

RESEARCH ARTICLE

Genome-wide analysis reveals selection for Chinese Rongchang pigs

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Abstract Livestock have undergone domestication and consequently strong selective pressure on genes or genomic regions that control desirable traits. To identify selection signatures in the genome of Chinese Rongchang pigs, we generated a total of about 170 Gb of DNA sequence data with about 6.4-fold coverage for each of six female individuals. By combining these data with the publically available genome data of 10 Asian wild boars, we identified 449 protein-coding genes with selection signatures in Rongchang pigs, which are mainly involved in growth and hormone binding, nervous system development, and drug metabolism. The accelerated evolution of these genes may contribute to the dramatic phenotypic differences between Rongchang pigs and Chinese wild boars. This study illustrated how domestication and subsequent artificial selection have shaped patterns of genetic variation in Rongchang pigs and provides valuable genetic resources that can enhance the use of pigs in agricultural production and biomedical studies.

Keywords domestication, genome, pig, re-sequencing, selection

ments^[2]. Modern domestic pigs have undergone strong genetic selection in specialized commercial populations; this has led to remarkable phenotypic changes and genetic adaptation, which makes these breeds an important world heritage and scientific resource for comparative genomic studies^[3,4]. Recently, a list of ‘domestication genes’ has been compiled for silkworms^[5], chickens^[6], pigeons^[7], rabbits^[8], dogs^[9], cattle^[10] and pigs^[11–14] by genomic sequencing. Rongchang pigs, a Chinese indigenous breed raised only in Southwest China with a center of production in the Sichuan basin, have been intensively selected for efficient accumulation of muscle and highly prized pork traits (i.e., juiciness, flavor, tenderness, pink hue and heavy marbling). The phenotypes of Rongchang pigs, characterized by their average-size head, concave and wrinkled face, well-developed limbs, concave back, tilted haunch and a big belly, are remarkably different from wild boars.

To identify genomic selection signatures in Rongchang pigs, we performed whole-genome resequencing of six female Rongchang pigs (about 170 Gb in total) and evaluated the genomic regions under selection.

1 Introduction

Genome sequencing and assembly for European domestic Duroc pigs has greatly improved the genetic resources available for this important livestock species^[1], and has enhanced the potential of this pig as a model organism for biomedical studies. Biological adaptability has enabled development of over 730 current pig breeds or lines that are distributed globally across a wide range of environ-

2 Materials and methods

2.1 Animals and genome sequencing

Genomic DNA was extracted from the blood of six female Rongchang pigs from a nucleus herd in Chongqing Municipal Breeding Pig Farm, with no direct and collateral blood relationship within the last three generations among the individuals selected. Sequencing was performed on a HiSeq 2000 platform (Illumina, San Diego, CA, USA). In addition, we downloaded genomic data of 101 pigs worldwide from the EMBL-EBI database (<http://www.ebi.ac.uk/>) under accession number ERR173 and the NCBI sequence read archive (SRA) under accession number

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SRA065461, which included 30 European domestic pigs, 20 Chinese domestic pigs, 30 Tibetan wild pigs from China, 10 Asian wild boars, six European wild boars, four other species in the genus *Sus*, and an African warthog^[1,11,15,16].

2.2 SNP calling

We first filtered low-quality paired reads, which mainly resulted from base-calling duplicates and adapter contamination. The qualified paired-end reads were mapped to the pig reference genome assembly (Sscrofa10.2)^[1] using BWA software^[17]. After alignment, we performed SNP calling on a population-scale for two groups (56 domestic pigs and 51 other pigs as detailed above and an African warthog) using a Bayesian approach implemented in SAMtools^[18]. The genotype likelihoods from reads for each individual at each genomic location were calculated, and the allele frequencies were estimated. Only the high-quality SNPs (coverage depth ≥ 4 and ≤ 1000 , RMS (root mean square) mapping quality ≥ 20 , distance of adjacent SNPs ≥ 5 bp and missing ratio of samples within each group $< 50\%$) were kept for the subsequent analysis.

2.3 Functional enrichment analysis

Functional enrichment analysis of Gene Ontology (GO) terms, pathway, and InterPro domains were identified using the DAVID web server^[19]. Genes were mapped to their respective human orthologs, and the lists were submitted to DAVID for enrichment analysis of the significant overrepresentation of GO biological processes (GO-BP), molecular function (GO-MF) terms, and KEGG pathway and InterPro categories. In all tests, all the known genes were assigned as the background, and P values (i.e., EASE score), which indicated significance of overlap between various gene sets, were calculated using Benjamini-corrected modified Fisher's exact test. Only terms with $P < 0.05$ were considered significant.

2.4 Phylogenetic analyses

Phylogenetic relationships were inferred using the package TreeBeST (<http://treesoft.sourceforge.net/treebest.shtml>) under the p -distance model using SNPs at a population-scale. We performed principle component analysis (PCA) with population-scale SNPs using EIGENSOFT4.2^[20]. The significance level of eigenvectors was determined using the Tracy-Widom test^[20].

