevalence in Swiss foxes. *Veterinary Parasitology*. 2020; 281: 109103. <https://doi.org/10.1016/j.vetpar.2020.109103>.
- [63] Huang T, Zhao CC, Xue M, Cao YF, Chen LK, Chen JX, *et al.* Current Progress and Outlook for Agrimonolide: A Promising Bioactive Compound from *Agrimonia pilosa* Ledeb. *Pharmaceuticals (Basel, Switzerland)*. 2023; 16: 150. <https://doi.org/10.3390/ph16020150>.
- [64] Roy A. Plumbagin: A Potential Anti-cancer Compound. *Mini Reviews in Medicinal Chemistry*. 2021; 21: 731–737. <https://doi.org/10.2174/1389557520666201116144421>.
- [65] Chicowski AS, Bredow M, Utiyama AS, Marcelino-Guimarães FC, Whitham SA. Soybean-Phakopsora pachyrhizi interactions: towards the development of next-generation disease-resistant plants. *Plant Biotechnology Journal*. 2024; 22: 296–315. <https://doi.org/10.1111/pbi.14206>.
- [66] He L, Pan Y, Yu J, Wang B, Dai G, Ying X. Decursin alleviates the aggravation of osteoarthritis via inhibiting PI3K-Akt and NF- $\kappa$ B signal pathway. *International Immunopharmacology*. 2021; 97: 107657. <https://doi.org/10.1016/j.intimp.2021.107657>.
- [67] Zamani Taghizadeh Rabe S, Iranshahi M, Mahmoudi M. In vitro anti-inflammatory and immunomodulatory properties of umbelliprenin and methyl galbanate. *Journal of Immunotoxicology*. 2016; 13: 209–216. <https://doi.org/10.3109/1547691X.2015.1043606>.
- [68] Yang Y, Sheng Q, Nie Z, Liu L, Zhang W, Chen G, *et al.* Daphnetin inhibits spinal glial activation via Nrf2/HO-1/NF- $\kappa$ B signaling pathway and attenuates CFA-induced inflammatory pain. *International Immunopharmacology*. 2021; 98: 107882. <https://doi.org/10.1016/j.intimp.2021.107882>.
- [69] Lu J, Fang K, Wang S, Xiong L, Zhang C, Liu Z, *et al.* Anti-Inflammatory Effect of Columbianetin on Lipopolysaccharide-Stimulated Human Peripheral Blood Mononuclear Cells. *Mediators of Inflammation*. 2018; 2018: 9191743. <https://doi.org/10.1155/2018/9191743>.
- [70] Siano F, Picariello G, De Paola M, Caruso T, Iannece P, Vasca E. Combined coulometric and voltammetric analytical approach provides novel insights into the antioxidant capacity of natural coumarins and furanocoumarins. *Talanta*. 2025; 287: 127579. <https://doi.org/10.1016/j.talanta.2025.127579>.
- [71] Taşdemir E, Atmaca M, Yıldırım Y, Bilgin HM, Demirtaş B, Obay BD, *et al.* Influence of coumarin and some coumarin derivatives on serum lipid profiles in carbon tetrachloride-exposed rats. *Human & Experimental Toxicology*. 2017; 36: 295–301. <https://doi.org/10.1177/0960327116649675>.
- [72] Venkata Sairam K, Gurupadaya BM, Chandan RS, Nagesha DK, Vishwanathan B. A Review on Chemical Profile of Coumarins and their Therapeutic Role in the Treatment of Cancer. *Current Drug Delivery*. 2016; 13: 186–201. <https://doi.org/10.2174/1567201812666150702102800>.
- [73] Rajabi M, Hossaini Z, Khalilzadeh MA, Datta S, Halder M, Mousa SA. Synthesis of a new class of furo[3,2-c]coumarins and its anticancer activity. *Journal of Photochemistry and Photobiology B: Biology*. 2015; 148: 66–72. <https://doi.org/10.1016/j.jphotobiol.2015.03.027>.
