

















Original Research

Metagenomic Composition and Predicted Metabolic Pathway Analyses of the Endometrial and Rectal Microbiota in Dairy Cows Following the Introduction of a Complex Feed Additive

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Abstract

Background: The microbiome composition in dairy cows (*Bos taurus*) directly impacts on health and reproductive performance. This study aimed to determine the metagenomic composition and predicted microbial community functions in the endometrium and rectal chyme of cows fed a complex feed additive (CFA). The latter included the *Bacillus mucilaginosus* 159 strain, a short-chain fatty acid, plus essential oils. **Methods:** Clinically healthy cows were divided into two groups ($n = 15$ in each): (I) a control group fed the standard diet, and (II) an experimental group. CFA was introduced into the diet of Group II during the entire transit period at a dose of 50 g per animal per day; moreover, all animals received Pen-Strep 400 antibiotics to prevent endometritis and other pathologies. The microbial community composition from the endometrium and rectal chyme biotopes was assessed using targeted next-generation sequencing. **Results:** Significant changes were observed in the composition and predicted metabolic pathways due to the CFA administration, with the endometrial microbiota being more responsive to CFA than the intestinal chyme microbiome. Remarkably, the Actinobacteriota representatives disappeared in the endometrium of Group II animals compared to controls, whose content ranged from 0.34 to 3.3%. The use of CFA also resulted in a less pronounced effect in four predicted metabolic pathways for microbial degradation of catechol in the endometrium compared to controls ($p < 0.05$). **Conclusions:** Our findings support the concept of a relationship between the gut microbiome and the reproductive system microflora of cows, as we observed changes in the composition and predicted metabolic pathways of the endometrial microbiota after orally administering CFA. This emphasizes the need for an integrated approach combining the correction of microecological disorders in the intestines and the reproductive system simultaneously.

Keywords: *Bos taurus*; dairy cows; microbiota; metabolic pathways; endometrium; rectal chyme; complex feed additive; next-generation sequencing

1. Introduction

Managed livestock represents an essential cornerstone for human nutrition, with cattle (*Bos taurus*) being used both for meat and milk [1]. In recent years, much attention has been paid to the comfort of the animals' habitat during the transit period, i.e., the 21 days immediately preceding and the 21 days following calving. This is the most critical period in the life of a productive dairy cow, shaping a state of homeostasis of the animal throughout the next physiological cycle and is the time of greatest susceptibility to disease

[2]. A disparity between the need for energy, the level of its supply through feed and the failure of internal pathogenetic mechanisms can inhibit the functionality of the reproductive system [3]. It can lead to disruption of the delicate balance of the microbiota, including microbial populations living in both the digestive and reproductive systems [4].

Modern treatment of postpartum endometritis is based on complex treatment regimens that include antibacterial drugs, often administered in combination [5]. The use of antibiotics, however, while suppressing the symptoms of



endometritis, often has no effect on improving the reproductive capability of the cow, and can often lead to a recurrence of infection. In our opinion, this may be due to the suppression of the normal microbiota composition of the reproductive organs, thereby perpetuating the initial problem it was designed to improve [6,7]. Treatment regimens for endometritis thus require adjustments that include the development of new treatment regimes, alongside effective preventive measures. The concomitant use of biological products to restore the microbiota before, during and after antibiotic treatment [8,9] can help normalize the microecological status of the cow's reproductive system and increase the effectiveness of standard therapy.

In terms of improving the gut health of livestock and poultry, therapeutic and nutritional interest in commercially produced biological feed additives, such as prebiotics [10,11] and probiotics [9,12–15], is rapidly growing. The microbiome of the cattle digestive system has been repeatedly confirmed to be associated with health and productivity of animals [16]. Probiotics have been shown to have significant potential in altering the composition of the rumen microbiota beneficially, with some probiotics colonizing the vagina after oral administration [17]. In a human study, a meta-analysis of 22 randomized controlled trials found that probiotics are very effective when used in combination with antibiotics to treat and prevent relapses of bacterial vaginosis [8]. When conducting studies of the vaginal mucosa of cows using quantitative PCR (qPCR), we discovered a relationship between the rumen microbiota and the reproductive system [18,19]. Based on the results obtained, the spectrum of action of the probiotic Provitol composed of a strain of the microorganism *Enterococcus faecium* and essential oils for oral administration has been expanded. This enabled us to recommend this developed biotechnological product for regulating the composition of the vaginal microbiota of cows through optimizing the composition of the rumen microbiome [20]. Previously, we [16,21] also used metagenomic next-generation sequencing (NGS) to explore the functional potential of metabolic pathways, α - and β -diversity and gene expression of the rumen microbiome in lactating cows fed an *E. faecium*-based probiotic Cellobacterin+. Using a PCR testing of samples from the lactating cows' rectum, rumen chyme, milk and feed, Yildirim *et al.* [22] reported a potential transmission of certain enterobacteria and clostridia endotoxins across the feed–cow–milk chain.

In addition to microorganisms with probiotic properties, essential oils, which are natural substances produced by plants, are widely used in human and veterinary medicine due to their antioxidant, antiviral, and antibacterial properties [23]. The efficacy of essential oils against mastitis [24], respiratory diseases [25], and reproductive pathologies in cattle [26] has been reported. For example, a recent study showed that cinnamon essential oil had high antibacterial activity against *Trueperella pyogenes* and *Es-*

cherichia coli that are pathogens associated with inflammatory diseases of the uterus in dairy cows [27]. Short- and medium-chain fatty acids are also considered as an effective means for restoring the composition of the microbiome and reducing inflammatory reactions in the body [28]. Antimicrobial activity is manifested due to the ability of these substances to penetrate the bacterial cell wall and inhibit enzymatic reactions and nutrient transport systems [29]. Moreover, the anti-inflammatory, antioxidant and immunomodulatory properties of organic acids are manifested both at the local and systemic levels [30]. This demonstrates the high potential of using essential oils and fatty acids as alternative antibacterial agent for endometritis treatment in dairy cows.

It has been shown that disturbances in the intestinal microbiota can cause diseases of the extraintestinal system, such as diabetes, mastitis, and reproductive pathologies [31]. On the other hand, immune homeostasis is fundamentally maintained by the immune system's interaction with the gut bacteria [31]. Therefore, further in-depth studies are required to explore the relationship between the composition of the intestinal microbiota and endometrium of cows [32] when administering oral preventive supplements.

The purpose of this study was to examine the metagenomic composition and predicted functions of the microbial communities of the endometrium and rectal chyme biotopes of cows. We specifically investigated the effect of a complex feed additive (CFA) containing the microorganism strain *Bacillus mucilaginosus* 159, a short-chain fatty acid and essential oils was introduced into the cow diet during the transit period.

2. Materials and Methods

2.1 Experimental Animals

The experiment was carried out including a total of 2261 animals, including Holstein cows of the third lactation with an average milk production of 12,819 kg per lactation and an average live weight of 715 kg. Animals with similar characteristics, in terms of productivity during the previous lactation, age, live weight, and date of calving, were subjected to clinical and gynecological examination [33] during the transit period conforming to interlactation (i.e., 21 days before calving). The criteria for selecting animals for the experiment were the absence of treatment with antibiotics and hormonal drugs or other drugs in the previous lactation. The live weight of cows in the experiment and control averaged 712–715 kg. The animals were kept in the same free-stall conditions, had the same diet composition, and had good reproductive history. Rations were calculated according to the feeding standards recommended by the National Academies of Sciences, Engineering, and Medicine [34] taking into account the physiological needs of animals during the whole transit period (i.e., 21 days before calving and 21 days after calving). The standard (base) diets were completely balanced in nutrients, and all components