2.5 Identification of selected regions

A sliding window approach (100-kb windows with 10-kb steps) was applied to quantify the pooled heterozygosity (H_p), genetic differentiation (F_{ST}), and selection statistics (Tajima's D , which is a measure of selection in the

genome) between Rongchang pigs and Asian wild boars. To detect regions with significant selective sweep signatures, we Z -transformed the resultant distributions of H_p scores and F_{ST} values, and simultaneously selected windows with low $Z(H_p)$ (< -2) in Rongchang pigs and high $Z(F_{ST})$ (> 2) as genomic regions with strong selective sweep signals that could harbor genes under selection.

3 Results and discussion

3.1 Sequencing and SNP calling

We generated a total of about 170 Gb of paired-end reads, of which 85% (144.43 Gb) of high-quality reads were mapped to the pig reference genome assembly (Sscrofa10.2) with about 6.4-fold coverage for each individual (Table S1). In addition, we downloaded about 1037 Gb of genomic data from 101 publically available pig genomes in the EMBL-EBI database and about 659 Gb in the NCBI SRA database^[1,11,15,16] (Table S2).

We performed SNP calling on a population-scale and identified 10.13 M SNPs from 107 individuals (Table S3). We then separately pooled and obtained SNP sets for each of two groups, which included 6.74 M from the 56 domestic pigs and 7.76 M from the 51 other individuals. We identified 6.50 M SNPs from six Rongchang pigs, of which 47205 were coding SNPs included 15662 non-synonymous nucleotide substitutions (15540 missense, 102 stop-gain and 20 stop-loss mutations) that were detected in 6910 genes (Table S4). These nonsynonymous SNP-containing genes in Rongchang pigs were mainly related to the G-protein coupled receptor protein signaling pathway (97 genes, $P = 2.14 \times 10^{-10}$), especially sensory perception of chemical stimulus (68 genes, $P = 1.02 \times 10^{-9}$), smell (61 genes, $P = 1.02 \times 10^{-8}$), olfactory transduction (56 genes, $P = 1.92 \times 10^{-8}$) and olfactory receptor (59 genes, $P = 5.73 \times 10^{-8}$). Pigs have one of the largest repertoires of functional olfactory receptor genes that encode the G-protein coupled receptor superfamily^[21]. In previous reports^[1], similar rapid evolution of olfactory-related genes with extensive nucleotide variation have been found, reflecting the importance of smell in this scavenging animal and other odor-driven behaviors, such as individual recognition and mating preferences^[22,23].

3.2 Genome-wide selective sweep signals

It has been well documented that from about 1000000 years ago, European and Chinese pigs diverged from each other, originating independently from different subspecies of ancestral wild boars around 10000 years ago^[1,16,24]. To examine relatedness between Rongchang pigs and other pigs, we constructed a neighbor-joining tree (Fig. 1a), and conducted PCA (Fig. 1b; Table S5) using genomic SNPs, both of which revealed a deep phylogenetic split between

