MINI-REVIEW

Metabolic control of adult neural stem cell behavior

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Abstract   **Neural stem cells generate new neurons throughout life in distinct regions of the mammalian brain. This process, called adult neurogenesis, is important for tissue homeostasis and physiological brain function. In addition, failing or altered neurogenesis has been associated with a number of diseases such as major depression and epilepsy. Thus, understanding the molecular mechanisms governing the neurogenic process in the adult brain may enable future therapeutic approaches to target neural stem/progenitor cells (NSPCs) and their progeny to ameliorate disease symptoms and/or disease progression. Recently, the control of cellular metabolism has emerged as a regulator of NSPC activity in the adult brain. Here we review recent findings that attempt to describe stage-specific modulations of metabolism to ensure proper neurogenesis and suggest future avenues of research aiming to understand how metabolism affects NSPC behavior.**

Keywords   adult neurogenesis, metabolic switch, quiescence, proliferation, differentiation

Introduction

Contrary to the long-held belief that the neurogenic activity of NSPCs tapers off after embryonic and early postnatal development, it was first shown in the mid-1960s and accepted by the field in the late 1990s that NSPCs in distinct regions of the mammalian brain retain their neurogenic potential throughout life (Altman and Das, 1964; Kuhn et al., 1996; Eriksson et al., 1998). Thus, the adult mammalian brain retains the capacity to generate new neurons that over the course of several weeks mature and functionally integrate into the pre-existing circuitries (Braun and Jessberger, 2014; Christian et al., 2014). However, adult neurogenesis is not widespread but rather restricted to discrete areas. Whereas in the rodent brain a substantial number of NSPCs persist in the subventricular zone (SVZ), lining the lateral ventricles, that give rise to a diverse set of olfactory neurons (Doetsch et al., 1997; 1999), this site of neurogenesis appears to be absent in the human (Bergmann et al., 2012). However, accumulating evidence suggests that in both rodent and human brains a substantial number of neurons are generated in the second main neurogenic area: the dentate gyrus (DG), the part of the

hippocampus that is critically involved in certain forms of learning and memory (Kuhn et al., 1996; Eriksson et al., 1998; Spalding et al., 2013). Strikingly, failing or altered hippocampal neurogenesis has been associated with several neuropsychiatric diseases, among others major depression, epilepsy, cognitive aging, and a number of neurodegenerative diseases (Zhao et al., 2008). Support for a role of newborn dentate granule cells for proper hippocampal function came from a number of studies showing hippocampus-dependent behavioral alterations upon experimental decrease or increase of neurogenesis levels in rodents (Deng et al., 2010). Given the potential role of life-long neurogenesis for cognition and hippocampus-dependent emotional control (Deng et al., 2010; Kheirbek et al., 2012), tremendous efforts have been undertaken to understand the cellular and molecular mechanisms governing the neurogenic process in the brain. Thus, over the past 15 years a number of intrinsic and niche-dependent regulators of neurogenesis have been identified that regulate distinct steps during the neurogenic process (Lie et al., 2005; Ge et al., 2006; Zhao et al., 2008; Favaro et al., 2009; Ables et al., 2010; Kim et al., 2012; Song et al., 2012). Only very recently a critical role of cellular metabolism to regulate the activity of adult NSPCs has been identified (Knobloch et al., 2013), in line with the important role of metabolism in other somatic stem cell systems (Nakada et al., 2010; Gan et al., 2010; Gurumurthy et al., 2010; Folmes et al., 2012; Ito and Suda, 2014). Here we review recent findings

describing a role for metabolic regulation during distinct NSPC states and describe potential routes of future research.