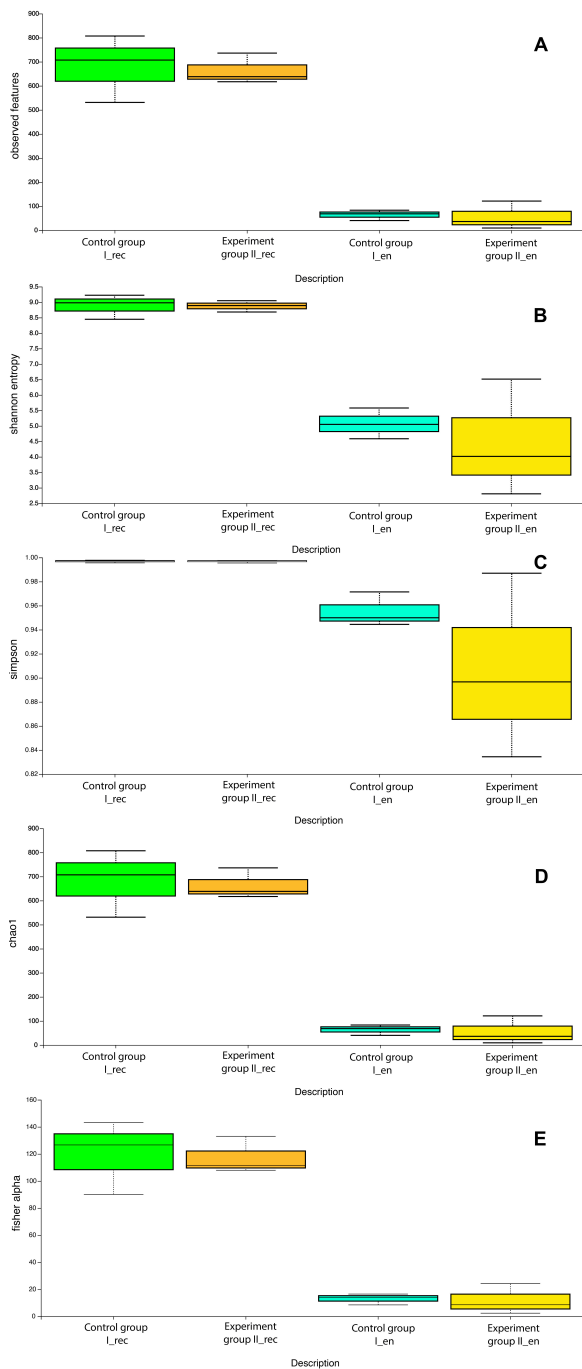


Fig. 1. Graphs of the microbial biodiversity distribution based on the NGS data for samples from the rectal chyme (_rec) and endometrium (_en) of dairy cows of the Holstein breed. Biodiversity indicators were as follows: (A) number of operational taxonomic units (OTUs), (B) Shannon entropy, (C) Simpson index, (D) Chao1 index, and (E) Fisher alpha index. The bottom of the horizontal bar represents the bottom value of the group, the top of the bar reflects the top value of the group, the dividing line within the bar conforms to the median, and the whiskers are the minimum and maximum values of the data. NGS, next-generation sequencing.

of the diet were studied for quality and safety. In particular, the formulation of the base diet in the second phase of the dry period included (in kg per cow per day): straw, 1.3; silage, 18.0; corn, 2.4; soybean meal, 0.2; rapeseed cake, 0.4; flattened barley, 1.5; beet pulp, 2.7; spent grain, 1.0; glycerin, 0.1; plus vitamin and mineral microsupplements. The composition of the base diet in the fresh calving period included (in kg per cow per day): hay, 0.9; silage, 27; wheat, 1.2; corn, 4.2; soybean meal, 1.3; rapeseed cake, 0.5; crushed barley, 1.6; beet pulp, 3.0; spent grain, 0.5; glycerin, 0.2; plus vitamin and mineral microsupplements. Daily monitoring of the correct loading of diet components, weekly assessment of dry matter intake and assessment of digestibility of diets were carried out.

2.2 Treatments

Based on the results of medical examination during the interlactation period (i.e., 21 days before calving), a total of 30 clinically healthy cows were selected and divided into two groups ($n = 15$ in each): control group (I), without the CFA (BIOTROF Ltd, St. Petersburg, Russia) introduction into the diet; and experiment group (II), with the CFA introduction into the diet at a dose of 50 g per animal per day during the entire transit period (i.e., 42 days). The CFA dosage was determined according to the manufacturer's instructions and recommendations, calculated earlier based on the initial concentration of active compounds, ease of administration when mixing in feed shops, cost of the additive, previous experience in developing similar supplements, and the results of preliminary experiments. In order to prevent endometritis and other possible pathological conditions, all animals from both groups, without exception, received the antibiotic Pen-Strep 400 (ImmCont GmbH, Eberswalde, Germany) based on benzylpenicillin-procaine and dihydrostreptomycin sulfate intramuscularly at the rate of 1 mL per 20 kg of animal weight once per day for 5 days, starting from the 5th day after calving. The animals did not receive any other drug or probiotic supplements.

The CFA included the following natural substances of antibacterial nature: the strain of probiotic bacteria *B. mucilaginosus* 159, a short-chain fatty acid, essential oils of plants eucalyptus and cinnamon, and wheat bran filler. The *B. mucilaginosus* 159 strain was obtained from the collection of BIOTROF LLC (St. Petersburg, Russia) and had been originally isolated from the chyme of the cow's rumen. The cells of the strain are motionless rods with rounded ends; it forms oval spores located subterminally and is not a genetically modified organism. The *B. mucilaginosus* 159 strain has no toxicogenic or virulent properties; the strain cell components are not toxic to laboratory animals [35]. The *B. mucilaginosus* 159 strain was cultured at 37 °C in a broth medium of the following composition: enzymatic peptone, pancreatic hydrolysate of fish meal, and sodium chloride, at $\text{pH} = 7.2 \pm 0.2$ (OOO Siana, Izhevsk, Russia).

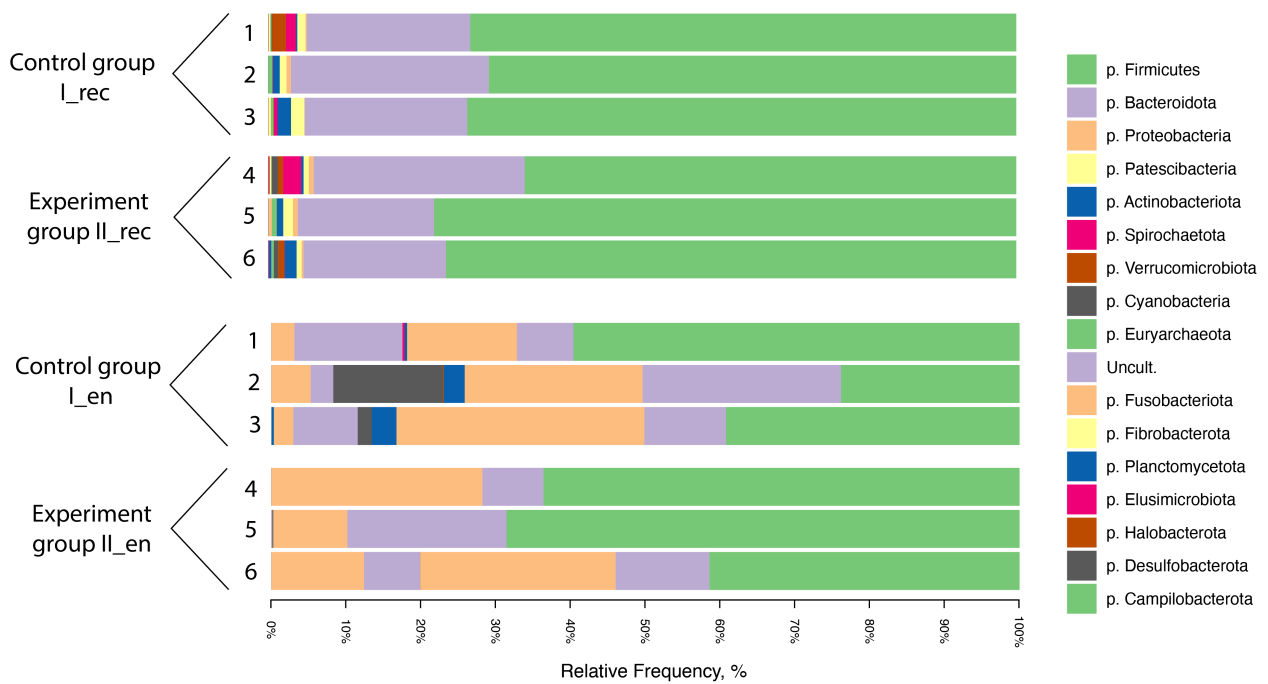


Fig. 2. Taxonomic composition of the microbiota at the phylum level based on the NGS data for samples from the rectal chyme (_rec) and endometrium (_en) of dairy cows of the Holstein breed. 1–6, animal IDs; p., phylum.; uncult., uncultured.