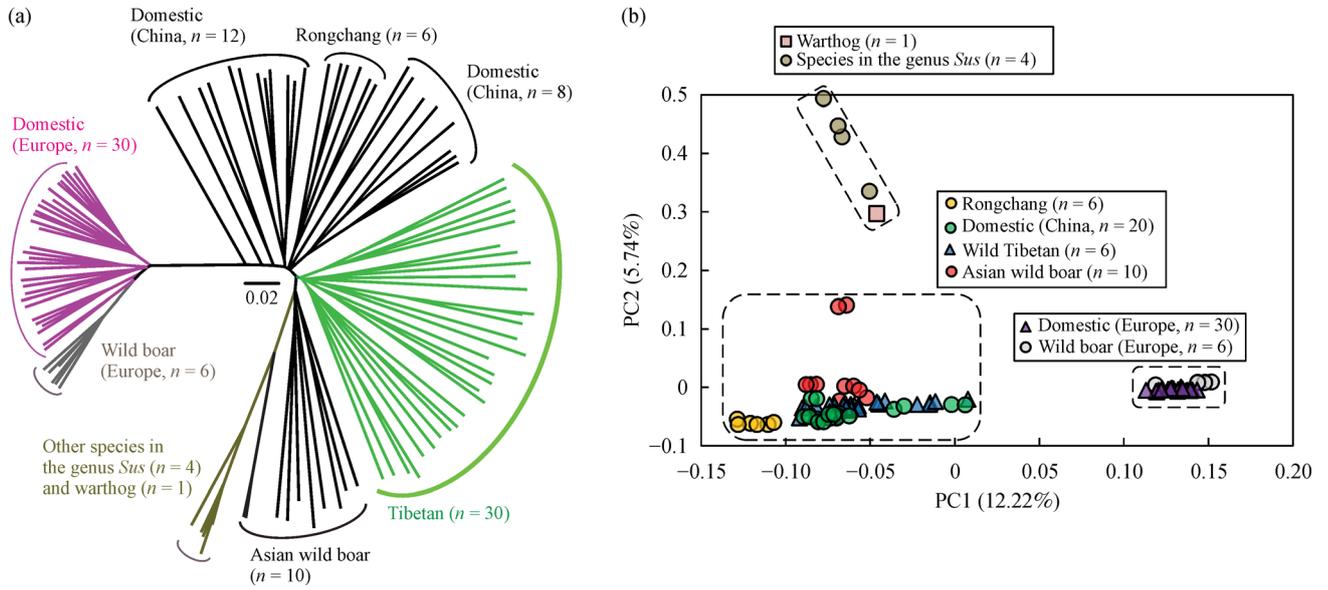


Fig. 1 Phylogenetic relationship of Rongchang pigs. (a) Neighbor-joining phylogenetic tree of pig breeds. The scale bar represents p -distance; (b) two-way principle component plot of pig breeds. The fractions of the variance explained are 12.2% and 5.74% for eigenvectors 1 and 2, respectively, with a Tracy-Widom P value $< 10^{-78}$ (Table S5).

European and Asian pigs. To accurately detect genomic footprints left by selection in Rongchang pigs and avoid genetic differences that resulted from geographic isolation of Europe from China, we specifically measured the genome-wide variations and frequency spectrum based on 8.32 M SNPs between six Rongchang pigs and 10 Asian wild boars (Table S6).

In total, 229772 100-kb windows with 10-kb steps across the pig genome contained ≥ 100 SNPs within each window and covered 84.4% of the genome and were used to identify the regions that may have been affected by selection during domestication. We empirically chose to set the thresholds at $Z(Hp)_{\text{Rongchang}} < -2$ and $Z(F_{ST}) > 2$, because they represent the extreme tails of the distributions and are hence likely enriched for strong selective sweep signals along the genome, which could harbor genes that underwent a selective sweep. From this we identified a total of 44.86 Mb of genomic data (1.61% of the genome containing 449 genes) with strong selective sweep signals in Rongchang pigs (Fig. 2a), which also exhibited significant differences ($P < 10^{-16}$, Mann-Whitney U test) based on $Z(Hp)$, $Z(F_{ST})$, and Tajima's D when compared with the genomic background (Fig. 2b).

We also constructed a phylogenetic neighbor-joining tree (Fig. 2c) and performed PCA (Fig. 2d) exclusively using the SNPs in regions with strong selective sweep signals. Although Rongchang pigs and Asian wild boars are genetically close, based on the 8.32 M SNPs across the whole genome (Fig. 1), they form two distinct clusters with respect to these SNPs (0.62 M, 7.45%), which are potentially under adaptive evolution resulting from industrial agriculture (Fig. 2c, Fig. 2d).

The 449 genes embedded in selected regions were analyzed using DAVID to examine whether these domestic genes were enriched for specific functional gene categories (Table 1). Our findings coincide with previous reports on genes related to pig domestication^[1-4,11-15]; genes related to growth and hormone binding, which included seven terms, were observed to be under strong selective sweep in Rongchang pigs, and may have contributed to the rapid growth and enhanced muscle development of domestic pigs.

Of note, 10 genes putatively under selection that are related predominantly to nervous system development, most with a single allele in Rongchang pigs, include *CNTN4*, *DLL3*, *GHSR*, *LHX5*, *MAP1B*, *MBP*, *METRN*, *NUMBL*, *TNFRSF12A* and *REST*, several of which affect brain development, neuronal functions, and behavior (Fig. 3). This result supports the view that altered behavior (such as reduced fear, higher levels of adult play, and tameness or aggression toward humans), in addition to the obvious dramatic changes in appearance and physiology, was also important during domestication, and that mutations affecting developmental genes may underlie these changes^[25-29]. In addition, four genes (*CYP2A6*, *GMPS*, *UPB1* and *UPP2*) in Rongchang pigs exhibited strong selective sweep signatures enriched for drug metabolism (Fig. S1). Given that alterations in these genes are associated with a variety of drug metabolism associated-diseases^[30], their positive selection may be attributed to constant exposure of domestic pigs in modern industry to much higher dosages of chemicals/drugs and an increased number of environmental xenobiotics, which could have accelerated evolution of drug metabolism.

