










Original Research

Serum lncRNA HOTAIRM1, but not miR-182-5p, as a Potential Non-Invasive Biomarker for Endometrial Cancer

Hevra Ekin Ulusoy^{1,*}, Yagmur Minareci¹, Canan Kucukgergin²,
Abdurrahman Fatih Aydin², Hamdullah Sozen¹, Mehtap Akin Efe¹,
Atacem Mert Aytakin¹, Mehmet Yavuz Salihoglu¹, Samet Topuz¹¹Department of Obstetrics and Gynecology, Istanbul Faculty of Medicine, Istanbul University, 34093 Istanbul, Türkiye²Department of Biochemistry, Istanbul Faculty of Medicine, Istanbul University, 34093 Istanbul, Türkiye*Correspondence: hevra.arpaci@istanbul.edu.tr (Hevra Ekin Ulusoy)

Academic Editor: Michael H. Dahan

Submitted: 14 September 2025 Revised: 17 November 2025 Accepted: 11 December 2025 Published: 11 February 2026

Abstract

Background: Endometrial cancer (EC) is the second most common gynecological malignancy and remains a major contributor to morbidity and mortality among women worldwide. Current diagnostic and prognostic tools have limited accuracy, highlighting the need for reliable, non-invasive biomarkers. Non-coding RNAs (ncRNAs), including long non-coding RNAs (lncRNAs) and microRNAs (miRNAs), have emerged as promising candidates. This study aimed to investigate serum lncRNA HOXA transcript antisense RNA myeloid-specific 1 (HOTAIRM1) and miR-182-5p expression levels in EC and evaluate their diagnostic value and association with clinicopathological features. **Methods:** This prospective cohort study analyzed serum samples that were prospectively collected from 73 women with histologically confirmed EC and 76 controls with non-malignant gynecological conditions. Quantitative real-time polymerase chain reaction (qRT-PCR) was used to quantify serum lncRNA HOTAIRM1 and miR-182-5p expression levels. Associations with clinicopathological parameters and diagnostic performance were evaluated. **Results:** Serum expression levels of lncRNA HOTAIRM1 were significantly higher in EC patients (median [interquartile range (IQR)]: 1.36 [0.74–3.37]) than in controls (1.03 [0.58–2.03]; $p = 0.043$), whereas miR-182-5p levels showed no significant difference ($p = 0.327$). Receiver operating characteristic (ROC) curve analysis demonstrated that serum lncRNA HOTAIRM1 had high specificity (93.2%) but low sensitivity (29.6%) for EC diagnosis. No significant associations were identified between either biomarker or clinicopathological variables ($p > 0.05$). **Conclusions:** This study provides one of the first evaluations of circulating lncRNA HOTAIRM1 and miR-182-5p in serum samples from patients with EC. Our findings suggest that serum lncRNA HOTAIRM1, but not miR-182-5p, may serve as a potential non-invasive biomarker for EC. Although its limited sensitivity restricts its role as a standalone diagnostic tool, its high specificity supports potential clinical relevance. These preliminary results should be interpreted with caution, as they represent an initial exploratory analysis of circulating lncRNA HOTAIRM1 and miR-182-5p in EC.

Keywords: endometrial cancer; lncRNA HOTAIRM1; miR-182-5p; non-coding RNAs; serum; biomarkers; diagnosis; prognosis

1. Introduction

Endometrial cancer (EC) is the second most common gynecological malignancy and the fourth most frequently diagnosed cancer among women worldwide, representing a significant public health concern with notable morbidity, mortality, and socioeconomic burden [1]. Despite advances in imaging and histopathology, there are currently no validated, non-invasive biomarkers in clinical practice for early diagnosis, post-treatment monitoring, or prognostic assessment of EC.

Non-coding RNAs (ncRNAs), transcribed from approximately 98% of the human genome that do not encode proteins, have emerged as critical regulators of gene expression. They are broadly classified into long non-coding RNAs (lncRNAs, >200 nucleotides) and small non-coding RNAs (sncRNAs, including microRNAs [miRNAs], ~22 nucleotides). Both lncRNAs and miRNAs modulate gene expression at transcriptional and post-transcriptional lev-

els and are increasingly recognized as key players in tumorigenesis, cancer progression, and metastasis, making them promising candidates for diagnostic and prognostic biomarkers, as well as therapeutic targets [2–11].

miRNAs implicated in cancer pathogenesis are generally categorized as oncogenes or tumor suppressors. Oncogenic miRNAs are overexpressed in tumor cells, whereas tumor suppressor miRNAs show reduced expression. More than 2000 miRNAs have been identified in humans, with 754 reported as potential biomarkers in EC in the past five years [12–14]. Several of these miRNAs are associated with key clinical features such as lymph node (LN) involvement, disease stage, metastatic status, and histological subtype [12–14]. Among these, miR-182-5p has shown variable expression across cancer types, functioning in key signaling pathways involved in carcinogenesis, tumor progression, metastasis, and treatment response in gynecological cancers [15]. In EC, miR-182-5p has been identified as an oncogenic miRNA [15,16].



Similarly, lncRNAs have been shown to regulate mRNAs, miRNAs, and proteins through various signaling pathways, thereby influencing tumor initiation, progression, cell differentiation, cell cycle regulation, and apoptosis [10]. The role of lncRNAs in EC has been highlighted in several studies. Among them, HOXA transcript antisense RNA myeloid-specific 1 (HOTAIRM1) has emerged as a cancer-related lncRNA that regulates *HOXA1* gene expression and promotes cell proliferation, migration, and invasion in type I EC cells [17,18]. Its expression, along with *HOXA1*, is significantly upregulated in type I EC tissues and correlates with FIGO (The International Federation of Gynecology and Obstetrics) stage and LN metastasis [17,18].

The interplay between lncRNAs and miRNAs—often via a “molecular sponge” mechanism—can disrupt tumor suppressor miRNA activity, leading to dysregulation of downstream mRNA targets and contributing to EC progression [19].

Previous studies have reported elevated tissue expression of both lncRNA HOTAIRM1 and miR-182-5p in EC, yet their serum levels remain largely unexplored [20,21]. The primary objective of this study was to evaluate the serum expression levels of lncRNA HOTAIRM1 and miR-182-5p as potential non-invasive diagnostic biomarkers for distinguishing EC from non-malignant gynecological conditions. The secondary objectives were to investigate their associations with key clinicopathological features and to explore potential correlations between lncRNA HOTAIRM1 and miR-182-5p expression levels.

2. Materials and Methods

2.1 Study Design and Ethical Approval

This prospective cohort study utilized serum samples that were prospectively collected from 73 patients with histologically confirmed EC and 76 controls with benign gynecological conditions. The study was conducted at the Department of Obstetrics and Gynecology, Division of Gynecologic Oncology, Istanbul Faculty of Medicine, Istanbul University, in collaboration with the Department of Biochemistry. Patient enrollment and serum collection took place between 2018 and 2019. The initial approval from the Institutional Review Board and Ethics Committee was obtained in 2018 (Approval No: 2018/1609). As the scientific scope of the research expanded to include additional ncRNAs, further approval was granted in 2023 (Approval No: 2023/119), supported by the Istanbul University Scientific Research Projects Unit (Project ID: TTU-2023-39847). This study was initiated prior to the requirement for registration of prospective cohort studies. All procedures were conducted in accordance with the Declaration of Helsinki, and written informed consent obtained at baseline included permission for long-term storage and future use of biospecimens.