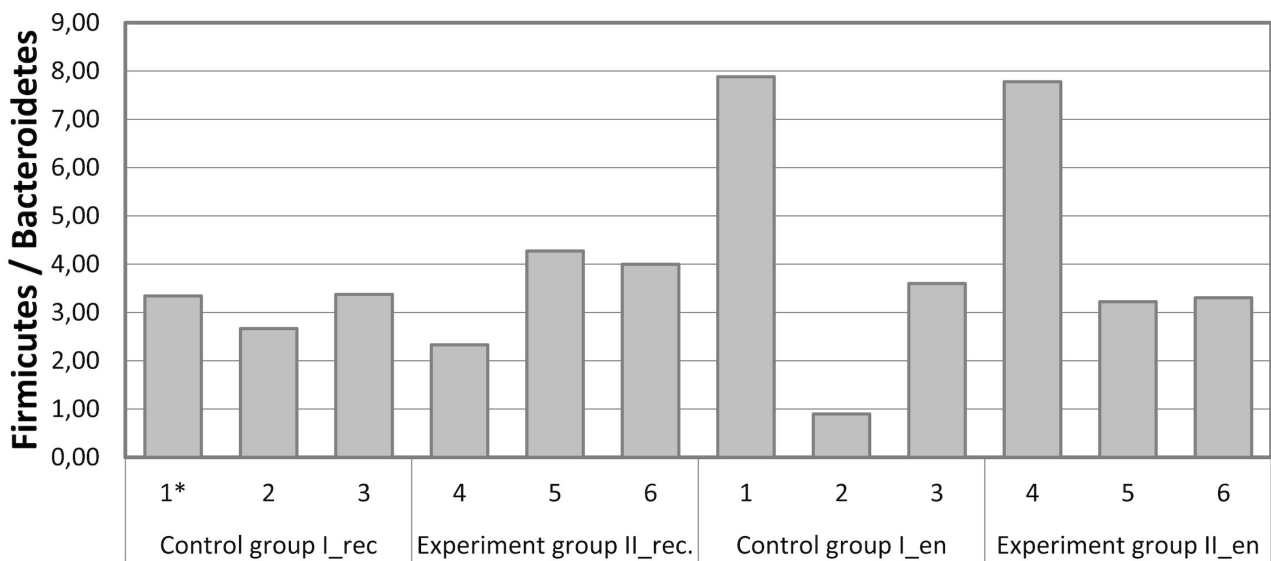


Fig. 3. The ratio of Firmicutes to Bacteroidetes in the microbiota at the phylum level based on the NGS data for samples from the rectal chyme (_rec) and endometrium (_en) of dairy cows of the Holstein breed. *1–6, animal IDs.

2.3 Samples

At the end of the experiment (i.e., on Day 21 after calving), endometrial and rectal chyme samples were taken from three clinically healthy cows from each group to analyze the composition and functions of the microbiota. Cows with abnormal calving, post-calving complications, or other clinical diseases before or during the study period were not included in the study. The collection of biological mate-

rial was carried out in compliance with aseptic conditions to ensure individuality and prevent cross-contamination of samples. The tail of the animals was covered with sterile gauze, and the perineum and vulva were washed with soap and water until completely clean, then treated with an antiseptic solution. Collection of scrapings from the surface of the endometrium was carried out using a cytobrush. Digestive chyme samples were collected directly from the rectum of each animal using disposable gloves. Overall,

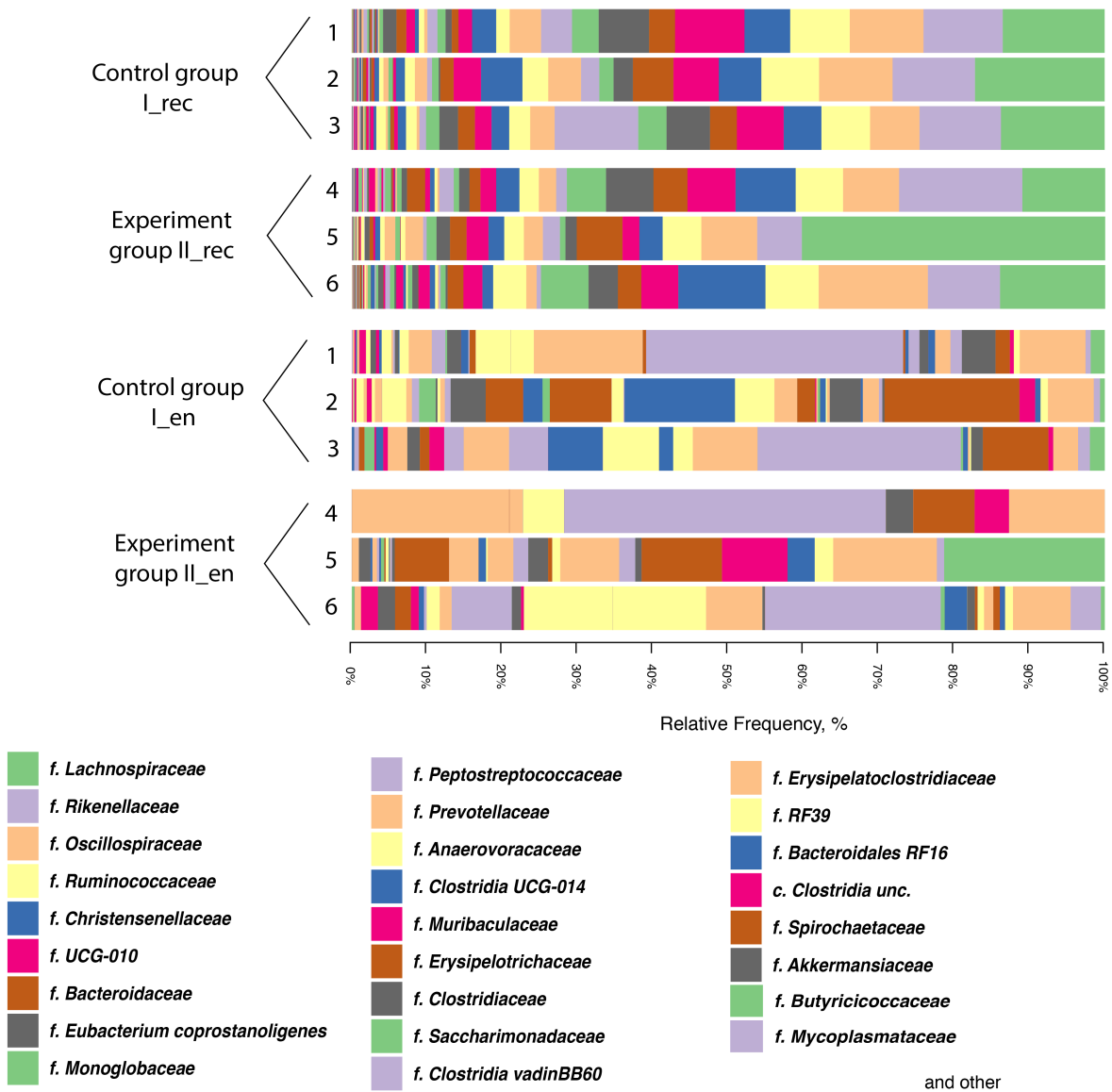


Fig. 4. Taxonomic composition of the microbiota at the family level based on the NGS data for samples from the rectal chyme (*_rec*) and endometrium (*_en*) of dairy cows of the Holstein breed. 1–6, animal IDs; f., family; c., class.

the sampling methods complied with the standards of humane treatment of animals. These methods were based on the use of a special veterinary probe and a specialist who was proficient in this technique. No painful manipulations were applied to the animals; therefore, the use of anesthesia was not required. A lubricative (vaseline oil), to facilitate the insertion of the probe, and an antiseptic were used as recommended and was successively washed off to prevent contamination.

Endometrial scraping samples for microbiota analysis were immediately frozen at $-20\text{ }^{\circ}\text{C}$ and sent to the laboratory for DNA extraction. Total DNA from the samples was isolated using the Genomic DNA Purification Kit (lot No. 10571031; Thermo Fisher Scientific, Inc., Carlsbad, CA, USA) according to the manufacturer's instructions.

2.4 Targeted Sequencing

The metagenomic composition of the microbial communities in scrapings from the cow endometrium and rectal chyme biotopes was assessed using the targeted next-generation sequencing (NGS). The latter was performed using the MiSeq platform (Illumina, Inc., San Diego, USA) and the following primers for the V3–V4 region of the 16S rRNA gene: forward, 5'-TCGTCGGCAGCGTCAG ATGTGTATAAGAGACAGCCTACGGGNGGCWGCAG-3'; reverse, 5'-GTCTCGTGGGCTCGGAGATGTGT ATAAGAGACAGGACTACHVGGGTATCTAATCC-3'. NGS was carried out using the respective reagents for preparation libraries (Nextera® XT Index Kit, lot No. FC-131-1001; Illumina), purification of qPCR products (Agen-court AMPure XP, lot No. A63881; Beckman Coulter Inc.,

Brea, CA, USA), and sequencing (MiSeq® Reagent Kit v2, 500 cycles, lot No. MS-102-2003; Illumina). Raw 16S rRNA sequence dataset was uploaded to the NCBI Sequence Read Archive (SRA; BioProject Number: PR-JNA1102864) [36].

2.5 Bioinformatics Procedures

The functional potential of the microbiota was assessed using the appropriate bioinformatics methods. In particular, bioinformatics analysis of NGS data was performed using a metagenomic analysis platform Quantitative Insights Into Microbial Ecology (QIIME2, Version 2022.2), version 2020.8 [37,38]. After the initial import of sequences into QIIME2 format, paired strings of reads were aligned. Sequences were then filtered by quality using the default settings. Noise sequences were filtered using the Deblur method. To construct a *de novo* phylogeny, the MAFFT software package, version 7 [39,40] was employed followed by the masked sequence alignment. The SILVA 138 reference database [41–43] was used for taxonomy analysis.

Based on the resultant table of operational taxonomic units (OTUs), biodiversity indices (Shannon, Simpson, Chao1, and Fisher alpha) were calculated using plugins of the QIIME2 software package [37,38]. For the statistical analysis of diversity indices, no additional transformation was performed. That is, no further standardization or scaling was applied to the calculated diversity data. The analysis result was presented in the respective graphs as actual values of diversity indices.