- [74] Garro HA, Pungitore CR. Coumarins as Potential Inhibitors of DNA Polymerases and Reverse Transcriptases. *Searching New Antiretroviral and Antitumoral Drugs. Current Drug Discovery Technologies*. 2015; 12: 66–79. <https://doi.org/10.2174/1570163812666150716111719>.
- [75] Klepka MT, Drzewiecka-Antonik A, Wolska A, Rejmak P, Ostrowska K, Hejchman E, *et al.* Synthesis, structural studies and biological activity of new Cu(II) complexes with acetyl derivatives of 7-hydroxy-4-methylcoumarin. *Journal of Inorganic Biochemistry*. 2015; 145: 94–100. <https://doi.org/10.1016/j.jinorgbio.2015.01.006>.
- [76] Mathieson S, Ferreira G, Jones CMP, Eyles J, Bowden JL, Sharma S, *et al.* The cost-effectiveness of guideline-recommended treatments for osteoarthritis: A systematic review. *Osteoarthritis and Cartilage*. 2025; 33: 1274–1292. <https://doi.org/10.1016/j.joca.2025.04.003>.
- [77] Hall M, van der Esch M, Hinman RS, Peat G, de Zwart A, Quicke JG, *et al.* How does hip osteoarthritis differ from knee osteoarthritis? *Osteoarthritis and Cartilage*. 2022; 30: 32–41. <https://doi.org/10.1016/j.joca.2021.09.010>.
- [78] Motaung A, Rants'o TA, Walvekar P, Luliński P, Choonara YE. Recent advances in intra-articular bioactive delivery for the

- treatment of osteoarthritis. *International Journal of Pharmaceutics*. 2026; 687: 126401. <https://doi.org/10.1016/j.ijpharm.2025.126401>.
- [79] Mabey T, Honsawek S. Cytokines as biochemical markers for knee osteoarthritis. *World Journal of Orthopedics*. 2015; 6: 95–105. <https://doi.org/10.5312/wjo.v6.i1.95>.
- [80] Sun Y, Zhou L, Lv D, Liu H, He T, Wang X. Poly(ADP-ribose) polymerase 1 inhibition prevents interleukin-1 $\beta$ -induced inflammation in human osteoarthritic chondrocytes. *Acta Biochimica et Biophysica Sinica*. 2015; 47: 422–430. <https://doi.org/10.1093/abbs/gmv033>.
- [81] Lin J, Li X, Qi W, Yan Y, Chen K, Xue X, *et al.* Isofraxidin inhibits interleukin-1 $\beta$  induced inflammatory response in human osteoarthritis chondrocytes. *International Immunopharmacology*. 2018; 64: 238–245. <https://doi.org/10.1016/j.intimp.2018.09.003>.
- [82] You JS, Cho IA, Kang KR, Oh JS, Yu SJ, Lee GJ, *et al.* Coumestrol Counteracts Interleukin-1 $\beta$ -Induced Catabolic Effects by Suppressing Inflammation in Primary Rat Chondrocytes. *Inflammation*. 2017; 40: 79–91. <https://doi.org/10.1007/s10753-016-0455-7>.
- [83] Gao L, Beninato R, Oláh T, Goebel L, Tao K, Roels R, *et al.* A Photopolymerizable Biocompatible Hyaluronic Acid Hydrogel Promotes Early Articular Cartilage Repair in a Minipig Model In Vivo. *Advanced Healthcare Materials*. 2023; 12: e2300931. <https://doi.org/10.1002/adhm.202300931>.
- [84] He Y, Zisan Z, Lu Z, Zheng L, Zhao J. Bergapten alleviates osteoarthritis by regulating the ANP32A/ATM signaling pathway. *FEBS Open Bio*. 2019; 9: 1144–1152. <https://doi.org/10.1002/2211-5463.12648>.
- [85] Chern CM, Zhou H, Wang YH, Chang CL, Chiou WF, Chang WT, *et al.* Osthole ameliorates cartilage degradation by down-regulation of NF- $\kappa$ B and HIF-2 $\alpha$  pathways in an osteoarthritis murine model. *European Journal of Pharmacology*. 2020; 867: 172799. <https://doi.org/10.1016/j.ejphar.2019.172799>.