Metabolic requirements of proliferating adult NSPCs

A tight regulation of NSPC quiescence and proliferation is crucial to ensure life-long neurogenesis and prevent exhaustion or uncontrolled growth of the stem cell pool. Most studies aimed at understanding the regulation of NSPC proliferation and quiescence have focused on the role of transcription factors and molecular pathways such as Wnt signaling and IGF signaling that have been previously implicated in growth regulation (Orford and Scadden, 2008; Suh et al., 2009). Many of these transcription factors and pathways ultimately influence cell metabolism, however, only over the last few years has the metabolic state of stem cells per se garnered attention (Varum et al., 2011; Folmes et al., 2012; Zhang et al., 2012), particularly in the field of hematopoietic stem cell (HSC) research (Gan et al., 2010; Nakada et al., 2010; Gurumurthy et al., 2010; Suda et al., 2011). So far, very little is known about the metabolic state of NSPCs and how it might influence NSPC behavior. Recently, we showed that *de novo* lipogenesis is crucial for NSPC proliferation (Knobloch et al., 2013). Similar to cancer cells, which produce the majority of their lipids *de novo* rather than taking them up from the environment (Menendez and Lupu, 2007), NSPCs also upregulate the lipogenic pathway. In addition, we showed that proliferating NSPCs have high activity levels of the key enzyme fatty acid synthase (Fasn) and that proliferation is significantly reduced upon pharmacological or genetic inhibition of Fasn. When Fasn is blocked in adult NSPCs *in vivo* using an inducible Fasn knockout mouse line, neurogenesis is dramatically inhibited. We have furthermore found that Spot14, a protein whose expression is largely restricted to quiescent NSPCs, acts as a regulator of Fasn by reducing Malonyl-CoA, which serves as a substrate for Fasn. When Spot14 is knocked down *in vivo*, the relatively quiescent Spot14 positive NSPCs increase proliferation, indicating that Spot14 might act as a molecular brake to regulate the activity of a specific metabolic program, namely *de novo* lipogenesis (Knobloch et al., 2013). The reasons for the importance of *de novo* fatty acid synthesis in proliferating NSPCs are not yet entirely understood. As most of the newly generated fatty acids end up in membrane lipids, it is likely that the NSPCs need this pathway to build new membranes for rapid proliferation. However, palmitic acid, the major fatty acid produced by Fasn, is also used for palmitoylation, a protein modification that has been shown to be important for many proteins involved in stem cell regulation such as for instance Wnt and Hedgehog proteins as well as for synaptic plasticity (Iwanaga et al., 2009; Fukata and Fukata, 2010). Furthermore, as fatty acids provide a means to efficiently store energy, these lipids might also be used by NSPCs for

energy production.

Interestingly, besides the lipogenic phenotype, proliferating NSPCs also share other metabolic pathways with cancer cells, and it has been proposed that such metabolic characteristics might be a universal feature of proliferating cells in general (Vander Heiden et al., 2009). An increase in (aerobic) glycolysis, whereby glucose is fermented into lactate despite the availability of oxygen, is common to many stem cells (reviewed by Folmes et al., 2012; Ito and Suda, 2014). This effect was first discovered in cancer cells by Otto Warburg and is known as the "Warburg effect." It is emerging that with such increased glycolytic flux and only partial breakdown of glucose, more metabolic intermediates are available for the biosynthesis of cellular building blocks, which are required for proliferation (Vander Heiden et al., 2009). The Warburg effect also seems to occur in proliferating NSPCs, as they have been shown to have high glycolytic flux and increased lactate production (reviewed by Kim et al., 2014). Furthermore, a recent study showed that NSPCs are more susceptible to glycolytic inhibition than primary neurons, even in the presence of alternative substrates for oxidative phosphorylation such as pyruvate (Candelario et al., 2013), implying that the glycolytic flux is not increased to fuel oxidative phosphorylation, but rather to provide metabolic intermediates. Taken together, many proliferating stem cells, including proliferating NSPCs, seem to share metabolic features that are similar in cancer cells.

Metabolism of quiescent adult NSPCs

It is now becoming apparent that stem cells are indeed in a metabolic state that is different from their progeny (Varum et al., 2011; Folmes et al., 2012; Knobloch et al., 2013) and that such metabolic differences influence cell behavior. A shared feature of many quiescent stem cells is their relative low levels of oxidative phosphorylation, probably due to the hypoxic environment in which they are residing. Low oxidative phosphorylation might also protect these quiescent stem cells from DNA damage caused by reactive oxygen species (ROS), which are produced during oxidative phosphorylation. The study of metabolic features of quiescent NSPCs is especially challenging, as they are activated and begin to proliferate when they are isolated from their *in vivo* niche and exposed to culture media containing growth factors (Costa et al., 2011), which likely alters their metabolism. In addition, the lack of good markers to prospectively isolate quiescent NSPCs and the relatively large numbers of cells required for metabolomics profiling has hindered the characterization of the metabolic state of quiescent NSPCs. Spot14, which is very selectively expressed in quiescent NSPCs and is functionally involved in regulating lipid metabolism, provides an interesting candidate (Knobloch et al., 2013) and metabolic profiling of Spot14 positive cells directly isolated from the brain might give further insight into