Reconstruction and prediction of the functional content of gene families and enzymes was carried out using the PICRUST2 software package, version 2.3.0 [44,45]. The MetaCyc database [46,47] was employed to analyze metabolic pathways and enzymes. Predicted MetaCyc metabolic pathway profiles were assessed by the amplicon sequence variants (ASV)-based abundance [48,49].

2.6 Statistical Analyses

Using Microsoft Excel XP/2003 (Microsoft Corporation, Redmond, WA, USA) and RStudio (version 1.1.453, Boston, MA, USA [50]), multivariate analysis of variance (multi-factor ANOVA) was used to process the results mathematically and statistically. The mean (M) and standard errors of the mean (\pm SEM) were displayed as the results. Student's *t*-test was used to determine the significance of the differences that were deemed statistically significant at $p < 0.05$. Tukey's Significant Difference Test (HSD) and the TukeyHSD function in the R Stats Package (version 4.5.0; [51]) were employed to compare mean values.

3. Results and Discussion

3.1 Biodiversity Analysis

As a result of the metagenomic NGS analysis, a total of 33,523 16S rRNA gene sequences were generated for the bovine intestinal chyme microbiome and 1367 for the endometrium microbiome. Bioinformatics data processing revealed differences ($p < 0.05$) in the number of OTUs, as well as the biodiversity indices (Shannon, Simpson, Chao1, and Fisher alpha) between the intestinal chyme of dairy cows and their endometrium (Fig. 1). Higher values of the studied indices in the intestinal chyme were indicative of a more diverse composition and less uniformity of the microbial population than in endometrial samples [52]. The Simpson index was significantly different ($p < 0.05$) between Group I and Group II for the endometrium (Fig. 1C), whereas other biodiversity indicators did not demonstrate significant between-groups differences for the endometrium microbial communities. Remarkably, no differences were found for intestinal chyme. This suggests that there might be no overall influence of CFA administered orally on the level of microbiota biodiversity within each of the two examined biotopes.

3.2 Structural Microbiome Examination

In the rectal chyme microbiota of the cows, 15 superphyla and phyla of microorganisms were discovered, and slightly less (12) in the endometrium (Fig. 2). Herewith, the phylum Firmicutes and superphylum Bacteroidota can be considered the dominating bacteria among both the rectum (respectively up to 73.4 and 26.4% in Group I and up to 77.8 and 28.2% in Group II) and endometrium (respectively up to 59.6 and 68.6% in Group I and up to 26.5 and 21.3% in Group II) biotopes. However, representatives of the phylum Proteobacteria in the endometrial microbiota were also represented in significant numbers (up to 33.1% in Group I and up to 28.2% in Group II). This was in contrast to the rectal chyme microbiota, where representatives of this phylum were detected in a trace amount not exceeding 0.64%.

Interestingly, the proportion of Firmicutes to Bacteroidetes both in the chyme of the digestive system and in the endometrium of cows and in both groups (Fig. 3) was quite high (up to 4.27 in the rectum and up to 7.9 in the endometrium). The exception was animal No. 2 in Group I, in whose endometrial microbiota this ratio was below 1. According to Miranda-CasoLuengo *et al.* [53], a low ratio of Firmicutes to Bacteroidetes in the reproductive system of cows was an early signal of the subsequent development of inflammatory diseases. Our data on the ratio of Firmicutes to Bacteroidetes, in general, coincided with the clinical observation of the absence of endometritis signs in the studied animals.

Another difference between the two studied biotopes was the established fact that the microbiota of the chyme of the rectum included representatives of the phyla Euryarchaeota, Patescibacteria and Fibrobacterota (Fig. 2) that



Fig. 5. Taxonomic composition of the microbiota at the genera level based on the NGS data for samples from the rectal chyme (_rec) and endometrium (_en) of dairy cows of the Holstein breed. 1–6, animal IDs; g., genus; f., family; unc., uncultured.

were not identified in the endometrium of cows. It is widely known that archaea are a specialized group of microorganisms including methanogenic euryarchaeota, one of the main habitats of which is the digestive system of ruminants [54,55]. Members of the superphylum Patescibacteria symbiotically interact with archaea, have been commonly found in anaerobic environments [56–58], including the digestive tract of ruminants [59], and are associated with anaerobic enzymatic metabolism [60]. Members of the superphy-

lum Fibrobacterota are cellulolytics in the herbivore gut and other cellulolytic ecosystems [61].

More pronounced changes ($p < 0.05$) under the influence of CFA were noted in the composition of the endometrial microbiota than in the intestinal chyme of cows. In particular, when administering CFA, representatives of the phylum Actinobacteriota were absent in the endometrium of cows of Group II, in contrast to Group I where their content ranged from 0.34 to 3.3%. According to some authors

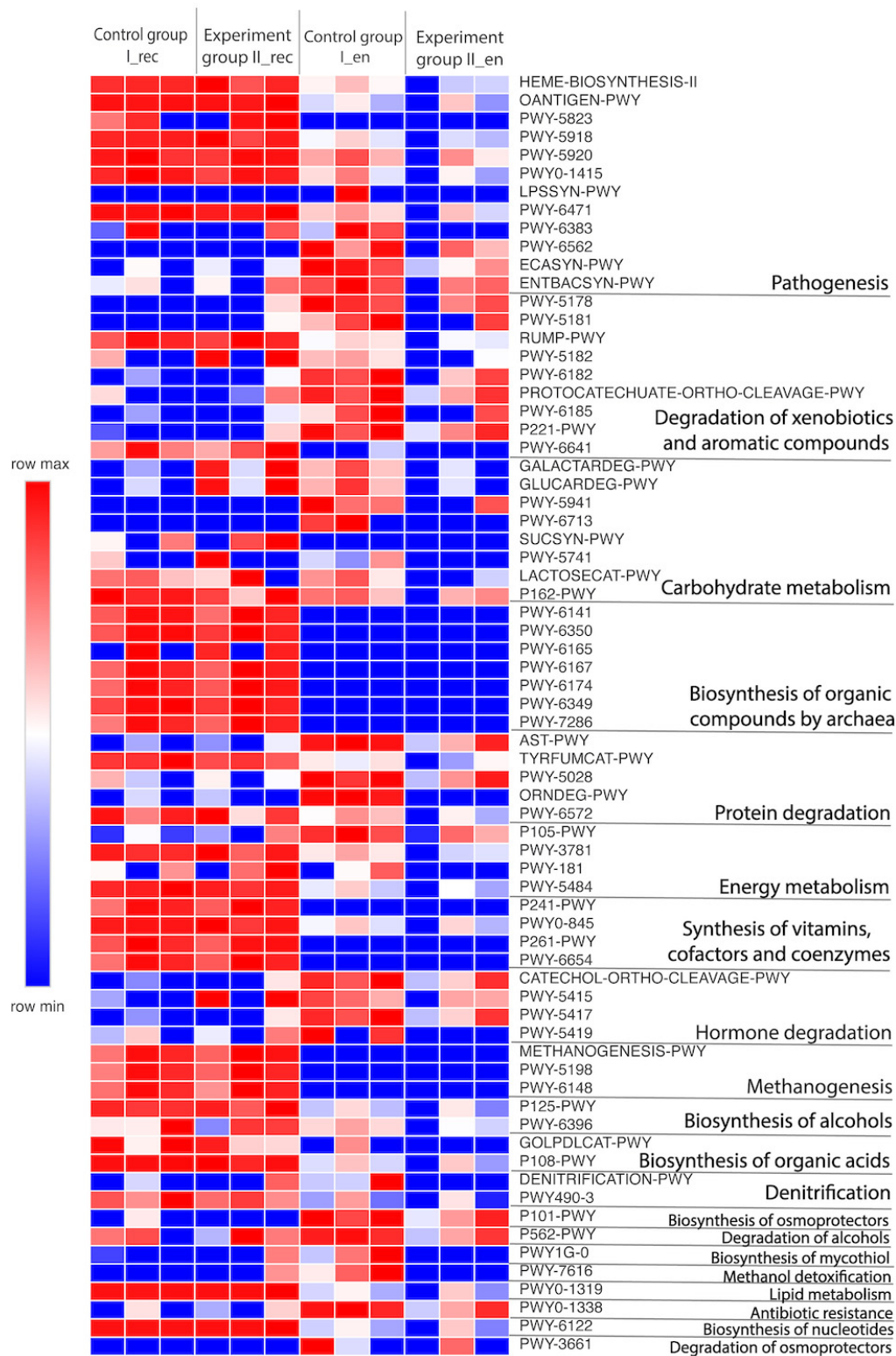


Fig. 6. Data on the functional annotation of the metabolic pathways based on the NGS data for samples from the rectal chyme (_rec) and endometrium (_en) of dairy cows of the Holstein breed as analyzed using the PICRUST2 software package.