2.2 Study Population

The study population included women with histologically confirmed EC, all diagnosed by endometrial biopsy and subsequently managed surgically. The control group consisted of women who presented to the general gynecology outpatient clinic for routine gynecological evaluation due to non-malignant indications and were confirmed to have no evidence of uterine or adnexal pathology on clinical, ultrasonographic, or laboratory assessment. To minimize potential confounding factors, women with uterine (cervical, myometrial, or endometrial) and/or ovarian/tubal pathologies—including benign gynecologic conditions such as uterine fibroids, ovarian cysts, endometrial polyps, and endometrial hyperplasia—were excluded. In particular, patients diagnosed with atypical and/or complex endometrial hyperplasia were explicitly excluded to avoid potential overlap with premalignant endometrial conditions. Exclusion criteria for the EC group included final histopathology not confirming EC, the presence of synchronous malignancies, recurrent disease, or prior oncology treatment (chemotherapy, radiotherapy, or immunotherapy).

All patients with EC were routinely followed up every three months after their initial surgery according to institutional protocol. Each follow-up visit included gynecological examination, transvaginal ultrasonography, and magnetic resonance imaging (MRI) to evaluate possible disease recurrence. Recurrence was defined as radiologically or histopathologically confirmed evidence of disease reappearance after initial treatment. In cases where MRI findings were unequivocally consistent with recurrent disease, diagnosis and treatment planning were based on radiological confirmation. For patients with indeterminate or atypical radiological or clinical findings, histopathological verification was obtained before initiating treatment. For recurrence-related analyses, only patients with complete recurrence data were included. Two patients had incomplete follow-up; their recurrence status was recorded as missing. We did not perform any imputation for missing data. A complete-case (available-case) analysis was therefore used for recurrence outcomes.

Both EC patients and controls were recruited consecutively during the predefined study period. No prior sample size calculation was performed; however, based on the total sample size, a post hoc power analysis was conducted using the G*Power statistical software (version 3.1.9.7; Heinrich-Heine-Universität Düsseldorf, Düsseldorf, Germany). For a two-tailed *t*-test with an alpha level of 0.05 and a moderate effect size ($d = 0.5$), the calculated statistical power of the study was 88%, indicating adequate power to detect clinically meaningful differences between groups. A *p*-value < 0.05 was considered statistically significant. This approach allows a post hoc assessment of whether the sample size was sufficient for the observed effect size.

2.3 Sample Collection and RNA Extraction

Venous blood samples were obtained from patients with EC at the time of diagnosis and from controls during outpatient visits. All samples were transferred to the Department of Biochemistry within 15 minutes of collection, centrifuged, aliquoted, and immediately stored at -80°C in RNase-free tubes to avoid repeated freeze–thaw cycles and minimize RNA degradation. The storage duration before RNA extraction ranged from 38 to 50 months. All EC and control samples were processed and analyzed simultaneously to minimize batch effects. Previous studies have demonstrated that circulating miRNAs remain stable for years when stored at ultra-low temperatures [22–24], and current biospecimen practice guidelines indicate that serum or plasma aliquots stored at $\leq -80^{\circ}\text{C}$ are suitable for cf-miRNA analysis over extended periods [25]. Because all specimens were collected, processed, and stored under identical conditions, any potential effect of storage duration on RNA integrity was expected to be consistent across groups. Total RNA was extracted from serum using the miRNeasy Serum/Plasma Advanced Kit (Cat. no. 217204; Qiagen, Germantown, MD, USA) according to the manufacturer's instructions. Complementary DNA (cDNA) synthesis was performed using the miRCURY LNA™ RT Kit (Cat. no. 339346; Qiagen, Germantown, MD, USA) and the RT² PreAMP cDNA Synthesis Kit (Cat. no. 330451; Qiagen, Germantown, MD, USA).

2.4 Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Expression levels of miR-182-5p and lncRNA HOTAIRM1 were quantified using a Rotor-Gene Q real-time PCR system (Qiagen, Germantown, MD, USA). Primers were purchased from Qiagen Technologies (Germantown, MD, USA): miRCURY LNA™ miRNA PCR Assay U6 (Cat. no. YP02119464), miRCURY LNA™ miRNA PCR Assay hsa-miR-182-5p (Cat. no. YP00206070), RT² lncRNA qPCR Assay (HOTAIRM1) (Cat. no. 330701LPH10483A), and RT² lncRNA qPCR Assay (GAPDH) (Cat. no. 330001PPH00150F). Relative expression levels were calculated using the $2^{-\Delta\Delta\text{CT}}$ method, normalizing to U6 (for miRNA) and GAPDH (for lncRNA). All qRT-PCR reactions were performed in triplicate (technical replicates) using the same instrument and reagent lots to ensure reproducibility. Negative (no-template) controls were included in every run. The average coefficient of variation (CV) for cycle threshold (Ct) values among replicates was below 2%, indicating high intra-assay precision. Moreover, all patient and control samples were analyzed within a single batch under identical reaction conditions to eliminate inter-assay variability.

2.5 Statistical Analysis

Data analysis was performed using NCSS (Number Cruncher Statistical System) 2007 software (Kaysville, UT, USA). Descriptive statistics were expressed as mean \pm SD,

median (interquartile range [IQR]), frequency, percentage, minimum, and maximum values. The normality of continuous variables was assessed using the Shapiro-Wilk test. Data with a normal distribution were presented as mean \pm SD, while non-normally distributed data were expressed as median (IQR). For comparisons between two groups, the Student's *t*-test or Mann-Whitney U test was used, depending on data distribution. The Shapiro-Wilk test revealed that serum lncRNA HOTAIRM1 and miR-182-5p expression levels were not normally distributed ($p < 0.05$); therefore, comparisons between groups were performed using the Mann-Whitney U test. For comparisons involving more than two groups, the one-way ANOVA or Kruskal-Wallis test was applied, followed by appropriate post-hoc analyses when significant differences were found. Associations between categorical variables were examined using the chi-square test or Fisher's exact test, as appropriate. Correlations between quantitative variables were analyzed using Spearman's rank correlation coefficient. Receiver operating characteristic (ROC) curve analysis was performed to evaluate diagnostic performance, including the determination of optimal sensitivity, specificity, and cut-off values. The optimal cut-off for serum lncRNA HOTAIRM1 was derived using the Youden index ($J = \text{sensitivity} + \text{specificity} - 1$), which identifies the maximum point of sensitivity and specificity on the ROC curve. Survival analyses were conducted using the Kaplan–Meier method, and all statistical tests were two-sided with a p -value < 0.05 considered statistically significant.