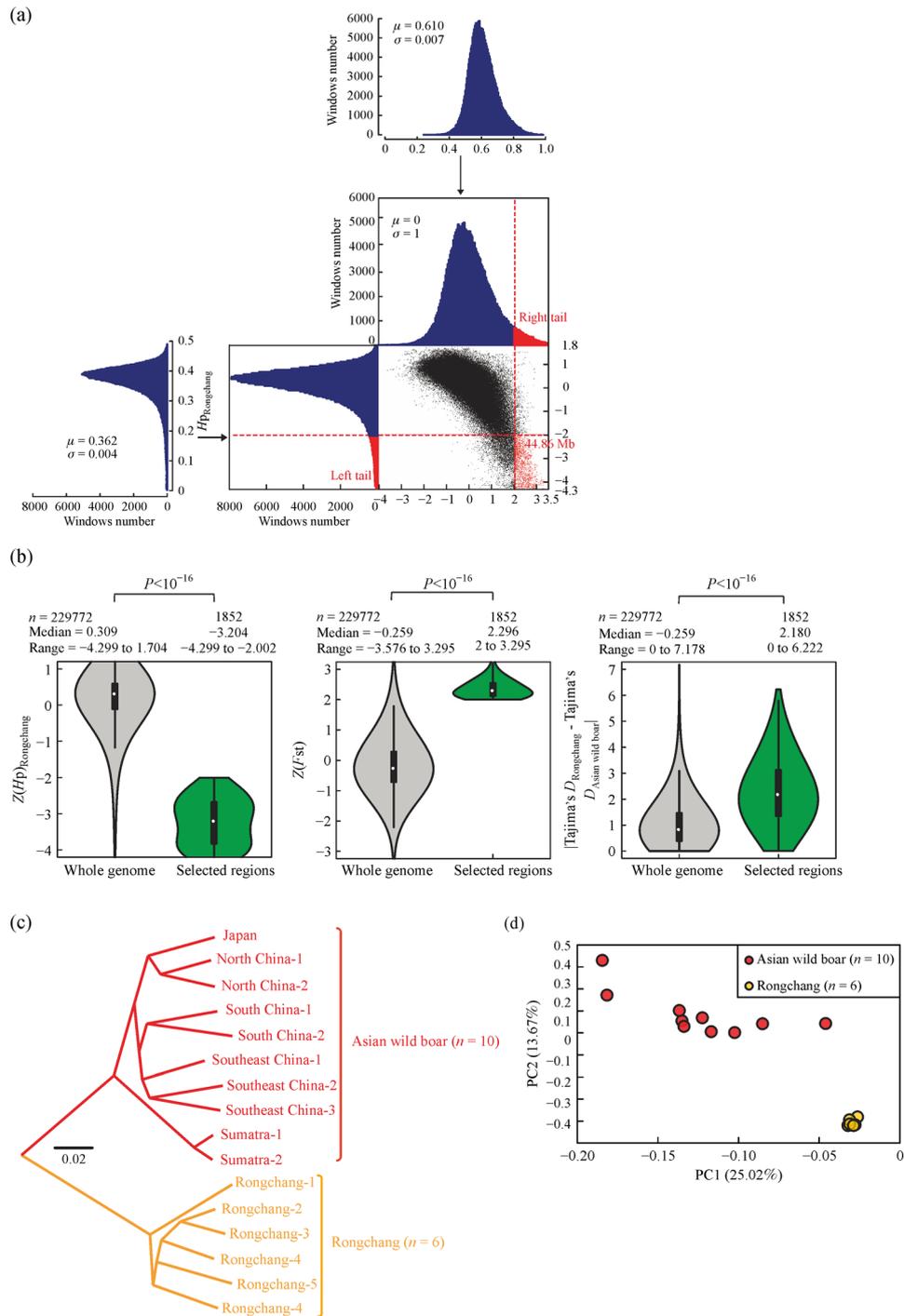


Fig. 2 Genomic regions with strong selective sweep signals in Rongchang pigs. (a) Genome-wide distribution of pooled heterozygosity values (H_p), genetic differentiation (F_{ST}), and corresponding Z transformations ($Z(H_p)$) and $Z(F_{ST})$, which were calculated in 100-kb windows with 10-kb steps ($n = 229772$, contain ≥ 100 SNPs). Data points located to the right of the vertical line (where $Z(F_{ST})$ is 2) and below the horizontal line (where $Z(H_p)$ is -2) were identified as selected regions in Rongchang pigs (red points). μ , mean; σ , standard deviation; (b) violin plot of $Z(H_p)_{\text{Rongchang}}$, $Z(F_{ST})$, and $|Tajima's D_{\text{Rongchang pigs}} - Tajima's D_{\text{Asian wild boars}}|$ in genomic regions with strong selective sweep signals for Rongchang pigs compared with the whole genome. Out of 229772 100-kb windows that contained ≥ 100 SNPs with 10-kb steps across the pig reference genome (gray violin), 1852 windows were picked out as regions with strong selective sweep signals (green violin). Each violin with the width depicting a 90°-rotated kernel density trace and its reflection. Vertical black boxes denote the interquartile range between the first and third quartiles (25th and 75th percentiles, respectively) and the white point inside denotes the median. Vertical black lines denote the lowest and highest values within a 1.5 times interquartile range from the first and third quartiles, respectively. The statistical significance was calculated by the Mann–Whitney U test; (c) phylogenetic tree (scale bar represents p -distance); (d) two-way principle component plot of Rongchang pigs ($n = 6$) and Asian wild boars ($n = 10$) based on SNPs in regions with strong selective sweep signals with 25.0% of variance explained for eigenvector 1, ($P = 0.030$, Tracy-Widom test) and 13.7% for eigenvector 2 ($P = 0.277$, Tracy-Widom test).