[62], an increase in the proportion of actinomycetes in the reproductive system of cows is an important bacteriological risk factor for the occurrence of inflammatory diseases of the uterus. Despite the fact that the studied cows were clin-

ically healthy, an increased proportion of actinomycetes in the endometrium of Group I cows may indicate a possible risk of developing subclinical endometritis or other inflammatory diseases of the reproductive system. However, we

were unable to establish significant differences in the abundance of Actinobacteriota for the intestinal chyme of the studied cows.

Interestingly, in the endometrium of two Group I cows and two Group II cows, a significant presence (up to 47.2%) of the *Mycoplasmataceae* family bacteria was revealed (Fig. 4). The data presented were in agreement with the results obtained by Knudsen *et al.* [63] who concluded that *Mycoplasmataceae* was one of the most abundant families in uterine lavage samples from clinically healthy cows.

The content of the *Lactobacillaceae* family bacteria in the intestinal chyme and endometrium of cows in both groups was at a very low level: no more than 0.08% in the chyme and no more than 2.6% in the endometrium (Fig. 4). On the one hand, the low number of normal flora representatives of lactobacilli in the endometrium of cows could be a consequence of antibiotic therapy, since antibiotics suppress some representatives of the normobiota [64]. On the other hand, the importance of lactobacilli in the reproductive tract of cows remains an open question. Their clear positive role in maintaining homeostasis and dominance in the reproductive system in healthy individuals has been established only in humans [65]. Indeed, a previous study of the vaginal microbiota of cows has demonstrated a unique composition with a low content of lactobacilli and a nearly neutral pH [66]. Unlike humans, the presence of the genera *Enterococcus* and *Streptococcus*, with a minimal proportion of *Lactobacillus* spp., has been demonstrated for cows [66], which is presumably associated with a higher (almost neutral) pH level compared to humans [67]. It is still not clear, however, whether this pH level is caused by a difference in the genotype or whether it is a consequence of the widespread reproductive system dysbiosis of cows. Indeed, based on human experiments, a decrease in the content of lactobacilli is usually accompanied by an increase in pH [68] — a common symptom of bacterial vaginosis.

While analyzing the effects of CFA, it is worth noting that the content of representatives of *Porphyromonas* spp. decreased in the endometrium of Group II cows by 5.3 times compared to Group I ($p < 0.05$; Fig. 5). Despite the fact that the animals studied were clinically healthy, the presence of *Porphyromonas* spp., primarily the species *P. levii*, in the reproductive system of cows was previously associated with metritis [69] and necrotic vulvovaginitis in cattle [70]. It is interesting that these microorganisms were absent in the intestinal chyme of the cows we examined, with the exception of animal No. 2 in Group I. In the endometrium of the cows studied, we did not identify bacteria of the genus *Fusobacterium*, while one of them, *F. necrophorum*, is a well-known pathogen causing metritis [71], as well as deep necrotizing inflammation in cattle [72].

Thus, the endometrial microflora composition was more sensitive to the introduction of CFA into the diet than the intestinal chyme microbiome. On the one hand, this may be due to the fact that the endometrial microbiota is

characterized by a high level of response to changes in exogenous factors [73]. The composition of the reproductive system microbiota is extremely dynamic and varies significantly with age, genotype, and physiological factors such as monthly hormonal fluctuations and the immune system [74]. At the same time, the bacterial community of the intestinal chyme is more stable. Species-rich communities have been shown to be less susceptible to invasion because they use limited resources more efficiently, with different species specializing in a potentially limited resource [75]. It has been found that the gut microbiota often exhibits “colonization resistance”, i.e., the indigenous microbiota actively prevents the emergence of new potentially beneficial (probiotic) microorganisms [76]. On the other hand, changes in the composition of the gut microbiome under the influence of CFA could occur at the species level, affecting pathogenic forms present at relatively low concentrations, which is not always detectable by NGS sequencing. This could have a stimulating effect on the immune system, reduce the pathogen load, and have additional beneficial effects on endometrial health via the gut–vaginal axis.

Concerning possible adverse effects of the antibiotic treatment on the results in this experiment, the use of probiotic supplements along with antibiotic treatment is a fairly common practice. Concomitant use of biopreparations to restore the microbiota during antibiotic treatment helps normalize the microecological status of the body and increases the effectiveness of standard therapy. The chromosome of *Bacillus* spp. usually contains loci to control their resistance to antibiotics, including benzylpenicillin novocaine and dihydrostreptomycin sulfate [77]. In the case of intrinsic or acquired resistance (as a result of chromosomal mutation), the risk of transmitting antibiotic resistance genes is considered very low [78].

3.3 Functional Microbiome Analysis: Metabolic Pathways

To establish whether alterations in the microbiota composition of the intestinal chyme and endometrium in cows were associated with changes in its functionality, we reconstructed and predicted the functional content of the microbial community using the PICRUSt2 software package [44,45]. As a result, 380 predicted gut microbiome metabolic pathways and 370 endometrial microbiota pathways were annotated. In general, the level of the predicted functional potential of 70 pathways changed ($p < 0.05$) in cows depending on the experiment treatment variant or the studied biotope of the body, i.e., intestinal chyme and endometrium (Fig. 6).

When comparing the expression of the predicted metabolic pathways of the microbiome in different biotopes, it should be noted that some processes, such as methanogenesis (METHANOGENESIS-PWY, PWY-5198 and PWY-6148 pathways), were active only in the intestinal chyme, but not in the endometrium of cows. Indeed, methanogenesis is carried out by methanogenic ar-

chaea of the superphylum Euryarchaeota [79] that we did not identify in the endometrium of cows, unlike the digestive system. Therefore, it is natural that the biosynthetic pathways predicted in our study for the formation of archaetidylserine, archaetidylethanolamine, chorismite, flavin, mevalonate, cytidine diphosphate (CDP)-archaeol and 7-(3-amino-3-carboxypropyl)-wyosine, inherent in archaea, also showed activity only in the intestinal chyme, but not in the endometrium.

The most pronounced changes ($p < 0.05$) in the predicted metabolic potential of the microbiota under the influence of CFA were observed in the endometrial microbiota rather than in the intestinal chyme (Fig. 6), which echoed the data on the taxonomic composition of the microbiota (Figs. 2,3,4,5). For instance, it was demonstrated that the introduction of CFA into the diet of cows (Group II) produced a less pronounced effect in of predicted heme biosynthesis metabolic pathways (PWY-5918, PWY-5920, and PWY0-1415) in the endometrium from 22.8 to 42.3% compared to Group I ($p < 0.05$). This suggested a potential reduction in pathogenesis processes under the influence of CFA. The fact is that most pathogens have complete heme biosynthesis pathways [80], since heme is necessary for the functioning of hemoproteins involved in energy production by the electron transport chain, detoxification of host immune effectors and other pathogenesis processes [81].

It is known that in cows prone to inflammatory diseases of the uterus in the early postpartum period, there is a predominance of progesterone concentration in the blood serum, along with low estradiol levels [82]. On the other hand, postpartum uterine infections have previously been shown to disrupt the integrity of the endometrial epithelium by impairing the prostaglandin secretion, thereby compromising folliculogenesis and impairing fertility [83]. We established in the endometrium that the use of CFA (Group II) led to a less pronounced effect in of four predicted metabolic pathways (CATECHOL-ORTHO-CLEAVAGE-PWY, PWY-5415, PWY-5417, and PWY-5419) by 31–100% compared with Group I ($p < 0.05$). These pathways are related to the microbial degradation of catechol, an estrogen that is the main metabolite of estradiol, which increases the prostaglandin production in the body [84]. A previous *in vitro* study has demonstrated that some bacteria, such as *Pseudomonas* spp., are capable of metabolizing catechol to produce 2-hydroxymuconic semialdehyde transiently [85]. Catechol is also known to have antimicrobial properties against pathogens [86], i.e., it can increase the body's resistance to infectious diseases. Reproductive system dysbiosis has been repeatedly associated with endometritis, infertility, or adverse pregnancy outcomes in all mammalian species studied [87]. On the other hand, Carneiro *et al.* [88] have suggested that, in cows that develop endometritis, bacteria present in the uterus after calving may be both the etiological agents and secondary infections that developed due to reduced resistance. That is,

the observed change in the composition of the reproductive tract microbiota during the occurrence of inflammatory diseases can only be a consequence of alternations in hormonal status.

In addition, in the endometrium of all Group II cows, no activation of the predicted metabolic pathway PWY1G-0 (mycothiol biosynthesis) was observed. In contrast, we observed the activity of this pathway in the endometrium of all Group I animals. This was probably due to the absence of representatives of the phylum Actinobacteriota in the endometrium of Group II cows (in contrast to Group I), as mentioned above, since mycothiol is a thiol found only in the taxon Actinomycetales (representatives of the phylum Actinobacteriota) [89]. The main role of mycothiol is to maintain intracellular redox homeostasis; it acts as an electron acceptor/donor and serves as a cofactor in detoxification reactions of alkylating agents, free radicals and xenobiotics. In addition, like glutathione, mycothiol may be involved in catabolic processes that play an important role in growth when consuming recalcitrant chemicals such as aromatic compounds [90].