3. Results

3.1 Clinical and Pathological Characteristics of Patients With EC and Control Subjects

A total of 73 patients with EC and 76 control subjects were included in the study. Age (mean \pm SD: 57.84 ± 11.19 vs. 55.44 ± 12.09 years, $p = 0.171$) and body mass index (BMI) (33.20 ± 6.64 vs. 33.75 ± 6.38 kg/m², $p = 0.563$) did not differ significantly between groups (Table 1). Similarly, no significant differences were observed regarding obesity ($p = 0.841$), smoking status ($p = 0.210$), hypertension ($p = 0.124$), or diabetes mellitus ($p = 0.099$), indicating that the groups were demographically comparable (Table 1).

Among patients with EC ($n = 73$), preoperative serum CA-125 levels ranged from 5 to 745 U/mL (mean \pm SD: 42.44 ± 131.34), with elevated values detected in 4 patients (5.5%). The majority were postmenopausal ($n = 58$, 79.5%) and parous ($n = 55$, 75.3%). According to the FIGO classification, 58 patients (79.5%) had early-stage disease (stages IA–IB), whereas 15 (20.5%) presented with advanced-stage disease (stages II–IV). Histologically, 58 patients (79.5%) had low-grade endometrioid adenocarcinoma (grades 1–2), and 15 (20.5%) had high-grade tumors (grade 3 endometrioid or non-endometrioid), including 9 grade 3 endometrioid adenocarcinomas, 2 serous carcinomas, 1 mixed serous and clear cell carcinoma, and 3 carcinosarcomas.

Table 1. Comparison of demographic and clinical characteristics between patients with EC and control subjects.

Variable	EC Patients (n = 73)	Controls (n = 76)	p-value
	Mean ± SD	Mean ± SD	
Age (years)	57.84 ± 11.19	55.44 ± 12.09	0.171 (ns)
BMI (kg/m ²)	33.20 ± 6.64	33.75 ± 6.38	0.563 (ns)
	n (%)	n (%)	
Obesity			0.841 (ns)
Non-obese (BMI <30 kg/m ²)	21 (28.8)	23 (30.3)	
Obese (BMI ≥30 kg/m ²)	52 (71.2)	53 (69.7)	
Smoking status			0.210 (ns)
Non-smoker	54 (74.0)	49 (64.5)	
Smoker	19 (26.0)	27 (35.5)	
Hypertension			0.124 (ns)
No	37 (50.7)	48 (63.2)	
Yes	36 (49.3)	28 (36.8)	
Diabetes mellitus			0.099 (ns)
No	50 (68.5)	61 (80.3)	
Yes	23 (31.5)	15 (19.7)	

Data are presented as mean ± SD or number (%) of subjects. EC, endometrial cancer; ns, not significant; BMI, body mass index.

$p < 0.05$ was considered statistically significant.

Myometrial invasion was <50% in 49 cases (67.1%) and ≥50% in 24 (32.9%). Lymphovascular space invasion (LVSI) was present in 21 patients (28.8%), and microcystic, elongated and fragmented (MELF)-type invasion was observed in 13 (17.8%). Cervical and isthmic involvement occurred in 7 (9.6%) and 13 (17.8%) cases, respectively. LN metastasis was detected in 7 (9.6%), and distant metastasis in 2 (2.7%) patients.

Regarding surgical management, 9 patients (12.3%) underwent no LN dissection (LND), 14 (19.2%) had pelvic–paraortic LND, 50 (68.5%) had pelvic or sentinel LND, and 31 (42.5%) underwent omental sampling or omentectomy. Among surgically staged patients, LN counts ranged from 0 to 51 (mean ± SD: 13.74 ± 11.14). Postoperatively, 40 patients (54.8%) received no adjuvant therapy, 12 (16.4%) underwent brachytherapy, 7 (9.6%) received pelvic radiotherapy (RT), and 14 (19.2%) received combined chemotherapy (CT) and pelvic RT.

Recurrence data were available for 71 of the 73 patients; two patients were excluded from recurrence analysis due to incomplete follow-up. Among the evaluable cohort, recurrence was identified in 9 patients (12.7%) based on radiological or histopathological confirmation—managed with CT in 5 cases, surgery with hyperthermic intraperitoneal chemotherapy (HIPEC) plus CT in 1, and combined CT and RT in 3.

3.2 Serum lncRNA HOTAIRM1 and miR-182-5p Expression Levels

Serum lncRNA HOTAIRM1 expression levels were significantly higher in EC patients compared to controls (median [IQR]: 1.36 [0.74–3.37] vs. 1.03 [0.58–2.03]; $p = 0.043$) (Fig. 1 and Table 2). In contrast, serum miR-182-

5p expression levels did not differ significantly between groups (1.26 [0.09–4.39] vs. 1.71 [0.21–3.58]; $p = 0.327$) (Fig. 2 and Table 2).

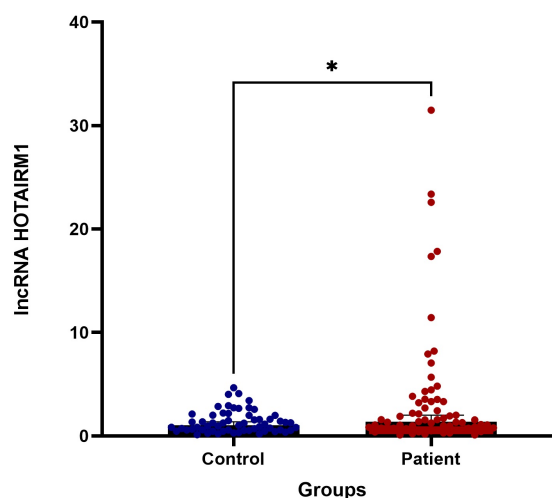


Fig. 1. Comparison of serum lncRNA HOTAIRM1 expression between EC patients and controls. * indicates statistical significance, defined as $p < 0.05$. lncRNA, long non-coding RNA; HOTAIRM1, HOXA transcript antisense RNA myeloid-specific 1.

3.3 Correlation Between Serum lncRNA HOTAIRM1 and miR-182-5p Expression

Spearman’s correlation analysis demonstrated no significant relationship between serum lncRNA HOTAIRM1

Table 2. Comparison of relative serum expression levels of lncRNA HOTAIRM1 and miR-182-5p between patients with EC and controls.

Biomarker	Group	Median (IQR)	<i>p</i> -value
lncRNA HOTAIRM1	Control	1.03 (0.58–2.03)	0.043*
lncRNA HOTAIRM1	Patient	1.36 (0.74–3.37)	
miR-182-5p	Control	1.71 (0.21–3.58)	0.327
miR-182-5p	Patient	1.26 (0.09–4.39)	

Values are presented as median (IQR). Statistical analyses were performed using the Mann-Whitney U test.

$p < 0.05$ was considered statistically significant. Bold values and “*” indicate statistically significant results. IQR, interquartile range.