the metabolic pathways active in quiescent NSPCs. As Spot14 positive NSPCs have lower levels of Malonyl-CoA (Knobloch et al., 2013), and as Malonyl-CoA is a negative regulator of fatty acid oxidation, it is possible that these quiescent Spot14 positive NSPCs utilize fatty acid oxidation. This is an intriguing concept, as a recent publication in the field of hematopoiesis showed that the most primitive HSC population uses fatty acid oxidation to maintain self-renewal (Ito et al., 2012). We are currently investigating the role of fatty acid oxidation in NSPCs and have observed that this pathway is indeed required, especially for the most quiescent NSPC population (Knobloch et al., unpublished). The detailed mechanisms remain to be elucidated.

Metabolic influence on NSPC behavior

Given the differences in metabolism between quiescent and proliferating stem cells, the question is whether these metabolic differences are a consequence of altered cell behavior or a prerequisite for it. To answer this, specific manipulation of distinct metabolic pathways are required, however this is extremely challenging as there is a complex interplay between different metabolic pathways due to shared metabolites, and the downregulation of a specific pathway often affects other metabolic pathways. Thus, great care has to be taken in interpreting results. Metabolic flux analyses, used to determine the actual rate of metabolite turnover, might provide a tool to address this complex interplay (reviewed by Sims et al., 2013). By using specific pathway inhibitors in combination with metabolic flux analyses, it should be possible to dissect the role of different metabolic pathways on stem cell behavior *in vitro*. Addressing their effects on proliferation and fate decision will provide important insights into basic stem cell biology. Strong evidence that the metabolic state is indeed a prerequisite and not a mere consequence of stem cell behavior comes from the induced pluripotent stem cell (iPSC) field. During reprogramming of fibroblasts into iPSCs, the somatic cells displayed a switch in their metabolic profile from an oxidative to a glycolytic phenotype prior to expression of pluripotent genes (Folmes et al., 2011). Blockade of glycolytic enzyme activity blunted reprogramming efficiency, indicating that a certain metabolic state is required to facilitate reprogramming. How exactly metabolism influences reprogramming remains to be elucidated, but it has been suggested that histone modifying enzymes can directly link metabolism and gene transcription/epigenetic modifications through their need of metabolic co-enzymes (Teperino et al., 2010).

Whether a metabolic switch occurs in NSPCs prior to fate changes, i.e. from quiescence to proliferation or from proliferation to differentiation, remains to be addressed. Studying neural stem cells in *Drosophila* (called neuroblasts), Homem and colleagues (2014) have recently published the first evidence that a metabolic switch indeed precedes cell-cycle exit and subsequent differentiation. *Drosophila* neuroblasts divide hundreds of times in a self-renewing mode before they shrink in size and undergo a symmetric terminal differentiation during metamorphosis. None of the common growth control pathways seem to be responsible for this phenomenon. Instead, the authors found a metabolic switch in neuroblasts from glycolysis to oxidative phosphorylation induced by the steroid hormone ecdysone and the Mediator complex, which lead to shrinkage and subsequent terminal differentiation. Importantly, blocking this switch by inhibiting the oxidative phosphorylation machinery prevented cell shrinkage and extended the life span of the pupal neuroblasts (Homem et al., 2014). Although the detailed mechanisms are not yet clear, increased catabolism through oxidative phosphorylation might prevent the accumulation of intermediates needed for the biosynthetic pathway, thus leading to reduced growth and cell-cycle exit. Whether this holds true for mammalian NSPCs remains to be determined. Further evidence that there is an increase in oxidative phosphorylation with neuronal differentiation comes from a recent study by Steib and colleagues, who have shown that the development of new neurons in adult mice is paralleled by extensive changes in mitochondrial mass, distribution, and shape (Steib et al., 2014). When inhibiting a mitochondrial fission factor necessary for increasing the numbers of mitochondria, maturation and survival of newborn neurons was strongly decreased. Although the authors did not directly address the metabolic properties of the newborn neurons, it is highly likely that the observed increase in mitochondrial mass is accompanied by an increase in oxidative metabolism.