4. Conclusions

The results of our metagenomic study provide additional information on the mechanisms of the possible influence of biological feed additives administered orally during the transit period on the intestinal and endometrial microbiota of cows (after calving) that are subject to antibiotic therapy. Intravaginal use of agents that affect the reproductive system microbiota usually does not influence the intestinal microbiota that is important for the homeostasis and resistance of the body as a whole. This effect can be achieved through the application of oral probiotics, such as CFA. The use of CFA made it possible to confirm the concept of an interconnection between the intestinal and uterine microbiota, since we observed changes in the composition and predicted metabolic pathways of the endometrial microbiota after oral administration of CFA. Previous studies have also shown that oral probiotics can help restore the reproductive system microbiota [17,91,92].

The composition and predicted metabolic pathways of the endometrial microbiota turned out to be more responsive to the introduction of CFA than the intestinal chyme microbiome. This is likely due to the fact that the endometrial microbiota is characterized by a high level of responsiveness to changes in external conditions [74], while the intestinal chyme bacterial community is characterized by a greater stability.

Our data suggest that the integrity of the microbiome of cows is essential for good reproductive health. They emphasize the need for an integrated approach that combines the correction of microecological disorders simultaneously in the intestines and in the reproductive system.

Availability of Data and Materials

This article contains a presentation of all the experiments and findings from the study. Further details can be obtained upon request from the corresponding author.

Author Contributions

Conceptualization, EAY and GYL; methodology, EAY, VAF and LAI; software, EAB and ESP; validation, EAB, ESP and EAY; formal analysis, EAB, ESP, MNR, DKG and EAY; investigation, TSS, AVD, ASD, KAK, IAK, VAZ; resources, TSS; data curation, EAY, NIN and DGT; visualization, ESP and AVD; supervision, GYL and DKG; project administration, EAY, MNR and GYL; funding acquisition, EAY and VAF; writing—original draft preparation, EAY; writing—review and editing, EAY, MNR and DKG. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

The study received ethical approval from the Bioethical Commission of the L.K. Ernst Federal Research Center for Animal Husbandry (Protocol No. 2023-10/1, dated October 31, 2023).

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Conflict of Interest

The authors declare no conflict of interest. EAY, GYL, LAI, EAB, TSS, VAF, KAK and IAK are employees of BIOTROF+ Ltd. TSS is an employee of JSC Gatchinskoe. The judgments in data interpretation and writing were not influenced by this relationship.

References

- [1] Zhu Y, Sun G, Cidan Y, Shi B, Tan Z, Zhang J, *et al.* Comprehensive Multi-Omic Evaluation of the Microbiota and Metabolites in the Colons of Diverse Swine Breeds. *Animals: an Open Access Journal from MDPI.* 2024; 14: 1221.
- [2] Podobed LI, Buryakov NP, Laptev GYu, Kurdeko AP, Aleksandrov SN, Buryakova MA, *et al.* The Growing Calf from Birth to High Yielding Cows: Technology, Feed and Veterinary Aspects. Right Print: St. Petersburg. 2017.
- [3] Temesgen MY, Assen AA, Gizaw TT, Minalu BA, Mersha AY. Factors affecting calving to conception interval (days open) in dairy cows located at Dessie and Kombolcha towns, Ethiopia. *PloS One.* 2022; 17: e0264029.
- [4] Wang J, Li Z, Ma X, Du L, Jia Z, Cui X, *et al.* Translocation of vaginal microbiota is involved in impairment and protection of uterine health. *Nature Communications.* 2021; 12: 4191.
- [5] Ghallab RS, El-Karim DRSG, Fayed AH, Rashad AMA. Efficiency of conventional and nanoparticle oxytetracycline in treatment of clinical endometritis in postpartum dairy cows. *Tropical Animal Health and Production.* 2023; 55: 118.
- [6] Patangia DV, Anthony Ryan C, Dempsey E, Paul Ross R, Stanton C. Impact of antibiotics on the human microbiome and consequences for host health. *MicrobiologyOpen.* 2022; 11: e1260.
- [7] Adnane M, Chapwanya A. Role of Genital Tract Bacteria in Promoting Endometrial Health in Cattle. *Microorganisms.* 2022; 10: 2238.
- [8] Chen R, Li R, Qing W, Zhang Y, Zhou Z, Hou Y, *et al.* Probiotics are a good choice for the treatment of bacterial vaginosis: a meta-analysis of randomized controlled trial. *Reproductive Health.* 2022; 19: 137.
- [9] Romanov MN, Grozina AA, Ilina LA, Laptev GY, Yildirim EA, Filippova VA, *et al.* From feed regulation to regulated feeding: intestinal microbiome and performance optimization in broiler chickens in response to antibiotic and probiotic treatment. In *Life of Genomes 2022: Abstracts of the International Conference* (pp. 44–45). Research Center “Regulatory Genomics”, Institute of Fundamental Medicine and Biology, Kazan (Volga Region) Federal University: Kazan. 2022.
- [10] Samal L, Behura NC. Probiotics: an emerging nutritional approach for improving gut health of livestock and poultry. *Asian Journal of Animal and Veterinary Advances.* 2015; 10: 724–739.
- [11] Kochish II, Romanov MN, Posyabin SV, Myasnikova OV, Korenyuga MV, Motin MS. The effect of the prebiotic Vetelact on the gut microbiota of chickens of the parent herd. *Rossiyskiy Zhurnal Problemy Veterinarnoy Sanitarii, Gigiyeny i Ekologii.* 2021; 2: 152–156. (In Russian)
- [12] Kamra DN, Pathak NN. Improvement in livestock productivity by use of probiotics: A review. *The Indian Journal of Animal Sciences.* 2005; 75: 128–134.
- [13] Grozina AA, Ilina LA, Laptev GY, Yildirim EA, Ponomareva ES, Filippova VA, *et al.* Probiotics as an alternative to antibiotics in modulating the intestinal microbiota and performance of broiler chickens. *Journal of Applied Microbiology.* 2023; 134: lxad213.
- [14] Laptev G, Turina D, Yildirim E, Ilina L, Gorfunkel E, Filippova V, *et al.* Analysis of changes in broiler microbiome biodiversity parameters due to intake of glyphosate and probiotic *Bacillus* sp. GI-8 using next-generation sequencing. *Smart Innovation, Systems and Technologies.* 2023; 362: 161–170.
- [15] Khatlab IM, Abdel-Wahed AM, Khatlab AS, Anele UY, El-Keredy A, Zaher M. Effect of dietary probiotics supplementation on intake and production performance of ewes fed Atriplex hay-based diet. *Livestock Science.* 2020; 237: 104065.
- [16] Yildirim EA, Laptev GYu, Ilyina LA, Dunyashev TP, Tyurina DG, Filippova VA, *et al.* The influence of a dietary *Enterococcus faecium* strain-based additive on the taxonomic and functional characteristics of the rumen microbiota of lactating cows. *Sel'skokhozyaistvennaya Biologiya.* 2020; 55: 1204–1219. (In Russian)
- [17] Reid G, Charbonneau D, Erb J, Kochanowski B, Beuerman D, Poehner R, *et al.* Oral use of *Lactobacillus rhamnosus* GR-1 and *L. fermentum* RC-14 significantly alters vaginal flora: randomized, placebo-controlled trial in 64 healthy women. *FEMS Immunology and Medical Microbiology.* 2003; 35: 131–134.
- [18] Laptev GYu, Novikova NI, Yildirim EA. Microbiome of Farm Animals: Connection with Health and Productivity. *Prospekt Nauki: Saint-Petersburg.* 2020.
- [19] Laptev GYu, Novikova NI, Ilyina LA, Yyldyrym EA, Dumova VA, Korochkina EA. Research of vaginal bacterial slime matrix