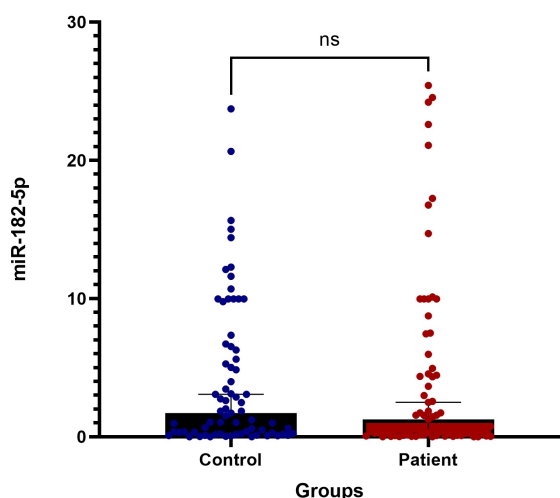


Fig. 2. Comparison of serum miR-182-5p expression between EC patients and controls. ns denotes not statistically significant.

and miR-182-5p expression in the EC groups ($p > 0.05$) (Fig. 3).

3.4 ROC and Survival Analysis of lncRNA HOTAIRM1 in EC

ROC curve analysis demonstrated that serum lncRNA HOTAIRM1 had an area under the curve (AUC) of 0.606 (95% CI: 0.507–0.706; $p = 0.037$), indicating a statistically significant yet modest discriminative ability to distinguish EC patients from controls (Fig. 4 and Table 3). The optimal diagnostic cut-off value was 3.06, corresponding to 29.6% sensitivity and 93.2% specificity (Table 3). These findings indicate that, although serum lncRNA HOTAIRM1 has high specificity—reflecting a strong capacity to correctly identify non-EC cases—its low sensitivity limits its ability to detect all true positives. Therefore, serum lncRNA HOTAIRM1 alone may not be sufficient as a screening biomarker; however, its high specificity supports its potential utility as a confirmatory diagnostic or prognostic adjunct when used in combination with other molecular or clinicopathological parameters.

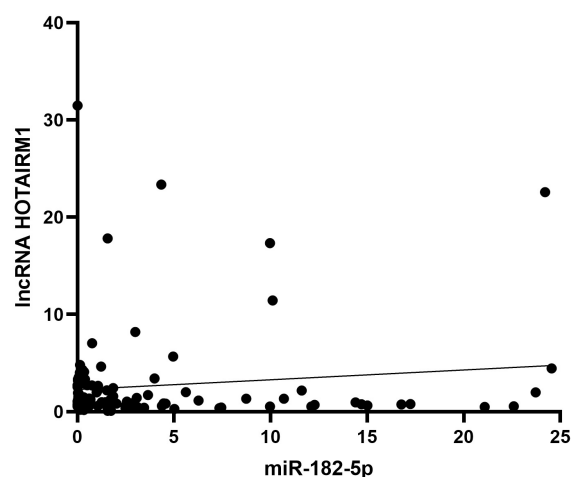


Fig. 3. Correlation between serum lncRNA HOTAIRM1 and miR-182-5p expression in patients with EC.

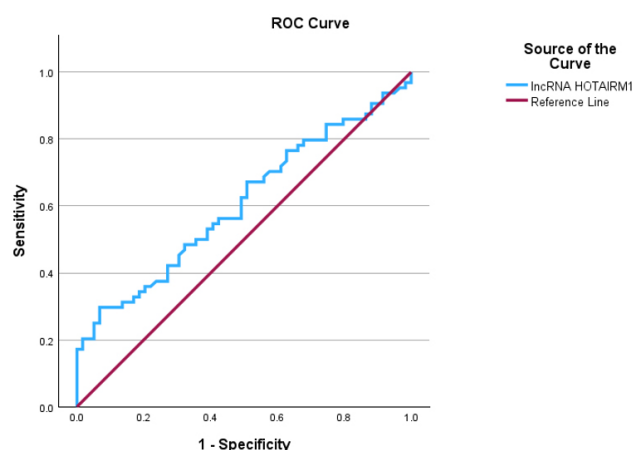


Fig. 4. ROC curve of serum lncRNA HOTAIRM1 for the diagnosis of EC. ROC, receiver operating characteristic.

Overall survival (OS) was defined as the interval from diagnosis to death from any cause, and disease-free survival (DFS) was defined as the interval from surgery to disease progression or recurrence. In the study cohort, the follow-up duration for DFS ranged from 0 to 75 months (mean \pm SD: 47.58 \pm 17.69), and the follow-up duration for OS ranged from 1 to 75 months (mean \pm SD: 48.49 \pm 15.42). Disease-related mortality was documented in 8 patients (10.9%). In survival analysis, neither serum lncRNA HOTAIRM1 nor miR-182-5p expression showed a significant association with DFS or OS ($p > 0.05$). For prognostic subgroup analyses, patients were stratified into “high” and “low” expression groups according to the optimal cut-off value (3.06) derived from the ROC curve using the Youden index ($J = \text{sensitivity} + \text{specificity} - 1$), as described previously. Kaplan-Meier survival analysis comparing patients with high vs. low serum lncRNA HOTAIRM1 expression revealed no statistically significant difference in OS (Log-rank $p > 0.05$) (Fig. 5), and a similar non-significant trend

Table 3. Diagnostic performance of serum lncRNA HOTAIRM1 in distinguishing patients with EC from controls.

Parameter	Sensitivity (%)	Specificity (%)	Cut-off value	AUC (95% CI)	<i>p</i> -value
lncRNA HOTAIRM1	29.6	93.2	3.06	0.606 (0.507–0.706)	0.037*

AUC, area under the ROC curve.

$p < 0.05$ was considered statistically significant. Bold values and “*” indicate statistically significant results.

was observed for DFS during the study period. These findings suggest that while lncRNA HOTAIRM1 and miR-182-5p may have limited prognostic value when considered individually, traditional survival endpoints such as DFS and OS continue to serve as the fundamental outcome measures in EC research. Given the relatively small sample size, multivariate analyses adjusting for clinicopathological parameters (such as age, FIGO stage, or adjuvant therapy) were not performed to prevent model overfitting. Future large-scale studies incorporating these factors are needed to confirm the independent prognostic value of circulating lncRNA HOTAIRM1 and miR-182-5p. This underscores the importance of integrating molecular biomarkers with established clinicopathological parameters to achieve a more comprehensive and reliable prognostic assessment.

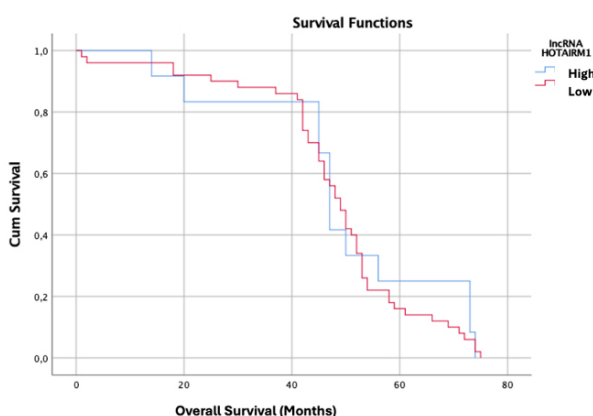


Fig. 5. Kaplan-Meier survival analysis of serum lncRNA HOTAIRM1 expression in EC.