- [86] Di Matteo A, Bathon JM, Emery P. Rheumatoid arthritis. *Lancet* (London, England). 2023; 402: 2019–2033. [https://doi.org/10.1016/S0140-6736\(23\)01525-8](https://doi.org/10.1016/S0140-6736(23)01525-8).
- [87] Rahmati M, Kwesiga MP, Lou J, Tan AL, McDermott MF. Novel Targeted Therapies for Rheumatoid Arthritis Based on Intracellular Signaling and Immunometabolic Changes: A Narrative Review. *Frontiers in Bioscience (Landmark Edition)*. 2024; 29: 42. <https://doi.org/10.31083/j.fbl2901042>.
- [88] Farzaei MH, Farzaei F, Abdollahi M, Abbasabadi Z, Abdolghafari AH, Mehraban B. A mechanistic review on medicinal plants used for rheumatoid arthritis in traditional Persian medicine. *The Journal of Pharmacy and Pharmacology*. 2016; 68: 1233–1248. <https://doi.org/10.1111/jphp.12606>.
- [89] Sahu D, Raghav SK, Gautam H, Das HR. A novel coumarin derivative, 8-methoxy chromen-2-one alleviates collagen induced arthritis by down regulating nitric oxide, NF $\kappa$ B and proinflammatory cytokines. *International Immunopharmacology*. 2015; 29: 891–900. <https://doi.org/10.1016/j.intimp.2015.08.012>.
- [90] An J, Hao D, Zhang Q, Chen B, Zhang R, Wang Y, *et al.* Natural products for treatment of bone erosive diseases: The effects and mechanisms on inhibiting osteoclastogenesis and bone resorption. *International Immunopharmacology*. 2016; 36: 118–131. <https://doi.org/10.1016/j.intimp.2016.04.024>.
- [91] Rzodkiewicz P, Gaśnińska E, Gajewski M, Bujalska-Zadrożny M, Szukiewicz D, Maśliński S. Esculetin reduces leukotriene B4 level in plasma of rats with adjuvant-induced arthritis. *Reumatologia*. 2016; 54: 161–164. <https://doi.org/10.5114/reum.2016.62469>.
- [92] Albiero LR, de Andrade MF, Marchi LF, Landi-Librandi AP, de Figueiredo-Rinhel ASG, Carvalho CA, *et al.* Immunomodulating action of the 3-phenylcoumarin derivative 6,7-dihydroxy-3-[3',4'-methylenedioxyphenyl]-coumarin in neutrophils from patients with rheumatoid arthritis and in rats with acute joint inflammation. *Inflammation Research*. 2020; 69: 115–130. <https://doi.org/10.1007/s00011-019-01298-w>.
- [93] Kong L, Ma R, Yang X, Zhu Z, Guo H, He B, *et al.* Psoralidin suppresses osteoclastogenesis in BMMs and attenuates LPS-mediated osteolysis by inhibiting inflammatory cytokines. *International Immunopharmacology*. 2017; 51: 31–39. <https://doi.org/10.1016/j.intimp.2017.07.003>.
- [94] Wang Q, Zhou X, Yang L, Zhao Y, Chew Z, Xiao J, *et al.* The Natural Compound Notopterol Binds and Targets JAK2/3 to Ameliorate Inflammation and Arthritis. *Cell Reports*. 2020; 32: 108158. <https://doi.org/10.1016/j.celrep.2020.108158>.
- [95] Chen X, Kuang N, Zeng X, Zhang Z, Li Y, Liu W, *et al.* Effects of daphnetin combined with Bcl2 siRNA on antiapoptotic genes in synovial fibroblasts of rats with collagen induced arthritis. *Molecular Medicine Reports*. 2018; 17: 884–890. <https://doi.org/10.3892/mmr.2017.8008>.