- of high-yielding cows in postpartum period by Real-Time PCR-method. *Rossiiskij Veterinarnyj Zhurnal: Sel'skokožajstvennyye Životnye*. 2014; 3: 10–12. (In Russian)
- [20] Laptev GYu, Novikova NI, Ilina LA, Yildirim EA, Filippova VA, Korochkina EA. Phytobiotic Provitol for the treatment of cows diagnosed with endometritis. *Sel'skokhozyaystvennyye Vesti*. 2014; 4: 18–19. (In Russian)
- [21] Laptev G, Yildirim E, Ilina L, Ponomareva E, Kalitkina K, Turina D, *et al.* Effect of a probiotic strain administration in different feeding phases on α - and β -diversity and gene expression of the rumen microbiome in lactating cows. *Smart Innovation, Systems and Technologies*. 2023; 362: 181–191.
- [22] Yildirim EA, Ilina LA, Laptev GYu, Tyurina DG, Filippova VA, Dubrovin AV, *et al.* The search for sources of enterobacteria and clostridia endotoxins in Russian dairy farms: possible transfer of endotoxins through the feed–cow–milk chain. *Smart Innovation, Systems and Technologies*. 2024; 397: 325–335.
- [23] Bonilla J, Sobral PJA. Application of active films with natural extract for beef hamburger preservation. *Ciência Rural*. 2019; 49: e20180797. (In Portuguese)
- [24] Budri PE, Silva NCC, Bonsaglia ECR, Fernandes Júnior A, Araújo Júnior JP, Doyama JT, *et al.* Effect of essential oils of *Syzygium aromaticum* and *Cinnamomum zeylanicum* and their major components on biofilm production in *Staphylococcus aureus* strains isolated from milk of cows with mastitis. *Journal of Dairy Science*. 2015; 98: 5899–5904.
- [25] Amat S, Baines D, Timsit E, Hallewell J, Alexander TW. Essential oils inhibit the bovine respiratory pathogens *Mannheimia haemolytica*, *Pasteurella multocida* and *Histophilus somni* and have limited effects on commensal bacteria and turbinate cells in vitro. *Journal of Applied Microbiology*. 2019; 126: 1668–1682.
- [26] Pinedo PJ, Velez JS, Bothe H, Merchan D, Piñeiro JM, Risco CA. Effect of intrauterine infusion of an organic-certified product on uterine health, survival, and fertility of dairy cows with toxic puerperal metritis. *Journal of Dairy Science*. 2015; 98: 3120–3132.
- [27] Paiano RB, de Sousa RLM, Bonilla J, Moreno LZ, de Souza EDF, Baruselli PS, *et al.* In vitro effects of cinnamon, oregano, and thyme essential oils against *Escherichia coli* and *Trueperella pyogenes* isolated from dairy cows with clinical endometritis. *Theriogenology*. 2023; 196: 106–111.
- [28] Giorgino A, Raspa F, Valle E, Bergero D, Cavallini D, Gariglio M, *et al.* Effect of Dietary Organic Acids and Botanicals on Metabolic Status and Milk Parameters in Mid-Late Lactating Goats. *Animals: an Open Access Journal from MDPI*. 2023; 13: 797.
- [29] Mani-López E, García HS, López-Malo A. Organic acids as antimicrobials to control *Salmonella* in meat and poultry products. *Food Research International*. 2012; 45: 713–721.
- [30] Grilli E, Tugnoli B, Passey JL, Stahl CH, Piva A, Moeser AJ. Impact of dietary organic acids and botanicals on intestinal integrity and inflammation in weaned pigs. *BMC Veterinary Research*. 2015; 11: 96.
- [31] Qin R, Tian G, Liu J, Cao L. The gut microbiota and endometritis: From pathogenesis to diagnosis and treatment. *Frontiers in Cellular and Infection Microbiology*. 2022; 12: 1069557.
- [32] Herath S, Lilly ST, Fischer DP, Williams EJ, Dobson H, Bryant CE, *et al.* Bacterial lipopolysaccharide induces an endocrine switch from prostaglandin F2 α to prostaglandin E2 in bovine endometrium. *Endocrinology*. 2009; 150: 1912–1920.
- [33] Sotnikova ED, Vatnikov YA, Kulikov EV. Obstetric and gynecological clinical examination of breeding stock cattle. *Vestnik Rossijskogo universiteta družby narodov: Seria Agronomiâ i životnovodstvo*. 2014; 3: 60–68. (In Russian)
- [34] National Academies of Sciences, Engineering, and Medicine; Division on Earth and Life Studies; Board on Agriculture and Natural Resources; Committee on Nutrient Requirements of Dairy Cattle. *Nutrient Requirements of Dairy Cattle: Eighth Revised Edition*. 2021.
- [35] Methodological Guidelines of the USSR Ministry of Health MU 2620-82: Methodological Guidelines for the Hygienic Assessment of Microbial Plant Protection Products against Insects and Diseases Based on Non-spore-forming Microorganisms. USSR Ministry of Health: Kiev. 1982.
- [36] Biotrof+. Development of a new biotechnological approach to the prevention and treatment of endometritis in dairy cattle. BioProject Number: PRJNA1102864. NCBI Sequence Read Archive (SRA). 2024. Available at: <https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1102864> (Accessed: 14 July 2024).
- [37] QIIME 2 Development Team. QIIME 2 user documentation. Version: 2020.8. QIIME 2 Doc. 2016–2020. Available at: <https://docs.qiime2.org/2020.8/> (Accessed: 14 July 2024).
- [38] Fung C, Rusling M, Lampeter T, Love C, Karim A, Bongiorno C, *et al.* Automation of QIIME2 Metagenomic Analysis Platform. *Current Protocols*. 2021; 1: e254.
- [39] Katoh K. MAFFT version 7. Multiple alignment program for amino acid or nucleotide sequences. Computational Biology Research Consortium. 2013. Available at: <https://mafft.cbrc.jp/alignment/software/> (Accessed: 14 July 2024).
- [40] Katoh K, Rozewicki J, Yamada KD. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics*. 2019; 20: 1160–1166.
- [41] The SILVA Ribosomal RNA Database. Release information: SILVA 138 SSU. Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures GmbH. 2019. Available at: <https://www.arb-silva.de/documentation/release-138/> (Accessed: 14 July 2024).
- [42] Quast C, Priesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, *et al.* The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic Acids Research*. 2013; 41: D590–D596.
- [43] Dueholm MS, Andersen KS, McIlroy SJ, Kristensen JM, Yashiro E, Karst SM, *et al.* Generation of Comprehensive Ecosystem-Specific Reference Databases with Species-Level Resolution by High-Throughput Full-Length 16S rRNA Gene Sequencing and Automated Taxonomy Assignment (AutoTax). *mBio*. 2020; 11: e01557-20.
- [44] PICRUST 2.0 (Phylogenetic Investigation of Communities by Reconstruction of Unobserved States). The Huttenhower Lab, Department of Biostatistics, Harvard T.H. Chan School of Public Health. 2020. Available at: <https://huttenhower.sph.harvard.edu/picrust/> (Accessed: 14 July 2024).
- [45] Douglas GM, Maffei VJ, Zaneveld JR, Yurgel SN, Brown JR, Taylor CM, *et al.* PICRUST2 for prediction of metagenome functions. *Nature Biotechnology*. 2020; 38: 685–688.
- [46] MetaCyc Metabolic Pathway Database. SRI International. 2022. Available at: <https://metacyc.org/> (Accessed: 14 July 2024).
- [47] Krieger CJ, Zhang P, Mueller LA, Wang A, Paley S, Arnaud M, *et al.* MetaCyc: a multiorganism database of metabolic pathways and enzymes. *Nucleic Acids Research*. 2004; 32: D438–D442.
- [48] Joos L, Beirinckx S, Haegeman A, Debode J, Vandecasteele B, Baeyen S, *et al.* Daring to be differential: metabarcoding analysis of soil and plant-related microbial communities using amplicon sequence variants and operational taxonomical units. *BMC Genomics*. 2020; 21: 733.
- [49] Maruyama H, Masago A, Nambu T, Mashimo C, Okinaga T. Amplicon sequence variant-based oral microbiome analysis using QIIME 2. *Journal of Osaka Dental University*. 2020; 54: 273–281.
- [50] RStudio Team. RStudio: Integrated Development for R. Version 1.1.453. RStudio, Inc.: Boston. 2018.