3.5 Association Between Serum lncRNA HOTAIRM1 and miR-182-5p Expression Levels and Clinicopathological Features in EC

Subgroup analyses were performed to examine potential associations between serum lncRNA HOTAIRM1 and miR-182-5p expression levels and key clinicopathological parameters, including FIGO stage, tumor grade, recurrence status, disease-related mortality, LN involvement, deep myometrial invasion, LVSI, MELF pattern, cervical stromal or isthmic involvement, and distant metastasis.

When stratified by FIGO stage, no significant differences in serum lncRNA HOTAIRM1 or miR-182-5p expression were observed between early-stage (IA–IB; $n = 58$) and advanced-stage (II–IV; $n = 15$) groups ($p = 0.968$ and $p = 0.180$, respectively). Similarly, when categorized by

tumor grade, expression levels did not differ significantly between low-grade (grades 1–2 endometrioid; $n = 58$) and high-grade (grade 3 endometrioid or non-endometrioid; $n = 15$) tumors (lncRNA HOTAIRM1: median [IQR] 1.32 [0.74–3.44] vs. 2.01 [0.71–3.70]; miR-182-5p: 0.76 [0.09–4.36] vs. 1.64 [0.13–5.87]; $p = 0.606$ and $p = 0.728$, respectively).

Although median miR-182-5p levels were higher in patients with advanced-stage disease (2.99 [0.17–19.12] vs. 0.76 [0.09–3.82]), LN involvement (4.46 [0.03–23.40] vs. 1.01 [0.10–3.99]), and recurrence (1.55 [0.39–4.46] vs. 0.75 [0.09–4.34]), these differences were not statistically significant ($p = 0.180$, $p = 0.616$, and $p = 0.595$, respectively). Similarly, lncRNA HOTAIRM1 expression showed a non-significant increase in patients with recurrence (3.44 [0.85–7.04] vs. 1.28 [0.73–2.83]; $p = 0.087$).

Additional subgroup analyses, including disease-related mortality, deep myometrial invasion, LVSI, MELF pattern, cervical stromal or isthmic involvement, and distant metastasis, also revealed no statistically significant differences in serum lncRNA HOTAIRM1 or miR-182-5p expression levels.

Overall, serum lncRNA HOTAIRM1 and miR-182-5p expression levels showed variability across clinical and pathological subgroups; however, none of the observed differences reached statistical significance. Given the limited sample sizes within certain subgroups and the absence of confidence interval calculation due to small subgroup counts, these findings should be interpreted with caution. Taken together, the results suggest that although these biomarkers may hold potential clinical relevance, their current utility in predicting disease progression, recurrence, or prognosis in EC remains limited. Detailed subgroup data for serum lncRNA HOTAIRM1 and miR-182-5p expression are presented in Tables 4A and 4B.

4. Discussion

EC is primarily diagnosed through histopathological evaluation of biopsy specimens, and no non-invasive biomarkers are currently implemented in clinical practice. The rising incidence and mortality of EC underscore the need to better elucidate its underlying molecular mechanisms. Increasing evidence indicates that lncRNAs and miRNAs are dysregulated in EC and may serve as promising diagnostic and prognostic biomarkers with potential roles in predicting disease progression, LN involvement, metastasis, treatment response, and diverse pathological or molecular features. Moreover, ncRNAs may represent novel therapeutic targets in EC.

Table 4A. Association of serum lncRNA HOTAIRM1 expression levels with clinicopathological parameters in patients with EC.

Clinicopathological parameter	Group	n	Median (IQR)	p-value
FIGO stage	Early stage (IA–IB)	58	1.47 (0.73–3.33)	0.968
	Advanced stage (II–IV)	15	1.30 (0.77–4.23)	
Tumor grade	Low grade (G1–2)	58	1.32 (0.74–3.44)	0.606
	High grade (G3/non-endometrioid)	15	2.01 (0.71–3.70)	
LN involvement	Negative	66	1.39 (0.74–3.33)	0.880
	Positive	7	1.07 (0.70–13.06)	
Recurrence	Absent	62	1.28 (0.73–2.83)	0.087
	Present	9	3.44 (0.85–7.04)	

Values are presented as median (IQR). Statistical analyses were performed using the Mann-Whitney U test. $p < 0.05$ was considered statistically significant.

Note. $n = 71$ for recurrence analysis (two patients excluded due to incomplete follow-up; recurrence status missing). FIGO, The International Federation of Gynecology and Obstetrics; LN, lymph node.

Table 4B. Association of serum miR-182-5p expression levels with clinicopathological parameters in patients with EC.

Clinicopathological parameter	Group	n	Median (IQR)	p-value
FIGO stage	Early stage (IA–IB)	58	0.76 (0.09–3.82)	0.180
	Advanced stage (II–IV)	15	2.99 (0.17–19.12)	
Tumor grade	Low grade (G1–2)	58	0.76 (0.09–4.36)	0.728
	High grade (G3/non-endometrioid)	15	1.64 (0.13–5.87)	
LN involvement	Negative	66	1.01 (0.10–3.99)	0.616
	Positive	7	4.46 (0.03–23.40)	
Recurrence	Absent	62	0.75 (0.09–4.34)	0.595
	Present	9	1.55 (0.39–4.46)	

Values are presented as median (IQR). Statistical analyses were performed using the Mann-Whitney U test. $p < 0.05$ was considered statistically significant.

Note. $n = 71$ for recurrence analysis (two patients excluded due to incomplete follow-up; recurrence status missing).

ncRNAs can be detected not only in tissues but also in biological fluids such as serum, plasma, and urine, where expression levels differ between healthy individuals and cancer patients. Although tissue-based analyses often yield distinct expression profiles, the optimal biological source for reliable EC biomarkers remains unclear. Circulating ncRNAs are particularly attractive because of their accessibility and the advantages of non-invasive sampling. Previous tissue-based studies have consistently demonstrated upregulation of both lncRNA HOTAIRM1 and miR-182-5p in EC, supporting their potential oncogenic role at the tissue level [17,21,26]. However, their circulating expression patterns remain poorly defined and have not been systematically explored. In this study, we evaluated serum levels of lncRNA HOTAIRM1 and miR-182-5p using qRT-PCR, as these molecules have been previously studied in EC tissues but remain largely unexplored in circulation.