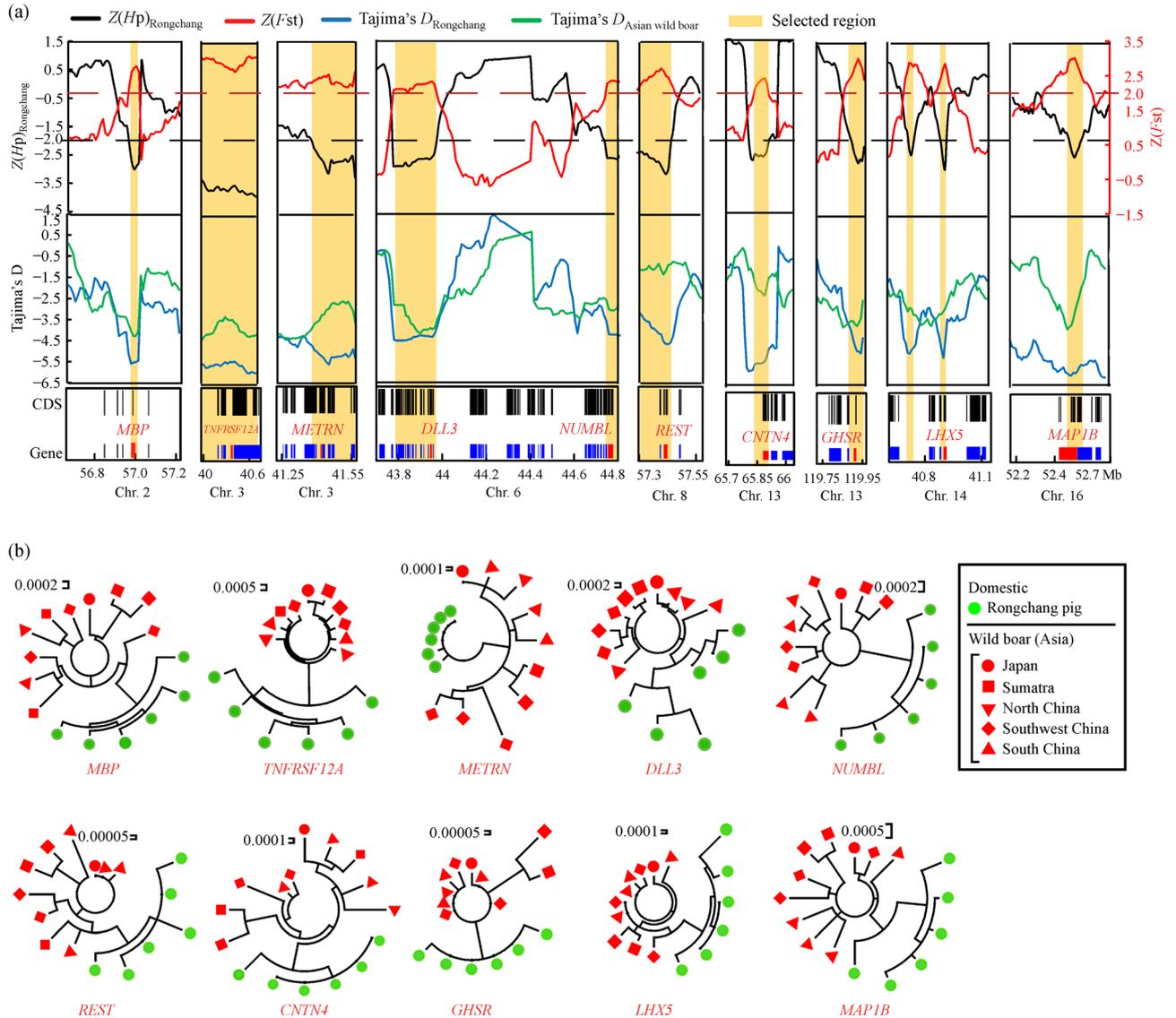


Fig. 3 Genes related to nervous system development that show selective sweep signatures in Rongchang pigs. (a) $Z(Hp)$, $Z(F_{ST})$, and Tajima's D values are plotted using a 10-kb sliding window. Genomic regions located above the upper horizontal dashed red line ($Z(F_{ST}) = 2$) and below the lower horizontal dashed black line ($Z(Hp) = -2$) were considered regions with strong selective sweep signals for Rongchang pigs (beige regions). Genome annotations are shown at the bottom (black bar: coding sequences, blue bar: genes). The boundaries of genes related to nervous system development are marked in red; (b) the gene trees for 10 genes related to nervous system development of 10 Asian wild boars and six Rongchang pigs.

Table 1 Functional gene categories enriched for genes affected by selection in Rongchang pigs

Category	Term description	Involved gene number	P value
GO-BP:0010648	Negative regulation of cell communication	13	0.007
GO-BP:0007242	Intracellular signaling cascade	40	0.011
GO-BP:0048009	Insulin-like growth factor receptor signaling pathway	3	0.015
GO-MF:0017046	Peptide hormone binding	4	0.018
GO-MF:0042562	Hormone binding	5	0.019
GO-MF:0005158	Insulin receptor binding	4	0.020
GO-BP:0051960	Regulation of nervous system development	10	0.022
GO-BP:0032868	Response to insulin stimulus	6	0.033

(Continued)