- [51] RDocumentation. TukeyHSD. 2019. Available at: <https://www.rdocumentation.org/packages/stats/versions/3.6.1/topics/TukeyHSD> (Accessed: 14 July 2024)
- [52] Shitikov VK, Rozenberg GS. Biodiversity assessment: An attempt of formal generalization. In Rozenberg GS (ed.) *Quantitative Methods of Ecology and Hydrobiology* (pp. 91–129). Samara Scientific Center of the Russian Academy of Sciences: Tolyatti. 2005.
- [53] Miranda-CasoLuengo R, Lu J, Williams EJ, Miranda-CasoLuengo AA, Carrington SD, Evans ACO, *et al.* Delayed differentiation of vaginal and uterine microbiomes in dairy cows developing postpartum endometritis. *PLoS One*. 2019; 14: e0200974.
- [54] Murray RM, Bryant AM, Leng RA. Rates of production of methane in the rumen and large intestine of sheep. *The British Journal of Nutrition*. 1976; 36: 1–14.
- [55] Thauer RK, Kaster AK, Seedorf H, Buckel W, Hedderich R. Methanogenic archaea: ecologically relevant differences in energy conservation. *Nature Reviews. Microbiology*. 2008; 6: 579–591.
- [56] Sorokin DY, Makarova KS, Abbas B, Ferrer M, Golyshin PN, Galinski EA, *et al.* Discovery of extremely halophilic, methyl-reducing euryarchaea provides insights into the evolutionary origin of methanogenesis. *Nature Microbiology*. 2017; 2: 17081.
- [57] Kuroda K, Yamamoto K, Nakai R, Hirakata Y, Kubota K, Nobu MK, *et al.* Symbiosis between *Candidatus* Patescibacteria and Archaea Discovered in Wastewater-Treating Bioreactors. *mBio*. 2022; 13: e0171122.
- [58] Vigneron A, Cruaud P, Guyoneaud R, Goñi-Urriza M. Into the darkness of the microbial dark matter *in situ* activities through expression profiles of *Patescibacteria* populations. *Frontiers in Microbiology*. 2023; 13: 1073483.
- [59] Qiu Q, Gao C, Aziz Ur Rahman M, Cao B, Su H. Digestive Ability, Physiological Characteristics, and Rumen Bacterial Community of Holstein Finishing Steers in Response to Three Nutrient Density Diets as Fattening Phases Advanced. *Microorganisms*. 2020; 8: 335.
- [60] Herrmann M, Wegner CE, Taubert M, Geesink P, Lehmann K, Yan L, *et al.* Predominance of *Cand.* Patescibacteria in Groundwater Is Caused by Their Preferential Mobilization From Soils and Flourishing Under Oligotrophic Conditions. *Frontiers in Microbiology*. 2019; 10: 1407.
- [61] Abdul Rahman N, Parks DH, Vanwonterghem I, Morrison M, Tyson GW, Hugenholtz P. A Phylogenomic Analysis of the Bacterial Phylum Fibrobacteres. *Frontiers in Microbiology*. 2016; 6: 1469.
- [62] Prunner I, Wagener K, Pothmann H, Ehling-Schulz M, Drillich M. Risk factors for uterine diseases on small- and medium-sized dairy farms determined by clinical, bacteriological, and cytological examinations. *Theriogenology*. 2014; 82: 857–865.
- [63] Knudsen LRV, Karstrup CC, Pedersen HG, Angen Ø, Agerholm JS, Rasmussen EL, *et al.* An investigation of the microbiota in uterine flush samples and endometrial biopsies from dairy cows during the first 7 weeks postpartum. *Theriogenology*. 2016; 86: 642–650.
- [64] Redondo-Lopez V, Cook RL, Sobel JD. Emerging role of lactobacilli in the control and maintenance of the vaginal bacterial microflora. *Reviews of Infectious Diseases*. 1990; 12: 856–872.
- [65] Gholfiof M, Adamson-De Luca E, Wessels JM. The female reproductive tract microbiotas, inflammation, and gynecological conditions. *Frontiers in Reproductive Health*. 2022; 4: 963752.
- [66] Swartz JD, Lachman M, Westveer K, O'Neill T, Geary T, Kott RW, *et al.* Characterization of the Vaginal Microbiota of Ewes and Cows Reveals a Unique Microbiota with Low Levels of Lactobacilli and Near-Neutral pH. *Frontiers in Veterinary Science*. 2014; 1: 19.
- [67] Appiah MO, Wang J, Lu W. Microflora in the reproductive tract of cattle: a review. *Agriculture*. 2020; 10: 232.
- [68] Zangl I, Pap IJ, Aspöck C, Schüller C. The role of *Lactobacillus* species in the control of *Candida* via biotrophic interactions. *Microbial Cell (Graz, Austria)*. 2019; 7: 1–14.
- [69] Santos TMA, Gilbert RO, Bicalho RC. Metagenomic analysis of the uterine bacterial microbiota in healthy and metritic postpartum dairy cows. *Journal of Dairy Science*. 2011; 94: 291–302.
- [70] Elad D, Friedgut O, Alpert N, Stram Y, Lahav D, Tiomkin D, *et al.* Bovine necrotic vulvovaginitis associated with *Porphyromonas levii*. *Emerging Infectious Diseases*. 2004; 10: 505–507.
- [71] Bicalho MLS, Machado VS, Oikonomou G, Gilbert RO, Bicalho RC. Association between virulence factors of *Escherichia coli*, *Fusobacterium necrophorum*, and *Arcanobacterium pyogenes* and uterine diseases of dairy cows. *Veterinary Microbiology*. 2012; 157: 125–131.
- [72] Nagaraja TG, Narayanan SK, Stewart GC, Chengappa MM. *Fusobacterium necrophorum* infections in animals: pathogenesis and pathogenic mechanisms. *Anaerobe*. 2005; 11: 239–246.
- [73] Eschenbach DA, Thwin SS, Patton DL, Hooton TM, Stapleton AE, Agnew K, *et al.* Influence of the normal menstrual cycle on vaginal tissue, discharge, and microflora. *Clinical Infectious Diseases: an Official Publication of the Infectious Diseases Society of America*. 2000; 30: 901–907.
- [74] Lehtoranta L, Ala-Jaakkola R, Laitila A, Maukonen J. Healthy Vaginal Microbiota and Influence of Probiotics Across the Female Life Span. *Frontiers in Microbiology*. 2022; 13: 819958.
- [75] Levine JM, D'antonio CM. Elton revisited: a review of evidence linking diversity and invasibility. *Oikos*. 1999; 87: 15–26.
- [76] McNulty NP, Yatsunenko T, Hsiao A, Faith JJ, Muegge BD, Goodman AL, *et al.* The impact of a consortium of fermented milk strains on the gut microbiome of gnotobiotic mice and monozygotic twins. *Science Translational Medicine*. 2011; 3: 106ra106.
- [77] Staal SP, Hoch JA. Conditional dihydrostreptomycin resistance in *Bacillus subtilis*. *Journal of Bacteriology*. 1972; 110: 202–207.
- [78] Gueimonde M, Sánchez B, G de Los Reyes-Gavilán C, Margolles A. Antibiotic resistance in probiotic bacteria. *Frontiers in Microbiology*. 2013; 4: 202.
- [79] Berghuis BA, Yu FB, Schulz F, Blainey PC, Woyke T, Quake SR. Hydrogenotrophic methanogenesis in archaeal phylum *Verstraetearchaeota* reveals the shared ancestry of all methanogens. *Proceedings of the National Academy of Sciences of the United States of America*. 2019; 116: 5037–5044.
- [80] Choby JE, Skaar EP. Heme Synthesis and Acquisition in Bacterial Pathogens. *Journal of Molecular Biology*. 2016; 428: 3408–3428.
- [81] Hederstedt L. Heme A biosynthesis. *Biochimica et Biophysica Acta*. 2012; 1817: 920–927.
- [82] Sugiura T, Akiyoshi S, Inoue F, Yanagawa Y, Moriyoshi M, Tajima M, *et al.* Relationship between bovine endometrial thickness and plasma progesterone and estradiol concentrations in natural and induced estrus. *The Journal of Reproduction and Development*. 2018; 64: 135–143.
- [83] Sheldon IM, Price SB, Cronin J, Gilbert RO, Gadsby JE. Mechanisms of infertility associated with clinical and subclinical endometritis in high producing dairy cattle. *Reproduction in Domestic Animals = Zuchthygiene*. 2009; 44: 1–9.
- [84] Pakrasi PL, Dey SK. Catechol estrogens stimulate synthesis of prostaglandins in the preimplantation rabbit blastocyst and endometrium. *Biology of Reproduction*. 1983; 29: 347–354.
- [85] Dagley S, Gibson DT. The bacterial degradation of catechol. *The Biochemical Journal*. 1965; 95: 466–474.
- [86] Razaviamri S, Wang K, Liu B, Lee BP. Catechol-Based An-

- timicrobial Polymers. *Molecules* (Basel, Switzerland). 2021; 26: 559.
- [87] Garcia-Garcia RM, Arias-Álvarez M, Jordán-Rodríguez D, Rebolgar PG, Lorenzo PL, Herranz C, *et al.* Female reproduction and the microbiota in mammals: Where are we? *Theriogenology*. 2022; 194: 144–153.
- [88] Carneiro LC, Cronin JG, Sheldon IM. Mechanisms linking bacterial infections of the bovine endometrium to disease and infertility. *Reproductive Biology*. 2016; 16: 1–7.
- [89] Spies HS, Steenkamp DJ. Thiols of intracellular pathogens. Identification of ovothiol A in *Leishmania donovani* and structural analysis of a novel thiol from *Mycobacterium bovis*. *European Journal of Biochemistry*. 1994; 224: 203–213.
- [90] Rawat M, Av-Gay Y. Mycothiol-dependent proteins in actinomycetes. *FEMS Microbiology Reviews*. 2007; 31: 278–292.
- [91] Anukam K, Osazuwa E, Ahonkhai I, Ngwu M, Osemene G, Bruce AW, *et al.* Augmentation of antimicrobial metronidazole therapy of bacterial vaginosis with oral probiotic *Lactobacillus rhamnosus* GR-1 and *Lactobacillus reuteri* RC-14: randomized, double-blind, placebo controlled trial. *Microbes and Infection*. 2006; 8: 1450–1454.
- [92] Vujic G, Jajac Knez A, Despot Stefanovic V, Kuzmic Vrbanovic V. Efficacy of orally applied probiotic capsules for bacterial vaginosis and other vaginal infections: a double-blind, randomized, placebo-controlled study. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*. 2013; 168: 75–79.