- [96] Cai L, Zong P, Zhou MY, Liu FY, Meng B, Liu MM, *et al.* 7-Hydroxycoumarin mitigates the severity of collagen-induced arthritis in rats by inhibiting proliferation and inducing apoptosis of fibroblast-like synoviocytes via suppression of Wnt/ $\beta$ -catenin signaling pathway. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2022; 94: 153841. <https://doi.org/10.1016/j.phymed.2021.153841>.
- [97] da Cunha MG, Ramos-Junior ES, Franchin M, Taira TM, Beutler JA, Franco GCN, *et al.* Effects of Cinnamoyloxy-mammeisin from Geopropolis on Osteoclast Differentiation and Porphyromonas gingivalis-Induced Periodontitis. *Journal of Natural Products*. 2017; 80: 1893–1899. <https://doi.org/10.1021/acs.jnatprod.7b00194>.
- [98] Zhang YY, Xie N, Sun XD, Nice EC, Liou YC, Huang C, *et al.* Insights and implications of sexual dimorphism in osteoporosis. *Bone Research*. 2024; 12: 8. <https://doi.org/10.1038/s41413-023-00306-4>.
- [99] Zhivodernikov IV, Kirichenko TV, Markina YV, Postnov AY, Markin AM. Molecular and Cellular Mechanisms of Osteoporosis. *International Journal of Molecular Sciences*. 2023; 24: 15772. <https://doi.org/10.3390/ijms242115772>.
- [100] Sun H, Wu J, Ding Y. Application progress of bone turnover biomarkers in early screening of osteoporosis. *Chinese Journal of Osteoporosis and Bone Mineral Diseases*. 2024; 17: 481–490. (In Chinese)
- [101] Zhang Y, Liang JQ, Liu PL, Wang Q, Liu L, Zhao HM. The RANK/RANKL/OPG system and tumor bone metastasis: Potential mechanisms and therapeutic strategies. *Frontiers in Endocrinology*. 2022; 13: 8. <https://doi.org/10.3389/fendo.2022.1063815>.
- [102] Abdulrahman SJ, Abdulhadi MA, Turki Jalil A, Falah D, Merza MS, Almulla AF, *et al.* Conjugated linoleic acid and glucosamine supplements may prevent bone loss in aging by regulating the RANKL/RANK/OPG pathway. *Molecular Biology Reports*. 2023; 50: 10579–10588. <https://doi.org/10.1007/s11033-023-08839-x>.
- [103] Wang Y, Liu H, Liu T, Yang H, He F. Insights into the Role of Macrophage Polarization in the Pathogenesis of Osteoporosis. *Oxidative Medicine and Cellular Longevity*. 2022; 2022: 2485959. <https://doi.org/10.1155/2022/2485959>.
- [104] Yu X, Huang Z, Zhang P. Research progress on the relationship between oxidative stress and osteoporotic fracture healing. *World Science and Technology - Modernization of Traditional Chinese Medicine*. 2024; 26: 3171–3179. (In Chinese)
- [105] LeBoff MS, Greenspan SL, Insogna KL, Lewiecki EM, Saag KG, Singer AJ, *et al.* The clinician's guide to prevention and treatment of osteoporosis. *Osteoporosis International: a Jour-*

- nal Established as Result of Cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA. 2022; 33: 2049–2102. <https://doi.org/10.1007/s00198-021-05900-y>.
- [106] Wang X, He Y, Guo B, Tsang MC, Tu F, Dai Y, *et al*. In vivo screening for anti-osteoporotic fraction from extract of herbal formula Xianlinggubao in ovariectomized mice. *PLoS One*. 2015; 10: e0118184. <https://doi.org/10.1371/journal.pone.0118184>.
- [107] Jia M, Li Y, Xin HL, Hou TT, Zhang ND, Xu HT, *et al*. Estrogenic activity of osthole and imperatorin in MCF-7 cells and their osteoblastic effects in Saos-2 cells. *Chinese Journal of Natural Medicines*. 2016; 14: 413–420. [https://doi.org/10.1016/S1875-5364\(16\)30037-1](https://doi.org/10.1016/S1875-5364(16)30037-1).