Category	Term description	Involved gene number	P value
GO-BP:0050769	Positive regulation of neurogenesis	5	0.037
GO-BP:0050767	Regulation of neurogenesis	8	0.040
GO-BP:0045664	Regulation of neuron differentiation	7	0.041
GO-BP:0010975	Regulation of neuron projection development	5	0.041
KEGG-Pathway: 00983	Drug metabolism	4	0.041
GO-BP:0006396	RNA processing	19	0.046
GO-BP:0010720	Positive regulation of cell development	5	0.047
GO-MF:0019899	Enzyme binding	18	0.049
GO-BP:0009725	Response to hormone stimulus	14	0.049

Note: *P* values (i.e., EASE scores), which indicate significance of the overlap between various gene sets, were calculated using a Benjamini-corrected modified Fisher's exact test. Only Gene Ontology (GO) biological process (GO-BP), GO-molecular function (GO-MF) and KEGG pathway terms with $P < 0.05$ were considered significant and listed.

4 Conclusions

This study examined the genetic relationships among Chinese Rongchang and other pigs, and uncovered genetic footprints of domestication and selection that provide an important resource for further improvements of this important livestock species. We envision that the data presented here will provide a representative example on which to base future deciphering of genomic footprints left by livestock domestication and selection.

Accession codes The genome resequencing reads of Rongchang pigs have been deposited into the NCBI SRA database under the accession SRP034675.

Supplementary materials The online version of this article at <http://dx.doi.org/10.15302/J-FASE-2017161> contains supplementary materials (Tables S1–S6; Fig. S1).

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Compliance with ethics guidelines Lei Chen, Shilin Tian, Long Jin, Zongyi Guo, Dan Zhu, Lan Jing, Tiandong Che, Qianzi Tang, Siqing Chen, Liang Zhang, Tinghuan Zhang, Zuohua Liu, Jinyong Wang, and Mingzhou Li declare that they have no conflicts of interest or financial conflicts to disclose.

All applicable institutional and national guidelines for the care and use of animals were followed.

References

- Groenen M A M, Archibald A L, Uenishi H, Tuggle C K, Takeuchi Y, Rothschild M F, Rogel-Gaillard C, Park C, Milan D, Megens H J, Li S, Larkin D M, Kim H, Frantz L A F, Caccamo M, Ahn H, Aken B L, Anselmo A, Anthon C, Auviel L, Badaoui B, Beattie C W, Bendixen C, Berman D, Blecha F, Blomberg J, Bolund L, Bosse M, Botti S, Bujie Z, Bystrom M, Capitanu B, Carvalho-Silva D, Chardon P, Chen C, Cheng R, Choi S H, Chow W, Clark R C, Clee C, Crooijmans R P M A, Dawson H D, Dehaes P, De Sapiro F, Dibbitts B, Drou N, Du Z Q, Eversole K, Fadista J, Fairley S, Faraut T, Faulkner G J, Fowler K E, Fredholm M, Fritz E, Gilbert J G R, Giuffra E, Gorodkin J, Griffin D K, Harrow J L, Hayward A, Howe K, Hu Z L, Humphray S J, Hunt T, Hornshøj H, Jeon J T, Jern P, Jones M, Jurka J, Kanamori H, Kapetanovic R, Kim J, Kim J H, Kim K W, Kim T H, Larson G, Lee K, Lee K T, Leggett R, Lewin H A, Li Y, Liu W, Loveland J E, Lu Y, Lunney J K, Ma J, Madsen O, Mann K, Matthews L, McLaren S, Morozumi T, Murtaugh M P, Narayan J, Truong Nguyen D, Ni P, Oh S J, Onteru S, Panitz F, Park E W, Park H S, Pascal G, Paudel Y, Perez-Enciso M, Ramirez-Gonzalez R, Reecy J M, Rodriguez-Zas S, Rohrer G A, Rund L, Sang Y, Schachtschneider K, Schraiber J G, Schwartz J, Scobie L, Scott C, Searle S, Servin B, Southey B R, Sperber G, Stadler P, Sweedler J V, Tafer H, Thomsen B, Wali R, Wang J, Wang J, White S, Xu X, Yerle M, Zhang G, Zhang J, Zhang J, Zhao S, Rogers J, Churcher C, Schook L B. Analyses of pig genomes provide insight into porcine demography and evolution. *Nature*, 2012, **491**(7424): 393–398
- Chen K, Baxter T, Muir W M, Groenen M A, Schook L B. Genetic resources, genome mapping and evolutionary genomics of the pig (*Sus scrofa*). *International Journal of Biological Sciences*, 2007, **3**(3): 153–165
- Rubin C J, Megens H J, Barrio A M, Maqbool K, Sayyab S, Schwochow D, Wang C, Carlborg O, Jern P, Jorgensen C B, Archibald A L, Fredholm M, Groenen M A M, Andersson L. Strong signatures of selection in the domestic pig genome. *Proceedings of the National Academy of Sciences of the United States of America*, 2012, **109**(48): 19529–19536
- Ai H, Fang X, Yang B, Huang Z, Chen H, Mao L, Zhang F, Zhang L, Cui L, He W, Yang J, Yao X, Zhou L, Han L, Li J, Sun S, Xie X, Lai B, Su Y, Lu Y, Yang H, Huang T, Deng W, Nielsen R, Ren J, Huang L. Adaptation and possible ancient interspecies introgression in pigs identified by whole-genome sequencing. *Nature Genetics*, 2015, **47**(3): 217–225
- Xia Q, Guo Y, Zhang Z, Li D, Xuan Z, Li Z, Dai F, Li Y, Cheng D, Li R, Cheng T, Jiang T, Becquet C, Xu X, Liu C, Zha X, Fan W, Lin