In a systematic review, Donkers *et al.* [21] analyzed 26 studies including 2110 women (1400 with EC) that assessed miRNA expression across tissue, serum, plasma, urine, and liquid-based cytology samples. Some studies did not stratify patients by subtype, whereas others conducted subgroup analyses or focused on type 1 and type 2 EC. A total of 106 miRNAs were investigated, with notable variability in expression profiles. Importantly, miR-182-

5p expression in EC was assessed in only one study analyzing both tissue and liquid-based cytology samples [21]. Kottaridi *et al.* [26] further demonstrated significantly elevated miR-182-5p expression, with a 27.3-fold increase in tissue, an 18.2-fold increase in liquid-based cytology, and a 22.7-fold overall increase in EC patients. Mechanistically, miR-182-5p is implicated in the regulation of the Wnt signaling pathway, which is essential for endometrial proliferation and differentiation; its dysregulation contributes to endometrial hyperplasia [27,28]. Specifically, downregulation of Wnt5a and Smad4 has been observed in EC tissues, and miR-182-5p may target both genes, thereby disrupting Wnt signaling and promoting disease progression [28–31]. Moreover, miRNA expression profiles differ across EC subtypes. Ratner *et al.* [32] identified subtype-specific miRNAs in endometrioid, serous, and carcinosarcoma subtypes, supporting the concept that EC exhibits distinct molecular behaviors. Additional studies have also suggested that certain miRNAs may predict LN involvement or LVSI, further underscoring their potential clinical relevance [33,34].

In our study, we evaluated the diagnostic potential of miR-182-5p, which has been only minimally explored in EC. Although prior studies indicated elevated expression, our analysis revealed no significant differences in serum

levels between patients and controls. These findings suggest that, while miR-182-5p may exert oncogenic effects within tumor tissue, its circulating levels have limited diagnostic value. Nonetheless, the trend toward higher expression in more aggressive disease warrants further investigation in larger cohorts.

Li *et al.* [17] demonstrated in tissue samples from 50 patients with type 1 EC that lncRNA HOTAIRM1 acts as an oncogene by regulating *HOXA1*, thereby promoting cell proliferation, migration, invasion, and epithelial–mesenchymal transition (EMT). Both HOTAIRM1 and *HOXA1* were significantly upregulated and correlated with FIGO stage and LN involvement [17]. These findings suggest that the HOTAIRM1/*HOXA1* axis plays a critical role in the initiation and progression of type 1 EC and may serve as both a prognostic biomarker and a therapeutic target [17].

In the present study, we demonstrated for the first time that serum lncRNA HOTAIRM1 expression levels were significantly elevated in EC patients compared with controls. While previous studies have focused primarily on tissue-based analyses, our findings provide novel evidence supporting its upregulation in serum, suggesting that serum lncRNA HOTAIRM1 may represent a promising non-invasive biomarker for EC. ROC curve analysis demonstrated high specificity but limited sensitivity, implying a strong positive predictive value but insufficient performance as a standalone diagnostic tool due to potential false negatives.

When compared with well-established serum biomarkers, cancer antigen 125 (CA125) and human epididymis protein 4 (HE4), our findings provide perspective on the relative diagnostic value of lncRNA HOTAIRM1. Both CA125 and HE4 are widely recognized markers in EC, consistently elevated in affected patients compared with healthy controls. Barr *et al.* [35] reported that the combination of CA125 and HE4 achieved an AUC of 0.77 (95% CI: 0.74–0.81) for EC detection, while HE4 alone demonstrated excellent diagnostic accuracy in premenopausal women (AUC = 0.91, sensitivity 84.5%, specificity 80.9%). Similarly, Bian *et al.* [36] found that HE4 outperformed CA125, CA19-9, and CA724 (AUC = 0.76, sensitivity 58%), and that combined testing (HE4 + CA125 + CA19-9 + CA724) improved discrimination (AUC = 0.82, sensitivity 59.1%, PPV 88%). These findings highlight HE4, particularly in combination with CA125, as a robust non-invasive biomarker correlated with advanced FIGO stage, deep myometrial invasion, and LVSI or LN involvement [35,36]. Compared with these validated markers, serum lncRNA HOTAIRM1 in our study exhibited comparable specificity but lower sensitivity, indicating limited diagnostic strength at this stage. While lncRNA HOTAIRM1 may reflect underlying tumor biology and hold potential for future biomarker development, its current clinical applicability remains preliminary and warrants confirmation through larger, multicenter investigations.

The molecular pathogenesis of EC has gained increasing attention, as an improved understanding of ncRNA expression may enhance risk stratification and disease classification. Conventional histopathological criteria alone are often insufficient to predict recurrence risk, leading to potential over- or under-treatment. This underscores the need for reliable molecular biomarkers to refine therapeutic decision-making.

In this study, subgroup analyses were performed to evaluate associations between serum lncRNA HOTAIRM1 and miR-182-5p expression levels and clinicopathological parameters. Although higher miR-182-5p levels were observed in patients with advanced-stage disease, LN involvement, or recurrence, and increased lncRNA HOTAIRM1 expression was noted in patients with recurrence, these differences were not statistically significant ($p > 0.05$). Similarly, no significant associations were found across other clinicopathological subgroups. While no definitive correlations were identified, the observed trends suggest that larger, well-powered cohorts are needed to further explore the potential roles of circulating ncRNAs in EC progression.

Molecular subtype data (POLEmut, dMMR, p53abn, NSMP classification) were unavailable for this cohort, as such, analyses were not routinely performed during the study period. Therefore, stratified evaluation of circulating lncRNA HOTAIRM1 across molecular subtypes could not be conducted. Future studies incorporating molecularly characterized cohorts are warranted to determine whether serum lncRNA HOTAIRM1 expression differs across EC subgroups, given the prognostic and therapeutic implications of molecular classification.

lncRNAs and miRNAs can act as oncogenes or tumor suppressors, and their dysregulation contributes to cancer initiation and progression. miRNAs regulate tumorigenesis by binding to target mRNAs and suppressing translation, while certain lncRNAs modulate miRNA function through competing endogenous mechanisms. Increasing evidence supports complex lncRNA–miRNA–mRNA networks shaping oncogenic pathways. For example, Sweef *et al.* [37] described the HOTAIRM1/miR-182-5p/ERO1A axis in lung cancer, where upregulation of HOTAIRM1 and ERO1A with concurrent miR-182-5p downregulation was associated with poor survival. In EC, independent studies have also reported increased lncRNA HOTAIRM1 and miR-182-5p expression in tumor tissue [17,21,26]. Consistent with these reports, we hypothesized a possible HOTAIRM1/miR-182-5p interaction in EC; however, our correlation analysis found no significant association between their serum levels ($p > 0.05$).

Taken together, these findings suggest that although circulating lncRNA HOTAIRM1 and miR-182-5p may reflect molecular alterations seen in EC tissues, their serum expression patterns show limited diagnostic and prognostic utility, suggesting that further exploration of these biomarkers should await stronger preliminary evidence.