- [108] An J, Yang H, Zhang Q, Liu C, Zhao J, Zhang L, *et al*. Natural products for treatment of osteoporosis: The effects and mechanisms on promoting osteoblast-mediated bone formation. *Life Sciences*. 2016; 147: 46–58. <https://doi.org/10.1016/j.lfs.2016.01.024>.
- [109] Kwak SC, Baek JM, Lee CH, Yoon KH, Lee MS, Kim JY. Umbelliferone Prevents Lipopolysaccharide-Induced Bone Loss and Suppresses RANKL-Induced Osteoclastogenesis by Attenuating Akt-c-Fos-NFATc1 Signaling. *International Journal of Biological Sciences*. 2019; 15: 2427–2437. <https://doi.org/10.7150/ijbs.28609>.
- [110] Ju Y, Xiong W, Pang Y, Chen Z, Hou J, Zheng S, *et al*. The Association between Dietary and Circulating Copper Levels and Osteoporosis: a Scoping Review. *Biological Trace Element Research*. 2025. <https://doi.org/10.1007/s12011-025-04899-1>. (online ahead of print)
- [111] Li S, Wen Q, Zhao M. Clinical study on the treatment of knee osteoarthritis of liver and kidney deficiency type with modified Duhuo Jisheng Decoction combined with sodium hyaluronate. *Hebei Traditional Chinese Medicine*. 2024; 46: 1475–1479. (In Chinese)
- [112] Guo N, Wang W. Clinical efficacy of Duhuo Parasitic Decoction combined with Methotrexate tablets in the treatment of rheumatoid arthritis. *World Traditional Chinese Medicine*. 2018; 13: 4. (In Chinese)
- [113] Tian D, Zhao H, Cao J, Zhang S, Wang W, Tang X, *et al*. Deciphering in vivo metabolic profile and pharmacological mechanisms of Jitongning Tablet for the treatment of Ankylosing spondylitis. *Journal of Pharmaceutical and Biomedical Analysis*. 2023; 227: 115271. <https://doi.org/10.1016/j.jpba.2023.115271>.
- [114] Zhao Y, Zhang Y, Xie Y, Wang Z. Analysis of Clinical Application Characteristics of Xianling Gubao Capsules in the Real World. *World Journal of Integrated Traditional Chinese and Western Medicine*. 2019; 14: 5. (In Chinese)
- [115] Zhan H, Zhang Z. Guidelines for clinical application of traditional Chinese patent medicines and simple preparations in the treatment of osteoporosis (2021). *Chinese Journal of Integrated Traditional Chinese and Western Medicine*. 2022; 42: 393–404. (In Chinese)
- [116] Fu Y, Shi D. The effect of Xianling Gubao capsule combined with zoledronic acid intravenous infusion and calcium carbonate D3 tablets on bone density and bone turnover in elderly patients with osteoporosis. *Chinese Journal of Gerontology*. 2021; 41: 3. (In Chinese)
- [117] Zheng M, Kuang N, Zeng X, Wang J, Zou Y, Fu Y. Daphnetin induces apoptosis in fibroblast-like synoviocytes from collagen-induced arthritic rats mainly via the mitochondrial pathway. *Cytokine*. 2020; 133: 155146. <https://doi.org/10.1016/j.cyto.2020.155146>.
- [118] Guo Q, Li L, Zheng K, Zheng G, Shu H, Shi Y, *et al*. Imperatorin and  $\beta$ -sitosterol have synergistic activities in alleviating collagen-induced arthritis. *Journal of Leukocyte Biology*. 2020; 108: 509–517. <https://doi.org/10.1002/JLB.3MA0320-440RR>.
- [119] Militaru FC, Vesa SC, Pop TR, Buzoianu AD. Pharmacogenetics aspects of oral anticoagulants therapy. *Journal of Medicine and Life*. 2015; 8: 171–175.