- Y, Shen Y, Jiang L, Jensen J, Hellmann I, Tang S, Zhao P, Xu H, Yu C, Zhang G, Li J, Cao J, Liu S, He N, Zhou Y, Liu H, Zhao J, Ye C, Du Z, Pan G, Zhao A, Shao H, Zeng W, Wu P, Li C, Pan M, Li J, Yin X, Li D, Wang J, Zheng H, Wang W, Zhang X, Li S, Yang H, Lu C, Nielsen R, Zhou Z, Wang J, Xiang Z, Wang J. Complete resequencing of 40 genomes reveals domestication events and genes in silkworm (*Bombyx*). *Science*, 2009, **326**(5951): 433–436
6. Rubin C J, Zody M C, Eriksson J, Meadows J R, Sherwood E, Webster M T, Jiang L, Ingman M, Sharpe T, Ka S, Hallböök F, Besnier F, Carlborg O, Bed'hom B, Tixier-Boichard M, Jensen P, Siegel P, Lindblad-Toh K, Andersson L. Whole-genome resequencing reveals loci under selection during chicken domestication. *Nature*, 2010, **464**(7288): 587–591
 7. Shapiro M D, Kronenberg Z, Li C, Domyan E T, Pan H, Campbell M, Tan H, Huff C D, Hu H, Vickrey A I, Nielsen S C, Stringham S A, Hu H, Willerslev E, Gilbert M T, Yandell M, Zhang G, Wang J. Genomic diversity and evolution of the head crest in the rock pigeon. *Science*, 2013, **339**(6123): 1063–1067
 8. Carneiro M, Rubin C J, Di Palma F, Albert F W, Alföldi J, Barrio A M, Pielberg G, Rafati N, Sayyab S, Turner-Maier J, Younis S, Afonso S, Aken B, Alves J M, Barrell D, Bolet G, Boucher S, Burbano H A, Campos R, Chang J L, Duranthon V, Fontanesi L, Garreau H, Heiman D, Johnson J, Mage R G, Peng Z, Queney G, Rogel-Gaillard C, Ruffier M, Searle S, Villafuerte R, Xiong A, Young S, Forsberg-Nilsson K, Good J M, Lander E S, Ferrand N, Lindblad-Toh K, Andersson L. Rabbit genome analysis reveals a polygenic basis for phenotypic change during domestication. *Science*, 2014, **345**(6200): 1074–1079
 9. Axelsson E, Ratnakumar A, Arendt M L, Maqbool K, Webster M T, Perloski M, Liberg O, Arnemo J M, Hedhammar A, Lindblad-Toh K. The genomic signature of dog domestication reveals adaptation to a starch-rich diet. *Nature*, 2013, **495**(7441): 360–364
 10. Daetwyler H D, Capitan A, Pausch H, Stothard P, van Binsbergen R, Brøndum R F, Liao X, Djari A, Rodriguez S C, Grohs C, Esquerré D, Bouchez O, Rossignol M N, Klopp C, Rocha D, Fritz S, Eggen A, Bowman P J, Coote D, Chamberlain A J, Anderson C, VanTassell C P, Hulsege I, Goddard M E, Guldbrandtsen B, Lund M S, Veerkamp R F, Boichard D A, Fries R, Hayes B J. Whole-genome sequencing of 234 bulls facilitates mapping of monogenic and complex traits in cattle. *Nature Genetics*, 2014, **46**(8): 858–865
 11. Li M, Tian S, Jin L, Zhou G, Li Y, Zhang Y, Wang T, Yeung C K, Chen L, Ma J, Zhang J, Jiang A, Li J, Zhou C, Zhang J, Liu Y, Sun X, Zhao H, Niu Z, Lou P, Xian L, Shen X, Liu S, Zhang S, Zhang M, Zhu L, Shuai S, Bai L, Tang G, Liu H, Jiang Y, Mai M, Xiao J, Wang X, Zhou Q, Wang Z, Stothard P, Xue M, Gao X, Luo Z, Gu Y, Zhu H, Hu X, Zhao Y, Plastow G S, Wang J, Jiang Z, Li K, Li N, Li X, Li R. Genomic analyses identify distinct patterns of selection in domesticated pigs and Tibetan wild boars. *Nature Genetics*, 2013, **45**(12): 1431–1438
 12. Li M, Chen L, Tian S, Lin Y, Tang Q, Zhou X, Li D, Yeung C K L, Che T, Jin L, Fu Y, Ma J, Wang X, Jiang A, Lan J, Pan Q, Liu Y, Luo Z, Guo Z, Liu H, Zhu L, Shuai S, Tang G, Zhao J, Jiang Y, Bai L, Zhang S, Mai M, Li C, Wang D, Gu Y, Wang G, Lu H, Li Y, Zhu H, Li Z, Li M, Gladyshev V N, Jiang Z, Zhao S, Wang J, Li R, Li X. Comprehensive variation discovery and recovery of missing sequence in the pig genome using multiple de novo assemblies. *Genome Research*, 2017, **27**(5): 865–874
 13. Fu Y, Li C, Tang Q, Tian S, Jin L, Chen J, Li M, Li C. Genomic analysis reveals selection in Chinese native black pig. *Scientific Reports*, 2016, **6**(1): 36354