Limitations

This study has several limitations that should be acknowledged. First, the sample size was relatively small and derived from a single institution, which may restrict the generalizability of the results. Second, despite standardized sample handling, potential pre-analytical variations—such as storage duration, RNA stability, or variability in internal reference gene expression—might have influenced measured expression levels. Although U6 and GAPDH were selected as internal controls based on prior validation in serum-based ncRNA studies and demonstrated consistent amplification efficiency in our dataset, cohort-specific stability testing was not performed. Third, external validation in an independent cohort and functional *in vitro* assays were beyond the scope of this investigation. Lastly, although the control group consisted of women with non-malignant gynecological conditions and no uterine or adnexal pathology, they may not fully represent healthy individuals, introducing minimal background variability.

5. Conclusions

This study demonstrated that serum lncRNA HOTAIRM1 is significantly elevated in patients with EC compared with controls, whereas serum miR-182-5p showed no diagnostic or prognostic value. Although lncRNA HOTAIRM1 exhibited relatively high specificity, its low sensitivity and lack of consistent association with clinicopathological parameters limit its clinical applicability. Importantly, this work represents one of the first investigations assessing the circulating expression of lncRNA HOTAIRM1 and miR-182-5p in EC, suggesting their potential as non-invasive biomarkers. While the findings provide valuable preliminary data supporting the exploration of circulating ncRNAs in EC, the current evidence indicates insufficient diagnostic or prognostic power to support clinical implementation. Further validation in larger, well-designed multicenter studies using standardized methodologies is required to confirm the diagnostic and prognostic potential of these biomarkers and to clarify their biological significance.

Availability of Data and Materials

The data supporting the findings of this study are available from the corresponding author upon reasonable request. These data are not publicly accessible due to privacy and ethical considerations regarding participant confidentiality.

Author Contributions

HEU contributed to the conceptualization, methodology development, data collection and formal analysis. YM contributed to methodology development and supervised the study. CK and AFA were responsible for data curation and assisted in methodology development. HS and MYS contributed to study design, coordination of data acquisi-

tion, and interpretation of the data. MAE and AMA performed data analysis. ST contributed to study conception, supervised data acquisition and management and participated in the interpretation of the results. All authors contributed to the critical revision of the manuscript for important intellectual content. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

This study protocol was reviewed and approved by the Institutional Review Board and Ethics Committee of Istanbul University (Initial Approval No: 2018/1609; Additional Approval No: 2023/119). All procedures were conducted in accordance with the ethical standards of the institutional and national research committees and with the Declaration of Helsinki. Written informed consent was obtained from all participants prior to inclusion in the study, covering both immediate participation and future use of stored biospecimens.

Acknowledgment

We gratefully acknowledge the Istanbul University Scientific Research Projects Unit for their generous funding and essential support in facilitating this study. We also thank the anonymous peer reviewers for their constructive feedback, which significantly improved the manuscript.

Funding

This study was supported by the Istanbul University Scientific Research Projects Unit (Project ID: TTU-2023-39847). No additional financial support was received from public, commercial, or not-for-profit funding agencies.

Conflict of Interest

The authors declare no conflict of interest.

Declaration of AI and AI-Assisted Technologies in the Writing Process

During the preparation of this work the authors used ChatGPT-5 in order to check spell and grammar. After using this tool, the authors reviewed and edited the content as needed and takes full responsibility for the content of the publication.

References

- [1] Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, *et al.* Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*. 2021; 71: 209–249. <https://doi.org/10.3322/caac.21660>.
- [2] Jarroux J, Morillon A, Pinskaya M. History, Discovery, and Classification of lncRNAs. *Advances in Experimental Medicine and Biology*. 2017; 1008: 1–46. https://doi.org/10.1007/978-981-10-5203-3_1.