- [120] Chen XL, Li SX, Ge T, Zhang DD, Wang HF, Wang W, *et al*. *Epimedium Linn: A Comprehensive Review of Phytochemistry, Pharmacology, Clinical Applications and Quality Control*. *Chemistry & Biodiversity*. 2024; 21: e202400846. <https://doi.org/10.1002/cbdv.202400846>.
- [121] Ahmad S, Ahsan F, Ansari JA, Mahmood T, Bano S, Shahanawaz M. Bioflavonoid Daidzein: Therapeutic Insights, Formulation Advances, and Future Directions. *Drug Research*. 2024; 74: 433–455. <https://doi.org/10.1055/a-2379-6849>.
- [122] Berezinskaja VV, Nikol'skaia BS. Pharmacology of sinomenin. *Farmakologija i Toksikologija*. 1956; 19: 13–14. (In Russian)
- [123] Cai XH, Zhang HY, Xie B. Matrine-Family Alkaloids: Versatile Precursors for Bioactive Modifications. *Medicinal Chemistry (Shariqah (United Arab Emirates))*. 2020; 16: 431–453. <https://doi.org/10.2174/1573406415666190507121744>.
- [124] Shaito A, Posadino AM, Younes N, Hasan H, Halabi S, Alhababi D, *et al*. Potential Adverse Effects of Resveratrol: A Literature Review. *International Journal of Molecular Sciences*. 2020; 21: 2084. <https://doi.org/10.3390/ijms21062084>.
- [125] Kotha RR, Luthria DL. Curcumin: Biological, Pharmaceutical, Nutraceutical, and Analytical Aspects. *Molecules (Basel, Switzerland)*. 2019; 24: 2930. <https://doi.org/10.3390/molecules24162930>.
- [126] Gao J, Chen F, Fang H, Mi J, Qi Q, Yang M. Daphnetin inhibits proliferation and inflammatory response in human HaCaT keratinocytes and ameliorates imiquimod-induced psoriasis-like skin lesion in mice. *Biological Research*. 2020; 53: 48. <https://doi.org/10.1186/s40659-020-00316-0>.
- [127] Liang Y, Xie L, Liu K, Cao Y, Dai X, Wang X, *et al*. Bergapten: A review of its pharmacology, pharmacokinetics, and toxicity. *Phytotherapy Research: PTR*. 2021; 35: 6131–6147. <https://doi.org/10.1002/ptr.7221>.
- [128] Sharifi-Rad J, Cruz-Martins N, López-Jornet P, Lopez EPF, Harun N, Yeskaliyeva B, *et al*. Natural Coumarins: Exploring the Pharmacological Complexity and Underlying Molecular Mechanisms. *Oxidative Medicine and Cellular Longevity*. 2021; 2021: 6492346. <https://doi.org/10.1155/2021/6492346>.
- [129] Jiang Q, Gong X, Jiao J, Song M, Han M, Zhang R, *et al*. Guidelines for Combined Diagnosis and Treatment of Rheumatoid Arthritis (2025). *Journal of Traditional Chinese Medicine*. 2025; 66: 1842–1856. <https://doi.org/10.13288/j.11-2166/r.2025.17.018>. (In Chinese)
- [130] Tu Y, Wang K, Tan L, Han B, Hu Y, Ding H, *et al*. Dolichosin A, a coumestan isolated from *Glycine tabacina*, inhibits IL-1 $\beta$ -induced inflammation in SW982 human synovial cells and suppresses RANKL-induced osteoclastogenesis: From network pharmacology to experimental pharmacology. *Journal of Ethnopharmacology*. 2020; 258: 112855. <https://doi.org/10.1016/j.jep.2020.112855>.
- [131] Aas Z, Babaei E, Hosseinpour Feizi MA, Dehghan G. Anti-proliferative and Apoptotic Effects of Dendrosomal Farnesiferol C on Gastric Cancer Cells. *Asian Pacific Journal of Cancer Prevention: APJCP*. 2015; 16: 5325–5329. <https://doi.org/10.7314/apjcp.2015.16.13.5325>.