Environmental impact on adult neural stem cell metabolism

Extrinsic stimuli having a positive or negative impact on adult NSPC behavior have been widely documented. Voluntary wheel running and enriched environment, for instance, lead to increased proliferation and survival of NSPCs and newborn neurons (Ma et al., 2009). On the contrary, aging negatively affects proliferation and neurogenesis (Knoth et al., 2010; Christian et al., 2014). Whether these extrinsic regulators directly alter the metabolic state of NSPCs is not clear. In a study addressing the effects of neurogenic modulators on Spot14 positive NSPCs *in vivo*, we showed that cells expressing this metabolically functional marker indeed react to extrinsic stimuli: running resulted in an increase in proliferating Spot14 positive NSPCs whereas aging reduced the number of Spot14 positive NSCPs (Knobloch et al., 2014).

Interestingly, a recent report by Chorna and colleagues (2013) showed that upon running, Fasn mRNA was significantly and specifically upregulated in the hippocampus of adult mice. The upregulation of Fasn mRNA was accompanied by higher palmitic acid levels and an increase

in proliferating NSPCs. Furthermore, when Fasn was chronically inhibited by intracerebroventricular micro infusion of a Fasn inhibitor, the increase in proliferation was prevented and the exercise-mediated cognitive enhancement was disrupted. These data provide evidence that the metabolic program of NSPCs can be directly influenced by extrinsic stimuli and confirm the important role of Fasn for NSPCs (Knobloch et al., 2013).

The age-related decline in neurogenesis has been robustly documented (Kuhn et al., 1996; Knoth et al., 2010; Ben Abdallah et al., 2010; Villeda et al., 2011), although it is currently under debate if this is due to an exhausted pool of NSPCs or whether the stem cells remain but stop proliferating (Lugert et al., 2010; Encinas et al., 2011; Bonaguidi et al., 2012).The decline in neurogenesis with aging has been associated with blood-borne factors present in the systemic milieu, such as various chemokines which are increased upon aging and might exert negative effects on NSPCs (Villeda et al., 2011), but the detailed mechanisms remain to be addressed. A proteomic comparison between NSPCs derived from young and aged mice revealed an altered metabolic phenotype upon aging including decreased mitochondrial quantity and lowered oxygen consumption rates and a shift toward more glycolytic metabolism (Stoll et al., 2011). As quiescent stem cells in general seem to have rather reduced oxidative phosphorylation along with having more primitive mitochondria and using the glycolytic pathway (see above), these data suggest an increased quiescence of NSPCs upon aging. However, whether such a metabolic shift is functionally responsible for the reduced neurogenesis needs to be tested experimentally. Metabolic changes have also been suggested as the underlying cause for the age-related decline in a recent study by Stein and Imai (Stein and Imai, 2014). The essential cofactor nicotinamide adenine dinucleotide (NAD^+) was markedly reduced in aged NSPCs due to a decline of the rate-limiting enzyme in the biosynthetic pathway of NAD^+, nicotinamide phosphoribosyl transferase (Nampt). Inhibition of Nampt *in vitro* and *in vivo* reduced proliferation of NSPCs and systemic administration of the Nampt product nicotinamide mononucleotide in aged mice prevented the radial glia like stem cell pool from diminishing. These data suggest that direct targeting of metabolic pathways might provide a novel avenue to treat the age-related decline in neurogenesis, for instance by providing essential metabolic intermediates via nutrition.

A few groups have studied the overall influence of nutrition on neurogenesis and NSPCs, specifically the role of a high fat diet and caloric restriction. Although there are some controversial results, it seems that a high fat diet leads to a decrease in proliferation of NSPCs (Lindqvist et al., 2006; Park et al., 2010; Boitard et al., 2012), whereas caloric restriction has beneficial effects on neurogenesis, probably due to increased survival of newborn neurons (Lee et al., 2000; 2002). However the mechanisms are poorly understood and whether NSPCs and their progeny are directly altering

their metabolic program upon major changes incirculating nutrients is not known.

The availability of peripheral nutrients, not only glucose levels but also the amounts of circulating lipids, is sensed in hypothalamic centers that regulate feeding and fasting behavior (Morton et al., 2006). Remarkably, this brain area has also been shown to have ongoing neurogenesis, mediated by so-called tanycytes that also show radial glia like phenotypes, and the formation of new neurons in the hypothalamus has been associated with energy balance (Kokoeva et al., 2005; Lee and Blackshaw, 2012; Li et al., 2012). Furthermore, a recent publication demonstrated that these stem cells, in contrast to NSPCs in the DG, increased their proliferation upon high-fat diet (Lee et al., 2012). These data suggest that there are indeed direct influences of circulating nutrients on NSPC behavior, a finding that might be of clinical relevance given the high fat diet of the Western World. The detailed mechanisms however remain to be elucidated.