- [3] Yates LA, Norbury CJ, Gilbert RJC. The long and short of microRNA. *Cell*. 2013; 153: 516–519. <https://doi.org/10.1016/j.cell.2013.04.003>.
- [4] Mattick JS, Makunin IV. Non-coding RNA. *Human Molecular Genetics*. 2006; 15: R17–R29. <https://doi.org/10.1093/hmg/dd1046>.
- [5] Banno K, Yanokura M, Kisu I, Yamagami W, Susumu N, Aoki D. MicroRNAs in endometrial cancer. *International Journal of Clinical Oncology*. 2013; 18: 186–192. <https://doi.org/10.1007/s10147-013-0526-9>.
- [6] McAlpine JN, Chiu DS, Nout RA, Church DN, Schmidt P, Lam S, *et al.* Evaluation of treatment effects in patients with endometrial cancer and POLE mutations: An individual patient data meta-analysis. *Cancer*. 2021; 127: 2409–2422. <https://doi.org/10.1002/cncr.33516>.
- [7] Talhouk A, McAlpine JN. New classification of endometrial cancers: the development and potential applications of genomic-based classification in research and clinical care. *Gynecologic Oncology Research and Practice*. 2016; 3: 14. <https://doi.org/10.1186/s40661-016-0035-4>.
- [8] Di Tucci C, Capone C, Galati G, Iacobelli V, Schiavi MC, Di Donato V, *et al.* Immunotherapy in endometrial cancer: new scenarios on the horizon. *Journal of Gynecologic Oncology*. 2019; 30: e46. <https://doi.org/10.3802/jgo.2019.30.e46>.
- [9] Bueno MJ, Pérez de Castro I, Malumbres M. Control of cell proliferation pathways by microRNAs. *Cell Cycle*. 2008; 7: 3143–3148. <https://doi.org/10.4161/cc.7.20.6833>.
- [10] Mestdagh P, Hartmann N, Baeriswyl L, Andreassen D, Bernard N, Chen C, *et al.* Evaluation of quantitative miRNA expression platforms in the microRNA quality control (miRQC) study. *Nature Methods*. 2014; 11: 809–815. <https://doi.org/10.1038/nmeth.3014>.
- [11] Rahimian N, Razavi ZS, Aslanbeigi F, Mirkhabbaz AM, Piroozmand H, Shahrzad MK, *et al.* Non-coding RNAs related to angiogenesis in gynecological cancer. *Gynecologic Oncology*. 2021; 161: 896–912. <https://doi.org/10.1016/j.ygyno.2021.03.020>.
- [12] Cancer Genome Atlas Research Network, Kandoth C, Schultz N, Cherniack AD, Akbani R, Liu Y, *et al.* Integrated genomic characterization of endometrial carcinoma. *Nature*. 2013; 497: 67–73. <https://doi.org/10.1038/nature12113>.
- [13] Vasilatou D, Sioulas VD, Pappa V, Papageorgiou SG, Vlahos NF. The role of miRNAs in endometrial cancer. *Epigenomics*. 2015; 7: 951–959. <https://doi.org/10.2217/epi.15.41>.
- [14] Höhn AK, Brambs CE, Hiller GGR, May D, Schmoedel E, Horn LC. 2020 WHO Classification of Female Genital Tumors. *Geburtshilfe Und Frauenheilkunde*. 2021; 81: 1145–1153. <https://doi.org/10.1055/a-1545-4279>.
- [15] Zehtabi M, Ghaedrahmati F, Dari MAG, Moramezi F, Kempisty B, Mozdziaik P, *et al.* Emerging biologic and clinical implications of miR-182-5p in gynecologic cancers. *Clinical & Translational Oncology*. 2025; 27: 2367–2382. <https://doi.org/10.1007/s12094-024-03822-9>.
- [16] Krishnan K, Steptoe AL, Martin HC, Wani S, Nones K, Waddell N, *et al.* MicroRNA-182-5p targets a network of genes involved in DNA repair. *RNA*. 2013; 19: 230–242. <https://doi.org/10.1261/rna.034926.112>.
- [17] Li X, Pang L, Yang Z, Liu J, Li W, Wang D. LncRNA HOTAIRM1/HOXA1 Axis Promotes Cell Proliferation, Migration And Invasion In Endometrial Cancer. *OncoTargets and Therapy*. 2019; 12: 10997–11015. <https://doi.org/10.2147/OTT.S222334>.
- [18] Zhao Y, Wang W, Guan C, Hu Z, Liu L, Li W, *et al.* Long non-coding RNA HOTAIRM1 in human cancers. *Clinica Chimica Acta*. 2020; 511: 255–259. <https://doi.org/10.1016/j.cca.2020.10.011>.
- [19] Shetty A, Venkatesh T, Kabbekodu SP, Tsutsumi R, Suresh PS. LncRNA-miRNA-mRNA regulatory axes in endometrial cancer: a comprehensive overview. *Archives of Gynecology and Obstetrics*. 2022; 306: 1431–1447. <https://doi.org/10.1007/s00404-022-06423-5>.
- [20] Vallone C, Rigon G, Gulia C, Baffa A, Votino R, Morosetti G, *et al.* Non-Coding RNAs and Endometrial Cancer. *Genes*. 2018; 9: 187. <https://doi.org/10.3390/genes9040187>.
- [21] Donkers H, Bekkers R, Galaal K. Diagnostic value of microRNA panel in endometrial cancer: A systematic review. *Oncotarget*. 2020; 11: 2010–2023. <https://doi.org/10.18632/oncotarget.27601>.
- [22] Kupec T, Bleilevens A, Iborra S, Najjari L, Wittenborn J, Maurer J, *et al.* Stability of circulating microRNAs in serum. *PLoS ONE*. 2022; 17: e0268958. <https://doi.org/10.1371/journal.pone.0268958>.
- [23] Sandau US, Wiedrick JT, McFarland TJ, Galasko DR, Fanning Z, Quinn JF, *et al.* Analysis of the longitudinal stability of human plasma miRNAs and implications for disease biomarkers. *Scientific Reports*. 2024; 14: 2148. <https://doi.org/10.1038/s41598-024-52681-5>.
- [24] Katayama ES, Hue JJ, Loftus AW, Ali SA, Graor HJ, Rothermel LD, *et al.* Stability of microRNAs in serum and plasma reveal promise as a circulating biomarker. *Non-Coding RNA Research*. 2025; 15: 132–141. <https://doi.org/10.1016/j.ncrna.2025.08.001>.
- [25] National Cancer Institute (NCI). Cell-free miRNA blood collection and processing: BEBP 508 compliant (Version 2–7). 2025. Available at: <https://biospecimens.cancer.gov/resources/bebp.asp> (Accessed: 21 January 2026).
- [26] Kottaridi C, Spathis A, Margari N, Koureas N, Terzakis E, Chrelias C, *et al.* Evaluation Analysis of miRNAs Overexpression in Liquid-Based Cytology Endometrial Samples. *Journal of Cancer*. 2017; 8: 2699–2703. <https://doi.org/10.7150/jca.19947>.
- [27] Wang Y, van der Zee M, Fodde R, Blok LJ. Wnt/B-catenin and sex hormone signaling in endometrial homeostasis and cancer. *Oncotarget*. 2010; 1: 674–684. <https://doi.org/10.18632/oncotarget.201>.
- [28] Tang S, Dai Y. RNA sequencing reveals significant miRNAs in Atypical endometrial hyperplasia. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*. 2018; 225: 129–135. <https://doi.org/10.1016/j.ejogrb.2018.03.025>.
- [29] Iozzo RV, Eichstetter I, Danielson KG. Aberrant expression of the growth factor Wnt-5A in human malignancy. *Cancer Research*. 1995; 55: 3495–3499.
- [30] Bui TD, Zhang L, Rees MC, Bicknell R, Harris AL. Expression and hormone regulation of Wnt2, 3, 4, 5a, 7a, 7b and 10b in normal human endometrium and endometrial carcinoma. *British Journal of Cancer*. 1997; 75: 1131–1136. <https://doi.org/10.1038/bjc.1997.195>.
- [31] Liu FS, Chen JT, Hsieh YT, Ho ESC, Hung MJ, Lu CH, *et al.* Loss of Smad4 protein expression occurs infrequently in endometrial carcinomas. *International Journal of Gynecological Pathology*. 2003; 22: 347–352. <https://doi.org/10.1097/01.pgp.0000092131.88121.0a>.
- [32] Ratner ES, Tuck D, Richter C, Nallur S, Patel RM, Schultz V, *et al.* MicroRNA signatures differentiate uterine cancer tumor subtypes. *Gynecologic Oncology*. 2010; 118: 251–257. <https://doi.org/10.1016/j.ygyno.2010.05.010>.
- [33] Concin N, Matias-Guiu X, Vergote I, Cibula D, Mirza MR, Marantz S, *et al.* ESGO/ESTRO/ESP guidelines for the management of patients with endometrial carcinoma. *International Journal of Gynecological Cancer*. 2021; 31: 12–39. <https://doi.org/10.1136/ijgc-2020-002230>.
- [34] de Foucher T, Sbeih M, Uzan J, Bendifallah S, Lefevre M, Chabbert-Buffet N, *et al.* Identification of micro-RNA expression profile related to recurrence in women with ESMO low-risk endometrial cancer. *Journal of Translational Medicine*. 2018; 16: 131. <https://doi.org/10.1186/s12967-018-1515-6>.
- [35] Barr CE, Njoku K, Jones ER, Crosbie EJ. Serum CA125 and

- HE4 as Biomarkers for the Detection of Endometrial Cancer and Associated High-Risk Features. *Diagnostics*. 2022; 12: 2834. <https://doi.org/10.3390/diagnostics12112834>.
- [36] Bian J, Sun X, Li B, Ming L. Clinical Significance of Serum HE4, CA125, CA724, and CA19-9 in Patients With Endometrial Cancer. *Technology in Cancer Research & Treatment*. 2017; 16: 435–439. <https://doi.org/10.1177/1533034616666644>.
- [37] Sweef O, Yang C, Wang Z. The Oncogenic and Tumor Suppressive Long Non-Coding RNA-microRNA-Messenger RNA Regulatory Axes Identified by Analyzing Multiple Platform Omics Data from Cr(VI)-Transformed Cells and Their Implications in Lung Cancer. *Biomedicines*. 2022; 10: 2334. <https://doi.org/10.3390/biomedicines10102334>.