- [132] Wang Q, Jiang J, Chen W, Jiang H, Zhang Z, Sun X. Targeted delivery of low-dose dexamethasone using PCL-PEG micelles for effective treatment of rheumatoid arthritis. *Journal of Controlled Release: Official Journal of the Controlled Release Society*. 2016; 230: 64–72. <https://doi.org/10.1016/j.jconrel.2016.03.035>.
- [133] Wang K, Yin C, Ye X, Chen Q, Wu J, Chen Y, *et al*. A

- Metabolic Driven Bio-Responsive Hydrogel Loading Psoralen for Therapy of Rheumatoid Arthritis. *Small* (Weinheim an Der Bergstrasse, Germany). 2023; 19: e2207319. <https://doi.org/10.1002/sml.202207319>.
- [134] Murayama N, Yamazaki H. Metabolic activation and deactivation of dietary-derived coumarin mediated by cytochrome P450 enzymes in rat and human liver preparations. *The Journal of Toxicological Sciences*. 2021; 46: 371–378. <https://doi.org/10.2131/jts.46.371>.
- [135] Wang Q, Qiao X, Qian Y, Liu CF, Yang YF, Ji S, *et al.* Metabolites identification of glycyrin and glycyrol, bioactive coumarins from licorice. *Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences*. 2015; 983-984: 39–46. <https://doi.org/10.1016/j.jchromb.2014.12.028>.
- [136] Gomez-Verjan JC, Estrella-Parra E, Vazquez-Martinez ER, Gonzalez-Sanchez I, Guerrero-Magos G, Mendoza-Villanueva D, *et al.* Risk assessment of Soulatrolide and Mamea (A/BA+A/BB) coumarins from *Calophyllum brasiliense* by a toxicogenomic and toxicological approach. *Food and Chemical Toxicology: an International Journal Published for the British Industrial Biological Research Association*. 2016; 91: 117–129. <https://doi.org/10.1016/j.fct.2016.03.010>.
- [137] Chik WI, Zhu L, Fan LL, Yi T, Zhu GY, Gou XJ, *et al.* *Saussurea involucreata*: A review of the botany, phytochemistry and ethnopharmacology of a rare traditional herbal medicine. *Journal of Ethnopharmacology*. 2015; 172: 44–60. <https://doi.org/10.1016/j.jep.2015.06.033>.
- [138] Yang L, Hou A, Wang S, Zhang J, Man W, Guo X, *et al.* A Review of the Botany, Traditional Use, Phytochemistry, Analytical Methods, Pharmacological Effects, and Toxicity of *Angelicae Pubescentis Radix*. *Evidence-based Complementary and Alternative Medicine: ECAM*. 2020; 2020: 7460781. <https://doi.org/10.1155/2020/7460781>.
- [139] Tian-Liang, Xi-Gu-Ri-Gan, Yu J, Qu S, Xie Q, Shama R, *et al.* *Ardisia gigantifolia* stapf (Primulaceae): A review of ethnobotany, phytochemistry, pharmacology, clinical application, and toxicity. *Journal of Ethnopharmacology*. 2023; 305: 116079. <https://doi.org/10.1016/j.jep.2022.116079>.
- [140] Tufano A, Coppola A, Contaldi P, Franchini M, Minno GD. Oral anticoagulant drugs and the risk of osteoporosis: new anticoagulants better than old? *Seminars in Thrombosis and Hemostasis*. 2015; 41: 382–388. <https://doi.org/10.1055/s-0034-1543999>.
- [141] Veselinović JB, Kocić GM, Pavic A, Nikodinovic-Runic J, Senerovic L, Nikolić GM, *et al.* Selected 4-phenyl hydroxycoumarins: in vitro cytotoxicity, teratogenic effect on zebrafish (*Danio rerio*) embryos and molecular docking study. *Chemicobiological Interactions*. 2015; 231: 10–17. <https://doi.org/10.1016/j.cbi.2015.02.011>.