Conclusions

Over the last few years metabolic control has been identified as an important regulator of somatic stem cell activity in a variety of tissue-specific stem cells, including adult NSPCs. However, we are just beginning to understand how switches between proliferating and quiescent NSPC states are orchestrated. Could it be that a plethora of signaling pathways converge on control of distinct metabolic states? This hypothesis is supported by the findings that a number of key regulators of neurogenesis such as BMPs, Shh, and FoxOs have been implicated in controlling metabolic states outside of the brain (Favaro et al., 2009; Paik et al., 2009; Renault et al., 2009; Schulz and Tseng, 2009; Mira et al., 2010; Teperino et al., 2012; Eijkelenboom and Burgering, 2013; Schulz et al., 2013). Moreover, future studies will test if metabolic changes simply occur secondary to fate switches or if altering metabolic states may be instructive for NSPC behavior. In addition, it will be of interest to test how similar or dissimilar distinct somatic stem cell types are with regards to their metabolic state. Can we identify a metabolic code of somatic stem cell multipotency, quiescence or activation?

On a more translational level it will be of interest to analyze how identified human mutations in genes encoding for metabolic regulators may contribute to human disease phenotypes due to NSPC-associated alterations. For example, recent work identified a point mutation in the human Fasn gene that is associated with non-syndromic cognitive impairment, even though it remains unclear at this time if this can be attributed to altered stem cell-associated plasticity (Najmabadi et al., 2011).

Although we do not fully understand the mechanisms of metabolic control of NSPC behavior yet, there is strong evidence that the metabolic state is a novel key player
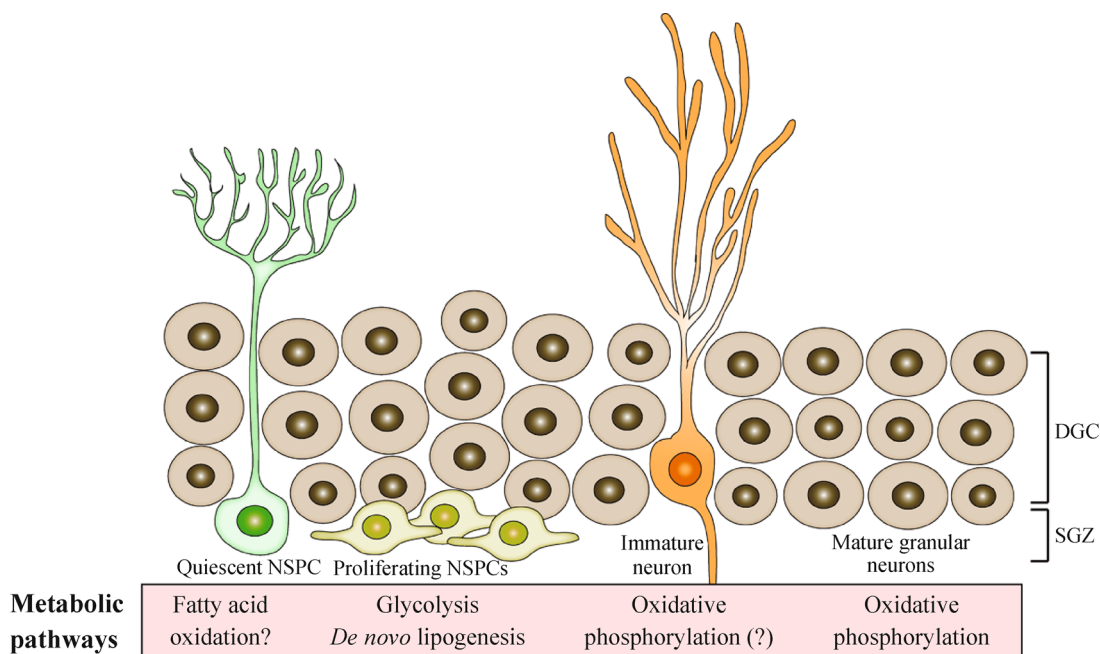
Figure 1   Schematic representation of metabolic pathways active in adult NSPCs and their progeny. Question marks indicate putative active pathways that need to be confirmed. SGZ, subgranular zone; DGC, dentate granule cell layer.

regulating the balance between stem cell quiescence/activation and subsequent differentiation (Fig. 1).

Compliance with ethics guidelines

Marlen Rnobloch and Sebastian Jessoerger declare that they have no conflict of interest.

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