 14. Li M, Tian S, Yeung C K, Meng X, Tang Q, Niu L, Wang X, Jin L, Ma J, Long K, Zhou C, Cao Y, Zhu L, Bai L, Tang G, Gu Y, Jiang A, Li X, Li R. Whole-genome sequencing of Berkshire (European native pig) provides insights into its origin and domestication. *Scientific Reports*, 2014, **4**(4): 4678
 15. Bosse M, Megens H J, Frantz L A, Madsen O, Larson G, Paudel Y, Duijvesteijn N, Harlizius B, Hagemeyer Y, Crooijmans R P, Groenen M A. Genomic analysis reveals selection for Asian genes in European pigs following human-mediated introgression. *Nature Communications*, 2014, **5**: 4392
 16. Frantz L A, Schraiber J G, Madsen O, Megens H J, Bosse M, Paudel Y, Semiadi G, Meijaard E, Li N, Crooijmans R P, Archibald A L, Slatkin M, Schook L B, Larson G, Groenen M A. Genome sequencing reveals fine scale diversification and reticulation history during speciation in *Sus*. *Genome Biology*, 2013, **14**(9): R107
 17. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*, 2009, **25**(14): 1754–1760
 18. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R. The sequence alignment/map format and SAMtools. *Bioinformatics*, 2009, **25**(16): 2078–2079
 19. Huang W, Sherman B T, Lempicki R A. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nature Protocols*, 2009, **4**(1): 44–57
 20. Patterson N, Price A L, Reich D. Population structure and eigenanalysis. *PLoS Genetics*, 2006, **2**(12): e190
 21. Nguyen D T, Lee K, Choi H, Choi M K, Le M T, Song N, Kim J H, Seo H G, Oh J W, Lee K, Kim T H, Park C. The complete swine olfactory subgenome: expansion of the olfactory gene repertoire in the pig genome. *BMC Genomics*, 2012, **13**(1): 584
 22. Marchese S, Pes D, Scaloni A, Carbone V, Pelosi P. Lipocalins of boar salivary glands binding odours and pheromones. *European Journal of Biochemistry*, 1998, **252**(3): 563–568
 23. Mak G K, Enwere E K, Gregg C, Pakarainen T, Poutanen M, Huhtaniemi I, Weiss S. Male pheromone-stimulated neurogenesis in the adult female brain: possible role in mating behavior. *Nature Neuroscience*, 2007, **10**(8): 1003–1011
 24. Larson G, Dobney K, Albarella U, Fang M, Matisoo-Smith E, Robins J, Lowden S, Finlayson H, Brand T, Willerslev E, Rowley-Conwy P, Andersson L, Cooper A. Worldwide phylogeography of wild boar reveals multiple centers of pig domestication. *Science*, 2005, **307**(5715): 1618–1621
 25. Albert F W, Somel M, Carneiro M, Aximu-Petri A, Halbwax M, Thalmann O, Blanco-Aguiar J A, Plyusnina I Z, Trut L, Villafuerte R, Ferrand N, Kaiser S, Jensen P, Pääbo S. A comparison of brain gene expression levels in domesticated and wild animals. *PLoS Genetics*, 2012, **8**(9): e1002962
 26. Amaral A J, Ferretti L, Megens H J, Crooijmans R P, Nie H, Ramos-Onsins S E, Perez-Enciso M, Schook L B, Groenen M A. Genome-wide footprints of pig domestication and selection revealed through massive parallel sequencing of pooled DNA. *PLoS One*, 2011, **6**(4): e14782

27. Li Y, Vonholdt B M, Reynolds A, Boyko A R, Wayne R K, Wu D D, Zhang Y P. Artificial selection on brain-expressed genes during the domestication of dog. *Molecular Biology and Evolution*, 2013, **30** (8): 1867–1876
28. Hare B, Plyusnina I, Ignacio N, Schepina O, Stepika A, Wrangham R, Trut L. Social cognitive evolution in captive foxes is a correlated by-product of experimental domestication. *Current Biology*, 2005, **15**(3): 226–230
29. Topál J, Gergely G, Erdohegyi A, Csibra G, Miklósi A. Differential sensitivity to human communication in dogs, wolves, and human infants. *Science*, 2009, **325**(5945): 1269–1272
30. Meyer U A, Zanger U M, Schwab M. Omics and drug response. *Annual Review of Pharmacology and Toxicology*, 2013, **53**(53): 475–502